

RESEARCH ARTICLE

Nosocomial Outbreak of New Delhi Metallo- β -Lactamase-1-Producing Gram-Negative Bacteria in South Africa: A Case-Control Study

Pieter de Jager^{1,2*}, Tobias Chirwa³, Shan Naidoo², Olga Perovic^{4,5}, Juno Thomas⁶

1 Epidemiology and Surveillance Unit, National Institute for Occupational Health, National Health Laboratory Service, Johannesburg, South Africa, **2** Department of Community Health, School of Public Health, Faculty of Health Sciences, University of the Witwatersrand, Johannesburg, South Africa, **3** Department of Epidemiology and Biostatistics, School of Public Health, Faculty of Health Sciences, University of the Witwatersrand, Johannesburg, South Africa, **4** Centre for Opportunistic, Tropical and Hospital Infections, National Institute for Communicable Diseases, National Health Laboratory Service, Johannesburg, South Africa, **5** Department of Clinical Microbiology and Infectious Diseases, School of Pathology, Faculty of Health Science, University of Witwatersrand, Johannesburg, South Africa, **6** Outbreak Response Unit, National Institute for Communicable Diseases, National Health Laboratory Service, Johannesburg, South Africa

* Pieter.deJager@wits.ac.za



OPEN ACCESS

Citation: de Jager P, Chirwa T, Naidoo S, Perovic O, Thomas J (2015) Nosocomial Outbreak of New Delhi Metallo- β -Lactamase-1-Producing Gram-Negative Bacteria in South Africa: A Case-Control Study. PLoS ONE 10(4): e0123337. doi:10.1371/journal.pone.0123337

Academic Editor: Matthew E. Falagas, Alfa Institute of Biomedical Sciences (AIBS), GREECE

Received: July 19, 2014

Accepted: March 2, 2015

Published: April 24, 2015

Copyright: © 2015 de Jager et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper.

Funding: This work was supported by the School of Public Health, Faculty of Health Sciences, University of the Witwatersrand, Johannesburg. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

Abstract

Objective

New Delhi metallo- β -lactamase (NDM)-producing Gram-negative bacteria have spread globally and pose a significant public health threat. There is a need to better define risk factors and outcomes of NDM-1 clinical infection. We assessed risk factors for nosocomial infection with NDM-1-producers and associated in-hospital mortality.

Methods

A matched case-control study was conducted during a nosocomial outbreak of NDM-1-producers in an adult intensive care unit (ICU) in South Africa. All patients from whom NDM-1-producers were identified were considered (n=105). Cases included patients admitted during the study period in whom NDM-1 producing Gram-negative bacteria were isolated from clinical specimens collected ≥ 48 hours after admission, and where surveillance definitions for healthcare-associated infections were met. Controls were matched for age, sex, date of hospital admission and intensive-care admission. Conditional logistic regression was used to identify risk factors for NDM-1 clinical infection and associated in-hospital mortality.

Findings

38 cases and 68 controls were included. *Klebsiella pneumoniae* was the most common NDM-1-producer (28/38, 74%). Cases had longer mean hospital stays (44.0 vs. 13.3 days; $P < 0.001$) and ICU stays (32.5 vs. 8.3 days; $P < 0.001$). Adjusting for co-morbid disease,

the in-hospital mortality of cases was significantly higher than controls (55.3% vs. 14.7%; AOR, 11.29; $P < 0.001$). Higher Charlson co-morbidity index score (5.2 vs. 4.1; AOR, 1.59; $P = 0.005$), mechanical ventilation days (7.47 vs. 0.94 days; AOR, 1.32; $P = 0.003$) and piperacillin/tazobactam exposure (11.03 vs. 1.05 doses; AOR, 1.08; $P = 0.013$) were identified as risk factors on multivariate analysis. Cases had a significantly higher likelihood of in-hospital mortality when the NDM-1-producer was *Klebsiella pneumoniae* (AOR, 16.57; $P = 0.007$), or when they had a bloodstream infection (AOR, 8.84; $P = 0.041$).

Conclusion

NDM-1 infection is associated with significant in-hospital mortality. Risk factors for hospital-associated infection include the presence of co-morbid disease, mechanical ventilation and piperacillin/tazobactam exposure.

Introduction

Resistance to β -lactams is a long recognised problem in Gram-negative bacteria[1] and with the introduction of new classes of β -lactams, novel β -lactamases have emerged.[1,2] Carbapenem resistance has become a growing problem over the last decade with the emergence of readily transferable plasmid mediated carbapenem-hydrolysing β -lactamases. [3,4] These carbapenemases constitute a heterogeneous and versatile group of enzymes hydrolysing β -lactams and also exhibit resistance to β -lactamase inhibitors, making them exceedingly difficult to treat.[4,5]

In 2008 a novel metallo- β -lactamase designated New Delhi metallo- β -lactamase (NDM-1) was identified in a Swedish patient returning from India.[6] The first case of NDM-1 in South Africa was identified in September 2011.[7] The bla_{NDM-1} gene is plasmid mediated and associated with numerous other resistance determinants conferring resistance to β -lactams, fluoroquinolones and aminoglycosides resulting in significant treatment option limitations.[4,8] Sensitivity to tigecycline and polymyxins are typically reserved although the efficacy of these treatment options have not been established and drug toxicity, particularly with colistin, poses further clinical challenges.[9] Compared to other carbapenemase types, NDM-1 displays a broader spectrum of antimicrobial resistance and its global spread has been singularly rapid; notably, it has been detected in diverse species and genera of Gram-negative bacteria.[10,11] NDM-1-producers have been documented on every continent except Antarctica,[12–14] with increasing reports of transmission and acquisition of NDM-1-producers both in healthcare facilities and in the community.[15,16]

With limited treatment options available, slowing and preventing the spread of bla_{NDM-1} through an understanding of risk factors for its acquisition is essential. However, there is a paucity of published epidemiological studies reporting on risk factors for NDM-1 clinical infection. In order to evaluate risk factors for NDM-1 clinical infection and associated in-hospital mortality, we conducted a matched case-control study during a prolonged outbreak of NDM-1 producing Gram-negative bacteria in a South African hospital. We hypothesised that exposure to antibiotics and medical devices would be risk factors for NDM-1 clinical infection and would be associated with greater in-hospital mortality.

