# Systematic revision of the genus Isodacrys Sharp, 1911 (Coleoptera: Curculionidae: Entiminae: Tanymecini) 

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#### Abstract

The genus Isodacrys Sharp, 1911 is revised. Twenty species of the genus are recognized ranging from south United States of America, Mexico, Guatemala and Honduras of which eight are herein described as new. These species are Isodacrys antrum Cortés-Hernández, new species (Mexico: Tamaulipas, Chiapas; Guatemala: Baja Verapaz); Isodacrys carlae Cortés-Hernández, new species (Mexico: Coahuila, Hidalgo, Nuevo León, San Luis Potosí, Tamaulipas); Isodacrys confusum Cortés-Hernández, new species (Mexico: Tamaulipas); Isodacrys fasciatum Cortés-Hernández, new species (Mexico: Coahuila, Durango, Nuevo León); Isodacrys frontalis Cortés-Hernández, new species (Mexico: Oaxaca; Guatemala: Sacatepéquez, Guatemala); Isodacrys kuchii Cortés-Hernández, new species (Mexico: Puebla); Isodacrys obrienorum Cortés-Hernández, new species (Guatemala: Totonicapán, Jalapa, San Marcos); and Isodacrys okuiltontli Cortés Hernández, new species (Mexico: Oaxaca). Insights into the monophyly of Isodacrys and its phylogenetic relationships with other Tanymecini based on adult morphology are given by implementing a phylogenetic analysis of 43 terminals ( 21 ingroup, 22 outgroup) coded for 72 adult morphological characters. Characters were discussed and highlighted for the inclusion in the phylogenetic analysis. Final analysis yielded two most-parsimonious cladograms of 242 steps, which support the monophyly of Isodacrys. Isodillex Cortés-Hernández, new genus is here described to accommodate Isodillex minutum (Sharp, 1911), new combination and Isodillex plumosum Cortés-Hernández, new species (Mexico: Zacatecas). Isodillex was recovered as sister group of Isodacrys. Key to separate Isodacrys species, occurrence map and habitus photographs are also provided.


Subjects Biodiversity, Entomology, Taxonomy, Zoology
Keywords Coleoptera, Curculionidae, Systematics, Phylogenetics, New species

## INTRODUCTION

Tanymecini (Curculionidae: Entiminae) is a cosmopolitan tribe of broad-nosed weevils with nearly 96 genera, including fossils (Alonso-Zarazaga \& Lyal, 1999). The tribe has been traditionally characterized by the presence of modified setae at the anterolateral margin of the prothorax called "postocular vibrissae" (Lacordaire, 1863; LeConte \& Horn, 1876; Van Emden, 1944; Howden, 1970), which are long, stout setae projecting towards eyes.

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They are commonly set on a tuft (as opposed to a fringe) and may be reduced in number or size. In addition to the presence of postocular vibrissae, which is not unique to this weevil tribe, there are additional characters to identify them: eyes in lateral view nearly round; antennal scape usually not reaching anterior margin of prothorax; anterior margin of prothorax in lateral view straight to dorsally produced anteriorly, never forming postocular lobes; mesepimeron not ascending, similar in size to mesepisternum; and metapisternum dilated at its anterior end, with acute process projected inwards between the mesepimeron and the metasternum (Lacordaire, 1863; LeConte \& Horn, 1876).

Only the subtribe Tanymecina is represented in the New World (Del Río \& Lanteri, 2019b), with 16 genera and approximately 311 described species. The study of the Tanymecini of the Americas has been greatly advanced by the contributions of Anne Howden, who studied them for over 50 years (Anderson \& Smith, 2017; Anderson, 2019). During that time, Howden described several genera and numerous species (Howden, 1959, 1961, 1966, 1970, 1982, 2011), highlighting adult characters potentially phylogenetically informative that had not been considered before. Nonetheless, the relationships and boundaries among New World tanymecines remain poorly understood (Howden, 1993a).

Based on Howden's remarkable contributions, one of the logical and subsequent steps to contribute to the knowledge of the American Tanymecini is the discussion and evaluation of the boundaries among these lineages and discussion of their evolutionary history through phylogenetic analyses (Gillet et al., 2018). As taxonomic concepts, relationships and classifications within Curculionidae are frequently tentative or poorly supported, the choice among mid-level classifications generally remains a matter of author preferences (Oberprieler, 2014). Nevertheless, focusing on a taxonomically manageable lineage such as a genus may yield rather compelling results and contribute to our understanding of weevil evolution (Franz \& Engel, 2010). In this context, some recent advances towards understanding the phylogenetic relationships among New World Tanymecini have been made (Jansen \& Franz, 2015, 2018). The present study focuses on the delimitation of Isodacrys and its phylogenetic relationships with other Tanymecini.

Isodacrys Sharp, 1911 is a genus of flightless tanymecines comprised of 13 described species, distributed from southern United States of America to Honduras. Little is known about their natural history: adults have been found on at least nine plant families, feeding on leaves (Burke, 1959) and stems, or in leaf litter and under rocks. Isodacrys species have been found in a variety of environments from xerophytic vegetation to pine-oak forests. Because males are unknown in six of the thirteen described species, it has been proposed that they may reproduce by parthenogenesis (Howden, 1961), as it has been confirmed to be the case in other broad-nosed weevils (Lanteri \& Normark, 1995). Immature stages remain unknown, but it is assumed that they feed in the soil on roots as other tanymecines do (Howden, 1993a).

The history of the classification of Isodacrys is summarized as follows. The genus was described and initially placed within Sciaphilina by Sharp (1911), based on the absence of postocular lobes and its apterous condition, even though it does not have connate tarsal claws as in other Sciaphilina. It included five species: Isodacrys guatemalenum Sharp, 1911, I. minutum Sharp, 1911, I. mexicanum Sharp, 1911, I. orizabae Sharp, 1911 and
I. schwarzi Champion, 1911. Pierce (1913) subsequently designated I. guatemalenum as the type species of the genus and transferred Pandeleteius ovipennis Schaeffer, 1908 to Isodacrys. Apparently based on Sharp's observations, Pierce included Isodacrys, Pandeleteius Schönherr, 1834, Polydacrys Schönherr, 1834 and Isodrusus Sharp, 1911 in a new tribe named Pandeleteini, within the subfamily Tanymecinae (Mitchell \& Pierce, 1911), which was characterized by the frequent reduction or absence of postocular vibrissae and separated procoxae. Posteriorly, Van Emden (1944) omitted Pierce's amendments and included all Tanymecinae within Tanymecini (sensu LeConte \& Horn, 1876). Howden (1961) reviewed Isodacrys and supported its inclusion within Tanymecini, highlighting its affinities with Pandeleteinus Champion, 1911, Isodrusus and Minyomerus Horn, 1876, and posteriorly with the West Indian genera Paululusus Howden, 1970 and Paradacrys Howden, 1970 (Howden, 1970).

Regardless of efforts to elucidate the boundaries of Isodacrys based on adult morphology, many of the character states among putatively related genera overlap or are poorly understood, requiring further discussion of putative homologies that remain obscure. Following a phylogenetic analysis using explicitly defined morphological adult character states, the circumscription of Isodacrys is herein reexamined and insights into its generic interrelationships are discussed. Although Isodacrys seems to be seldom collected and therefore uncommon in collections, an extensive number of specimens were obtained for study from various museums. As a result, eight Isodacrys species new to science are described here. A comprehensive phylogenetic analysis was conducted for 43 terminals ( 21 outgroups, 22 ingroups) accompanied by a list of the characters included in the analysis and a brief discussion of them, when necessary. From the 72 characters included ( 53 binary and 19 multistate), 63 comprised external morphology and nine from male and female genitalia. The monophyly of Isodacrys is supported by one synapomorphy and two homoplastic characters. In addition, Isodillex Cortés-Hernández, new genus is hereby erected to accommodate Isodillex minutum (Sharp, 1911), new combination and Isodillex plumosum Cortés-Hernández, new species. Isodillex is recovered as the sister group to Isodacrys.

## MATERIALS AND METHODS

## Taxon sampling

Tribal concepts follow Alonso-Zarazaga \& Lyal (1999), except for the genus Platyaspistes Schönherr, 1840 which is considered to belong to Leptopiini Oke, 1951 instead of Tropiphorini Marseul, 1863 (see Marvaldi et al., 2018). An exemplar approach based on morphology was followed (Prendini, 2001). Given that generic relationships among New World Tanymecini remain obscure and that there has only been one specific attempt to estimate the boundaries of one genus (Jansen \& Franz, 2015), a comprehensive taxon sampling (Maddison, Donoghue \& Maddison, 1984; Nixon \& Carpenter, 1993) of putatively related genera (Howden, 1959, 1961, 1963, 1969, 1970) was intended. A total of 43 species was included in order to test the monophyly of Isodacrys and elucidate its relationships with other members of the tribe.

The outgroup comprised 21 species representing eight tanymecine genera (Table 1), and one member of the tribes Leptopiini (Platyaspistes prasinus [Erichson, 1834], recently transferred from Piazomiina Reitter (1913) by Marvaldi et al. (2018) and Naupactini Gistel (1856) (Megalostylus albicans [Lacordaire]), all distributed in the Americas. Specimens of the type species of each outgroup genus were included when available. The ingroup contains 22 species, including the 13 previously described Isodacrys species and eight additional herein described. Isodillex plumosum new species and I. minutum new combination (see results) were also included as part of the ingroup. Specimens of Isodacrys schwarzi Champion, 1911 were not available for study, nonetheless, the species was scored and included in the analysis based on habitus photographs of a "cotype" provided by the United States National Museum and Howden's (1961) redescription. A list of the 43 species included in the analysis is provided (Table 1).

The specimens examined are deposited in the following institutions: MZFC, Museo de Zoología "Alfonso L. Herrera", Facultad de Ciencias, Universidad Nacional Autónoma de México, Mexico City, Mexico (Juan J. Morrone); IEXA, Colección Entomológica del Instituto de Ecología A.C., Xalapa, Veracruz, Mexico (Leonardo Delgado); CNIN, Colección Nacional de Insectos, Instituto de Biología, Universidad Nacional Autónoma de México, Mexico City, Mexico (Alejandro Zaldívar); UAQE, Colección de Insectos de la Licenciatura en Biología, Facultad de Ciencias Naturales de la Universidad Autónoma de Querétaro, Campus Juriquilla, Querétaro, Mexico (Robert W. Jones); ICZ, Insect Collection of Zacapa, Instituto de Investigaciones del Centro Universitario de Zacapa, Universidad de San Carlos de Guatemala, Zacapa, Guatemala (Manuel A. Barrios-Izás); CMNC, Canadian Museum of Nature Collection, Ottawa, Ontario, Canada (François Génier); TAMUIC, Texas A\&M University Insect Collection, Department of Entomology, Texas A\&M University, College Station, Texas, United States of America (Karen W. Wright); CAS, California Academy of Sciences, Department of Entomology, San Francisco, California, United States of America (Christopher C. Grinter); ASUCOB, Arizona State University Charles O'Brien Collection, Tempe, Arizona, United States of America (Emmy Engasser); and CNC, Canadian National Collection of Insects, Arachnids and Nematodes, Agriculture and Agri-Food Canada, Ottawa, Canada (Patrice Bouchard).

## Morphological analysis

The analysis was based on 72 discrete characters ( 53 binary and 19 multistate) of the adults, 63 from external morphology, five from female genitalia and four from male genitalia. Observations and measurements of externally visible and dissected structures were made with a Leica MZ6 stereomicroscope. Habitus photographs were taken using a Leica Z16 APOA stereomicroscope and multilayer images were processed using Leica Application Suite version 4.3 .0 software. Photographs of head, ventrites and legs were taken with a Hitachi SU3500 II Scanning Electron Microscope. Genitalia photographs were taken using an AXIO Zoom.V16 stereomicroscope equipped with an AxioCam MRc5 and images were processed using ZEN 2012 software. Illustrations were prepared to exemplify multiple character states, which are highlighted with arrows, with an indication of character numbers and applicable states given in parentheses.

| Taxon | Distribution |
| :---: | :---: |
| Tribe Naupactini Gistel |  |
| Megalostylus albicans (Lacordaire) | Mexico |
| Tribe Leptopiini Oke |  |
| Platyaspistes prasinus (Erichson, 1834)* | Chile |
| Tribe Tanymecini Lacordaire |  |
| Hadromeropsis brevicoma Howden, 1982 | Mexico |
| Minyomerus microps (Say, 1831)* | Canada, United states of America |
| Minyomerus laticeps (Casey, 1888) | United states of America, Mexico |
| Pandeleteius hilaris (Herbst, 1797)* | Canada, United states of America |
| Pandeleteius rotundicollis (Fall, 1907) | United states of America, Mexico |
| Pandeleteius inflatus Champion, 1911 | Mexico |
| Scalaventer cyrillae Howden, 1970* | Jamaica |
| Scalaventer jamaicensis Howden, 1970 | Jamaica |
| Scalaventer subtropicus (Fall, 1907) | United states of America, Cuba |
| Paululusus hispaniolae Howden, 1970 | Dominican Republic, Haiti |
| Paululusus constanzae Howden, 1970 | Dominican Republic |
| Isodrusus debilis Sharp, 1911* | United States of America, Mexico, Guatemala, Honduras |
| Isodrusus guajavus, Howden, 1970 | Jamaica |
| Aff. Isodrusus sp. | Colombia |
| Pandeleteinus submetallicus (Schaeffer, 1908)* | United states of America, Mexico |
| Pandeleteinus subcancer Howden, 1969 | Mexico |
| Pandeleteinus elytroplanatus Howden, 1959 | United States of America, Mexico |
| Aff. Pandeleteinus sp. | Brazil |
| Paradacrys ensiformis Howden, 1970 | Bahamas |
| Isodillex minutum (Sharp, 1911) new combination | Mexico |
| Isodillex plumosum new species | Mexico |
| Isodacrys antrum new species | Mexico, Guatemala |
| Isodacrys apicale Howden, 1961 | Mexico |
| Isodacrys brevirostre Howden, 1961 | Mexico |
| Isodacrys buchanani Howden, 1961 | United States of America, Mexico |
| Isodacrys burkei Howden, 1961 | United States of America |
| Isodacrys carlae new species | Mexico |
| Isodacrys confusum new species | Mexico |
| Isodacrys crispum Howden, 1961 | Mexico |
| Isodacrys ellipticum Howden, 1961 | Guatemala, Honduras |
| Isodacrys fasciatum new species | Mexico |
| Isodacrys frontalis new species | Mexico, Guatemala |
| Isodacrys geminatum Howden, 1961 | Mexico |
| Isodacrys guatemalenum Sharp, 1911* | Guatemala, Honduras |
| Isodacrys kuchii new species | Mexico |

(Continued)

| Table 1 (continued) |  |
| :--- | :--- |
| Taxon | Distribution |
| Isodacrys mexicanum Sharp, 1911 | Mexico |
| Isodacrys obrienorum new species | Guatemala |
| Isodacrys okuiltontli new species | Mexico |
| Isodacrys orizabae Sharp, 1911 | Mexico |
| Isodacrys ovipennis (Schaeffer, 1908) | United States of America, Mexico |
| Isodacrys schwarzi Champion, 1911 | Mexico |

Selection of characters was based on previous literature as noted in the character discussion or based on original observations and comparison of the studied species. Terminology for the external morphology was mainly in accordance with Torre-Bueno (Nichols, 1989) and Howden (1959, 1961, 1969, 1970). Additional specialized terms were used for female (Howden, 1995, 2011; Lanteri \& Del Río, 2008) and male genitalia (Bruhn, 1947; Wanat, 2007; Oberprieler, Anderson \& Marvaldi, 2014). Specifically, we use the term temones for the apodemes of the median lobe, the term manubrium for the apodeme of tegmen and the term spiculum gastrale for the apodeme of sternite IX (sensu Oberprieler, Anderson \& Marvaldi, 2014).

## Phylogenetic analyses

Character statements (i.e., characters and character states) are in accordance with Sereno (2007). An iterative approach was applied to reevaluate and improve homology statements (Hennig, 1966; Franz, 2005). As part of the phylogenetic research cycle, excessively vague and homoplastic characters were reformulated to reflect more accurately synapomorphic conditions in particular lineages (Jenner, 2004; Franz, 2012, 2014). Character states of species that could not be observed were treated as missing data (Maddison, 1993), scored with a "?". Character states with inapplicable entries on various terminals were scored with a "-", based on absence statements of the structures under study (Maddison, 1993; Hawkins, Hughes \& Scotland, 1997).

The data matrix of 43 terminal taxa and 72 morphological characters (Table 2; Supplemental Information 1) was compiled and edited using WINCLADA version 1.00.08 (Nixon, 2002). Phylogenetic analyses were performed under maximum parsimony as optimality criterion in TNT v1.5 (Goloboff \& Catalano, 2016) using heuristic traditional search under equal weights. Cladograms obtained were rooted with Megalostylus albicans Champion, 1911 (Entiminae: Naupactini) based on recent analyses of Entiminae weevils depicting the phylogenetic closeness of Naupactini and Tanymecini (Marvaldi et al., 2018; contra Gillet et al., 2018). Most parsimonious trees (MPT's) were consistently obtained through several rounds of analyses modifying the number of replicates and trees retained per replicate. Final trees were obtained with the following commands: random seed $=1,300$ random addition sequences, swapping algorithm tree bisection and reconnection (TBR), holding 200 trees per replication. From the equally most parsimonious trees, a strict consensus was calculated. The resulting cladograms and

Table 2 Data matrix of 43 taxa and 72 morphological characters of Isodacrys and outgroups, used for the cladistic analysis. Inapplicable and missing character states are indicated with a "-" and a "?", respectively. Numbers inside square brackets represent polymorphisms.