Methods

Ethics statement

Study participant age ranged from 20 years to 90 years, with a mean age of 61.3 and a median age of 64. Verbal informed consent was obtained from all patients or their next of kin prior to conducting telephonic interviews which collected information on past hospitalization/chronic care admission and travel history. Verbal consent was obtained as this was a retrospective study and patients had subsequently relocated to various parts of the country. Consent was captured on a consent form by the researchers. Consent to review clinical records were obtained from the hospital and all patient data were anonymized and de-linked from unique identifiers prior to analysis. Ethics approval for this study, including the consent procedure, was obtained from the Human Research Ethics Committee (Medical) at the University of the Witwatersrand, Johannesburg. (M130248)

Study setting and laboratory methods

The outbreak occurred across three private hospitals in South Africa with strong referral links. This study was confined to the hospital where the majority of cases (90/105, 86%) were detected, and all cases and controls were from the adult ICU. The hospital is located in the greater Johannesburg area, has a total of 322 beds including a 37-bed ICU, and offers tertiary-level specialist care.

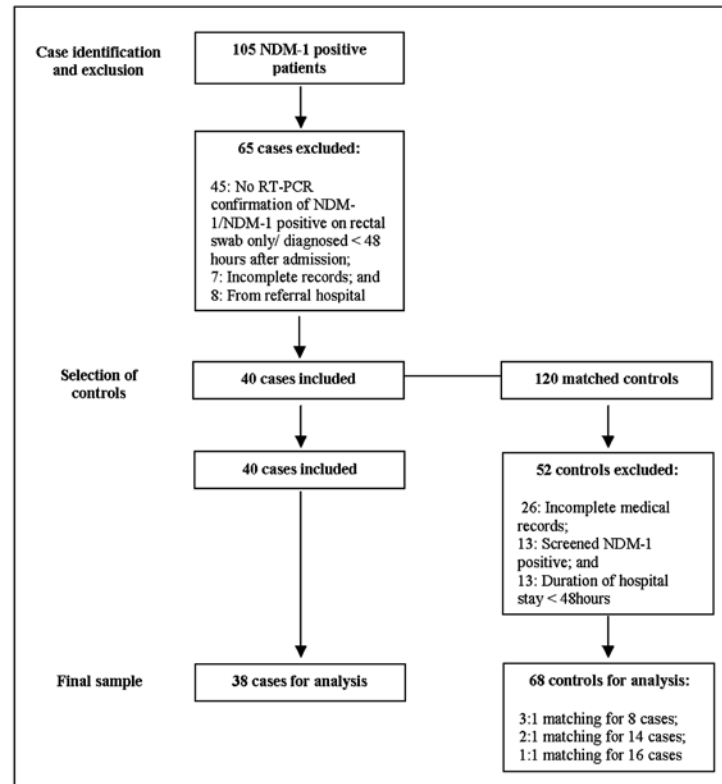
In early August 2011 *Klebsiella pneumoniae* isolated from an 86-year-old male admitted following a hip fracture was found to harbour bla_{NDM-1} . In response to this, the first case of NDM-1 both in the hospital and the country, a rectal screening programme was instituted to identify patients colonised with NDM-1-producers, with screening criteria revisions throughout the course of the outbreak. The method of screening employed by all diagnostic laboratories throughout the outbreak was direct real-time polymerase chain reaction (RT-PCR) testing for bla_{NDM-1} on dry rectal swabs. Clinical isolates demonstrating phenotypic resistance to carbapenems were further tested for bla_{NDM-1} using RT-PCR. The LightCycler 480 II (Roche Applied Science) instrument was used for the RT-PCR assay. The bla_{NDM-1} gene was amplified by a real time polymerase chain reaction (PCR) using the LightCycler 480 Probes Master kit (Roche Diagnostics, IN, USA) and the LightMix Modular NDM (ESBL) kit (Roche Diagnostics, IN, USA). The positive control was provided with the LightMix Modular kits and sterile water was used as a negative control. An internal control, the LightMix Modular PhHV Internal Control kit (Roche Diagnostics, IN, USA) was also included in the run.

All microbiological testing was conducted in routine private diagnostic laboratories servicing the private healthcare sector. Thirteen NDM-positive isolates (seven case isolates and six epidemiologically linked environmental specimens collected in the adult ICU) were subjected to DNA fingerprinting by macro-restriction analysis on pulsed-field gel electrophoresis (PFGE) at the Infection Control Services Laboratory, National Health Laboratory Services. PFGE was performed as described previously.[17]

Cases and Controls

The study design is summarized in [Fig 1](#).

All patients admitted between 1 July 2011 and 31 October 2012 were eligible for inclusion. We included cases where bla_{NDM-1} was detected on an isolate from a specimen collected at least 48 hours after admission and the infection was categorised as a healthcare-associated infection as per the Centers for Disease Control and Prevention/National Healthcare Safety Network definitions.[18] Potential cases were excluded if bla_{NDM-1} was detected on rectal



NDM-1 = New Delhi metallo-β-lactamase; RT-PCR = real-time polymerase chain reaction testing

Fig 1. Study design and selection of cases and controls. NDM-1 = New Delhi metallo-β-lactamase; RT-PCR = real-time polymerase chain reaction testing.

doi:10.1371/journal.pone.0123337.g001

screening alone, where rectal screening was positive within the first 48 hours of admission, or where clinical records were incomplete. After exclusion of cases not fulfilling the inclusion criteria, 40 cases remained and three controls were matched to each case for sex (male/female), age (+/- 5 years), date of hospital admission (+/- 14 days) and ICU admission (yes/no). Where more than three eligible controls were identified on the hospital’s electronic database, three controls were randomly selected. Controls were excluded if they had *bla*_{NDM-1} detected on any sample during the hospitalisation period, if patient records were incomplete or missing, or if the patient was admitted for less than 48 hours.

No controls could be found meeting the matching criteria for two cases and for three cases only two matching controls could be identified. Another 52 controls were excluded for missing/incomplete medical records (n = 26), record of screening NDM-1 positive on dry rectal swab (n = 13) or being admitted for less than 48 hours (n = 13). The final sample consisted of 38 cases and 68 controls.

Data collection

Clinical data were collected for cases and controls from clinical records, laboratory results and hospital billing data. Exposure to antibiotics (carbapenems, aminoglycosides, fluoroquinolones, third- and fourth-generation cephalosporins, and piperacillin/tazobactam) and corticosteroids were recorded as total number of doses received. Exposure to medical devices (central venous line and indwelling urinary catheter) as well as selected medical interventions (haemodialysis, mechanical ventilation and parenteral nutrition) was recorded as total number of days

exposed. Patients who underwent laparotomy or thoracotomy were grouped and compared to patients who received other (mainly orthopaedic) or no surgery. Co-morbid disease and severity of illness on admission was measured by calculating Charlson co-morbidity index and Mortality Probability Model III (MPM III) scores respectively.[19,20] Exposure data for cases were collected from the date of admission until the date of collection of the first sample yielding an NDM-1-producing isolate (time at risk). For controls, exposure data were collected from the date of admission until the date of discharge or death (time at risk). Beyond the time at risk, total length of hospital and total length of intensive care unit stay were also collected.

Data on past hospital or long-term care facility admission and international travel history in the year leading up to the admission of interest were collected through telephonic interviews for both cases and controls. All data were collected between November 2012 and November 2013 by trained professional nurses and medical doctors.

Statistical analysis

We evaluated risk factors associated with case status and compared in-hospital mortality between cases and controls. Except for MPM-III scores, where its calculation would have been invalid, there was no missing clinical data in the final sample used for analysis. Where past admission, travel history or MPM-III scores were missing, observations were excluded from the analysis.

Data were entered into Epi-Info version 7 and exported to Excel 2007 where it was inspected for errors before being imported to Stata Version 12 for statistical analysis. Continuous variables such as length of hospital stay, MPM-III and Charlson scores, are described through the reporting of means and standard deviations. Two sided t-test for two groups (cases and controls) was used to compare means of continuous variables with normal distributions. Where data were not normally distributed Mann-Whitney U test was used. For differences in proportions such as previous hospitalisation or travel history, Mantel-Haenszel Chi square test was used. Bivariate conditional logistic regression analysis was undertaken to calculate crude odds ratio's for exposure to medical devices and interventions, antibiotics and duration of stay. Stepwise conditional logistic regression was conducted to identify factors associated with case status. All exposure variables with a $P < 0.20$ at the univariate level were considered in the final multiple regression model. Significance was taken at a level of 0.05. Conditional logistic regression was further undertaken to calculate the odds of in-hospital mortality for cases and controls as well as for different sites of infection and clinical isolates. Adjusted odds ratios were calculated using multivariable conditional logistic regression.

Results

The most common NDM-1-producing isolate among the 38 cases was *Klebsiella pneumoniae* (28/38, 74%) followed by *Enterobacter cloacae* (5/38, 13%), *Klebsiella oxytoca* (2/38, 5%), *Serratia marcescens* (2/38, 5%) and *Citrobacter amalonaticus* (1/38, 3%). The most common clinical specimen types yielding NDM-1 were sputum (16/38, 42%), blood (12/38, 32%) and urine (5/38, 13%) followed by pus (2/38, 5%), broncho-alveolar lavage (2/38, 5%) and pleural fluid (1/38, 3%).

PFGE showed two closely related clusters: cluster A comprised three case isolates and six environmental isolates, whilst cluster B comprised three case isolates. Given the protracted course of the outbreak, this suggests that these isolates are all related.[21]

Cases had a longer mean total length of hospital stay (44.0 vs 13.3 days, $P < 0.001$) and longer mean durations of time at risk, particularly mean ICU time at risk (18.9 vs 8.3 days, $P < 0.001$)

Table 1. Duration of stay, time at risk and co-morbid status for cases and controls.

| Variable | Cases (n = 38) Mean (SD) | Controls(n = 68) Mean (SD) | p-value |
|-------------------------------------|--------------------------|----------------------------|------------------|
| Time at risk (total, days) | 22.2 (±15.8) | 13.3 (±9.5) | 0.004 |
| Time at risk (intensive care, days) | 18.9 (±13.7) | 8.3 (±7.2) | <0.001 |
| Total length of stay (days) | 44.0 (±28.2) | 13.3 (±9.5) | <0.001 |
| Total length of ICU stay (days) | 32.5 (±27.0) | 8.3 (±7.2) | <0.001 |
| MPM III Score (%) | 11.5 (±7.1) | 8.3 (±6.8) | 0.072 |
| Age Adjusted Charlson Score | 5.2 (±3.1) | 4.1 (±2.2) | 0.032 |

SD = standard deviation; time at risk: from admission to discharge/death (controls) or NDM-1 diagnosis (cases); MPM-III = Mortality Probability Model III; total length of stay: time from admission to discharge/death; p-values calculated using Mann-Whitney U test.

doi:10.1371/journal.pone.0123337.t001

than controls (Table 1). Charlson co-morbidity index scores were, on average, significantly higher in cases than controls (5.2 vs 4.1, P = 0.032).

Risk factors associated with case status

Cases had significantly higher odds of having been hospitalised or admitted to a long-term care facility in the previous year (OR 6.83; 95% CI 2.32–20.16) or being transferred from a referral hospital (OR 4.98; 95% CI 1.56–15.93) compared to controls (Table 2). No association was found between travel history and case status. Although total time at risk was not associated with case status, an ICU stay of longer than seven days was associated with a significant risk of infection with NDM-1-producers (OR 4.82; 95% CI 1.80–12.91). Exposure to any antibiotics (carbapenem, fluoroquinolone, aminoglycoside, third- or fourth-generation cephalosporins, or piperacillin/tazobactam) was also significantly associated with case status (OR 4.77; 95% CI 1.38–16.48). No association between HIV status or surgery (laparotomy or thoracotomy) and infection with NDM-1-producers was found.