| Taxon/character | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | 16 | 17 | 18 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Megalostylus albicans | 0 | 0 | 0 | 0 | 0 | 0 | 0 | - | - | - | 0 | 0 | 0 | 0 | 0 | 0 | - | - |
| Platyaspistes prasinus | 0 | 0 | 1 | 0 | 0 | 0 | 0 | - | - | - | 0 | 0 | 0 | 0 | 0 | 0 | - | - |
| Hadromeropsis brevicoma | 1 | 0 | 1 | 0 | 0 | 0 | 0 | - | - | - | 1 | 0 | 0 | 1 | 0 | 0 | - | - |
| Minyomerus microps | 2 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | - | - |
| Minyomerus laticeps | 2 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | - | - |
| Pandeleteius hilaris | 0 | 1 | 1 | 2 | - | 1 | 1 | 1 | 0 | 0 | 1 | 0 | 1 | 1 | 1 | 0 | - | - |
| Pandeleteius rotundicollis | 0 | 1 | 1 | 0 | 0 | 1 | 1 | 0 | 1 | 0 | 1 | 0 | 0 | 1 | 1 | 0 | - | - |
| Pandeleteius inflatus | 0 | 1 | 1 | 2 | - | 1 | 1 | 1 | 0 | 0 | 1 | 0 | 0 | 1 | 1 | 0 | 0 | 1 |
| Scalaventer cyrillae | 0 | 1 | 2 | 0 | 1 | 1 | 1 | 0 | 1 | 1 | 0 | 0 | 1 | 1 | 2 | 1 | 2 | 1 |
| Scalaventer jamaicensis | 0 | 1 | 2 | 0 | 1 | 1 | 1 | 0 | 1 | 1 | 0 | 0 | 1 | 1 | 2 | 0 | 2 | 1 |
| Scalaventer subtropicus | 0 | 1 | 2 | 0 | 1 | 1 | 1 | 0 | 1 | 1 | 0 | 0 | 1 | 1 | 2 | 1 | 2 | 1 |
| Paululusus hispaniole | 0 | 1 | 2 | 0 | 1 | 1 | 1 | 2 | 1 | 1 | 0 | 0 | 1 | 1 | 2 | 0 | 1 | 0 |
| Paululusus constanzae | 0 | 1 | 2 | 0 | 1 | 1 | 1 | 2 | 1 | 1 | 0 | 0 | 1 | 1 | 2 | 0 | 1 | 0 |
| Isodrusus debilis | 1 | 1 | 1 | 1 | - | 1 | 1 | 2 | 0 | 0 | 0 | 0 | 0 | 1 | 4 | 1 | 2 | 2 |
| Isodrusus guajavus | 0 | 1 | 2 | 1 | - | 1 | 1 | 2 | 0 | 0 | 0 | 0 | 1 | 1 | 4 | 1 | 2 | 2 |
| aff. Isodrusus sp. | 0 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 0 | 0 | 0 | 1 | 1 | 3 | 1 | 2 | 2 |
| Pandeleteinus submetallicus | 1 | 1 | 1 | 0 | 0 | 0 | 1 | 2 | 0 | 0 | 1 | 0 | 0 | 1 | 2 | 0 | 2 | 0 |
| Pandeleteinus subcancer | 1 | 1 | 1 | 0 | 0 | 1 | 1 | 2 | 0 | 0 | 0 | 0 | 1 | 1 | 2 | 1 | 2 | 1 |
| Pandeleteinus elytroplanatus | 1 | 1 | 1 | 0 | 0 | 1 | 1 | 2 | 0 | 0 | 1 | 0 | 0 | 1 | 2 | 1 | 2 | 0 |
| aff Pandeleteinus sp. | 0 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 0 | 0 | 0 | 1 | 1 | 3 | 1 | 2 | 2 |
| Paradacrys ensiformis | 0 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 1 | 0 | 0 | 1 | 1 | 3 | 0 | 2 | 1 |
| Isodillex minutum | 0 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 0 | 0 | 0 | 0 | 1 | 2 | 1 | 1 | 2 |
| Isodillex plumosum | 0 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 0 | 0 | 0 | 0 | 1 | 2 | 1 | 1 | 2 |
| Isodacrys antrum | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 0 | 0 | 1 | 0 | 1 | 3 | 1 | 2 | 2 |
| Isodacrys apicale | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 0 | 0 | 0 | 1 | 1 | 2 | 1 | 1 | 0 |
| Isodacrys brevirostre | [01] | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 0 | 0 | 1 | 0 | 1 | 2 | 1 | 1 | 1 |
| Isodacrys buchanani | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 0 | 0 | 0 | 1 | 1 | 2 | 1 | 2 | 1 |
| Isodacrys burkei | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 1 | 0 | 0 | 0 | 1 | 2 | 1 | 2 | 0 |
| Isodacrys carlae | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 0 | 0 | 0 | 0 | 1 | 2 | 1 | 2 | 0 |
| Isodacrys confusum | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 0 | 0 | 0 | 0 | 1 | 2 | 1 | 2 | 0 |
| Isodacrys crispum | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 0 | 0 | 0 | 0 | 1 | 3 | 1 | 2 | 1 |
| Isodacrys ellipticum | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 1 | 0 | 0 | 1 | 1 | 2 | 1 | 1 | 1 |
| Isodacrys fasciatum | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 0 | 0 | 0 | 0 | 1 | 2 | 1 | 2 | 0 |
| Isodacrys frontalis | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 0 | 0 | 1 | 1 | 1 | 2 | 1 | 1 | 0 |
| Isodacrys geminatum | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 0 | 0 | 1 | 0 | 1 | 3 | 1 | 1 | 2 |
| Isodacrys guatemalenum | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 1 | 0 | 0 | 1 | 1 | 2 | 1 | 1 | 1 |
| Isodacrys kuchii | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 0 | 0 | 0 | 1 | 1 | 2 | 1 | 2 | 1 |
| Isodacrys mexicanum | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 0 | 0 | 0 | 1 | 1 | 2 | 1 | 1 | 0 |
| Isodacrys obrienorum | 0 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 0 | 0 | 1 | 1 | 1 | 3 | 1 | 2 | 2 |

(Continued)

| Table 2 (continued) |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Taxon/character | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | 16 | 17 | 18 |
| Isodacrys okuiltontli | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 0 | 0 | 0 | 1 | 1 | 2 | 1 | 1 | 2 |
| Isodacrys orizabae | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 0 | 0 | 0 | 0 | 1 | 2 | 1 | 1 | 0 |
| Isodacrys ovipennis | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 2 | 0 | 1 | 0 | 0 | 1 | 1 | 2 | 1 | 1 | 0 |
| Isodacrys schwarzi | 1 | 1 | 1 | ? | ? | ? | 1 | 2 | 0 | 0 | 0 | 0 | ? | 1 | 2 | 1 | 2 | 0 |
| Taxon/character | 19 | 20 | 21 | 22 | 23 | 24 | 25 | 26 | 27 | 28 | 29 | 30 | 31 | 32 | 33 | 34 | 35 | 36 |
| Megalostylus albicans | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | - | - | 0 | - |
| Platyaspistes prasinus | 0 | 1 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | - | - | - | 0 | - |
| Hadromeropsis brevicoma | 1 | 1 | 0 | 0 | 1 | 1 | - | 1 | 1 | 1 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 0 |
| Minyomerus microps | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 2 | 2 | 0 | 1 | 1 | 1 | 1 | 2 | 0 | 0 |
| Minyomerus laticeps | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 2 | 2 | 0 | 1 | 1 | 1 | 0 | 0 | 0 | 0 |
| Pandeleteius hilaris | 1 | 1 | 0 | 1 | 1 | 1 | - | 1 | 2 | 1 | 1 | 1 | 1 | 1 | 0 | 0 | 1 | 0 |
| Pandeleteius rotundicollis | 1 | 1 | 0 | 1 | 1 | 1 | - | 1 | 2 | 1 | 0 | 1 | 1 | 1 | 0 | 0 | 1 | 0 |
| Pandeleteius inflatus | 1 | 1 | 0 | 1 | 1 | 1 | - | 1 | 2 | 1 | 1 | 1 | 1 | 1 | 0 | 0 | 1 | 0 |
| Scalaventer cyrillae | 1 | 1 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 1 | 1 | 1 | 0 | - | - | 1 | 1 |
| Scalaventer jamaicensis | 1 | 0 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 1 | 1 | 1 | 0 | - | - | 1 | 0 |
| Scalaventer subtropicus | 1 | 0 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 1 | 1 | 1 | 0 | - | - | 1 | 1 |
| Paululusus hispaniole | 1 | 0 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 1 | 1 | 1 | 0 | - | - | 1 | 0 |
| Paululusus constanzae | 1 | 0 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 1 | 1 | 1 | 0 | - | - | 1 | 0 |
| Isodrusus debilis | 1 | 0 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 2 | 1 | 1 | 1 | 1 | 1 | 1 | 0 |
| Isodrusus guajavus | 1 | 0 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 2 | 1 | 1 | 0 | - | - | 1 | 0 |
| aff. Isodrusus sp. | 1 | 0 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 1 | 1 | 1 | [01] | 1 | 2 | 1 | 0 |
| Pandeleteinus submetallicus | 1 | 1 | 0 | 0 | 1 | 1 | - | 1 | 2 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 |
| Pandeleteinus subcancer | 1 | 0 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 2 | 1 | 1 | 1 | 1 | 1 | 1 | 0 |
| Pandeleteinus elytroplanatus | 1 | 1 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 1 | 1 | 1 | 1 | 0 | 0 | 1 | 0 |
| aff Pandeleteinus sp. | 1 | 0 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 |
| Paradacrys ensiformis | 1 | 0 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 1 | 1 | 1 | 0 | - | - | 1 | 0 |
| Isodillex minutum | 1 | 1 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 1 | 1 | 1 | 0 | - | - | 1 | 0 |
| Isodillex plumosum | 1 | 1 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 1 | 1 | 1 | 0 | - | - | 1 | 0 |
| Isodacrys antrum | 1 | 1 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 1 | 1 | 1 | 0 | - | - | 1 | 0 |
| Isodacrys apicale | 1 | 1 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 1 | 1 | 1 | 1 | 0 | 1 | 1 | 0 |
| Isodacrys brevirostre | 1 | 1 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 2 | 1 | 1 | 0 | - | - | 1 | 0 |
| Isodacrys buchanani | 1 | 0 | 0 | 1 | 1 | 0 | 1 | 1 | 2 | 2 | 2 | 1 | 1 | 1 | 1 | 2 | 1 | 0 |
| Isodacrys burkei | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 1 | 2 | 2 | 2 | 1 | 1 | 0 | - | - | 1 | 0 |
| Isodacrys carlae | 1 | 0 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 2 | 1 | 1 | 0 | - | - | 1 | 0 |
| Isodacrys confusum | 1 | 0 | 0 | 1 | 1 | 0 | 1 | 1 | 2 | 2 | 1 | 1 | 1 | 0 | - | - | 1 | 0 |
| Isodacrys crispum | 1 | 0 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 2 | 1 | 1 | 1 | 1 | 2 | 1 | 0 |
| Isodacrys ellipticum | 1 | 0 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 2 | 1 | 1 | 0 | - | - | 1 | 0 |
| Isodacrys fasciatum | 1 | 0 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 1 | 1 | 1 | 0 | - | - | 1 | 0 |
| Isodacrys frontalis | 1 | 1 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 1 | 1 | 1 | 0 | - | - | 1 | 0 |
| Isodacrys geminatum | 1 | 1 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 2 | 1 | 1 | 0 | - | - | 1 | 0 |


| Taxon/character | 19 | 20 | 21 | 22 | 23 | 24 | 25 | 26 | 27 | 28 | 29 | 30 | 31 | 32 | 33 | 34 | 35 | 36 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Isodacrys guatemalenum | 1 | 0 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 2 | 1 | 1 | 0 | - | - | 1 | 0 |
| Isodacrys kuchii | 1 | 1 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 2 | 1 | 1 | 0 | - | - | 1 | 0 |
| Isodacrys mexicanum | 1 | 0 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 2 | 1 | 1 | 1 | 1 | 2 | 1 | 0 |
| Isodacrys obrienorum | 1 | 1 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 1 | 1 | 1 | [01] | 1 | 2 | 1 | 0 |
| Isodacrys okuiltontli | 1 | 0 | 0 | 1 | 1 | 1 | - | 1 | 2 | 2 | 2 | 1 | 1 | 0 | - | - | 1 | 0 |
| Isodacrys orizabae | 1 | 1 | 1 | 1 | 1 | 1 | - | 1 | 2 | 2 | 2 | 1 | 1 | [01] | 1 | 2 | 1 | 0 |
| Isodacrys ovipennis | 1 | 0 | 1 | 1 | 1 | 0 | 1 | 1 | 2 | 2 | 2 | 1 | 1 | 1 | 1 | 1 | 1 | 0 |
| Isodacrys schwarzi | 1 | ? | 0 | 1 | ? | 0 | 1 | 1 | 2 | 2 | 1 | 1 | 1 | 1 | 0 | ? | 1 | 0 |
| Taxon/character | 37 | 38 | 39 | 40 | 41 | 42 | 43 | 44 | 45 | 46 | 47 | 48 | 49 | 50 | 51 | 51 | 53 | 54 |
| Megalostylus albicans | 0 | 0 | 0 | - | 0 | - | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Platyaspistes prasinus | 0 | 0 | 0 | - | 0 | - | 0 | 0 | 0 | 0 | 0 | 0 | - | 0 | 0 | 0 | 0 | 1 |
| Hadromeropsis brevicoma | 0 | 1 | 0 | - | 2 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 2 | 1 | 1 | 1 |
| Minyomerus microps | 0 | 2 | 1 | 1 | 1 | - | 1 | - | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Minyomerus laticeps | 0 | 2 | 1 | 1 | 1 | - | 1 | - | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Pandeleteius hilaris | 0 | 1 | 0 | - | 2 | 0 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 1 | 1 | 1 |
| Pandeleteius rotundicollis | 0 | 1 | 0 | - | 2 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 2 | 1 | 1 | 1 |
| Pandeleteius inflatus | 0 | 3 | 0 | - | 2 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | - | 0 | 1 | 1 | 1 | 1 |
| Scalaventer cyrillae | 0 | 1 | 0 | - | 2 | 0 | 0 | 1 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 1 | 1 | 1 |
| Scalaventer jamaicensis | 0 | 1 | 0 | - | 2 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 1 | 1 | 1 |
| Scalaventer subtropicus | 0 | 1 | 0 | - | 2 | 0 | 0 | 1 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 1 | 1 | 1 |
| Paululusus hispaniole | 0 | 1 | 0 | - | 2 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | 1 | 2 | 1 | 1 | 1 |
| Paululusus constanzae | 1 | 3 | 1 | 0 | 2 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 2 | 1 | 1 | 1 |
| Isodrusus debilis | 0 | 1 | 0 | - | 2 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 |
| Isodrusus guajavus | 0 | 1 | 0 | - | 2 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 1 | 1 |
| aff. Isodrusus sp. | 0 | 3 | 1 | 1 | 2 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | - | 0 | 1 | 1 | 1 | 1 |
| Pandeleteinus submetallicus | 0 | 1 | 0 | - | 2 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 1 | 1 |
| Pandeleteinus subcancer | 0 | 1 | 0 | - | 2 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 1 | 0 |
| Pandeleteinus elytroplanatus | 0 | 1 | 0 | - | 2 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 1 |
| aff Pandeleteinus sp. | 0 | 1 | 0 | - | 2 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 1 | 1 |
| Paradacrys ensiformis | 0 | 1 | 0 | - | 2 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 1 | 1 |
| Isodillex minutum | 0 | 3 | 1 | 0 | 2 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | - | 0 | 1 | 1 | 1 | 1 |
| Isodillex plumosum | 0 | 3 | 1 | 0 | 2 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 1 | 1 |
| Isodacrys antrum | 0 | 3 | 1 | 1 | 2 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 1 |
| Isodacrys apicale | 0 | 2 | 1 | 0 | 2 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 | 1 |
| Isodacrys brevirostre | 0 | 3 | 1 | 1 | 2 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 |
| Isodacrys buchanani | 0 | 3 | 1 | 0 | 2 | 2 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | [01] | 1 | 0 | 0 | 0 |
| Isodacrys burkei | 0 | 4 | 1 | 0 | 2 | 1 | 0 | 1 | 0 | 1 | 1 | 0 | 0 | 0 | 1 | 1 | 1 | 1 |
| Isodacrys carlae | 0 | 2 | 1 | 1 | 2 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 |
| Isodacrys confusum | 0 | 2 | 1 | 1 | 2 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 |
| Isodacrys crispum | 0 | 3 | 1 | 0 | 2 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 |
| Isodacrys ellipticum | 1 | 3 | 1 | 0 | 2 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 1 | 1 |

(Continued)

| Table 2 (continued) |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Taxon/character | 37 | 38 | 39 | 40 | 41 | 42 | 43 | 44 | 45 | 46 | 47 | 48 | 49 | 50 | 51 | 51 | 53 | 54 |
| Isodacrys fasciatum | 0 | 2 | 1 | 1 | 2 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 |
| Isodacrys frontalis | 0 | 3 | 1 | 1 | 2 | 1 | 0 | 1 | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 |
| Isodacrys geminatum | 0 | 2 | 1 | 1 | 2 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 |
| Isodacrys guatemalenum | 1 | 3 | 1 | 0 | 2 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 1 | 1 |
| Isodacrys kuchii | 0 | 2 | 1 | 1 | 2 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 0 | 0 |
| Isodacrys mexicanum | 0 | 2 | 1 | 0 | 2 | 1 | 0 | 1 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | 1 | 1 | 1 |
| Isodacrys obrienorum | 0 | 2 | 1 | 1 | 2 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 |
| Isodacrys okuiltontli | 0 | 2 | 1 | 0 | 2 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 1 |
| Isodacrys orizabae | 0 | [23] | 1 | 0 | 2 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 1 | 1 | 1 |
| Isodacrys ovipennis | 0 | 4 | 1 | 0 | 2 | 1 | 0 | 1 | 0 | 1 | 1 | 0 | 0 | 0 | 1 | 1 | 1 | 1 |
| Isodacrys schwarzi | 0 | 3 | 1 | 0 | 2 | 2 | 0 | 1 | 0 | 0 | 0 | 0 | 1 | ? | 1 | 0 | 0 | 0 |
| Taxon/character | 55 | 56 | 57 | 58 | 59 | 60 | 61 | 62 | 63 | 64 | 65 | 66 | 67 | 68 | 69 | 70 | 71 | 72 |
| Megalostylus albicans | 0 | 0 | - | 0 | 0 | 0 | 0 | - | - | 0 | - | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Platyaspistes prasinus | 0 | 0 | - | 1 | 0 | 1 | 0 | - | - | ? | ? | ? | ? | ? | ? | ? | ? | ? |
| Hadromeropsis brevicoma | 0 | 1 | 0 | 2 | 0 | 0 | 0 | - | - | 2 | 1 | ? | 2 | 0 | 0 | 0 | 0 | 0 |
| Minyomerus microps | 1 | 0 | - | 0 | 1 | 0 | 0 | - | - | 1 | - | 0 | 2 | 0 | - | - | - | - |
| Minyomerus laticeps | 1 | 0 | - | 0 | 1 | 0 | 0 | - | - | 1 | - | 0 | 2 | 0 | ? | ? | ? | ? |
| Pandeleteius hilaris | 0 | 1 | 0 | 2 | 0 | 0 | 0 | - | - | 2 | 0 | 1 | 1 | 1 | 1 | 0 | 0 | 0 |
| Pandeleteius rotundicollis | 0 | 1 | 0 | 2 | 0 | 0 | 0 | - | - | 2 | 0 | 1 | 1 | 1 | 0 | 1 | 0 | 0 |
| Pandeleteius inflatus | 0 | 1 | 0 | 2 | 0 | 0 | 0 | - | - | 2 | 0 | 1 | 1 | 1 | ? | ? | ? | ? |
| Scalaventer cyrillae | 0 | 1 | 1 | 2 | 0 | 0 | 1 | 1 | 0 | 2 | 1 | 1 | 1 | 1 | 0 | 1 | 0 | 0 |
| Scalaventer jamaicensis | 0 | 1 | 1 | 2 | 0 | 0 | 1 | 1 | 0 | 2 | 1 | 1 | 1 | 1 | 0 | 1 | 0 | 0 |
| Scalaventer subtropicus | 0 | 1 | 1 | 2 | 0 | 0 | 1 | 1 | 0 | 2 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 0 |
| Paululusus hispaniole | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 0 |
| Paululusus constanzae | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | ? |
| Isodrusus debilis | 0 | 1 | 0 | 2 | 0 | 1 | 1 | 1 | 0 | 2 | 0 | 1 | 1 | 1 | - | - | - | - |
| Isodrusus guajavus | 0 | 1 | 0 | 2 | 0 | 1 | 1 | 1 | 0 | ? | ? | ? | ? | ? | ? | ? | ? | ? |
| aff. Isodrusus sp. | 0 | 1 | 0 | 2 | 0 | 1 | 1 | 1 | 0 | ? | ? | ? | ? | ? | 1 | ? | 0 | ? |
| Pandeleteinus submetallicus | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 0 | 1 | 1 | 1 | 1 | 1 | 0 | 2 |
| Pandeleteinus subcancer | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 0 | 2 | 0 | 1 | 1 | 1 | 1 | 1 | 0 | 2 |
| Pandeleteinus elytroplanatus | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 0 | 1 | 1 | 1 | 1 | 1 | 0 | 2 |
| aff Pandeleteinus sp. | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 0 | 1 | 1 | 1 | 1 | 1 | 1 | 0 |
| Paradacrys ensiformis | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 0 | 1 | 1 | 1 | 0 | 1 | 1 | 0 |
| Isodillex minutum | 0 | 1 | 0 | 2 | 0 | 0 | 0 | - | - | 2 | 1 | 1 | 1 | 1 | - | - | - | - |
| Isodillex plumosum | 0 | 1 | 0 | 2 | 0 | 0 | 0 | - | - | 2 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 0 |
| Isodacrys antrum | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | - | - | 1 | 1 | 1 | - | - | - | - |
| Isodacrys apicale | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | ? | ? | ? | ? | ? | ? | ? | ? | ? |
| Isodacrys brevirostre | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 0 | 1 | 1 | 1 | - | - | - | - |
| Isodacrys buchanani | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 0 | 1 | 1 | 1 | 1 | 1 | 0 | 1 |
| Isodacrys burkei | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 0 | 1 | 1 | 1 | 1 | 0 | 1 | 1 |
| Isodacrys carlae | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 0 | 1 | 1 | 1 | - | - | - | - |

Table 2 (continued)

| Taxon/character | $\mathbf{5 5}$ | $\mathbf{5 6}$ | $\mathbf{5 7}$ | $\mathbf{5 8}$ | $\mathbf{5 9}$ | $\mathbf{6 0}$ | $\mathbf{6 1}$ | $\mathbf{6 2}$ | $\mathbf{6 3}$ | $\mathbf{6 4}$ | $\mathbf{6 5}$ | $\mathbf{6 6}$ | $\mathbf{6 7}$ | $\mathbf{6 8}$ | $\mathbf{6 9}$ | $\mathbf{7 0}$ | $\mathbf{7 1}$ | $\mathbf{7 2}$ |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| Isodacrys confusum | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 0 | 1 | 1 | 1 | 1 | 1 | 0 | 2 |
| Isodacrys crispum | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 0 | 2 | 0 | 1 | 1 | 1 | - | - | - | - |
| Isodacrys ellipticum | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 0 | 1 | 1 | 1 | - | - | - | - |
| Isodacrys fasciatum | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 0 | 1 | 1 | 1 | - | - | - | - |
| Isodacrys frontalis | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 0 | 1 | 1 | 1 | - | - | - | - |
| Isodacrys geminatum | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 1 | 1 | 1 | 1 | - | - | - | - |
| Isodacrys guatemalenum | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| Isodacrys kuchii | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 0 | 1 | 1 | 1 | 1 | 1 | 1 | 2 |
| Isodacrys mexicanum | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 1 | 1 | 1 | 1 | 1 | 0 | 0 | 1 |
| Isodacrys obrienorum | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 2 |
| Isodacrys okuiltontli | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | $?$ |
| Isodacrys orizabae | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 0 | 1 | 1 | 1 | 1 | 0 | 1 | 0 |
| Isodacrys ovipennis | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | 2 | 0 | 1 | 1 | 1 | 1 | 0 | 1 | 1 |
| Isodacrys schwarzi | 0 | 1 | 0 | 2 | 0 | 0 | 1 | 0 | 1 | $?$ | $?$ | $?$ | $?$ | $?$ | $?$ | $?$ | $?$ | $?$ |

character state transformations were examined in WINCLADA under various optimizations. Bremer branch support and parsimony Jackknife values were also calculated in TNT. For Bremer support (Bremer, 1994), 10,000 suboptimal trees up to 15 steps longer than MPT's were retained. Jackknife values (Farris et al., 1996; Davis, 2011) were computed using 1,000 replications.