On univariate analysis exposure to aminoglycosides, piperacillin/tazobactam and corticosteroids were significantly associated with case status (Table 3). Each additional dose of piperacillin/tazobactam or a corticosteroid was associated with a 5% increase in risk of developing infection with a NDM-1-producer, while each additional dose of an aminoglycoside was associated with a 3% increase in risk. Although exposure to fluoroquinolones, carbapenems and third-/fourth-generation cephalosporins were associated with an increased risk of case status, none of these showed statistical significance at the 5% level. Each additional day of exposure to a central venous line or indwelling urinary catheter was associated with an 8% and 7% increased risk of case status on univariate analysis respectively. Selected medical interventions were significantly associated with NDM-1-producer infection, with a 16% and 27% increased risk for each additional day of haemodialysis and mechanical ventilation respectively. As summarised in Table 4, multivariate analysis showed mechanical ventilation and exposure to piperacillin/tazobactam to be significantly associated with case status.

Of the 68 controls 10 died in hospital (14.7%), while 21 of the 38 cases died in hospital (55.3%); this translates to an attributable mortality of 47.5% (Table 5). After adjusting for co-morbid disease, case status was associated with an eleven-fold higher risk of in-hospital mortality (AOR 11.29; 95% CI 2.57–49.60) compared to controls. Cases with bloodstream infections due to NDM-1-producers (AOR 8.84; 95% CI 1.09–71.55), or where the organism harbouring the *bla*_{NDM-1} was *Klebsiella pneumoniae* (AOR 16.57; 95% CI 2.12–129.6) had a significantly higher likelihood of in-hospital mortality.

Table 2. Univariate analysis of pre-hospital factors, HIV status, time at risk, surgery and antibiotic exposure among cases and controls.

| Exposure Variable | Case patient (n = 38) with exposure | | Control patient (n = 68) with exposure | | Unadjusted OR(95%CI) | p-value |
|---------------------------------------|-------------------------------------|----|--|----|----------------------|------------------|
| | Number | % | Number | % | | |
| Previous Hospitalization/Chronic care | | | | | | |
| No | 10 | 29 | 40 | 83 | 1 | |
| Yes | 24 | 71 | 8 | 17 | 6.83 (2.32–20.16) | <0.001 |
| Travel outside South Africa | | | | | | |
| No | 30 | 94 | 47 | 98 | 1 | |
| Yes | 2 | 6 | 1 | 2 | 3.24 (0.29–36.63) | 0.343 |
| Transfer from referral hospital | | | | | | |
| No | 23 | 61 | 60 | 88 | 1 | |
| Yes | 15 | 39 | 8 | 12 | 4.98 (1.56–15.93) | 0.007 |
| HIV Status | | | | | | |
| HIV negative | 34 | 89 | 63 | 93 | 1 | |
| HIV positive | 4 | 11 | 5 | 7 | 1.53 (0.29–8.11) | 0.615 |
| Time at risk (total) | | | | | | |
| ≤ 14 days | 17 | 45 | 44 | 65 | 1 | |
| > 14 days | 21 | 55 | 24 | 35 | 2.12 (0.97–4.62) | 0.059 |
| Time at risk (intensive care) | | | | | | |
| 1–7 days | 9 | 24 | 40 | 59 | 1 | |
| >7 days | 29 | 76 | 28 | 41 | 4.82 (1.80–12.91) | 0.002 |
| Surgery* | | | | | | |
| No | 14 | 37 | 33 | 49 | 1 | |
| Yes | 24 | 63 | 35 | 51 | 1.60 (0.72–3.56) | 0.254 |
| Exposure to antibiotics** | | | | | | |
| No | 5 | 13 | 27 | 40 | 1 | |
| Yes | 33 | 87 | 41 | 60 | 4.77 (1.38–16.48) | 0.014 |

*Refers to laparotomy or thoracotomy;

**Refers to receiving any dose or either a carbapenem or fluoroquinolone or aminoglycoside or third/fourth generation cephalosporin or piperacillin/tazobactam;

OR = odds ratio

doi:10.1371/journal.pone.0123337.t002

Discussion

To our knowledge this is the largest epidemiological study investigating risk factors and in-hospital mortality associated with clinical infection during an outbreak of NDM-1-producers, and adds evidence to support rational preventive and control measures. We found that higher Charlson co-morbidity scores, mechanical ventilation and piperacillin/tazobactam exposure were independently associated with infection with NDM-1-producers. Secondly, in-hospital mortality was found to be significantly higher in patients with clinical infection due to NDM-1-producers. Molecular strain typing of NDM-1-producing *Klebsiella pneumoniae* isolates from cases and the environment supported the hypothesis of horizontal transmission occurring in the ICU.

We identified three previously published papers reporting on risk factors for infection with NDM-1-producers. The first was a review of reported cases (n = 77) across the European Union which found travel to India, Pakistan or the Balkans to be associated with NDM-1

Table 3. Univariate analysis of exposure to antibiotics (aminoglycosides, fluoroquinolones, carbapenems, third/fourth generation cephalosporins and piperacillin/tazobactam), corticosteroids, invasive medical devices and selected medical interventions among cases and controls.

| Exposure Variable | Case patient (n = 38) with exposure mean (SD) | Control patient (n = 68) with exposure mean (SD) | Crude OR (95%CI) | p-value |
|---------------------------------|---|--|------------------|--------------|
| Aminoglycosides (dose, any) | 10.42 (±22.53) | 2.43 (±10.23) | 1.03 (1.00–1.06) | 0.043 |
| Gentamycin | 0.97 (±5.35) | 0.25 (±1.74) | 1.07 (0.93–1.23) | 0.320 |
| Amikacin | 7.29 (±18.79) | 2.17 (±10.05) | 1.02 (0.99–1.06) | 0.125 |
| Tobramycin | 2.16 (±13.30) | 0 (±0) | - | - |
| Fluoroquinolone (dose, any) | 1.53 (±3.75) | 0.91(±2.76) | 1.09 (0.96–1.24) | 0.162 |
| Ciprofloxacin | 0.71 (±3.02) | 0.16 (±1.00) | 1.19 (0.90–1.57) | 0.234 |
| Levofloxacin | 0.66 (±2.33) | 0.49 (±2.32) | 1.07 (0.91–1.26) | 0.429 |
| Moxifloxacin | 0.15 (±0.97) | 0.26 (±1.32) | 0.96 (0.67–1.38) | 0.830 |
| Carbapenem (dose, any) | 16.08(±29.93) | 5.59(±11.97) | 1.02 (1.00–1.05) | 0.062 |
| Doripenem | 6.16 (±18.43) | 0.15(±1.21) | 1.18 (0.96–1.46) | 0.117 |
| Ertapenem | 1.39 (±4.03) | 1.22 (±3.56) | 0.99 (0.88–1.12) | 0.930 |
| Meropenem | 8.52 (±16.74) | 4.22 (±11.17) | 1.02 (0.99–1.05) | 0.175 |
| Cephalosporin (dose, any) | 2.5 (±7.07) | 2.19 (±6.0) | 1.00 (0.94–1.06) | 0.992 |
| Cefepime | 1.68 (±6.43) | 0.51 (±3.07) | 1.06 (0.96–1.16) | 0.240 |
| Ceftriaxone | 0.82 (±2.82) | 1.67 (±4.93) | 0.93 (0.83–1.04) | 0.201 |
| Pip-tazobactam (dose) | 11.03 (±12.10) | 6.17 (±10.31) | 1.05 (1.02–1.10) | 0.015 |
| Steroids (dose, any) | 23.5 (±23.93) | 7.22 (±12.96) | 1.05 (1.02–1.09) | 0.003 |
| Invasive Medical Devices | | | | |
| Central venous line (days) | 15.42 (±14.66) | 6.51 (±6.71) | 1.08 (1.03–1.13) | 0.003 |
| Urinary catheter (days) | 18.61 (±15.92) | 7.35 (±7.93) | 1.07 (1.03–1.12) | 0.001 |
| Medical Interventions | | | | |
| Mechanical Ventilation (days) | 7.47 (±8.55) | 0.94 (±2.34) | 1.27 (1.10–1.48) | 0.001 |
| Parental Nutrition (days) | 2.53 (±3.40) | 1.40 (±3.83) | 1.07 (0.96–1.20) | 0.217 |
| Haemodialysis (days) | 6.03 (±14.3) | 0.68 (±2.74) | 1.16 (1.01–1.33) | 0.030 |

SD = standard deviation; OR = odds ratio.

doi:10.1371/journal.pone.0123337.t003

acquisition.[22] The second study was a case series (n = 5) of a nosocomial outbreak of carbapenem resistant enterobacteriaceae harbouring *bla*_{NDM-1} in Canada.[23] The third study by Lowe *et al.*[24] investigated nosocomial transmission of NDM-1 to seven patients from two index cases and found exposure to fluoroquinolones, trimethoprim-sulfamethoxazole and carbapenems to be possible risk factors for NDM-1 acquisition.[24] We found exposure to both carbapenems and fluoroquinolones to be associated, albeit not significantly, with subsequent infection due to a NDM-1-producer. We did not assess trimethoprim-sulfamethoxazole exposure in our study as it was not commonly prescribed in the setting of this outbreak. Our

Table 4. Multiple conditional logistic regression analysis for factors associated with case status.

| Exposure Variable | Adjusted OR (95% CI)* | p-value |
|-----------------------------------|-----------------------|--------------|
| Charlson co-morbidity index score | 1.59 (1.15–2.18) | 0.005 |
| Mechanical Ventilation (days) | 1.32 (1.10–1.59) | 0.003 |
| Piperacillin/tazobactam (dose) | 1.08 (1.02–1.15) | 0.013 |

* Adjusted for Charlson co-morbidity index score, mechanical ventilation and piperacillin/tazobactam; OR = odds ratio.

doi:10.1371/journal.pone.0123337.t004

Table 5. Risk factors associated with in-hospital mortality.

| Variable | Death (n = 31) n(%) | Unadjusted OR (95% CI) | p-value | Adjusted* OR (95% CI) | p-value |
|------------------------------|---------------------|------------------------|--------------|-----------------------|--------------|
| Case—Control | | | | | |
| Control | 10 (32) | 1 | | 1 | |
| Case | 21 (68) | 12.81 (2.94–55.82) | 0.001 | 11.29 (2.57–49.60) | 0.001 |
| Site of Infection | | | | | |
| None | 10 (32) | 1 | | 1 | |
| Pneumonia | 11 (36) | 5.5e (-) | 0.994 | 3.54e(-) | 0.993 |
| Blood stream infection | 8 (26) | 9.03 (1.10–74.21) | 0.041 | 8.84 (1.09–71.55) | 0.041 |
| Other | 2 (6) | 4.37 (0.37–51.24) | 0.240 | 3.51 (0.28–44.71) | 0.333 |
| Isolate | | | | | |
| None | 10 (32) | 1 | | 1 | |
| <i>Klebsiella pneumoniae</i> | 16 (52) | 19.30 (2.50–148.83) | 0.005 | 16.57 (2.12–129.6) | 0.007 |
| Other GNB | 5 (16) | 6.36 (0.72–56.51) | 0.097 | 6.08 (0.69–53.90) | 0.105 |

*Adjusted for Charlson co-morbidity index; OR = odds ratio; GNB = Gram-negative bacteria.

doi:10.1371/journal.pone.0123337.t005

analysis shows aminoglycoside and piperacillin/tazobactam exposure to be significantly associated with case status at the univariate level, and piperacillin/tazobactam was found to be associated with clinical infection with NDM-1-producers after adjusting for co-morbid disease.

Our findings that an increased duration of exposure to central venous lines, urinary catheters, mechanical ventilation and haemodialysis were associated with an increased risk of infection with NDM-1-producers are consistent with risk factors for the acquisition of carbapenemase-producers identified by previous investigators. Medical devices such as urinary catheters[25,26] and central venous lines[25–27] as well as interventions such as mechanical ventilation[25,27,28] and haemodialysis[25] are well-established risk factors. These risk factors have also been found consistent in the acquisition of IMP-type metallo-β-lactamase producing Gram-negatives.[29,30] This is the first study that identifies and quantifies these exposures for NDM-1-producers.