## Taxa description and nomenclature

The genus-level diagnoses of Isodacrys and Isodillex new genus highlight characters present in all their members and accounts for their variability. Species-level diagnoses, although similarly structured, represent unique complementary accounts of character states observed in each species. For ease of comparison, characters given in descriptions of new taxa follow the sequence of characters included in the phylogenetic analysis. Additional characters not included in the data matrix are displayed accordingly with the body part sequence that is presented as follows: Size, dorsal coverture of integument, rostrum, head, prothorax, elytra, legs, abdomen and genitalia. Body length and width were measured in dorsal view from the frons to the apex of the elytra and at the widest point of the elytra, respectively. An identification key to the species of Isodacrys is arranged with emphasis being placed on the most readily observable characters.

The electronic version of this article in Portable Document Format (PDF) will represent a published work according to the International Commission on Zoological Nomenclature (ICZN), and hence the new names contained in the electronic version are effectively published under that Code from the electronic edition alone. This published work and the nomenclatural acts it contains have been registered in ZooBank, the online registration system for the ICZN. The ZooBank LSIDs (Life Science Identifiers) can be resolved and the associated information viewed through any standard web browser by appending the LSID
to the prefix http://zoobank.org/. The LSID for this publication is: [urn:lsid:zoobank.org: pub:F5D5C6F7-3FB4-4D1F-91F3-0D6380C8FF59]. The online version of this work is archived and available from the following digital repositories: PeerJ, PubMed Central and CLOCKSS.

Isodacrys antrum: Isodacrys carlae: Isodacrys confusum; Isodacrys fasciatum: Isodacrys frontalis: Isodacrys kuchii: Isodacrys obrienorum: Isodacrys okuiltontli: Isodillex: Isodillex plumosum:

## Distribution of Isodacrys species

For the material examined for each species of Isodacrys, all localities and sampling points were georeferenced and included in the occurrence maps done in QGIS Version 3.12.3București (Quantum GIS Development Team, 2020).

## RESULTS

## Phylogenetic analysis

The heuristic search for most parsimonious trees of 43 terminals and 72 discrete characters under equal weights yielded two equally most parsimonious trees ( $L=242, \mathrm{CI}=0.39$, $\mathrm{RI}=0.70)$. The strict consensus adds two steps to the tree length $(L=244, \mathrm{CI}=0.38$, $\mathrm{RI}=0.69)$, depicting one collapsed node regarding the position of Pandeleteinus submetallicus and P. elytroplanatus relative to the Pandeleteinus subcancer-Isodacrys antrum clade (Fig. 1). Bremer and Jackknife values are mapped on internal nodes of the strict consensus tree (Fig. 1). Branch support values were higher in basal clades and particular West Indian genera but lower on deeper clades related with Isodacrys species relationships. One of the two most parsimonious trees (Fig. 2) was chosen to illustrate character state optimizations ( 57 unambiguous, 6 ACCTRAN and 9 DELTRAN, see Agnarsson \& Miller, 2008).

Isodacrys was recovered as monophyletic, including a total of 20 species in five main clades (Fig. 1). Isodacrys is supported by one synapomorphy and two homoplastic character states (Fig. 2). The Isodillex clade, which is sister to Isodacrys and supported by four homoplastic character states (Fig. 2), is proposed to accommodate Isodillex minutum (Sharp, 1911), new combination and Isodillex plumosum Cortés-Hernández, new species. Isodillex was excluded from the present circumscription of Isodacrys based on the combination of five homoplastic character states supporting the new taxon (Fig. 2). Some of these character states were considered relevant for taxonomic amendments. The clade Isodacrys-Isodillex resulted sister to the clade Paradacrys-Scalaventer, which occur in the West Indies, based on one synapomorphy and one homoplastic character state.

Regardless of the lack of diagnostic character states supporting the monophyly of Isodacrys, the depicted combination of character states supporting it plus more inclusive congruent information on basal clades are relevant enough to circumscribe the genus as here presented (see diagnosis of Isodacrys). The exclusion of the Isodillex is considered necessary based on the not sulcate condition of the anterior margin of ventrites III, IV and V (char. 61:0 and thus inapplicable entries in chars. 62 and 63; chars. 61:1, 62:0 and 63:1 in


Figure 1 Strict consensus of two most parsimonious trees obtained from the analysis under equal weights $(L=244, C I=0.38, R I=0.69)$. Node regarding the position of Pandeleteinus submetallicus and P. elytroplanatus is collapsed. Numbers above and below branches indicate Bremer support and Jackknife values (cut-off at 63\%), respectively. A lateral habitus photograph of one species from each clade within Isodacrys is provided as it is for Isodillex, its sister taxon (see "Discussion").

Full-size DOI: 10.7717/peerj.10191/fig-1

Isodacrys). Secondary characters that can also be helpful in separating Isodillex from Isodacrys are dorsal scales contiguous, not overlapping (char. 1:0; char. 1:1 in Isodacrys, with reversal in I. obrienorum), pronotum in dorsal view as wide as long (char. 29:1; char. 29:2 in Isodacrys, with reversals to state 1 in I. schwarzi, I. apicale, I. confusum-I. fasciatum, I. frontalis and I. antrum-I. obrienorum) and manubrium shorter than median lobe (char. 72:0; char. 72:1 synapomorphy for Isodacrys with reversal to state 0 in I. orizabae and with evolutionary transition to state 2 in the I. kuchii-I. obrienorum clade).
Accordingly, Isodillex is considered a different taxon, sister group of Isodacrys. Otherwise, the circumscription of Isodacrys would be vague, needing further ad hoc hypothesis of character state evolution.

Given the terminals and characters included in the present analysis, there are five main clades within Isodacrys (Fig. 1). The basal division within the genus is between the I. ellipticum clade (Fig. 1), supported by two homoplastic character states (presence of longitudinal carina on the epistome 10:1 and setae of elytra strongly modified, conspicuously longer 37:1), and the I. crispum-I. antrum clade, which incorporates the


Figure 2 One of the two most parsimonious cladograms ( $L=242, \mathrm{CI}=0.39, \mathrm{RI}=0.70$ ) with preferred character state optimization (ACCTRAN: chars. $7,15,16,31,47$ and 70; DELTRAN: chars. 8, 13, 23, 32, 41, 44, 54, 64 and 71 ). Black circles represent non-homoplastic character state transformations, whereas white circles represent homoplastic character state transformations. Numbers on and below each circle correspond to character and state codes, respectively. Colored stars indicate which species is illustrating the highlighted character states.

Full-size DOI: 10.7717/peerj.10191/fig-2
remaining Isodacrys species, also supported by two homoplastic character states related to the size of the prolegs relative to the metalegs (chars. 52:0 and 53:0).

The second division is within the Isodacrys crispum-I. antrum clade, divided into two clades, the I. crispum clade (Fig. 1) and the clade I. okuiltontli-I. antrum, supported respectively by two (anterior portion of scrobe at least one fifth shorter than posterior portion 17:2 and presence of postocular vibrissae 32:1) and one (elytra in dorsal view obovate $38: 2$, but character states 3 and 4 also present within the clade) homoplastic character states. In the Isodacrys crispum clade, there is a basal division splitting I. crispum from I. buchanani-I. schwarzi, whereas in the clade I. okuiltontli-I. antrum there is a split
depicting I. okuiltontli as the sister taxon of the I. apicale-I. antrum clade. Isodacrys buchanani-I. schwarzi is well supported by one unreversed character state (basal margin of elytra angularly emarginated 42:2) and three homoplastic characters (antennae with scape covered with scales 24:0, apical declivity of elytra concave 49:1 and inner margin of protibiae without teeth 54:0).

Isodacrys apicale-I. antrum clade, supported by one homoplastic character (presence of fovea between eyes 20:1), includes most of the Isodacrys species in two main clades. The first clade is I. apicale (Fig. 1), supported by the presence of postocular vibrissae (char. 32:1, with reversal in I. burkei) and by outer corner of the hemisternites VIII truncate (char. 70:0), while I. kuchii-I. antrum clade by three homoplastic character states (five visible intervals at base of elytra in dorsal view 40:1, inner margin of protibiae without teeth 54:0, with reversal in I. antrum, and manubrium of tegmen longer than median lobe 72:2).

Finally, the Isodacrys kuchii-I. antrum clade is divided into two clades, I. kuchii and I. brevirostre (Fig. 1). The Isodacrys kuchii clade is supported by two homoplastic character states (anterior portion of scrobe at least one fifth shorter than posterior portion 17:2 and profemora at least one fifth wider than metafemora 52:1). The Isodacrys brevirostre clade is supported by one synapomorphy (postrostrum with longitudinal sulci mesad of dorsolateral margins 12:1).

## Character discussion (Figs. 2-7)

In the following section, characters included in the phylogenetic analysis are presented and discussed. The characters are divided in subheadings: general appearance; rostrum; antennal scrobes; head; mouthparts; antenna; prothorax; elytra; legs; abdomen; and terminalia. An introductory section for each subheading is presented with highlights and explanations (when necessary) of the characters and the character states of either Isodacrys and/or outgroups included. The characters are presented in the following sequence: (1) character number, (2) character description, (3) character statement (character states, see Sereno, 2007), (4) additiveness (for multistate characters only), (5) applicability (when necessary; e.g., reductive coding), (6) similar characters in other studies,
(7) optimization of character states in the MPT's or/and synapomorphies, (8) character statistics (length, consistency index, retention index), and (9) additional information (particular comments for the character). See Fig. 2 for preferred character state optimizations. See Fig. 3 for an overview of several adult morphological features in Isodacrys, which may apply to other genera; Figs. 4-7 are referred for character states illustrated.

## General appearance (Figs. 4-5)

The arrangement and shape of the scales in New World Tanymecini are complex along the body, varying even within the same structure (e.g., along elytra, see Howden, 1959).
Variation is more evident along fasciae or vittae, when present, and along the apical declivity of the elytra. Considering scales as serialogs (see Ochoterena et al., 2019), character states proposed are based on the dominant patterns presented in Jansen \& Franz (2015).


Figure 3 Overview of various adult characters in Isodacrys. (A) Isodacrys brevirostre, rostrum in dorsal view; small triangles indicate anterior and posterior end of epifrons' median sulcus; (B) I. apicale, rostrum and head in lateral view; acronyms AP and PP refer to anterior and posterior portion of the scrobe, respectively; (C) I. fasciatum new species, head, prothorax and base of elytra in dorsal view; (D) I. schwarzi lateral habitus taken by Ashton Smith (Usage Rights: CC0 1.0, Public-domain. Available at http://scan-bugs.org/portal/collections/individual/index.php?occid=37439808); (E) I. kuchii new species, aedeagus in lateral view; (F) I. fasciatum new species, ovipositor in lateral view; (G) I. frontalis new species, spermatheca.

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The shape of the scales is defined as subcircular when their margin is evenly rounded, whereas polygonal refers to scales with angulate margins, composed by at least three sides. Polygonal scales vary along the body in form and number of sides.


Figure 4 Photographs illustrating some of the characters included in the analysis. (A) Scalaventer cyrillae, prothorax in dorsal view; (B) Isodacrys carlae new species, rostrum in dorsal view; (C) S. cyrillae, rostrum in dorsal view. (D) I. brevirostre, rostrum in dorsal view; (E) I. apicale, rostrum and head in lateral view; acronyms AP and PP refer to the anterior and posterior portion of the scrobe, respectively; (F) Paululusus hispaniole, prementum. Numbers and numbers between parentheses refer to characters and their states, respectively.

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1. Dorsal habitus, scales, arrangement: (0) Contiguous, not overlapping (Fig. 4A); (1) variously overlapping non-linearly (Fig. 5E); (2) only overlapping posteriorly. Non-additive. See character 1 by Jansen \& Franz (2015). State 0 convergently present in Megalostylus albicans-Pandeleteius inflatus, the clade aff. Pandeleteinus sp.-Isodillex and in Isodacrys obrienorum; state 1 convergently present in Hadromeropsis brevicoma, the Pandeleteinus sumetallicus-P. subcancer clade, Isodrusus debilis and Isodacrys; state 2 synapomorphy for Minyomerus (l: 7, ci: 0.28 , ri: 0.70 ).
2. Habitus dorsal, scales, shape: (0) Subcircular; (1) polygonal (Fig. 4A). Character state 1 synapomorphic for the Pandeleteius-Isodacrys clade (1: 1, ci: 1, ri:1).


Figure 5 Photographs illustrating some of the characters included in the analysis. (A) Isodacrys brevirostre, antenna; (B) Hadromeropsis brevicoma, dorsal habitus; (C) Isodacrys apicale, rostrum, head and prothorax in lateral view; (D) I. guatemalenum, dorsal habitus; (E) I. fasciatum new species, head, prothorax and base of elytra in dorsal view; (F) I. ovipennis, dorsal habitus. Numbers and numbers between parentheses refer to characters and their states, respectively.
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## Rostrum (Figs. 3-4)

The dorsal area of the rostrum in tanymecine weevils can be generally divided into three more or less defined regions from the apex to the frons (Fig. 3A): nasal plate, epistome and epifrons (postrostrum). These regions are herein described to propose homology statements.

The nasal plate consists of a defined, bare area located at the tip of the rostrum, between or slightly in front of the insertion of the antennae, at the apical margin of the epistome (Pierce, 1913; Vaurie, 1963; Franz, 2012). We agree with this perspective and therefore


Figure 6 Photographs illustrating some of the characters included in the analysis. (A) Isodacrys geminatum, lateral habitus; (B) I. apicale, apex of elytra; (C) I. geminatum, dorsolateral habitus showing femora in dorsal view; (D) I. frontalis, apex of metatibia from tarsal socket view; (E) I. mexicanum, apex of metatibia; (F) I. buchanani, ventrites two, three, four and five. Numbers and numbers between parentheses refer to characters and their states, respectively. Full-size DOI: 10.7717/peerj.10191/fig-6
it is treated as a structure different from the epistome (Girón \& Franz, 2010; Lanteri \& Del Río, 2017; Girón \& Howden, 2019). It can be delimited posteriorly from the remainder of the rostrum by the epistomal setae, which indicate the anterior end of the epistome and the beginning of the nasal plate (Fig. 3A). In many Neotropical entimines, the posterior margin of the nasal plate can be elevated, forming a carina (Vaurie, 1963). Howden (1959) first used the term nasal plate for Tanymecini as "the sclerite that is within the apical emargination or extend forward from the apex of the beak", considering it as a synonym of the epistome. In the literature, the terms nasal plate and epistome have been used to refer to the same structure or different parts of the apical region of the rostrum (Pierce, 1913; Blatchley \& Leng, 1916; Howden, 1959; Anderson, 2002; Franz, 2012;


Figure 7 Photographs illustrating some of the characters included in the analysis. (A) Isodacrys confusum new species, female sternite VIII in dorsal view; (B) I. kuchii new species, ovipositor in dorsal view; (C) I. kuchii new species, aedeagus in lateral view; (D) I. kuchii new species, male hemisternites VIII and sternite IX in dorsal view. Numbers and numbers between parentheses refer to characters and their states, respectively.

Full-size DOI: 10.7717/peerj.10191/fig-7

Oberprieler, Anderson \& Marvaldi, 2014, Jansen \& Franz, 2015, 2018; Marvaldi et al., 2018; Girón \& Howden, 2019). In the present study, the apical emargination refers to the posterior margin of the nasal plate. Regarding the issue as to whether they are either the same or different structures, Howden (1966) avoided the term nasal plate. The epistome constitutes the reduced frontoclypeal margin or sclerite directly behind the labrum (Nichols, 1989). Although the labrum is considered absent in Curculionidae, it is unclear whether the absent condition represents a complete loss or variable fusions of the labrum and clypeus, and whether the epistomal lobe is a derived modification of the frons or a remnant of the clypeus or labrum (Davis, 2017). Herein, based on the studied species, the epistome is considered as the dorsal area of the rostrum located between the antennal insertions, distinguished by the presence of modified scales (smaller, of different shape and color and sometimes more scattered compared to remaining scales on the rostrum). Additionally, the epistome is usually irregularly covered by erect to semierect pale setae which are set in shallow foveae (Fig. 3A). Finally, the epifrons comprises the proximal region of the rostrum, extending to the anterior margin of the eyes (Oberprieler, Anderson \& Marvaldi, 2014). In some tanymecines, there is a fovea at the posterior margin of the epistome and beginning of the epifrons (at anterior end of the median sulcus [median line in Howden's terminology], when present, Fig. 3A). In several West Indian representatives this fovea is anteriorly bifurcated, forming a Y -shaped depression (Howden, 1970). This fovea could represent a rudimentary/vestigial suture delimiting the typically modified apical region of the rostrum (constituted by the epistome and the nasal plate) in Tanymecini.