Of the early NDM-1 cases detected in the United States and United Kingdom, many had epidemiological links to India and Pakistan.[4,15] We found no association between international travel and case status. Despite not being able to complete the telephonic interview for all cases (32 completed/38, 84%) or controls (48 completed/68, 71%), we would argue that international travel was not a risk factor for NDM-1 acquisition in the cases linked to this nosocomial outbreak. Of the first five cases identified in the outbreak, none reported any travel history in the year preceding admission, and none of the cases interviewed telephonically reported travel to India, Pakistan or the Balkans, which had been identified as high NDM-1-transmission regions at the time of the outbreak.[4,15] In India, Gram-negative bacteria surveillance isolates collected two years prior to the first identification of NDM-1 has subsequently been shown to harbour *bla*_{NDM-1}. [31] Similarly, given the lack of standardised surveillance in South Africa, it is likely that *bla*_{NDM-1} had been present in clinically-relevant bacteria for some time before the index case was identified.

In-hospital mortality for extended-spectrum beta-lactamase producers has been reported at around 37%[32] and amongst patients with carbapenem-resistant *Klebsiella pneumoniae* at between 44% and 48%. [25,28] Crude mortality in patients with bloodstream infections caused by KPC-producing *Klebsiella pneumoniae* is estimated at 53%. [33] Given these reported mortality rates and the limited treatment options available for NDM-1-producers, our finding of a 55.3% crude in-hospital mortality rate was to be expected. However, considering this outbreak

occurred in a well-resourced private sector hospital, mortality rates in patients with similar infections cared for in public sector hospitals in South Africa would be expected to be higher due to limited available antibiotics and ICU facilities. This would likely be the case in many under-resourced healthcare facilities worldwide, which further underscores the importance of taking preventive action to reduce transmission of such multidrug-resistant organisms in the hospital setting, thereby preventing nosocomial outbreaks and limiting dissemination into the community.

The hospital undertook a range of interventions to control the outbreak. Patients found to be colonized with NDM-1-producers through the rectal screening program were cohorted and assigned dedicated nursing staff and medical equipment. Healthcare workers and cleaning staff received targeted education about the importance of hand hygiene, and stringent hand washing protocols were instituted throughout the hospital. Hand hygiene practice in the adult ICU was monitored for compliance through closed circuit television. Infrastructural alterations to the adult ICU increased the ICU's capacity to effectively isolate patients. Weekly meetings were attended by multidisciplinary role-players, including the hospital infection prevention and control practitioners, hospital management staff, medical microbiologists, hospital clinicians, and members of the National Institute for Communicable Diseases' Outbreak Response Unit. Controlling the outbreak was resource intensive and demanded a concerted effort from all role-players, with critical review of the outbreak situation and re-evaluation of interventional strategies throughout. Although sporadic cases of colonisation or infection with NDM-1-producers continue to be reported, no clusters or epidemiologically-linked cases have been identified since the end of the outbreak.

This study has a number of limitations. Due to the inherent nature of outbreak investigations, there were a limited number of potential cases. All potential cases were reviewed and as many matching controls as were available were included. However, the small sample size limits the study's power to detect other antimicrobial agents as risk factors for infection with NDM-1-producers. The outbreak was confined to the adult ICU, limiting generalisability to a paediatric population. Missing clinical records and missing data on international travel and previous admissions in the year leading up to the admission of interest reduced our sample size and ability to evaluate pre-hospitalization risk factors. The fluctuating point prevalence of NDM-1-producers and the clinicians' enhanced diagnostic suspicion of infection with NDM-1-producers as the outbreak evolved may bias findings. We addressed this, however, by matching controls for date of hospital admission.

Given the dearth of new antimicrobials in the drug development pipeline, the burgeoning threat of conquer by virtually untreatable multidrug-resistant organisms of clinical relevance is becoming realised thanks to the emergence and rapid spread of, amongst others, the carbapenemases.^[34,35] Through a better understanding of the risk factors and epidemiological characteristics of patients developing clinical infection with NDM-1-producers, infection prevention and control practice and antimicrobial stewardship programs can be tailored to identify vulnerable patients and prioritise areas for risk reduction, both in an outbreak situation and beyond. This study contributes to a growing body of knowledge for action by identifying risk factors for infection with NDM-1-producers, and highlights the 'bottom line'—such infections exact significant mortality and swift, effective action is needed.

Acknowledgments

We thank the following colleagues for their kind support and valuable assistance: Dr Trevor Frankish, Dr Steve Taylor, Trisha Fourie, Mariaan Greese, Joy Cleghorn, Chrismar Hattingh,

Melanie Janse van Vuuren, Dr Victor Matabane, Rob Stewart, Dr Leandra Blann, Dr Singh-Moodley and Professor Adriano Duse.

Author Contributions

Conceived and designed the experiments: PDJ JT. Performed the experiments: PDJ JT. Analyzed the data: PDJ TC. Contributed reagents/materials/analysis tools: OP. Wrote the paper: PDJ JT OP TC SN.