The rostrum is always directed ventrally in Isodacrys, forming an angle between the ventral outline of the rostrum and the head, varying from obtusely angled to almost right angled. In the outgroups the rostrum can be directed ventrally or virtually in line with the main body axis.
3. Rostrum, orientation of dorsolateral margins: (0) Convergent anteriorly; (1) subparallel (Fig. 4B); (2) divergent towards apex. Additive. See character 22 in

Franz (2012), character 8 in Girón \& Franz (2012), character 9 in Lanteri \& Del Río (2017) and character 0 in Del Río et al. (2018). State 2 is convergently present in Isodrusus guajavus and the Paululusus-Scalaventer clade (l: 3, ci: 0.66, ri: 0.80). In Isodacrys the dorsolateral margins of the rostrum are subparallel. Nonetheless, Howden (1961) considered that some Isodacrys species have dorsolateral margins of the rostrum slightly convergent anteriorly. According to our observations, this varies intraspecifically and in a way almost imperceptible to consider the dorsolateral margins convergent anteriorly in Isodacrys.
4. Rostrum, anterior margin of nasal plate, shape: (0) Emarginate (Fig. 4B); (1) straight; (2) produced anteriorly. Non-additive. See character 3 in Marvaldi et al. (2018). Character state 1 synapomorphy for the Isodrusus debilis-I. guajavus clade; character state 2 synapomorphy for the Pandeleteius hilaris-P. inflatus clade (l: 2, ci: 1, ri: 1). Homology statement among these character states is based on topological correspondence.
5. Emargination of nasal plate, degree: (0) Strongly emarginate; (1) slightly indented (Fig. 4B). Character state 1 synapomorphy for the aff. Pandeleteinus sp.-Isodacrys clade (l: 1, ci: 1, ri: 1). The anterior margin of the nasal plate in Isodacrys is almost straight, at most slightly directed inward medially, where it bears a small indentation, sometimes irregular.
6. Rostrum, posterior margin of nasal plate, carina: (0) Absent, posterior margin flat; (1) present (Fig. 4C). See character 10 in Jansen \& Franz (2015) and character 16 in Lanteri \& Del Río (2017). Character state 1 synapomorphy for the Pandeleteius-Isodacrys clade, with reversal in Pandeleteinus submetallicus (l: 2, ci: 0.50, ri: 0.80). Howden (1961) described several species of Isodacrys as having the apical emargination (which herein is termed posterior margin of the nasal plate) not carinate, but according to our observations, it is actually carinate in all Isodacrys species. The degree to which the posterior margin is carinate varies among individuals and species.
7. Rostrum, epistome, degree of development: (0) Indistinct; (1) distinct (Fig. 4C). ACCTRAN optimization is preferred, as DELTRAN suggests parallel origins of the epistome. Thus, character state 1 synapomorphy for the Tanymecini clade, with reversal in Hadromeropsis brevicoma (1: 2, ci: 0.50, ri: 0.50).
8. Rostrum, epistome, constitution relative to remainder of rostrum: (0) Strongly concave (Fig. 4C); (1) obliquely angled; (2) in a continuous plane with (Fig. 4D). Non-additive. Coded as inapplicable when the epistome is indistinct (see character 7, state 0). See character 10 in Franz (2012). DELTRAN optimization preferred in accordance with the assumption of non-additiveness of character states. Thus, state 0 present in Minyomerus and Pandeleteius rotundicollis; state 1 synapomorphy for the Pandeleteius hilaris-P. inflatus clade; state 2 synapomorphy for the Pandeleteinus-Isodacrys clade, with reversal to state 0 in Scalaventer (1: 3, ci: 0.66, ri: 0.83). Character state 2, present in Isodacrys and relatives, can be slightly depressed. This varies among individuals and species.
9. Rostrum, epistome, posterior margin: (0) Indistinct; (1) keeled (Fig. 4C). Coded as inapplicable when the epistome is indistinct (see character 7, state 0). See character 14 in

Franz (2012). Character state 1 convergently present in Pandeleteius rotundicollis and the Paululusus-Scalaventer clade (l: 2, ci: 0.50, ri: 0.80).
10. Rostrum, epistome, longitudinal carina extending along epistome, reaching posterior margin of nasal plate: (0) Absent; (1) present (Fig. 4C). Coded as inapplicable when the epistome is indistinct (see character 7 , state 0 ). State 1 convergently present in the Paradacrys-Scalaventer clade, the Isodacrys ellipticum-I. guatemalenum clade and the I. burkei-I. ovipennis clade (l: 3, ci: 0.33 , ri: 0.77).
11. Constitution of epifrons: (0) Flat to slightly depressed mesally (Fig. 4D); (1) concave. State 1 synapomorphy for the Hadromeropsis brevicoma-Pandeleteinus submetallicus clade, with reversal to state 0 for the Pandeleteinus subcancer-Isodacrys clade (l: 2, ci: 0.50, ri: 0.80). In Isodacrys the epifrons is flat to slightly depressed mesally when the median sulcus and/or foveae are present. This varies among individuals and species.
12. Epifrons, longitudinal sulci mesad of dorsolateral margins: (0) Absent; (1) present (Fig. 4D). Character state 1 synapomorphy for the Isodacrys brevirostre-I. antrum clade (l: $1, \mathrm{ci}: 1$, ri: 1). These longitudinal sulci are short, extending from near the antennal insertion to the deflection of the scrobe.
13. Rostrum, epifrons, fovea: (0) Absent; (1) present (Fig. 4C). DELTRAN optimization is preferred because ACCTRAN postulates several origins of the fovea in Isodacrys. Thus, state 1 convergently present in Minyomerus, Pandeleteius hilaris and the Pandeleteinus subcancer-Isodacrys clade, with subsequent reversals in Isodrusus debilis, Isodillex, Isodacrys crispum, I. orizabae, I. burkei, I. carlae-I. fasciatum clade, I. brevirostre, I. geminatum and I. antrum (l: 12, ci: 0.08, ri: 0.38).

## Antennal scrobes (Figs. 3-4)

The scrobes can be curved or angled in the species included in this analysis (Figs. 3B and $4 \mathrm{E})$. When angled, two discernible parts of the scrobe can be distinguished relative to orientation of the rostrum: an anterior and a posterior part (Fig. 3B, AP and PP respectively; called horizontal and vertical portion, respectively, by Howden, 1961, 1970). In some taxa the scrobe can reach the ventral part of the rostrum. The dorsal margin of the scrobe was taken as reference guide to determine the shape of the scrobe; distance from the posterior portion of the scrobe to the eye was considered at the nearest point between them.
14. Rostrum, scrobe, vestiture: (0) Present; (1) absent, scrobe glabrous (Fig. 4E). Character state 1 synapomorphy for Hadromeropsis-Isodacrys clade (l: 1, ci: 1: ri: 1). The scrobes are always glabrous in Isodacrys, whereas in the outgroups it varies from glabrous to covered by different types of vestiture (setae, scales, plumose scales).
15. Rostrum, scrobe in lateral view, dorsal margin, shape: (0) Evenly curved, directed to compound eye; (1) strongly curved, directed to ventral surface of rostrum; (2) obtusely angled (Fig. 4E); (3) right-angled; (4) acutely angled. Additive. See characters 14 and 12 in Lanteri \& Del Río (2017) and in Marvaldi et al. (2018), respectively. ACCTRAN optimization is preferred because DELTRAN postulates two origins of character state 1. Thus, character state 1 synapomorphy for the Tanymecini clade with reversal to state 0 in Hadromeropsis brevicoma and subsequent evolutionary transition to state 2 in the

Pandeleteinus-Isodacrys clade; state 3 convergently present in the aff. Pandeleteinus sp.-aff. Isodrusus sp. clade, Paradacrys ensiformis, Isodacrys crispum and the I. geminatumI. obrienorum clade; character state 4 synapomorphy for the Isodrusus debilis-I. guajavus clade (1: 8, ci: 0.50 , ri: 0.77 ).
16. Rostrum, ventral margin of scrobe, length relative to ventral surface of rostrum: (0) Ending before ventral surface; (1) reaching ventral surface (Fig. 4E). ACCTRAN optimization is preferred to preserve homology of character state 1 . Thus, character state 1 synapomorphy for the Pandeleteinus-Isodacrys clade with reversals in Pandeleteinus submetallicus and the Paradacrys ensiformis-Scalaventer jamaicensis clade (1: 4, ci: 0.25 , ri: 0.75).
17. Rostrum, scrobe, anterior portion, length relative to posterior portion: (0) At least one fifth longer; (1) subequal (Fig. 4E); (2) at least one fifth shorter. Non-additive. Coded as inapplicable when scrobes are curved (see character 15 , states 0 and 1 ). State 0 present in Pandeleteius inflatus; state 1 convergently present in Paululusus and the Isodillex-Isodacrys geminatum clade; state 2 convergently present in the PandeleteinusScalaventer clade, the Isodacrys crispum-I. schwarzi clade, I. burkei, the I. kuchiiI. fasciatum clade and the I. antrum-I. obrienorum clade (1: 7, ci: 0.28 , ri: 0.61 ).
18. Rostrum, scrobe, posterior portion, distance to eye: (0) At least one fifth longer than width of scrobe (Fig. 4E); (1) as wide as width of scrobe; (2) at least one fifth narrower than width of scrobe. Non-additive. Coded as inapplicable when scrobes are curved (see character 15, states 0 and 1). State 0 convergently present in Pandeleteinus elytroplanatus-P. submetallicus, Paululusus, Isodacrys schwarzi, I. apicale-I. ovipennis, I. carlae-I. fasciatum and I. frontalis; state 1 convergently present in Pandeleteius inflatus, Pandeleteinus subcancer-Isodacrys buchanani, I. kuchii and I. brevirostre; state 2 convergently present in the aff. Pandeleteinus sp.-Isodrusus guajavus clade, Isodillex, Isodacrys okuiltontli and the I. geminatum-I. obrienorum clade (1: 11, ci: 0.18, ri: 0.55 ).
19. Rostrum in lateral view, angle relative to head: (0) Directed anteriorly, in line with main body axis; (1) directed ventrally. Character state 1 synapomorphy for the Tanymecini clade (1: 1, ci: 1: ri: 1).

## Head (Fig. 4)

A fovea can be present between the eyes (at posterior end of the median sulcus, when present). The convexity of eyes is variable within Isodacrys, from slightly protruding to strongly convex, always more convex posteriorly.
20. Head, frons, fovea between eyes: (0) Absent; (1) present (Fig. 4D). State 1 convergently present in Platyaspistes prasinus, Hadromeropsis brevicoma-Pandeleteinus submetallicus, Scalaventer cyrillae, Isodillex and the Isodacrys apicale-I. obrienorum clade, with subsequent reversals in Minyomerus, the Pandeleteinus subcancer-Isodacrys okuiltontli clade, I. mexicanum, I. ovipennis and the I. carlae-I. fasciatum clade (1: 9, ci: 0.11 , ri: 0.55).
21. Head, eyes, convexity: (0) Strongly convex (Fig. 4B); (1) flat to slightly convex. See Marvaldi \& Lanteri (2005), character 3 in Girón \& Franz (2010), character 36 in Franz (2012), character 4 in Del Río et al. (2018) and character 27 in Marvaldi et al. (2018). State 1
convergently present in Platyaspistes prasinus and the Isodacrys orizabae-I. ovipennis clade (l: 2, ci: 0.50 , ri: 0.66 ).

## Mouthparts (Fig. 4)

The mandibles of Entimine weevils can be covered by setae and scales, the latter covering either partially or apparently the entire surface of the mandibles. In Isodacrys (as well as in other tanymecines), the mandibles are covered by setae, without scales. The presence of scales in the mandibles has been highlighted to separate some genera of New World Tanymecini (Howden, 1982).

Exposure of the maxillary palpi within Entiminae has been observed in several tribes (Anderson, 2002; Girón \& Franz, 2010; Marvaldi et al., 2014) that are not closely related. Therefore, this condition within Tanymecini as in other tribes could represent isolated cases of reversals, if adelognathy (prementum completely covering maxillae) evolved early in the subfamily and was reversed secondarily (Marvaldi et al., 2014). Nonetheless, we do not follow the traditional nomenclature (adelognathous and phanerognathous conditions, respectively) to refer to these states because there are different opinions about the limits between them (Thompson, 1992). Howden (1959) had already noticed that the lower part of the maxillae (i.e., cardo, stipes) is visible for Pandeleteius and Pandeleteinus and this is also applicable to Isodacrys and other tanymecine genera (see also Howden, 1993b).
22. Mouthparts, mandibles, scales: (0) Present; (1) absent. Character state 1 synapomorphy for Pandeleteius-Isodacrys clade, with reversal in Pandeleteinus submetallicus (1: 2, ci: 0.50 , ri: 0.80 ).
23. Mouthparts, prementum relative to maxillae: (0) Prementum completely concealing maxillae; (1) prementum incompletely concealing maxillae (Fig. 4F). See character 19 in Marvaldi et al. (2018). DELTRAN optimization is preferred based on the assumption of no immediate relationship with Platyaspistes prasinus (Leptopiini). Thus, character state 1 convergently present in P. prasinus and the Hadromeropsis-Isodacrys clade (1: 2, ci: 0.50, ri: 0.50).

## Antenna (Figs. 3, 5)

The antennal scape in New World Tanymecini is covered by interspersed setae along its surface (Fig. 5A). Additionally, some clades within Isodacrys also have appressed scales, restricted to the apical half of the scape. In the outgroups when scales are present, they evenly cover the scape. The shape of the scape in the studied Tanymecini is consistently capitate, abruptly thickened at apex (Nichols, 1989; Fig. 3A). Additionally, funicular antennomeres I and II are usually of different shape and longer than remaining funicular antennomeres (Fig. 3A).
24. Antenna, scape, squamose vestiture: (0) Present; (1) absent (Fig. 5A). See character 2 in Lanteri \& Del Río (2017). State 0 convergently present in Megalostylus albicans-Minyomerus, the Isodacrys buchanani-I. schwarzi clade, the I. burkei-I. ovipennis clade and I. confusum; state 1 synapomorphy for the Hadromeropsis brevicoma-Isodacrys clade (1: 4, ci: 0.25 , ri: 0.62 ).
25. Antenna, scape, squamose vestiture, arrangement: (0) Scape evenly covered by scales; (1) scape with scales restricted to dorsal surface or apex (Fig. 3D). State 1 convergently present in Isodacrys buchanani-I. schwarzi, I. burkei-I. ovipennis and I. confusum (1: 1, ci: 1, ri: 1).
26. Antenna, scape, shape: (0) Clavate, gradually thickening towards apex; (1) capitate, abruptly thickened at apical region (Fig. 5A). See character 30 in Franz (2012), character 12 in Girón \& Franz (2012), character 29 in Lanteri \& Del Río (2017), character 6 in Del Río et al. (2018) and character 30 in Marvaldi et al. (2018). Character state 1 synapomorphy for the Tanymecini clade (l: 1 , ci: 1 , ri: 1 ).
27. Antenna, funicular antennomere I, length relative to funicular antennomere II: (0) Equal; (1) shorter; (2) longer (Fig. 5A). Non-additive. See character 8 in Del Río et al. (2018) and character 34 in Marvaldi et al. (2018). Character state 1 autapomorphy for Hadromeropsis brevicoma; character state 2 synapomorphy for the Tanymecini clade (l: 2, ci: 1, ri: 1).

## Prothorax (Figs. 3-5)

In Isodacrys, the shape of the pronotum is always subcylindrical, with constrictions near anterior and posterior margins (Fig. 3C), making the sides of the pronotum between these constrictions sinuate (Howden, 1961).

The presence of postocular vibrissae has been a traditional character for distinguishing Tanymecini from other tribes (Lacordaire, 1863; LeConte \& Horn, 1876; Van Emden, 1944; Howden, 1970; Fig. 3B). Nevertheless, this character is evidently homoplastic because there is no trace of postocular vibrissae in several genera and species of New World Tanymecini, and vibrissae are also present in other tribes of broad-nosed weevils (e.g., Anomophthalmini; Morrone, 1998). These are likely used to clean the eyes of debris by pulling the rostrum down and back. Thus, it may have significant adaptive value and be subject to convergent evolution (Del Río \& Lanteri, 2019b) or retentions. Notwithstanding, we consider that there may be a latent phylogenetic signal at this level. Postocular vibrissae characters follow observations made in previous works (Howden, 1961, 1969; Franz, 2012; Jansen \& Franz, 2015; Marvaldi et al., 2018). Postocular vibrissae are considered vestigial when their length does not surpass at least half length of the compound eye. Additionally, vestigial postocular vibrissae are thin and usually only visible with high magnification. The well-developed ones are at least two thirds length of the eye. Franz (2012) considered the postocular vibrissae in members of Eustylini, which are set inward in a postocular lobe and as a fringe (see Franz, 2012, character 48) as non-homologous from members of Pandeleteius, which are finer, fewer, more arched, and more externally situated. Following Franz's reasoning, characters of the postocular vibrissae were coded as inapplicable for Platyaspistes prasinus, which presents postocular lobes. According to the most parsimonious trees, there is a trend to the reduction/loss of the postocular vibrissae within New World Tanymecini.
28. Pronotum in dorsal view, shape: (0) Trapezoidal, sides convergent from the base towards the apex; (1) subglobose, sides strongly curved, maximum width near midlength (Fig. 5B); (2) subcylindrical, sides at most slightly sinuate (Fig. 5F). Additive.

See Howden (1959, 1961), character 38 in Lanteri \& Del Río (2017), character 10 in Del Río et al. (2018) and character 38 in Marvaldi et al. (2018). State 1 synapomorphy for Tanymecini (although only present in the Hadromeropsis brevicoma-Pandeleteinus submetallicus clade), with subsequent convergent transition to state 2 in Minyomerus, Pandeleteinus elytroplanatus and the P. subcancer-Isodacrys clade (1: 4, ci: 0.50, ri: 0.71 ).
29. Pronotum in dorsal view, length/width ratio: (0) Wider than long (Fig. 5B); (1) as wide as long (Fig. 5E); (2) longer than wide (Fig. 5F). Additive. Character state 1 convergently present in the Pandeleteius hilaris-Isodillex clade, Isodacrys schwarzi, I. apicale, the I. confusum-I. fasciatum clade, I. frontalis and the I. antrum-I. obrienorum clade; character state 2 convergently present in Pandeleteinus subcancer, the Isodrusus debilis-I. guajavus clade and the Isodacrys clade (l: 9, ci: 0.22 , ri: 0.65 ). As wide as long is limited to ratios of length/width not surpassing one tenth longer/wider.
30. Pronotum in dorsal view, constrictions near anterior and posterior margins: (0) Absent; (1) present (Figs. 3C and 5B). Character state 1 synapomorphy for the Tanymecini clade (l: 1, ci: 1, ri: 1 ).
31. Prothorax in lateral view, anterior margin, shape: (0) Nearly straight, at level with ventral margin; (1) oblique, dorsally produced anteriorly (Fig. 5C). ACCTRAN optimization is preferred because it preserves ancestral homology of character state 1 for the Tanymecini clade, with reversal in Hadromeropsis brevicoma (l: 2, ci: 0.50, ri: 0.50).
32. Prothorax, postocular vibrissae: (0) Absent (Fig. 2); (1) present (Fig. 5C). See Howden (1959, 1961, 1970), character 48 in Franz (2012) and character 37 in Marvaldi et al. (2018). DELTRAN optimization is preferred as it posits the presence of postocular vibrissae as ancestral state in the Tanymecini clade. Thus, state 1 convergently present in the Minyomerus-Isodrusus debilis clade, the Isodacrys crispum-I. schwarzi clade and the I. apicale-I. ovipennis clade, the latter with reversal in I. burkei (l: 6, ci: 0.16 , ri: 0.68 ). According to our MPT's the most plausible scenario within Tanymecini is that absence of postocular vibrissae is a derived condition in Isodacrys, Isodillex, and the West-Indian clade.
33. Prothorax, postocular vibrissae, number: (0) More than six postocular vibrissae (Fig. 5C); (1) from one to six postocular vibrissae. Coded as inapplicable when the postocular vibrissae are absent (see character 32, state 0 ). State 0 present in Minyomerus laticeps-Pandeleteinus elytroplanatus; state 1 convergently present in Minyomerus microps and the Pandeleteinus submetallicus-Isodacrys clade, with subsequent reversals in Isodacrys schwarzi and I. apicale (1: 4, ci: 0.25, ri: 0.57).
34. Prothorax, postocular vibrissae, number of well-developed postocular vibrissae: (0) More than five well-developed postocular vibrissae; (1) from one to five well-developed postocular vibrissae (Fig. 5C); (2) all postocular vibrissae short, vestigial. Additive. Coded as inapplicable when postocular vibrissae are absent (see character 32, state 0 ). State 0 present in Minyomerus laticeps-Pandeleteinus elytroplanatus; state 1 convergently present in the Pandeleteinus submetallicus-Isodrusus debilis clade, Isodacrys apicale and I. ovipennis; state 2 convergently present in Minyomerus microps, aff. Isodrusus sp. and Isodacrys crispum-I. obrienorum (1: 7, ci: 0.28, ri: 0.54).
35. Prosternum, intercoxal process, degree of development: (0) Not continuous, divided in two halves (anterior and posterior) not contiguous to each other; (1) continuous. See characters 18 and 19 in Jansen \& Franz (2015). Character state 1 synapomorphy for the Pandeleteius-Isodacrys clade (1: 1, ci: 1, ri: 1). Isodacrys species always have the intercoxal process continuous and therefore the procoxae are separated (Howden, 1961).

## Elytra (Figs. 3, 5, 6)

The vestiture of the elytra in Isodacrys is evenly distributed in single rows on the intervals or interspersed irregularly. There are some outgroups where setae are interspersed or absent on some intervals (e.g., Scalaventer, see Howden, 1970).

In entimines, humeri absence has been proposed to be associated with thickening and merging of the elytra along the elytral suture, and reductions in the metendosternite and wings (Franz, 2012). Absence of humeri is considered herein when humeral lobes sensu Torre-Bueno (Nichols, 1989) cannot be distinguished. The apterous condition, reflected by the lack of humeri, is constant in Isodacrys whereas in other New World Tanymecini it varies interspecifically (Howden, 1970; original observations, e.g., aff. Isodrusus sp.). Additionally, in wingless Tanymecini the number of visible intervals at the base of the elytra can vary from five to seven (Figs. 3C, 5E, 5F). This may be related to the development of the humeral angles. Howden (1961) considered that several Isodacrys species have partially developed/vestigial humeri because the elytra can be slightly produced at the very base of intervals seven to nine, where humeri take place in winged weevils. These slight elevations are not considered proper humeri herein. The basal margin of the elytra is usually emarginate in Neotropical tanymecines (e.g., Fig. 3C). Although Howden (1961) stated that some Isodacrys species have the basal margin of the elytra straight, the epipleural intervals of the elytra always extend more anteriorly than the dorsal ones. The shape of the emargination of the basal margin of the elytra is usually more clear-cut in winged genera. Finally, elytral intervals and the apical declivity (Figs. 3D, 5F, 6 A) can provide additional characters. In some tanymecines, the odd-numbered intervals can be convex relative to the even-numbered intervals (Howden, 1970; Lanteri \& Del Río, 2017). Within Isodacrys the intervals are evenly flat, but some species display basal or apical elevations and/or depressions (Howden, 1961). The elytra can be evenly arcuate from base to apex or deflected along apical third, forming a discernible declivity (Howden, 1961). When the apical declivity is evident, the shape below summit of the declivity can be oblique to different degrees to almost straight (Fig. 6A) or strongly concave (Fig. 3D) in some Isodacrys species as in other tanymecines (Howden, 1961, 1970).
36. Elytra, setae, arrangement: (0) In single rows, present in all intervals; (1) in single rows, not present in all intervals. Inapplicable when setae are interspersed. Character state 1 synapomorphy for the Scalaventer cyrillae-S. subtropicus clade (1: 1, ci: 1, ri: 1).
37. Elytral setae relative to remaining setae of dorsal surface of head and pronotum: (0) At most slightly longer; (1) strongly modified, conspicuously longer (Fig. 5D). State 1 convergently present in Paululusus constanzae and the Isodacrys ellipticum-I. guatemalenum clade (1: 2, ci: 0.50, ri: 0.50 ).
38. Elytra in dorsal view, shape: (0) Oblong, sides converging to apex from midlength; (1) subcylindrical, sides converging to apex near apical fourth (Fig. 5B); (2) obovate, widest point before midlength; (3) subelliptical, widest point near midlength (Fig. 5D); (4) ovate, widest point after midlength (Fig. 5F). Non-additive. State 1 present in the Hadromeropsis brevicoma-Scalaventer subtropicus clade; state 2 convergently present in Minyomerus and the Isodacrys okuiltontli-I. obrienorum clade; state 3 convergently present in Pandeleteius inflatus, aff. Isodrusus sp., Paululusus constanzae, the Isodillex-Isodacrys schwarzi clade, the I. brevirostre-I. frontalis clade and I. antrum (1: 10, ci: 0.40 , ri: 0.76 ).
39. Elytra, humeri: (0) Present (Fig. 5B); (1) absent (Fig. 5D). See character 62 in Franz (2012), character 19 in Girón \& Franz (2012), character 51 in Lanteri \& Del Río (2017) and character 22 in Del Río et al. (2018). Character state 1 convergently present in Minyomerus, aff. Isodrusus sp., Paululusus constanzae and the Isodillex-Isodacrys clade (l: 4, ci: 0.25 , ri: 0.81 ).
40. Elytra in dorsal view, visible intervals at base: (0) Seven (Fig. 5F); (1) five (Fig. 5E). Coded as inapplicable when humeri are present (Character 39, state 0 ). State 0 synapomorphy for the Paululusus constanzae-Isodacrys ovipennis clade; state 1 convergently present in Minyomerus, aff. Isodrusus sp. and the Isodacrys kuchiiI. obrienorum clade. (1: 2, ci: 0.50 , ri: 0.90 ).
41. Elytra, base, shape: (0) Bisinuate; (1) straight; (2) emarginate (Fig. 5E). Non-additive. See character 50 in Lanteri \& Del Río (2017) and character 21 in Del Río et al. (2018). DELTRAN optimization is preferred as it states a common origin of character state 2 for Hadromeropsis-Isodacrys clade. ACCTRAN optimization implies an ancestral state 1 for Tanymecini clade, which could be problematic because Minyomerus has several psammophilic traits (adaptations to sandy environments) (Jansen \& Franz, 2015). Thus, character state 1 synapomorphy for Minyomerus; character state 2 synapomorphy for the Hadromeropsis-Isodacrys clade (1: 2, ci: 1, ri: 1).
42. Elytra, base, emargination, shape: (0) Strongly emarginate medially (Fig. 5B); (1) roundly emarginate (Fig. 5E); (2) angularly emarginate. Non-additive. State 0 present in Hadromeropsis brevicoma-Scalaventer subtropicus; state 1 convergently present in Pandeleteius inflatus, aff. Isodrusus sp., Paululusus constanzae and the Isodillex-Isodacrys clade; state 2 synapomorphy for the Isodacrys buchanani-I. schwarzi clade (1: 5, ci: 0.40, ri: 0.78).
43. Elytra, strial punctures, degree of development: (0) Conspicuous, deep (Fig. 5E); (1) inconspicuous. Character state 1 synapomorphy for Minyomerus (l: 1, ci: 1, ri: 1). In some species of Minyomerus the strial punctures are faint or not evident beneath the appressed scales (Jansen \& Franz, 2015).
44. Elytra, striae, type: (0) Striate-punctate; (1) punctate (Fig. 5E). Coded as inapplicable when strial punctures are indistinct (see character 43, state 1). DELTRAN optimization is preferred, which preserves common origin of character state 1 . Thus, state 1 synapomorphy for the Pandeleteinus-Isodacrys clade, with subsequent reversals in aff. Pandeleteinus sp., aff. Isodrusus sp. and Paradacrys ensiformis (1: 4, ci: 0.25 , ri: 0.62 ).
45. Elytra, intervals, convexity: (0) Evenly flat; (1) alternate intervals convex. See character 63 in Franz (2012) and character 59 in Lanteri \& Del Río (2017).

Character state 1 convergently present in Pandeleteius hilaris and the Scalaventer cyrillae-S. subtropicus clade (1: 2, ci: 0.50 , ri: 0.50 ).
46. Elytra, intervals, swelling at apical junction of intervals two and ten; five and six: (0) Absent; (1) present (Fig. 5F). Character state 1 synapomorphy for the Isodacrys burkeiI. ovipennis clade (l: 1, ci: 1, ri: 1).
47. Elytra, intervals three to five at base: (0) Not elevated; (1) elevated. ACCTRAN optimization is preferred, which preserves homology statement within Isodacrys. Thus, state 1 convergently present in Pandeleteius rotundicollis, P. hilaris, Scalaventer and in the Isodacrys mexicanum-I. ovipennis clade, the latter with reversal in I. orizabae (1: 5, ci: 0.20, ri: 0.42).
48. Elytra, intervals seven to nine at basal sixth, depression: (0) Absent; (1) present. Character state 1 convergently present in the Isodacrys brevirostre-I. frontalis clade and I. antrum (1: 2, ci: 0.50 , ri: 0.50 ).
49. Elytra in lateral view, apical declivity, shape: (0) Oblique to straight (Fig. 6A); (1) concave (Fig. 3D). Coded as inapplicable in species with elytra evenly arcuate. See character 66 in Franz (2012) and character 56 in Lanteri \& Del Río (2017). State 1 convergently present in Paululusus hispaniole and the Isodacrys buchanani-I. schwarzi clade (l: 2, ci: 0.50 , ri: 0.50 ).
50. Elytra, apical margins, shape: (0) Conjointly rounded; (1) bisinuate (Fig. 6B). See Howden (1961), character 58 in Lanteri \& Del Río (2017) and character 43 in Marvaldi et al. (2018). State 1 convergently present in Paululusus and Isodacrys apicale (1: 2, ci: 0.50, ri: 0.50).

## Legs (Figs. 5-6)

Within Tanymecini (as in other entimines), the relative size of the legs varies in different ways, from prolegs distinctly larger than metalegs to prolegs shorter than metalegs. The relative size of mesolegs is not considered because the mesotibiae are always the smallest and the mesofemora are of the same size of the non-enlarged pair. In some species the prolegs can be larger in males than in females (Howden, 1959, 1961, 1970, 1982; Franz, 2012; Jansen \& Franz, 2015, 2018). The width of the metafemora was taken from maximum width of the lateral outer face, as many Curculionoidea have the metafemora (and sometimes meso and profemora) with its inner face flattened and curved when seen in dorsal view (see metaleg in Fig. 6C).

Commonly in weevils, there are denticles, often called "teeth", along the inner margin of the protibiae, which may vary in number and degree of development among individuals or species (Howden, 1959, 1961, 1963, 1970). Some clades within Isodacrys may have the protibial inner margin smooth, without teeth. Terminology of the metatibial apex (true corbel, false corbel and simple metatibial apex) follow Oberprieler (2010) and Marvaldi et al. (2014), incorporating the observations made by Thompson (1992) and therefore it is presented as in Cortes-Hernández \& Morrone (2019, see figures 1-6). In Isodacrys species the metatibial apex is simple. Additionally, the metatibial apex in some Entiminae can bear two different combs of bristles: a distal comb across the apex, oriented transversally to the main axis of tibia, and an ascending dorsal comb on the outer margin of metatibiae,
oriented longitudinally to the main axis of tibia (Buchanan, 1939; Anderson, 2002; Marvaldi et al., 2018). Nonetheless, the shape of the distal comb varies; in Isodacrys and other Neotropical Tanymecini the distal comb is obliquely arcuate, with two discernible regions: the innermost bears short bristles while the outermost bears long bristles (Fig. 6E). This is apparently related to the shape of the outer apical edge of the tibiae.
51. Legs, profemora, length relative to metafemora: (0) At least one fifth shorter; (1) subequal (Fig. 6C); (2) at least one fifth longer (Fig. 5B). Non-additive. See character 51 in Franz (2012). State 1 synapomorphy for the Pandeleteius hilaris-Isodacrys clade; state 2 convergently present in Hadromeropsis brevicoma-Pandeleteius rotundicollis and Paululusus (l: 3, ci: 0.66, ri: 0.83).
52. Legs, profemora, width relative to metafemora: (0) Subequal; (1) at least one fifth wider (Fig. 5B). See character 51 in Franz (2012), character 20 in Jansen \& Franz (2015), character 66 in Lanteri \& Del Río (2017) and character 46 in Marvaldi et al. (2018). State 0 convergently present in Megalostylus albicans-Minyomerus, Isodrusus debilis and the Isodacrys crispum-I. obrienorum clade; state 1 convergently present in the Hadromeropsis brevicoma-Isodacrys guatemalenum clade, the I. mexicanum-I. ovipennis clade and the I. kuchii-I. fasciatum clade (l: 5, ci: 0.20, ri: 0.71 ).
53. Legs, protibiae, length relative to metatibiae: (0) Subequal (Fig. 6A); (1) at least one fifth longer (Fig. 5B). State 0 convergently present in Megalostylus albicans-Minyomerus, Pandeleteinus elytroplanatus, Isodrusus debilis and the Isodacrys crispum-I. obrienorum clade; state 1 convergently present in the Hadromeropsis brevicoma-Isodacrys guatemalenum clade and the I. mexicanum-I. ovipennis clade (l: 5, ci: 0.20, ri: 0.78).
54. Legs, inner margin of protibiae, teeth: (0) Absent (Fig. 2); (1) present. See character 56 in Franz (2012), character 70 in Lanteri \& Del Río (2017), character 31 in Del Río et al. (2018) and character 48 in Marvaldi et al. (2018). DELTRAN optimization is preferred based on the assumption of no immediate relationship with Platyaspistes prasinus (Leptopiini). Thus, state 1 convergently present in P. prasinus, the Hadromeropsis brevicoma-Isodacrys ovipennis clade and I. antrum, with subsequent reversals in Pandeleteinus subcancer, Isodrusus debilis, the Isodacrys buchanani-I. schwarzi clade and the I. kuchii-I. obrienorum clade (1: 7, ci: 0.14 , ri: 0.57).
55. Legs, metatibial apex, vestiture of distal comb: (0) Bristles setiform, shorter or as long as surrounding setae (Fig. 6E); (1) bristles conical, thickened, shorter than surrounding setae. See character 22 in Jansen \& Franz (2015). Character state 1 synapomorphy for Minyomerus (see figures 5 and 6 in Cortes-Hernández \& Morrone, 2019; 1: 1, ci: 1, ri: 1).
56. Legs, metatibial apex, corbel: (0) Present; (1) absent (Fig. 6D). See character 58 in Franz (2012), character 71 in Lanteri \& Del Río (2017) and character 50 in Marvaldi et al. (2018). Character state 1 synapomorphy for the Hadromeropsis-Isodacrys clade (l: 1, ci: 1, ri: 1 ).
57. Legs, metatibial apex, false corbel: (0) Absent (Fig. 6D); (1) present. Coded as inapplicable when true corbel is present (see character 56 , state 0 ). See character 58 in Franz (2012) and character 51 in Marvaldi et al. (2018). Character state 1 synapomorphy for Scalaventer (see figures 3 and 4 in Cortes-Hernández \& Morrone, 2019; 1: 1, ci: 1, ri: 1).
58. Legs, metatibial apex, distal comb, arrangement: (0) Straight to slightly arcuate, at most with bristles progressively longer posteriorly; (1) restricted to outermost region of tibiae; (2) obliquely arcuate, bristles in two discernible parts, the outermost bearing longer bristles (Fig. 6E). Non-additive. Character state 0 present in Megalostylus albicans and Minyomerus; character state 1 autapomorphy for Platyaspistes prasinus; character state 2 synapomorphy for the Hadromeropsis brevicoma-Isodacrys clade (1: 2, ci: 1, ri: 1).
59. Legs, tarsomeres 1,2 and 3, ventral surface, vestiture: (0) With pads of setiform setae (Fig. 6E); (1) with stout, spiniform setae. See character 25 in Jansen \& Franz (2015). Character state 1 synapomorphy for Minyomerus (l: 1, ci: 1, ri: 1).
60. Legs, tarsal claws, arrangement: (0) Free (Fig. 6E); (1) connate. See characters 61 and 53 in Franz (2012) and in Marvaldi et al. (2018), respectively. Character state 1 convergently present in Platyaspistes prasinus and the aff. Isodrusus sp.-Isodrusus guajavus clade (1: 2, ci: 0.50 , ri: 0.66 ).

Abdomen (Fig. 6)
Howden (1969) observed that several genera of Neotropical Tanymecini have the anterior margin of ventrites III, IV and V modified, each with a transverse sulcus (Fig. 6F). The sulci of ventrites IV and $V$ are each progressively wider. These sulci can be extended across the entire width of each ventrite or medially only (Anderson, 2002), enclosed by the lateral margins of the ventrites, where they are squamose (Howden, 1970). The posterior margin of the sulcus can be evenly rounded relative to the posterior surface of the ventrite or carinate. These sulci were highlighted as potentially valuable characters on a generic and a specific level in Tanymecini (Howden, 1969).
61. Abdomen, anterior margin of ventrites III, IV and V, sulcus: (0) Absent, ventrites flat; (1) present (Fig. 6F). Character state 1 synapomorphy for the Pandeleteinus-Isodacrys clade, with reversal in Isodillex (1: 2, ci: 0.50 , ri: 0.88 ).
62. Abdomen, anterior margin of ventrites III, IV and V, degree of development: (0) Enclosed by lateral margins of ventrites (Fig. 6F); (1) along entire width. Coded as inapplicable when sulci are absent (see character 61, state 0 ). Character state 1 convergently present in the aff. Isodrusus sp.-Isodrusus guajavus clade and Scalaventer (1: 2, ci: 0.50 , ri: 0.80 ).
63. Abdomen, anterior margin of ventrites III, IV and V , posterior margin of sulcus: (0) Present in all, sharply delimiting sulcus; (1) obliterate in ventrites IV and V, sulcus becoming flat posteriorly (Fig. 6F). Coded as inapplicable when sulci are absent (see character 61, state 0). State 0 convergently present in Pandeleteinus subcancer, the aff. Isodrusus sp.-Isodrusus guajavus clade, Scalaventer and Isodacrys crispum; state 1 present in the Pandeleteinus-Isodacrys clade (1: 4, ci: 0.25 , ri: 0.57 ).

## Terminalia (Figs. 3 and 7)

Characters of the female terminalia are explained and discussed in Howden (1995) and Lanteri \& Del Río (2008). Particular character states for Isodacrys and other New World Tanymecini are herein highlighted. The ovipositor in Isodacrys is membranous and flexible, with slightly sclerotized regions, including the gonocoxites, baculi and proximal
rods (Fig. 3F). The distal gonocoxites are usually moderately sclerotized laterally and the styli are reduced to a single thickened seta (Figs. 3F and 7B). Although Howden (1995) stated that Pandeleteius hilaris has a single ventral baculus, there are two baculi very close to each other, diverging posteriorly just before the distal gonocoxites. This condition is also present in Isodacrys species (Fig. 7B) as in other Neotropical tanymecines.

Although Howden pointed out the relevance of the aedeagus to separate some genera of Neotropical Tanymecini (Howden, 1961, 1970) she did not establish discrete characters or clear differences among aedeagus types. Following her observations, we consider the following aedeagal characters into account. The penis of some entimines has a sclerotized structure in the proximal portion of the internal sac within the median lobe, referred as genital sclerite (Bruhn, 1947; Oberprieler, 2014; Jansen \& Franz, 2015, 2018). In Isodacrys the genital sclerite within the median lobe is absent, but a proximal flagellum can be present (Fig. 3E). In the pedal type of aedeagus, the sternite VIII is divided into two hemisternites (Oberprieler, Anderson \& Marvaldi, 2014). These hemisternites are crescentic or teardrop shaped in the terminals included in this study, with their outer corners acute or truncate. Howden (1961) stated that the median lobe in Isodacrys is generally short and stout, but males remain unknown in eight species. The length/width ratio of the median lobe was considered in preliminary analyses, depicting a high level of incongruence (homoplasy). For that reason, it was excluded from subsequent analyses. Length relative to other genital structures resulted in more informative homology hypotheses.
64. Female terminalia, sternite VIII, lamina, shape: (0) Subrhomboidal; (1) subcordiform; (2) triangular (Fig. 7A). Non-additive. See character 78 in Lanteri \& Del Río (2017). DELTRAN optimization is preferred, which establishes common origin of character state 1 in Minyomerus and not in Platyaspistes prasinus (Leptopiini), where genital characters were not observed. Thus, state 1 synapomorphy for Minyomerus; state 2 for the Hadromeropsis brevicoma-Isodacrys clade (1: 2, ci: 1, ri: 1).
65. Female terminalia, sternite VIII, apex of lamina, shape: (0) Acute (Fig. 7A); blunt (1). Coded as inapplicable when the lamina of sternite VIII is not triangular (see character 64, state 2). See character 79 in Lanteri \& Del Río (2017). State 1 convergently present in Hadromeropsis brevicoma, the Paululusus-Scalaventer clade, Isodillex, Isodacrys guatemalenum, I. okuiltontli, I. mexicanum and the I. geminatum-I. obrienorum clade (l: 7, ci: 0.14, ri: 0.50 ).
66. Female terminalia, ovipositor, baculi, arrangement: (0) Distinctly separated by distance greater than width of one baculus; (1) almost contiguous, separated by distance similar to width of one baculus (Fig. 7B). Character state 0 present in Megalostylus albicans-Minyomerus; state 1 present in Pandeleteius-Isodacrys (1: 1, ci: 1, ri: 1).
67. Female terminalia, ovipositor, distal gonocoxites, degree of sclerotization: (0) Not sclerotized; (1) slightly to moderately sclerotized laterally (Fig. 7B); (2) strongly sclerotized. Non-additive. See character 84 in Lanteri \& Del Río (2017). Character state 1 synapomorphy for the Pandeleteius-Isodacrys clade; character state 2 present in Minyomerus-Hadromeropsis brevicoma (1: 2, ci: 1, ri: 1).
68. Female terminalia, ovipositor, styli : (0) Present, strongly sclerotized; (1) vestigial, represented by single thickened seta (Fig. 7B). Character state 1 synapomorphy for the Pandeleteius-Isodacrys clade (l: 1, ci: 1, ri: 1).
69. Male terminalia, genital sclerite: (0) Present; (1) absent (Fig 7C). See character 37 in Jansen \& Franz (2015) and character 99 in Lanteri \& Del Río (2017). Character state 0 convergently present in Megalostylus albicans-Pandeleteius rotundicollis, Paradacrys ensiformis, Scalaventer jamaicensis and S. cyrillae; character state 1 present in the Pandeleteius hilaris-Isodacrys clade (1: 4, ci: 0.25, ri: 0.40 ).
70. Male terminalia, sternite VIII, outer corner, shape: (0) Truncate; (1) not truncate (Fig. 7D). ACCTRAN optimization preferred because it posits a common origin of character state 1 . Thus, character state 0 convergently present in Megalostylus albicans, Hadromeropsis brevicoma, Pandeleteius hilaris and Isodacrys apicale-I. ovipennis clade; character state 1 synapomorphy for the Pandeleteius-Isodacrys clade (1: 3, ci: 0.33 , ri: 0.66 ).
71. Male terminalia, median lobe, length relative to spiculum gastrale: (0) About as long as spiculum gastrale; (1) shorter. DELTRAN optimization is preferred based on the absence of males in Isodacrys ellipticum, therefore character state 1 is considered as independent in I. guatemalenum. Thus, state 1 convergently present in aff. Pandeleteinus sp., Paradacrys ensiformis, Isodacrys guatemalenum, the I. orizabae-I. ovipennis clade and I. kuchii (1: 6, ci: 0.16, ri: 0.28 ).
72. Male terminalia, tegmen, manubrium, length relative to median lobe: (0) Shorter; (1) as long as median lobe; (2) longer (Fig. 7C). Non-additive. State 0 convergently present in Megalostylus albicans-Pandeleteius hilaris and the aff. Pandeleteinus sp.-Isodillex plumosum clade; state 1 synapomorphy for Isodacrys (although with an evolutionary transition to state 2 in the I. kuchii-I. obrienorum clade); state 2 convergently present in Pandeleteinus and the Isodacrys kuchii-I. obrienorum clade (1: 5, ci: 0.40 , ri: 0.66 ).