References

1. Bradford PA. Extended-Spectrum β -Lactamases in the 21st Century: Characterization, Epidemiology, and Detection of This Important Resistance Threat. *Clin Microbiol Rev.* 2001; 14: 933–951. doi: [10.1128/CMR.14.4.933-951.2001](https://doi.org/10.1128/CMR.14.4.933-951.2001) PMID: [11585791](https://pubmed.ncbi.nlm.nih.gov/11585791/)
2. Jacoby GA, Munoz-Price LS. The new β -lactamases. *New England Journal of Medicine.* 2005; 352: 380–391. PMID: [15673804](https://pubmed.ncbi.nlm.nih.gov/15673804/)
3. Nordmann P, Poirel L. Emerging carbapenemases in Gram-negative aerobes. *Clinical Microbiology and Infection.* 2002; 8: 321–331. doi: [10.1046/j.1469-0691.2002.00401.x](https://doi.org/10.1046/j.1469-0691.2002.00401.x) PMID: [12084099](https://pubmed.ncbi.nlm.nih.gov/12084099/)
4. Gupta N, Limbago BM, Patel JB, Kallen AJ. Carbapenem-resistant enterobacteriaceae: epidemiology and prevention. *Clinical infectious diseases.* 2011; 53: 60. doi: [10.1093/cid/cir202](https://doi.org/10.1093/cid/cir202) PMID: [21653305](https://pubmed.ncbi.nlm.nih.gov/21653305/)
5. Arias CA, Murray BE. Antibiotic-Resistant Bugs in the 21st Century—A Clinical Super-Challenge. *New England Journal of Medicine.* 2009; 360: 439–443. doi: [10.1056/NEJMp0804651](https://doi.org/10.1056/NEJMp0804651) PMID: [19179312](https://pubmed.ncbi.nlm.nih.gov/19179312/)
6. Yigit H, Queenan AM, Anderson GJ, Domenech-Sanchez A, Biddle JW, Steward CD, et al. Novel Carbapenem-Hydrolyzing β -Lactamase, KPC-1, from a Carbapenem-Resistant Strain of *Klebsiella pneumoniae*. *Antimicrob Agents Chemother.* 2001; 45: 1151–1161. doi: [10.1128/AAC.45.4.1151-1161.2001](https://doi.org/10.1128/AAC.45.4.1151-1161.2001) PMID: [11257029](https://pubmed.ncbi.nlm.nih.gov/11257029/)
7. Brink AJ, Coetzee J, Clay CG, Sithole S, Richards GA, Poirel L, et al. Emergence of New Delhi metallo-beta-lactamase (NDM-1) and *Klebsiella pneumoniae* carbapenemase (KPC-2) in South Africa. *Journal of Clinical Microbiology.* 2012; 50: 525–527. doi: [10.1128/JCM.05956-11](https://doi.org/10.1128/JCM.05956-11) PMID: [22116157](https://pubmed.ncbi.nlm.nih.gov/22116157/)
8. Nordmann P, Poirel L, Walsh TR, Livermore DM. The emerging NDM carbapenemases. *Trends Microbiol.* 2011; 19: 588–595. doi: [10.1016/j.tim.2011.09.005](https://doi.org/10.1016/j.tim.2011.09.005) PMID: [22078325](https://pubmed.ncbi.nlm.nih.gov/22078325/)
9. Livermore DM. Has the era of untreatable infections arrived? *J Antimicrob Chemother.* 2009; 64: i29–i36. doi: [10.1093/jac/dkp255](https://doi.org/10.1093/jac/dkp255) PMID: [19675016](https://pubmed.ncbi.nlm.nih.gov/19675016/)
10. Bonomo RA. New Delhi Metallo- β -Lactamase and Multidrug Resistance: A Global SOS? *Clin Infect Dis.* 2011; 52: 485–487. doi: [10.1093/cid/ciq179](https://doi.org/10.1093/cid/ciq179) PMID: [21258101](https://pubmed.ncbi.nlm.nih.gov/21258101/)
11. Molton JS, Tambyah PA, Ang BSP, Ling ML, Fisher DA. The Global Spread of Healthcare-Associated Multidrug-Resistant Bacteria: A Perspective From Asia. *Clin Infect Dis.* 2013; 56: 1310–1318. doi: [10.1093/cid/cit020](https://doi.org/10.1093/cid/cit020) PMID: [23334810](https://pubmed.ncbi.nlm.nih.gov/23334810/)
12. Moellering RC Jr. NDM-1—a cause for worldwide concern. *New England Journal of Medicine.* 2010; 363: 2377–2379. doi: [10.1056/NEJMp1011715](https://doi.org/10.1056/NEJMp1011715) PMID: [21158655](https://pubmed.ncbi.nlm.nih.gov/21158655/)
13. Walsh TR. Emerging carbapenemases: a global perspective. *International Journal of Antimicrobial Agents.* 2010; 36, Supplement 3: S8–S14. doi: [10.1016/S0924-8579\(10\)70004-2](https://doi.org/10.1016/S0924-8579(10)70004-2)
14. Nordmann P, Poirel L, Toleman MA, Walsh TR. Does broad-spectrum β -lactam resistance due to NDM-1 herald the end of the antibiotic era for treatment of infections caused by Gram-negative bacteria? *J Antimicrob Chemother.* 2011; 66: 689–692. doi: [10.1093/jac/dkq520](https://doi.org/10.1093/jac/dkq520) PMID: [21393184](https://pubmed.ncbi.nlm.nih.gov/21393184/)
15. Kumarasamy KK, Toleman MA, Walsh TR, Bagaria J, Butt F, Balakrishnan R, et al. Emergence of a new antibiotic resistance mechanism in India, Pakistan, and the UK: a molecular, biological, and epidemiological study. *The Lancet Infectious Diseases.* 2010; 10: 597–602. doi: [10.1016/S1473-3099\(10\)70143-2](https://doi.org/10.1016/S1473-3099(10)70143-2) PMID: [20705517](https://pubmed.ncbi.nlm.nih.gov/20705517/)
16. Nordmann P, Couard J-P, Sansot D, Poirel L. Emergence of an Autochthonous and Community-Acquired NDM-1–Producing *Klebsiella pneumoniae* in Europe. *Clin Infect Dis.* 2012; 54: 150–151. doi: [10.1093/cid/cir720](https://doi.org/10.1093/cid/cir720) PMID: [21960718](https://pubmed.ncbi.nlm.nih.gov/21960718/)
17. Marais E, Moodley A, Govender N, Kularatne R, Thomas J, Duse A. Clusters of *Klebsiella pneumoniae* infection in neonatal intensive care units in Gauteng: scientific letter. *South African Medical Journal.* 2006; 96: p–813. PMID: [17068651](https://pubmed.ncbi.nlm.nih.gov/17068651/)
18. Horan TC, Andrus M, Dudeck MA. CDC/NHSN surveillance definition of health care–associated infection and criteria for specific types of infections in the acute care setting. *American journal of infection control.* 2008; 36: 309–332. doi: [10.1016/j.ajic.2008.03.002](https://doi.org/10.1016/j.ajic.2008.03.002) PMID: [18538699](https://pubmed.ncbi.nlm.nih.gov/18538699/)