Systematic revision of the genus Isodacrys Sharp, 1911
(Figs. 1, 2, 3, 4B, 4D, 4E, 5A, 5C-5F, 6-25)
Isodacrys Sharp, 1911: 175-177; Champion, 1911: 341. Gender masculine.
Type species. Isodacrys guatemalenum Sharp, 1911; subsequently designated by Pierce, 1913: 401.
 scales variously overlapping non-linearly or contiguous, polygonal (chars.1:0,1 and 2:1). Dorsolateral margins of rostrum subparallel (char. 3:1), dorsal surface flat, nasal plate with anterior margin medially indented (chars. 4:0 and 5:1), posterior margin carinate (char. 6:1); epistome in continuous plane with remainder of rostrum (chars. 7:1 and 8:2), or slightly depressed, covered by smaller, shinier scales of different shape than remainder of rostrum, posterior margin of epistome indistinct (char. 9:0); scrobes glabrous (char. 14:1), angled, from obtusely-angled to right-angled (char. 15:2,3), reaching ventral surface of rostrum (char. 16:1); head deflexed, directed ventrally (char. 19:1); mandibles covered with setae, without scales (char. 22:1); prementum incompletely concealing maxillae (char. 23:1); antennal scape capitate, abruptly thickened at apical region


Figure 8 Occurrence map of nine Isodacrys species, ordered alphabetically.

> Full-size DOI: 10.7717/peerj.10191/fig-8


Figure 9 Occurrence map of the remaining eleven Isodacrys species, ordered alphabetically.

> Full-size DOI: 10.7717/peerj.10191/fig-9


Figure 10 Isodacrys antrum new species. (A) Dorsal habitus; (B) lateral habitus; (C) ventral habitus; (D) head in anterior view. Full-size DOI: 10.7717/peerj.10191/fig-10


Figure 11 Isodacrys antrum new species, female genitalia. (A) Ovipositor in dorsal view; (B) ovipositor in lateral view; (C) sternite VIII in dorsal view; (D) spermatheca.

Full-size DOI: 10.7717/peerj.10191/fig-11
(char. 26:2), funicular antennomere I longer than funicular antennomere II (char. 27:2); pronotum in dorsal view subcylindrical (char. 28:2), from as wide as long to longer than wide (char. 29:1,2), with constrictions near anterior and posterior margins (char. 30:1),


Figure 12 Isodacrys carlae new species. (A) Dorsal habitus; (B) lateral habitus; (C) ventral habitus; (D) head in anterior view.

Full-size DOI: 10.7717/peerj.10191/fig-12
anterolateral margin of prothorax with postocular vibrissae or not (char. 32:0,1), when present usually vestigial (very short and thin) or with less than five well developed postocular vibrissae (char. 34:1,2), intercoxal process of prosternum continuous, separating procoxae (char. 35:1); humeri absent (char. 39:1), basal margin of elytra roundly or angularly emarginate (chars. 41:2 and 42:2); profemora as long as metafemora (char. 51:1), wider or not (char. 52:0,1), protibiae longer than metatibiae or not (char. 53:0,1), apex of metatibiae simple (chars. 56:1 and 57:0), distal comb of metatibial apex obliquely arquate (char. 58:2), tarsal claws free (char. 60:0); anterior margins of ventrites III, IV and V sulcate, sulci enclosed by lateral margins of ventrites (chars. 61:1 and 62:0); manubrium usually as long as or longer than median lobe, rarely shorter (char. 72:0,1,2).

The genus Isodacrys can be confused with other wingless tanymecines (Minyomerus, Isodillex, Paululusus in part). Isodacrys is easily distinguished from Minyomerus by the metatibial apex simple, covered by setae, whereas Minyomerus possesses a true corbel, covered by conical thickened bristles. Isodacrys can be separated from Paululusus by the scrobe reaching the ventral surface of the rostrum and the profemora as long as the metafemora, whereas in Paululusus the scrobe ends before the ventral surface of the rostrum and the profemora are longer than the metafemora. Finally, Isodacrys is easily separated from Isodillex by the sulcate anterior margins of ventrites III, IV and V, whereas in Isodillex the anterior margins of ventrites III, IV and V are flat, not sulcate.


Figure 13 Isodacrys carlae new species, female genitalia. (A) Ovipositor in dorsal view; (B) ovipositor in lateral view; (C) sternite VIII in dorsal view; (D) spermatheca.

Full-size DOI: 10.7717/peerj.10191/fig-13


Figure 14 Isodacrys confusum new species. (A) Dorsal habitus; (B) lateral habitus; (C) ventral habitus; (D) head in anterior view. Full-size DOI: 10.7717/peerj.10191/fig-14

Remarks. Isodacrys species are distributed from south of the United States of America to Honduras (Figs. 8-9), mainly across Mexican and Central American mountain ranges of $\sim 1500-3100 \mathrm{~m}$ above sea level; some species also occur in lowlands. Adults have been found on nine plant families (Asteraceae, Betulaceae, Cucurbitaceae, Fabaceae, Fagaceae, Malvaceae, Pinaceae, Rhamnaceae and Solanaceae), leaf litter, unidentified mushrooms and under rocks.


Figure 15 Isodacrys confusum new species, genitalia. (A) Median lobe in dorsal view; (B) median lobe and tegmen in lateral view; (C) male hemisternites VIII and sternite IX in dorsal view; (D) ovipositor in dorsal view; (E) female sternite VIII in dorsal view; (F) spermatheca; (G) ovipositor in lateral view.

Full-size DOI: 10.7717/peerj.10191/fig-15

## Isodacrys antrum Cortés-Hernández, new species

(Figs. 8, 10-11)
urn:lsid:zoobank.org:act:FBE7F34A-9C93-4BDC-9423-3D3A915B926A
Diagnosis. Setae lanceolate, completely arched, mostly inconspicuous on temple and disc of pronotum; scrobe with dorsal margin right-angled, posterior portion of scrobe separated from eye by distance shorter than width of scrobe; head in lateral view with dorsal outline continuously curved towards rostrum (Fig. 10B); anterolateral margin of prothorax without postocular vibrissae (Fig. 10B); elytra with surface of intervals seven to nine strongly concave near base (Fig. 10B); protibiae slightly bent inward at apical fourth, inner margin with three small teeth; lamina of female sternite VIII oval (Fig. 11C).

Isodacrys antrum can be confused with I. obrienorum, I. geminatum, I. brevirostre and I. frontalis. Isodacrys antrum can be readily separated from I. obrienorum and I. geminatum by the concavity near the base of the elytral intervals seven to nine and the protibiae with three small teeth along the inner margin. Neither Isodacrys obrienorum nor I. geminatum present any concavity near the base of the elytra and the inner margins of the protibiae are smooth, without teeth. In I. brevirostre and I. frontalis the concavity near base of the elytral intervals seven to nine is shallower and therefore not as conspicuous as in I. antrum. Additionally, in Isodacrys brevirostre and I. frontalis the inner margins of the protibiae are smooth, without teeth.


Figure 16 Isodacrys fasciatum new species. (A) Dorsal habitus; (B) lateral habitus; (C) ventral habitus; (D) head in anterior view. Full-size DOI: 10.7717/peerj.10191/fig-16


Figure 17 Isodacrys fasciatum new species, female genitalia. (A) Ovipositor in dorsal view; (B) ovipositor in lateral view; (C) sternite VIII in dorsal view; (D) spermatheca.
Full-size DOI: 10.7717/peerj.10191/fig-17

Description - Habitus. $\odot 1.9-2.1 \mathrm{~mm}$ long, $0.8-1.0 \mathrm{~mm}$ wide. Scales polygonal, granulate, mainly overlapping irregularly, from dark brown to cinereous in coloration. Head and rostrum with irregular pattern of brown and cinereous scales; scales surrounding


Figure 18 Isodacrys frontalis new species. (A) Dorsal habitus; (B) lateral habitus; (C) vental habitus; (D) head in anterior view.

Full-size DOI: 10.7717/peerj.10191/fig-18


Figure 19 Isodacrys frontalis new species, female genitalia. (A) Ovipositor in dorsal view; (B) ovipositor in lateral view; (C) sternite VIII in dorsal view; (D) spermatheca.
Full-size DOI: 10.7717/peerj.10191/fig-19
eyes lighter, cinereous. Disc of pronotum with brown scales, with narrow median longitudinal vitta of dark brown scales, conspicuous or not; sides of prothorax with irregular, broad, longitudinal vittae of cinereous scales. Base of elytral intervals three to


Figure 20 Isodacrys kuchii new species. (A) Dorsal habitus; (B) lateral habitus; (C) ventral habitus; (D) head in anterior view.

Full-size DOI: 10.7717/peerj.10191/fig-20
seven with cinereous scales, disc of elytra with irregular pattern of brown and cinereous scales. Setae lanceolate, completely arched, mostly inconspicuous on temple and disc of pronotum. Rostrum. Short, dorsolateral margins of rostrum subparallel, with inconspicuous longitudinal sulci mesad of dorsolateral margins at bent of scrobes (Fig. 10D); dorsal surface flat; median sulcus indistinct. Nasal plate with surface shiny, rugulose; anterior margin medially indented; posterior margin V-shaped, carinate. Epistome in continuous plane with remainder of rostrum, covered by two or three rows of shiny, small scales (Fig. 10D); anterior margin bearing six epistomal setae on each side; posterior margin fuzzy, distinguishable by presence of greater, not modified scales. Scrobe deep, bent, reaching ventral surface of rostrum; dorsal margin right-angled; anterior portion of scrobe about $2 / 3$ length of posterior portion; posterior portion of scrobe separated from eye by distance shorter than width of scrobe (Fig. 10B). Antenna setose, without scales; scape capitate (Fig. 10D); funicular antennomere I clavate, wider and longer than remaining antennomeres; funicular antennomere II clavate, longer than remaining funicular antennomeres; funicular antennomeres III to VII moniliform, distal antennomeres becoming broader. Head. In lateral view with dorsal outline continuously curved to rostrum, frons not prominent(Fig. 10B); eyes moderate in size, suboval, prominent, lateral, separated from outline of frons by $1 / 2-2 / 3$ diameter of eye. Prothorax. Pronotum in dorsal view subcylindrical, as wide as long, lateral outlines sinuate; in lateral view dorsal outline almost straight, anterior and posterior constrictions inconspicuous.


Figure 21 Isodacrys kuchii new species, genitalia. (A) Median lobe and tegmen in dorsal view; (B) median lobe and tegmen in lateral view; (C) male hemisternites VIII and sternite IX in dorsal view; (D) ovipositor in lateral view; (E) spermatheca; (F) ovipositor in dorsal view.

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Full-size DOI: 10.7717/peerj.10191/fig-21
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Anterolateral margin of prothorax without postocular vibrissae or scales projected anteriorly (Fig. 10B). Elytra. 1.9-2.1 times longer than pronotum, at base as wide as base of prothorax. Basal margin of elytra roundly emarginate. Elytra in dorsal view subelliptical, widest at midlength (Fig. 10B), five intervals visible at base. Elytra in lateral view with dorsal outline evenly curved towards summit of apical declivity; apical declivity oblique. Surface of intervals seven to nine strongly concave near base (Fig. 10B). Legs. Procoxae narrowly separated (Fig. 10C), intercoxal process $1 / 7-1 / 6$ width of procoxa; profemora fusiform, subequal in length and width to metafemora; protibiae slightly bent inward at apical fourth, as long as metatibiae, with three small teeth along inner margin. Dorsal surface of tarsi setose, first tarsomere with few appressed scales or not (probably lost by abrasion). Tarsal claws free. Abdomen. Ventrites III, IV and V with anterior margin narrowly sulcate, progressively wider and deeper (Fig. 10C); sulci enclosed by lateral margins of ventrites; posterior margin of sulci obliterated, ventrites becoming flat posteriorly. Genitalia. $\uparrow$ : Lamina of sternite VIII oval (Fig. 11C); spiculum ventrale 2.0-2.2 times length of lamina. Ovipositor with distal gonocoxites slightly sclerotized laterally, styli represented by single thickened seta; two ventral baculi close to each other,


Figure 22 Isodacrys obrienorum new species. (A) Dorsal habitus; (B) lateral habitus; (C) ventral habitus; (D) head in anterior view. Full-size DOI: 10.7717/peerj.10191/fig-22


Figure 23 Isodacrys obrienorum new species, genitalia. (A) Median lobe in dorsal view; (B) median lobe in lateral view; (C) male hemisternites VIII and sternite IX in dorsal view; (D) ovipositor in dorsal view; (E) female sternite VIII in dorsal view; (F) spermatheca; (G) ovipositor in lateral view.

Full-size DOI: 10.7717/peerj.10191/fig-23


Figure 24 Isodacrys okuiltontli new species. (A) Dorsal habitus; (B) lateral habitus; (C) ventral habitus; (D) head in anterior view.

Full-size DOI: 10.7717/peerj.10191/fig-24
separated by distance similar to width of one baculus; vagina with two pairs of lightly sclerotized proximal rods. Spermatheca u-shaped; corpus slightly wide; collum produced, conical; ramus broadly cylindrical, short; cornu strongly curved near corpus, then gently sinuate, elongate, reaching apex of ramus, apically gradually narrowed (Fig. 11D).

Type material. Holotype $\%$ : [MEXICO: Tamaulipas/El Cielo, 3 km SE Alta Cima/ 23.03440-99.186967 $\pm 20 \mathrm{~m} / 860 \mathrm{~m}$, 24.VIII.2009/L.D. Sáenz LSD-370/cloud forest, nest under rock] [WORLD/WEEVIL/DATABASE/WWD0133374] [Isodrusus/or/Isodacrys/ det. R.S. Anderson, 20] (CMNC). Paratypes ( 5 ¢ $\uparrow$ ): [GUAT.: BAJA VERAPAZ/8.6 km. W. Chilascó, 1,560 m./24.V.1991, R. Anderson/oak|pine|liquidambar/forest litter, 91-18] (2, CMNC). [MEXICO: Chiapas/Mpio: Tapalapa/12 Km N Coapilla/12-III-94, R. Jones] [Isodacrys/Det. R.S. Anderson, 1995] (1, CMNC); [MEX.: CHIAPAS, 3 km./W. San Cristobal, 2,000 m./18.IX.1991, R. Anderson/oak|pine forest, 91-108] (1, MZFC); [MEXICO: Tamaulipas/same data as holotype] [WORLD/WEEVIL/DATABASE/ WWD0133381] (1, CMNC).

Etymology. From the Latin noun antrum, meaning cavity, referring to the concavities on intervals seven to nine, stronger and more conspicuous than elytral depressions in other Isodacrys species.


Figure 25 Isodacrys okuiltontli new species, genitalia. (A) Median lobe in dorsal view; (B) median lobe in lateral view; (C) male hemisternites VIII and sternite IX in dorsal view; (D) ovipositor in dorsal view; (E) female sternite VIII in dorsal view; (F) spermatheca; (G) ovipositor in lateral view.

Full-size DOI: 10.7717/peerj.10191/fig-25

Remarks. Adults have been collected in an ant nest under a rock and in leaf litter, in cloud and oak-pine forests, between 860 and $2,000 \mathrm{~m}$ above sea level. Males remain unknown.

Comments. The shape of the lamina of female sternite VIII is uniquely shaped among Isodacrys species.

## Isodacrys apicale Howden, 1961

(Figs. 3B, 4E, 5C, 6B, 8)
Isodacrys apicale Howden, 1961: 91-92. Holotype: Ơ, examined; labeled as [151| 35/Choix, SIN./13-VIII-35] [HOLOTYPE/Isodacrys/apicale/A.T.Howden/No.7385] [CNC/379712]. O’Brien \& Wibmer, 1982: 46 (checklist); Morrone, 1999: 145 (checklist); Cortes-Hernández \& Morrone, 2019: 50.

Diagnosis. Setae of elytra spatulate, incompletely arched, erect on declivity; scrobe with anterior portion as long as posterior portion, posterior portion separated from eye by distance 1.2 times width of scrobe (Fig. 4E); eyes large, subcircular; pronotum as wide as long, anterolateral margin of prothorax with more than six postocular vibrissae, some well-developed (Fig. 4E); elytra in dorsal view obovate, widest before midlength, with seven intervals visible at base, apical margins of elytra bisinuate (Fig. 6B); protibiae with teeth
along inner margin, prolegs not larger than metalegs; median lobe stout, with apex in dorsal view attenuated into acute point.

Isodacrys apicale can be readily separated from I. mexicanum, I. orizabae, I. burkei and I. ovipennis by the prothorax with well-developed postocular vibrissae, the prolegs not larger than the metalegs and apical margins of the elytra bisinuate. In the other mentioned species, the postocular vibrissae are reduced in number and usually reduced in size too (or absent as in I. burkei), the prolegs are larger than the metalegs and the apical margins of the elytra are conjointly rounded, not bisinuate.

Additional material examined. MEXICO: Sinaloa, same data as holotype (1 paratype, CMNC); Sonora, 1 mi . W Álamos, 16.VII.1964, H.R. Burke, J. Apperson (1, TAMUIC).

Remarks. The apex of ventrite V in the male is emarginate, whereas in the females, the apex of ventrite V is rounded (see also Howden, 1961).

Comments. Isodacrys apicale was described from a series of three specimens from Choix, Sinaloa, Mexico. The holotype, deposited in CNC, was examined. One paratype is deposited in CMNC and another paratype is deposited in Dirección General de Defensa Agrícola (SAGARPA, Mexico; Howden, 1961). The examined specimen from Sonora agrees with both the types examined and the original species description.

Photographs of the holotype were examined to complement the description of the male genitalia in diagnosis. Female genitalia were not observed.

## Isodacrys brevirostre Howden, 1961

(Figs. 1, 3A, 4D, 5A, 8)
Isodacrys brevirostre Howden, 1961:79-80. Holotype: $\ddagger$, examined; labeled as [Cuernavaca 6 Mi.,/N., 7500' Morelos/15-VIII-1954/J. G. Chillcott] [HOLOTYPE/Isodacrys/brevirostre/ A.T.Howden/No. 7383] [CNC/379713]. O’Brien \& Wibmer, 1982: 46 (checklist); Morrone, 1999: 145 (checklist); Cortes-Hernández \& Morrone, 2019: 50.

Diagnosis. Setae of elytra spatulate, semierect, becoming erect towards apex of elytra; rostrum short, with longitudinal sulci mesad of dorsolateral margins at bent of scrobes (Fig. 4D); scrobe with anterior portion as long as posterior portion, posterior portion separated from eye by distance equal to width of scrobe; anterolateral margin of prothorax without postocular vibrissae; elytra in dorsal view subelliptical, widest at midlength, with five intervals visible at base, intervals seven to nine slightly depressed at base; prolegs not larger than metalegs, inner margin of protibiae without teeth; spermatheca fishhook-shaped, corpus wide, collum produced, subcylindrical, bent apically towards ramus, ramus inconspicuous.

Isodacrys brevirostre can be confused with other species with non-denticulate protibiae: Isodacrys frontalis, I. geminatum and I. obrienorum. Isodacrys brevirostre can be easily separated from I. frontalis by its spatulate and semierect elytral setae, becoming erect towards the apex of the elytra, the frons not tumescent and posterior portion of the scrobe separated from eye by distance equal to width of the scrobe. In I. frontalis, elytral setae are
lanceolate, from completely arched to incompletely arched, the frons strongly prominent and posterior portion of the scrobe separated from eye by at least twice width of the scrobe. From Isodacrys geminatum and I. obrienorum, I. brevirostre can be distinguished also by the elytral setae, which are lanceolate in I. geminatum and I. obrienorum, and the right-angled scrobe (obtusely angled in I. brevirostre), with posterior portion of the scrobe separated from eye by a distance shorter than the width of the scrobe. Finally, it can be also confused with I. antrum, which possesses teeth along the inner margin of the protibiae and the head in lateral view with dorsal outline continuously curved towards the rostrum. In I. brevirostre inner margin of the protibiae is smooth, without teeth, and the dorsal outline of the head is at least slightly deflexed at the frons towards the rostrum.

Additional material examined. MEXICO: Estado de México, 6 km al SE de Atlautla, 27.IX.2017, Redeo en vegetación, B. Pino-Encino, 2450, $19^{\circ} 00^{\prime} 46^{\prime \prime} \mathrm{N}, 98^{\circ} 45^{\prime} 26^{\prime \prime} \mathrm{W}, \mathrm{K}$. Cortés, L. Delgado (1, IEXA); Estado de México, La Mirasol, 7 km SW Santiago de Tianguistengo, 2.XI.1973, 2,800 m, C.W. O'Brien (36, ASUCOB; 3, CASENT); Estado de México, Ocoyoacac, 28.X.1973, on misc. flowers, C.W. O’Brien (4, CASENT; 89, ASUCOB); Estado de México, Bejucos, Mex. Temascaltepec, 3.VII.1933, H.E. Hinton, R.L. Usinger (3 paratypes, CASENT); Estado de México, Real de Arriba, VII.1932, H. Hinton (1 paratype, TAMUIC); Estado de México, Hwy 190, 11 mi. W Río Frío, 5.VI.1983, 9800', C. \& L. O’Brien, G. Marshall (3, ASUCOB); Estado de México, Hwy15, 28 mi. W. Toluca, 6.VIII.1982, 9100', L. O’Brien, G. Wibmer (38, ASUCOB); Guanajuato, 3.6 mi. NE Guanajuato, 5.VII.1985, Woolley, Zolnerowich (1, TAMUIC); Guerrero, 6 km NE Ayotoxtla (Tlatlauquitepec Rd), 24.X.1973, A.N. García A. (2, ASUCOB); Guerrero, 9 km SE Tlatlauquitepec, 24.X.1974, A.N. García A. (1, ASUCOB); Hidalgo, Hwy105 16 mi. N Metzquititlán, 30.VII.1982, 7200', L. O'Brien, G. Wibmer (23, ASUCOB); Hidalgo, Hwy105 Mineral Real del Monte, 14.VI.1983, 9400', C. \& L. O'Brien, G. Marshall (1, ASUCOB); Hidalgo, Hwy105, 3 mi. S Zacualtipan, 3.VIII.1982, 6959', C. \& L. O’Brien, G. Wibmer (2, ASUCOB); Mexico City, Cañada Contreras, 5.X.1982, s/composit, 2,550 m, K. Luna (10, MZFC); Mexico City, Road from Mexico City P.N. Cumbres de Ajusco, 4.IX.1982, 2, 750 m, C. \& L. O’Brien, G. Wibmer (17, ASUCOB); Mexico City, Delegación Tlalpan, Tlalpuente, 4.XI.1993, L. Torres-Miller (1, CMNC); Mexico City, El Pedregal, 4.IX.1982, 2,340 m, C. \& L. O’Brien, G. Wibmer (1, ASUCOB); Mexico City, Unión Perif. Ajusco, 4.IX.1982, 2,340 m, K. Luna (1, MZFC); Mexico City, 7 mi . N Valle de Bravo, 7.VIII.1982, 8300', C. \& L. O’Brien, G. Wibmer (1, ASUCOB); Michoacán, 2 mi. S Carapan, 6.VII.1986, H. \& A. Howden (1, CMNC); Michoacán, 2 mi. S Carapan, 6.VII.1985, Woolley, Zolnerowich (1, TAMUIC); Michoacán, 28.5 mi. S Nueva Italia, 9.VII.1985, Woolley, Zolnerowich (1, TAMUIC); Michoacán, 17 mi . W Pátzcuaro, 4.VIII.1982, 7700', C. \& L. O'Brien, G. Wibmer (6, ASUCOB); Morelos, 8 km N Cuernavaca, Hwy 95, 5.IX.1982, C. \& L. O’Brien, G. Wibmer (5, ASUCOB); Morelos, Santa María Ahuacatitlán, 6.XI.1980, F. Aguirre (1, CNIN); Morelos, Tetela del Volcán, 23.VIII.1984, V. Butze (1, CNIN); Oaxaca, Hwy 175, 27 km NE Oaxaca, 29.VIII.1982, 8500', C. \& L. O’Brien, G. Wibmer (1, ASUCOB); Oaxaca, Hwy 175, 66 km NE Oaxaca, 29.VIII.1982, 8500', C. \& L. O'Brien, G. Wibmer (1, ASUCOB); Oaxaca, Carr.

Oaxaca-Tuxtepex, bosque de pino-encino, en hojarasca, 2,296 m, $17^{\circ} 20^{\prime} 57.8^{\prime \prime} \mathrm{N}, 96^{\circ} 30^{\prime}$ 57.5" W, 9.VI.2018, M. Barrios (6, MZFC); Oaxaca, 2 mi. north San José Pacífico, 16.VII.1974, Clark, Murray, Ashe, Schaffner (4, CMNC); Oaxaca, 2 miles north San José Pacífico, 17.VII.1974, Clark, Murray, Ashe, Schaffner (2, TAMUIC); Oaxaca, 8 km S Suchixtepec, Río Molino, 19.VI.1979, 2,200 m, H. \& A. Howden (30, CMNC; 2, ASUCOB); Oaxaca 8 km S Suchixtepec, Río Molino, 6.VIII.1986, H. \& A. Howden (1, CMNC); Oaxaca, 10 km S Suchixtepec, 24.VII.1992, Roadside beating, 2,000 m, R.S. Anderson (24, CMNC); Oaxaca, 1 mi. S Suchixtepec Hwy 175, 2.VI.1983, 9200', C. \& L. O’Brien, G. Marshall (2, ASUCOB); Puebla, 67 mi. E Puebla, 26.VI.1971, 7800', C.W. \& L. O’Brien, Marshall (1, CASENT); Puebla, 7 mi. N Zacapoaxtla, 12.VI.1983, 5000', C. \& L. O’Brien, G. Marshall (1, ASUCOB); Puebla, 6 mi. NE Zacatepec, 27.VI.1975, sifting leaf litter, D.S. Chanddler (1, ASUCOB); Puebla, 6 mi. NE Zacatepec, 27.VI.1975, L.E. Watrous (1, ASUCOB); Puebla, Zacatlán km 2 desv. a Piedras Encimadas, 6.VII.2006, 2,449 m, $20^{\circ} 01^{\prime} 07^{\prime \prime} \mathrm{N}, 98^{\circ} 04^{\prime} 43^{\prime \prime}$ W, L. Cervantes, D. Brzoska (1, CNIN); Veracruz, Hwy 140, 17 mi. NW Jalapa, 20.VIII.1982, 8000', C. \& L. O'Brien, G. Wibmer (1, ASUCOB).

Remarks. Adults of I. brevirostre have been found on Condalia sp. (Rhamnaceae; Howden, 1961), on Asteraceae, beating miscellaneous vegetation and in leaf litter, between $\sim 2,000$ and $2,800 \mathrm{~m}$ above sea level. Some were collected on pine-oak forests. Males remain unknown.

Comments. Isodacrys brevirostre was described based on 23 female specimens from several localities in central Mexico: Cuernavaca, Morelos; Toluca and Temascaltepec (misspelled as Temescaltepec), Estado de México; and Jacala, Hidalgo. The holotype, deposited in CNC, was examined. The 22 paratypes were originally placed in CASENT, CNC, Howdens' collection (now housed at the Canadian Museum of Nature), and USNM. Three paratypes from CASENT and one from TAMUIC were also examined. The TAMUIC paratype is probably a subsequent donation from Howden collection.

Among the specimens observed, there are at least three character states that vary intraspecifically: length of the cornu of spermatheca, arrangement of dorsal scales (recorded as polymorphic in character 1) and scales color (from light brown to dark brown; see also Howden, 1961). Regardless of this intraspecific variation, we were not able to establish a clear-cut division among these morphotypes and treat them as different terminals in the phylogenetic analyses.

## Isodacrys buchanani Howden, 1961

(Figs. 2, 6F, 8)
Isodacrys buchanani Howden, 1961: 82-83. Holotype: O', not examined. O’Brien \& Wibmer, 1982: 46 (checklist); Howden, 1993a: 2 (checklist); Morrone, 1999: 145 (checklist); Cortes-Hernández \& Morrone, 2019: 50.

Diagnosis. Setae lanceolate, small, completely arched; scrobe with anterior portion at least one fifth shorter than posterior portion, posterior portion separated from eye by distance equal to width of scrobe; antennal scape covered with scales at apex dorsally; anterolateral
margin of prothorax with one or two postocular vibrissae; basal margin of elytra angularly emarginate, sutural interval at summit of declivity strongly tumid, apical declivity strongly concave; prolegs not larger than metalegs, inner margin of protibiae without teeth; median lobe slender, as long as spiculum gastrale, nearly two times longer than temones, apex in dorsal view acute, manubrium as long as median lobe; spermatheca comma-shaped, corpus slightly subglobose, cornu and ramus inconspicuous.

Isodacrys buchanani can be readily separated from I. schwarzi by the prothorax with one or two reduced postocular vibrissae, which in the latter the postocular vibrissae are well developed, larger and more numerous.

Material examined. MEXICO: Puebla, 6 mi . SE Tehuacán, 7.VII.1973, Mastro, Schaffner (2, TAMUIC; 1, CMNC); Tamaulipas, 4 mi. S San Fernando, Hwy 101, 5.V.1983, C. \& L. O'Brien, G. Marshall (1, ASUCOB). UNITED STATES OF AMERICA: Texas, Duval Co., 1 mi. W San Diego, 10.IV.1973, W.E. Clark (1, TAMUIC); Texas, 15 mi . SW Jct FR 3073 y Hwy 16, 10.IV.1987, B.F. \& J.L. Carr (1, CMNC; 2, CNC); Texas, Jim Hogg Co., 5 mi. W Hebbronville, 3.XI.1990, T. Carlow (3, TAMUIC); Texas, 37 mi . N Laredo, Webb Co., 15.X.1970, C.W. O’Brien (53, ASUCOB; 6, CMNC).

Remarks. The holotype and the paratype were collected from cut flowers (Howden, 1961).
Comments. Isodacrys buchanani was described based on two specimens from Mexico, intercepted at Laredo, Texas, United States. As data in Howden (1961) indicate, since cut flowers are a relatively perishable commodity, it suggests that type locality is somewhere near Laredo in Tamaulipas or Nuevo León, Mexico. The holotype and the paratype are deposited in USNM and were not examined. Identity of material examined was corroborated from specimens identified by Anne Howden and by agreement the with original species description. Material examined confirm the occurrence of I. buchanani in Tamaulipas and also in Texas. Its southernmost occurrence is in Tehuacán, Puebla, México.

## Isodacrys burkei Howden, 1961

(Fig. 8)
Isodacrys burkei Howden, 1961: 88-90. Holotype: O', not examined. O’Brien \& Wibmer, 1982: 46 (checklist); Howden, 1993a: 2 (checklist); Anderson, 2002: 780.

Diagnosis. Setae lanceolate, small, completely arched, mostly inconspicuous on disc of pronotum; epistome with longitudinal carina joined with posterior margin of nasal plate (e.g., Fig. 4C); scrobe with anterior portion at least one fifth shorter than posterior portion, posterior portion separated from eye by distance nearly twice width of scrobe; antennal scape covered with scales at apex dorsally; eyes large, slightly convex, not prominent; anterolateral margin of prothorax without postocular vibrissae, with conspicuous tooth extending anteriorly to eyes; elytra in dorsal view ovate, widest after midlength, intervals two and ten, and intervals five and six tumescent at their apical
junctions, intervals three to five slightly elevated at base; profemora wider than metafemora, protibiae longer than metatibiae, inner margin of protibiae with teeth; median lobe shorter than spiculum gastrale, slightly longer than temones, apex in dorsal view acute, manubrium about as long as median lobe; spermatheca comma-shaped, corpus subglobose, collum produced, subconical, curved towards ramus, ramus broadly subcylindrical, short.

Isodacrys burkei can be readily separated from I. ovipennis, I. orizabae and I. mexicanum by anterolateral margin of the prothorax without postocular vibrissae, instead with a conspicuous tooth extending anteriorly towards the eyes. In I. ovipennis, I. orizabae and I. mexicanum the prothorax bears few postocular vibrissae and the anterolateral tooth of the prothorax is absent.

Material examined. UNITED STATES OF AMERICA: Texas, Brazos Co., 8.V.1956, H. R. Burke (1 paratype, TAMUIC); Texas, Brazos Co., 12.V.1960, H. R. Burke (1 paratype, TAMUIC); Texas, Brazos Co. Bastrop St. Pk., 24-27.V.1963, M. Kaulbars (2, CMNC); Texas, Brazos Co. Bastrop St. Pk., 5.VI.1989, Sweeping composites, R.S. Anderson (6, CMNC); Texas, Brazos Co., College Station, Lick Creek Park, 12.VI.1993, R. Jones (2, UAQE); Texas, College Station, Brazos Co., 15.IV.1963, H.R. Burke (20, CMNC); Texas, College Station, 3.V.1931, swept from weeds, H.J. Reinhard (1 paratype, TAMUIC); Texas, Brazos Co., on Hwy30 7.6 miles east jct. Hwy. 6, 7.V.1981, Sweeping Coreopsis sp., S. J. Merritt (1, TAMUIC); Texas, Brazos Co., Lick Ck. Pk., ca. 3 mi. S College Station, 18.X.1987, R.S. Anderson (4, CASENT; 50, CMNC); Texas, Brazos Co. Lick Creek Park ca. 5 mi. S College Station, 16.IV.1989, Sweeping flowers, R.S. Anderson (2, CMNC); Texas, Lee Co., 29.V.1959, S.D. \& R.H. Burke (1 paratype, TAMUIC); Texas, 0.5 mile west of Minter Springs, Brazos Co., 20.IV.1970, V.V. Board (1, TAMUIC); Texas, Leon Co. 9 km N Flynn, 28.V.1995, Quercus incanus, H. \& A. Howden (5, CMNC); Texas, Milano, Milam Co., 28.IV.1939, \#7315, On tomato plants, very numerous, eating stems particularly, causing plants to fall over (1 paratype, TAMUIC).

Remarks. Adults have been collected on Quercus sp. (Fagaceae), Coreopsis sp. (Asteraceae) and have been reported feeding on tomato plants (Solanaceae) and various herbaceous plants of different families (Fabaceae and Cucurbitaceae). They can be found in pest proportions eating the foliage (Burke, 1959; Howden, 1993a) or the stems.

Comments. Isodacrys burkei was described based on 91 specimens from five counties in Texas, United States of America: Brazos, Gonzales, Lee, Leon and Milam. The holotype, deposited in TAMUIC, was not examined. The paratypes are distributed in TAMUIC, BMNH, FMNH, CNC, CUIC, Dirección General de Defensa Agrícola (SAGARPA, Mexico), KU, USNM, Howden collection (now housed at the Canadian Museum of Nature) and Kissinger's collection. Species identity was corroborated by comparison with paratypes examined from TAMUIC and by agreement with the original species description.

The tumescence of elytral intervals two and ten, and intervals five and six at their apical junction varies in degree of development among individuals (Howden, 1961).

## Isodacrys carlae Cortés-Hernández, new species

(Figs. 4B, 8, 12-13)
urn:lsid:zoobank.org:act:146A1489-52AE-477D-A304-86136B5880BA
Diagnosis. Setae lanceolate, completely arched, mostly inconspicuous on temple; rostrum with median sulcus deep, without foveae at its endings (Fig. 12D); scrobe narrow, anterior portion of scrobe about $1 / 2$ length of posterior portion, posterior portion separated from eye by distance 1.5-1.6 times width of scrobe (Fig. 12B); antennal scape without scales (Fig. 12D) or with one or two scales near apex; anterolateral margin of prothorax without postocular vibrissae (Fig. 12B), pronotum with two narrow median longitudinal vittae of white scales (Fig. 12A); profemora 1.2-1.3 times wider than metafemora, inner margin of protibiae without teeth, dorsal surface of first and second or all tarsomeres with appressed scales; and spermatheca peanut-shaped (Fig. 13D).

Isodacrys carlae can be readily separated from I. kuchii by its narrow scrobe, anterior portion of the scrobe about $1 / 2$ length of posterior portion, and posterior portion separated from eye by distance 1.5-1.6 times width of the scrobe. In Isodacrys kuchii the scrobe is broader, anterior portion of the scrobe about $2 / 3$ length of posterior portion and posterior portion of the scrobe separated from eye by distance equal to width of the scrobe. Isodacrys carlae can be also confused with I. confusum and I. fasciatum which differ from I. carlae by the scale pattern on the pronotum (white scales medially instead of brown).

Description - Habitus. $\ddagger 2.4-3.0 \mathrm{~mm}$ long, $1.0-1.3 \mathrm{~mm}$ wide. Scales polygonal, granulate, mainly overlapping irregularly, from brown to white in coloration. Head from vertex to frons mainly covered by brown scales; rostrum and genae covered with white scales. Disc of pronotum covered with brown scales, with two narrow median longitudinal vittae of white scales, continuing along base of elytral intervals two and three (Fig. 12A); sides of prothorax with white scales, with irregular longitudinal brown vittae between anterior and posterior constrictions of prothorax. Basal tenth of elytral intervals two and three, and intervals seven to ten with white scales, epipleura irregularly covered with brown and white scales, interval 10 almost completely covered with white scales, disc of elytra mostly with brown scales, at apical fifth with white fasciae from interval five towards interval one at summit of apical declivity. Setae lanceolate, completely arched, mostly inconspicuous on temple. Rostrum. Dorsolateral margins of rostrum subparallel (Fig. 4B); dorsal surface flat, becoming slightly depressed near median sulcus; median sulcus deep, short, extending from posterior margin of epistome to near anterior margin of eyes, without foveae (Fig. 12D). Nasal plate with surface shiny; anterior margin medially indented; posterior margin U/V-shaped, carinate. Epistome in continuous plane with remainder of rostrum, covered by three rows of shiny, smaller scales (Fig. 12D); anterior margin bearing four epistomal setae on each side; posterior margin limiting with median sulcus. Scrobe narrow, deep, bent, reaching ventral surface of rostrum; dorsal margin obtusely angled; anterior portion of scrobe about $1 / 2$ length of posterior portion; posterior portion of scrobe separated from eye by distance $1.5-1.6$ times width of scrobe (Fig. 12B). Antenna setose, without scales (Fig. 12D) or with one or two scales near apex of scape; scape capitate;
funicular antennomere I clavate, wider and longer than remaining funicular antennomeres; funicular antennomere II clavate; funicular antennomeres III to VII moniliform, distal antennomeres becoming broader. Head. Head in lateral view with dorsal outline deflexed at frons towards rostrum; eyes moderate in size, subcircular, prominent, lateral, separated from outline of frons by $4 / 5$ diameter of eye. Prothorax. Pronotum in dorsal view subcylindrical, 1.1-1.2 times longer than wide, lateral outlines slightly sinuate (Fig. 12A); in lateral view dorsal outline slightly sinuate (Fig. 12B), anterior and posterior constrictions inconspicuous. Anterolateral margin of prothorax without postocular vibrissae, with row of scales slightly projected anteriorly (Fig. 12B). Elytra. 1.8-2.0 times longer than pronotum, at base as wide as base of prothorax. Basal margin of elytra roundly emarginate. Elytra in dorsal view obovate, widest before midlength (Fig. 12A), five intervals visible at base. Elytra in lateral view with dorsal outline evenly curved towards summit of apical declivity; apical declivity with upper two thirds straight, lower third slightly oblique (Fig. 12B). Elytral intervals with no elevations or depressions. Legs. Procoxae narrowly separated (Fig. 12C), intercoxal process $1 / 5-1 / 4$ width of procoxa; profemora fusiform, subequal in length to metafemora, 1.2-1.3 times wider than metafemora; protibiae straight, as long as metatibiae, inner margin without teeth. Dorsal surface of tarsi setose, first and second or all tarsomeres with appressed scales. Tarsal claws free. Abdomen. Ventrite III with anterior margin narrowly sulcate, sulcus shallow; sulcus of ventrites IV and V progressively wider and deeper (Fig. 12C); sulci enclosed by lateral margins of ventrites; posterior margin of sulci obliterated, ventrites becoming flat posteriorly. Genitalia. ©: Lamina of sternite VIII triangular, apex acute (Fig. 13C); spiculum ventrale $4.5-4.7$ times length of lamina. Ovipositor with distal gonocoxites slightly sclerotized laterally, styli represented by single thickened seta; two ventral baculi close to each other, separated by distance similar to width of one baculus; vagina with two pairs of lightly sclerotized proximal rods (Fig. 13B). Spermatheca peanut-shaped (Fig. 13D); corpus subglobose, narrowed to collum; collum produced, subglobose, funnel-shaped, directed to opposite direction of cornu; ramus broadly tumid, inconspicuous; cornu strongly bent near corpus, then straight, slightly sinuate, apically gradually narrowed.