19. Charlson ME, Pompei P, Ales KL, MacKenzie CR. A new method of classifying prognostic comorbidity in longitudinal studies: development and validation. *Journal of chronic diseases*. 1987; 40: 373–383. PMID: [3558716](#)
20. Higgins TL, Teres D, Copes WS, Nathanson BH, Stark M, Kramer AA. Assessing contemporary intensive care unit outcome: An updated Mortality Probability Admission Model (MPM0-III)*. *Critical care medicine*. 2007; 35: 827–835. PMID: [17255863](#)
21. Tenover FC, Arbeit RD, Goering RV, Mickelsen PA, Murray BE, Persing DH, et al. Interpreting chromosomal DNA restriction patterns produced by pulsed-field gel electrophoresis: criteria for bacterial strain typing. *Journal of clinical microbiology*. 1995; 33: 2233. PMID: [7494007](#)
22. Struelens MJ, Monnet DL, Magiorakos AP, Santos O'Connor F, Giesecke J. New Delhi metallo-beta-lactamase 1-producing Enterobacteriaceae: emergence and response in Europe. *Euro Surveill*. 2010; 15. Available: <http://www.ncbi.nlm.nih.gov/pubmed/21144431>
23. Borgia S, Lastovetska O, Richardson D, Eshaghi A, Xiong J, Chung C, et al. Outbreak of Carbapenem-Resistant Enterobacteriaceae Containing blaNDM-1, Ontario, Canada. *Clin Infect Dis*. 2012; doi: [10.1093/cid/cis737](#)
24. Lowe CF, Kus JV, Salt N, Callery S, Louie L, Khan MA, et al. Nosocomial Transmission of New Delhi Metallo- β -Lactamase-1-Producing *Klebsiella pneumoniae* in Toronto, Canada. *Infection Control and Hospital Epidemiology*. 2013; 34: 49–55. doi: [10.1086/668778](#) PMID: [23221192](#)
25. Schwaber MJ, Klarfeld-Lidji S, Navon-Venezia S, Schwartz D, Leavitt A, Carmeli Y. Predictors of Carbapenem-Resistant *Klebsiella pneumoniae* Acquisition among Hospitalized Adults and Effect of Acquisition on Mortality. *Antimicrob Agents Chemother*. 2008; 52: 1028–1033. doi: [10.1128/AAC.01020-07](#) PMID: [18086836](#)
26. Tuon FF, Rocha JL, Toledo P, Arend LN, Dias CH, Leite TM, et al. Risk factors for KPC-producing *Klebsiella pneumoniae* bacteremia. *Braz J Infect Dis*. 2012; 16: 416–419. doi: [10.1016/j.bjid.2012.08.006](#) PMID: [22980584](#)
27. Hyle EP, Ferraro MJ, Silver M, Lee H, Hooper DC. Ertapenem-Resistant Enterobacteriaceae: Risk Factors for Acquisition and Outcomes. *Infection Control and Hospital Epidemiology*. 2010; 31: 1242–1249. doi: [10.1086/653029](#) PMID: [21029005](#)
28. Gopi Patel M, Shirish Huprikar M, Stephanie H. Factor M MPH, Stephen G. Jenkins P, David P. Calfee M MS. Outcomes of Carbapenem-Resistant *Klebsiella pneumoniae* Infection and the Impact of Antimicrobial and Adjunctive Therapies. *Infection Control and Hospital Epidemiology*. 2008; 29: 1099–1106. doi: [10.1086/596025](#) PMID: [18973455](#)
29. Hirakata Y, Yamaguchi T, Nakano M, Izumikawa K, Mine M, Aoki S, et al. Clinical and Bacteriological Characteristics of IMP-Type Metallo- β -Lactamase-Producing *Pseudomonas aeruginosa*. *Clin Infect Dis*. 2003; 37: 26–32. doi: [10.1086/375594](#) PMID: [12830405](#)
30. Peleg AY, Franklin C, Bell JM, Spelman DW. Dissemination of the Metallo- β -Lactamase Gene blaIMP-4 among Gram-Negative Pathogens in a Clinical Setting in Australia. *Clin Infect Dis*. 2005; 41: 1549–1556. doi: [10.1086/497831](#) PMID: [16267725](#)
31. Castanheira M, Deshpande LM, Mathai D, Bell JM, Jones RN, Mendes RE. Early Dissemination of NDM-1- and OXA-181-Producing Enterobacteriaceae in Indian Hospitals: Report from the SENTRY Antimicrobial Surveillance Program, 2006–2007. *Antimicrob Agents Chemother*. 2011; 55: 1274–1278. doi: [10.1128/AAC.01497-10](#) PMID: [21189345](#)
32. Szilágyi E, Fűzi M, Damjanova I, Böröcz K, Szőnyi K, Tóth á., et al. Investigation of extended-spectrum beta-lactamase-producing <i>Klebsiella pneumoniae</i> outbreaks in Hungary between 2005 and 2008. *Acta Microbiologica et Immunologica Hungarica*. 2010; 57: 43–53. doi: [10.1556/AMicr.57.2010.1.4](#) PMID: [20350878](#)
33. Zarkotou O, Pourmaras S, Tselioti P, Dragoumanos V, Pitiriga V, Ranellou K, et al. Predictors of mortality in patients with bloodstream infections caused by KPC-producing *Klebsiella pneumoniae* and impact of appropriate antimicrobial treatment. *Clinical Microbiology and Infection*. 2011; 17: 1798–1803. doi: [10.1111/j.1469-0691.2011.03514.x](#) PMID: [21595793](#)
34. Boucher HW, Talbot GH, Bradley JS, Edwards JE, Gilbert D, Rice LB, et al. Bad Bugs, No Drugs: No ESKAPE! An Update from the Infectious Diseases Society of America. *Clin Infect Dis*. 2009; 48: 1–12. doi: [10.1086/595011](#) PMID: [19035777](#)
35. Freire-Moran L, Aronsson B, Manz C, Gyssens IC, So AD, Monnet DL, et al. Critical shortage of new antibiotics in development against multidrug-resistant bacteria—Time to react is now. *Drug Resistance Updates*. 2011; 14: 118–124. doi: [10.1016/j.drug.2011.02.003](#) PMID: [21435939](#)