Type material. Holotype Q : [7000' nr. Jame,/33 mi. S.E. Saltillo,/Coah. Mex. VII.18'63/A. Howden Acacia] [Isodacrys/n.sp.] (CMNC). Paratypes ( 45 ¢ $\uparrow$ ): [ $7000^{\prime} \mathrm{nr}$. Jame, $/ 33 \mathrm{mi}$. S.E. Saltillo,/Coah. Mex. VII.18'63/A. Howden Acacia] (1, CMNC); [7500', nr. Jame,/ 33 mi.S.E.Saltillo,/Coah. Mex. VII.18'63/A. T. Howden] (6, CMNC; 3, MZFC); [7500' nr. Jame,/33 mi. S.E. Saltillo,/Coah. Mex. VII.25'63/A. T. Howden] (7, CMNC; 1, CNIN; 1 IEXA); [20 mi. S.E. Saltillo,/6000', Rt. 015, Coah./Mex., VI.20-21, 1971/H. F. Howden]; [MEXICO: Coahuila/12.4 mi S Saltillo/4-VII-1985, J. Woolley/G. Zolnerowich 85/023] [TAMU-ENTO/X0725914] (1, TAMUIC); [MEXICO: Coahuila/12.4 mi S Saltillo/ 4-VII-1985, J. Woolley/G. Zolnerowich 85/023] [TAMU-ENTO/X0725888] (1, TAMUIC); [MEXICO, Coah., Hwy./57, 15 mi . SE. Saltillo,/7000' 12Sept. 1982 C.\&/L.O’Brien \& G. Wibmer] (1, ASUCOB; 1, MZFC); [MEX., Coah., 10 mi./E. Saltillo 7000 $/$ /VIII-14-1971 C\&L/O’Brien \& Marshall] (1, ASUCOB]; [MEXICO, Hidalgo,/Municipio Metztitlán/1 km
W. San Cristobal,/20.635 N, $98.846^{\circ} \mathrm{W}, / 15-\mathrm{VIII}-2017$, S.M. Clark] (1, ASUCOB); [MEXICO: Nuevo Leon/road to Galeana, 1/km. N jct. rte. 58/17 July 1988/R. Turnbow] (1, CMNC); [MEXICO: Nuevo Leon/9 mi. west Iturbide/July 3, 1974/Clark, Murray,/Ashe, Schaffner] [TAMU-ENTO/X0725413] (1, TAMUIC); [MEXICO: Nuevo Leon/9 mi. west Iturbide/July 3, 1974/Clark, Murray,/Ashe, Schaffner] [TAMU-ENTO/X0724919] (1, TAMUIC); [MEXICO: Nuevo Leon/9 mi. west Iturbide/July 3, 1974/Clark, Murray,/ Ashe, Schaffner] [TAMU-ENTO/X0725718] (1, IEXA); [MEXICO: Nuevo Leon/9 mi. west Iturbide/July 3, 1974/Clark, Murray,/Ashe, Schaffner] [TAMU-ENTO/X0726871] (1, TAMUIC); [MEXICO: Nuevo Leon/9 mi. west Iturbide/July 3, 1974/Clark, Murray,/ Ashe, Schaffner] [TAMU-ENTO/X0730294] (1, TAMUIC); [MEXICO: Nuevo Leon/9 mi. west Iturbide/July 3, 1974/Clark, Murray,/Ashe, Schaffner] [TAMU-ENTO/X0726000] (1, CNIN); [MEX. N.L. $40 \mathrm{mi} . / \mathrm{NW}$ Jcn. Hwy 57\&60/VI-22-1971 C.W./O’Brien \& Marshall] (1, ASUCOB; 1, MZFC); [MEXICO, N.L., Hwy./58, 32 mi. W. Linares,/6000' 11Sept. 1982 C. \&/L. O'Brien \& G. Wibmer] (3, ASUCOB); [MEXICO, N.L., 11 km./NE. S. Antonio Peña/Nevada, 2,000 m. Aug16/1977A.N.Garcia A.] [beating pine and/miscellaneous] (1, ASUCOB); [MEX., N.L., 29 mi./SW. Linares 5700'/VIII-15-1971 C\&L/O’Brien \& Marshall] (1, ASUCOB); [MEXICO: San Luis Potosi/4 km. W Guadalcazar/18 July 1988/ R. Turnbow] [Isodacrys/geminatum Howd./det. R.S. Anderson 90] (1, CMNC); [MEXICO: Tamaulipas/19.6 mi. sw. Jaumave/18-VII-1973/Gaumer and Clark] [TAMU-ENTO/ X0725801] [Isodacrys/sp./DET./A. Howden] (1, TAMUIC); [MEXICO: Tamaulipas/19.6 mi. sw. Jaumave/18-VII-1973/Gaumer and Clark] [TAMU-ENTO/X0727602] (1, TAMUIC); [MEXICO: Tamaulipas/19.6 mi. sw. Jaumave/18-VII-1973/Gaumer and Clark] [TAMU-ENTO/X0725553] (1, TAMUIC); [MEXICO, Tam., Hwy./101, 28 mi. NE. Tula/ 4500' 22July1982 CW\&/L. O’Brien \& G. Wibmer] (1, ASUCOB).

Etymology. This species is named after the first author's beloved mother, Carla Angélica Hernández-Olvera, for all her support and endless kindness.

Remarks. Adults have been collected on Acacia (Fabaceae) and beating miscellaneous vegetation between $\sim 1,370$ and $2,130 \mathrm{~m}$ above sea level. Males remain unknown.

Comments. This species had been previously recognized as an undescribed Isodacrys species by Anne Howden (as labels on the herein designated holotype indicate) and subsequently (perhaps following Howden) by Charles W. O'Brien in 1999 according to revised material at ASUCOB.

## Isodacrys confusum Cortés-Hernández, new species

(Figs. 7A, 8, 14-15)
urn:lsid:zoobank.org:act:3EDF5293-38FD-4217-8730-164074806BAA
Diagnosis. Setae lanceolate, completely arched; rostrum with median sulcus short, without foveae at its endings; scrobe deep, anterior portion of scrobe about $2 / 3$ length of posterior portion, posterior portion of scrobe separated from eye by distance 1.4-1.6 times width of scrobe; antennal scape covered with scales at apex dorsally (Fig. 14B); anterolateral margin of prothorax without postocular vibrissae (Fig. 14B), pronotum covered with white
scales medially, with two broad, lateral longitudinal vittae of brown scales (Fig. 14A); profemora 1.2-1.3 times wider than metafemora, inner margin of protibiae without teeth, all tarsomeres with appressed scales (Fig. 14D); male terminalia with manubrium longer than median lobe (Fig. 15B); spermatheca comma-shaped, collum produced, bent basally to ramus, contiguous with corpus (Fig. 15F).

Isodacrys confusum can be easily confused with I. carlae, I. kuchii or I. fasciatum, but it can be externally separated from I. carlae and I. kuchii by its scale pattern on the prothorax, which is covered with white scales medially (covered with brown scales medially instead of white in I. carlae and I. kuchii) and by the scape covered with scales dorsally (always lacking in I. kuchii and generally lacking in I. carlae). Finally, I. confusum can be separated from Isodacrys fasciatum by the scape covered with scales and by the spermatheca with collum directed towards ramus, contiguous to corpus. The scape is not covered with scales and the ramus is not contiguous to the corpus in I. fasciatum.

Description - Habitus. ơ $1.7-2.5 \mathrm{~mm}$ long, $0.7-1.1 \mathrm{~mm}$ wide; $\uparrow 2.4-2.6 \mathrm{~mm}$ long, $1.1-1.2 \mathrm{~mm}$ wide. Scales polygonal, granulate, mainly overlapping irregularly, from brown to white in coloration. Head and rostrum mainly covered by white scales, with irregular patches of brown scales on vertex and frons. Disc of pronotum covered with white scales medially, with two broad, lateral longitudinal vittae of brown scales (Fig. 14A); sides of prothorax with white scales, with irregular longitudinal brown vittae between anterior and posterior constrictions of prothorax. Disc of elytra irregularly with white and brown scales, intervals six to four at second fourth with brown fasciae, apical third with brown fascia from interval six towards interval one at summit of apical declivity; epipleura mostly with white scales. Setae lanceolate, completely arched. Rostrum. Dorsolateral margins of rostrum subparallel; dorsal surface flat, becoming slightly depressed near median sulcus; median sulcus short, extending from posterior margin of epistome to near anterior margin of eyes, without foveae (Fig. 14D). Nasal plate with surface shiny; anterior margin medially indented; posterior margin V-shaped, carinate. Epistome in continuous plane with remainder of rostrum, covered by three rows of shiny, smaller scales (Fig. 14D); anterior margin bearing four or five epistomal setae on each side; posterior margin limiting with median sulcus. Scrobe deep, bent, reaching ventral surface of rostrum; dorsal margin obtusely angled; anterior portion of scrobe about $2 / 3$ length of posterior portion; posterior portion of scrobe separated from eye by distance 1.4-1.6 times width of scrobe. Antenna setose, apex of scape covered with scales (Fig. 14B); scape capitate; funicular antennomere I clavate, wider and longer than remaining funicular antennomeres; funicular antennomere II clavate, longer than remaining funicular antennomeres; funicular antennomeres III-VII moniliform, distal antennomeres becoming broader. Head. Head in lateral view with dorsal outline deflexed at frons towards rostrum; eyes moderate in size, subcircular, prominent, lateral, separated from dorsal outline of frons by distance similar to diameter of eye. Prothorax. Pronotum in dorsal view subcylindrical, nearly as wide as long, lateral outlines slightly sinuate; in lateral view dorsal outline slightly sinuate, anterior and posterior constrictions inconspicuous. Anterolateral margin of prothorax without postocular vibrissae (Fig. 14B), with row of scales slightly projected anteriorly.

Elytra. 1.8-2.0 times longer than pronotum, at base as wide as base of prothorax. Basal margin of elytra roundly emarginate. Elytra in dorsal view obovate, widest before midlength (Fig. 14A), five intervals visible at base. Elytra in lateral view with dorsal outline gently evenly curved towards summit of apical declivity; apical declivity with upper two thirds straight, lower third slightly oblique. Elytral intervals with no elevations or depressions. Legs. Procoxae narrowly separated (Fig. 14C), intercoxal process 1/6-1/4 width of procoxa; profemora fusiform, subequal in length to metafemora, 1.2-1.3 times wider than metafemora; protibiae straight, as long as metatibiae, inner margin without teeth. Dorsal surface of tarsi setose, all tarsomeres with appressed scales (Fig. 14D). Tarsal claws free. Abdomen. Ventrite III with anterior margin narrowly sulcate, sulcus shallow; sulcus of ventrites IV and V progressively wider and deeper (Fig. 14C); sulci enclosed by lateral margins of ventrites; posterior margin of sulci obliterated, ventrites becoming flat posteriorly. Genitalia. © : hemisternites VIII teardrop-shaped, outer corner not truncate (Fig. 15C); basal plate of sternite IX lightly sclerotized, distally bifurcate, tips of bifurcations sclerotized; spiculum gastrale as long as median lobe, distally continuing through basal plate; ring of tegmen without parameres; manubrium 1.5 times longer than median lobe (Fig. 15B); temones $4 / 5$ length of median lobe; median lobe 12 times longer than wide in dorsal view, evenly curved in lateral view (Fig. 15B), apical opening elliptical, apex sharply acute. ọ: Lamina of sternite VIII triangular, apex acute (Fig. 15E); spiculum ventrale 4.3-4.6 times length of lamina. Ovipositor with distal gonocoxites slightly sclerotized laterally, styli represented by single thickened seta; two ventral baculi close to each other, separated by distance similar to width of one baculus; vagina with two pairs of lightly sclerotized proximal rods (Fig. 15G). Spermatheca comma-shaped; corpus slightly subglobose, narrowing towards cornu; collum produced, subcylindrical, bent basally to ramus, contiguous with corpus; ramus from indistinct to slightly tumid; cornu strongly curved near corpus, then evenly curved, elongate, reaching apex of ramus, apically gradually narrowed (Fig. 15F). Sexual dimorphism. Ventrites I and II slightly concave mesally in males, convex in females; apex of ventrite V slightly truncate in males, rounded in females. Males are usually distinctively smaller and thinner than females.

Type material. Holotype Ơ: [MEXICO: Tamaulipas/16.3 mi. ne. Jaumave/18-VII.1973/ Gaumer and Clark] [taken on/Mimosa/monancistra] [TAMU-ENTO/X0729035] (TAMUIC); paratypes ( 8 ƠƠ, 28 ¢ ¢ ): [MEXICO, Tam., Hwy 101/18 mi. SW. Cd. Victoria,/4300' 22July1982 C.W.\&/L.O’Brien \& G. Wibmer] ( 1 Ơ, 1 \& $\uparrow$, ASUCOB; 1 Ơ, MZFC); [MEXICO, Tam., Hwy 101/24 mi. SW. Cd. Victoria,/2150' 23July1982 C.W.\&/L. O’Brien \& G. Wibmer] ( 1 Ơ, 2 \& , ASUCOB); [MEXICO, Tam., Hwy 101/22 mi. SW. Cd. Victoria,/3000' 22July1982 C.W.\&/L. O’Brien \& G. Wibmer] (1 Ơ, 1 \&, ASUCOB); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0726618] (1 \&, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0727115] (1 \&, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0725169] (1 \&, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0724605] (1 \&, TAMUIC);
[MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0725410] (1 ¢, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0726220] (1 \& , TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0726502] (1 \&, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0727010] (1 Ó, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0726280] (1 \&, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0725437] (1 \&, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0725446] (1 \&, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0725407] (1 \& , TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0725141] (1 \&, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0725168] (1 \&, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0725412] (1 \&, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0726973] ( 1 O , IEXA); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0725112] (1 \& , IEXA); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0725393] (1 ○, CNIN); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/ X0729284] ( 1 \&, CNIN); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0728579] (1 Ơ, TAMUIC); [MEXICO: Tamaulipas/ same data as holotype] [taken on/same data as holotype] [Isodacrys/sp./DET./A. Howden] [TAMU-ENTO/X0728911] (1 O, MZFC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0728902] [ơ] (1 Ó, MZFC); [MEXICO: Tamaulipas/same data as holotype] [TAMU-ENTO/X0726515]; [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [Isodacrys/ geminatum/Howden/det. R.S. Anderson 1981] (1 \&, CMNC); [MEXICO: Tamaulipas/ same data as holotype] [taken on/same data as holotype] ( $10^{\circ}, 3 甲 9$, CMNC).

Etymology. This species is named with the Latin word confusum meaning confused, because of its external resemblance with several species of Isodacrys.

Remarks. Most of the type series was collected on Mimosa monancistra Benth. (Fabaceae). Adults have been collected between $\sim 650$ and $1,220 \mathrm{~m}$ above sea level.

## Isodacrys crispum Howden, 1961

(Figs. 1 and 8)
Isodacrys crispum Howden, 1961: 84-85. Holotype: $\uparrow$, not examined. O’Brien \& Wibmer, 1982: 46 (checklist); Morrone, 1999: 145 (checklist); Cortes-Hernández \& Morrone, 2019: 50.

Diagnosis. Scales with carinate margins; setae lanceolate, completely arched; rostrum short; scrobe right-angled, anterior portion shorter than posterior portion, posterior portion separated from eye by distance equal to width of scrobe; anterolateral margin of prothorax with one to three vestigial postocular vibrissae; sulci of ventrites III, IV and V with posterior margin sharp, conspicuously delimiting sulci; prolegs not larger than metalegs, inner margin of protibiae with teeth; spermatheca comma-shaped, corpus subglobose, cornu inconspicuous, ramus slightly tumid.

Isodacrys crispum can be readily separated from similar Isodacrys species by its scales with carinate margins, short rostrum, the prothorax with only one to three vestigial postocular vibrissae, sulci of ventrites III, IV and V with posterior margin sharp, conspicuously delimiting sulci and the protibiae with teeth along inner margin. Although Isodacrys crispum can be superficially confused with Isodrusus debilis, it is easily separated by the absence of humeri and the tarsal claws free. Isodrusus debilis has well-developed humeri and tarsal claws connate.

Material examined. MEXICO: Durango, 25 mi. W Durango, 10.VI.1964, Oak, H. Howden (5, CMNC); Durango, 25 mi. W Durango, 29.VI.1964, H. Howden (30, CMNC; 2 ASUCOB); Durango, 25 mi. W Durango, 3.VII.1964, H. \& A. Howden (3, CNC); Durango, 28 mi. W Durango, 9.VI.1967, Quercus clivicola F. consanguinea, C.H. Maen (1, CMNC); Durango, 25 mi. W Durango, 18.VIII.1974, on Quercus, 7800', C.W. \& L. O’Brien, Marshall (2, UAQE); Durango, 25 mi. W Durango, 18.VIII.1974, on pine/on Quercus, 7800 ', C.W. \& L. O’Brien, Marshall (34, ASUCOB); Durango, 26.5 mi . SW Durango, 21. VII.1982, General sweeping, Fred. G. Andrews (9, CMNC); Zacatecas, 27 mi NW Fresnillo, Hwy 45, 17.VIII.1974, on Quercus, 7800', C.W. \& L. O'Brien, Marshall (32, ASUCOB).

Remarks. Some adults have been collected on Quercus sp. (Fagaceae) and Pinus sp. (Pinaceae), at $\sim 2,370 \mathrm{~m}$ above sea level. Isodacrys crispum has a strong superficial resemblance to Isodrusus debilis (Howden, 1961). Males remain unknown.

Comments. Isodacrys crispum was described from only one specimen from Palos Colorados, Durango, Mexico. The holotype, deposited in AMNH, was not examined. Identity of material examined was corroborated from specimens identified by Anne Howden and by agreement with the original species description.

## Isodacrys ellipticum Howden, 1961

(Fig. 8)
Isodacrys ellipticum Howden, 1961: 86-87. Holotype: $\uparrow$, not examined. O’Brien \& Wibmer, 1982: 46 (checklist); Morrone, 1999: 145 (checklist).

Diagnosis. Setae of elytra spatulate, long, from semierect to erect; epistome with longitudinal carina joined with posterior margin of nasal plate (e.g., Fig. 4C); scrobe with anterior portion as long as posterior portion; anterolateral margin of prothorax without postocular vibrissae; elytra in dorsal view subelliptical, widest at midlength, with seven intervals visible at base; profemora wider than metafemora, protibiae longer than
metatibiae, inner margin of protibiae with teeth; and spermatheca $u$-shaped, corpus subcylindrical, slightly swollen, collum inconspicuous, ramus very short, broadly cylindrical.

Isodacrys ellipticum can be separated from I. guatemalenum by the spatulate shape of its elytral setae, from semierect to erect and spermatheca with collum inconspicuous, ramus very short. In I. guatemalenum the elytral setae are parallel-sided, usually longer and erect, and in the spermatheca both the collum and the ramus are inconspicuous.

Material examined. GUATEMALA: Zacapa, San Lorenzo Quarry Road, 3-7 km N Sta. Cruz, CA9, 17.VII.2008, Dry forest, C. \& L. O’Brien, F. Skillman (1, ASUCOB). HONDURAS: Comayagua, Comayagua, 2.VI.1978, Gary V. Manley (1, CMNC); Cortés, 5 km N Cofradía, 5.VIII.1977, C. \& L. O’Brien, G. Marshall (41, ASUCOB); Francisco Morazán, Distrito Central, 5 mi. S Tegucigalpa, 9.VI.1974, night, 4000', C. \& L. O'Brien, G. Marshall (3, CASENT); Francisco Morazán, 17 mi. NE Talanga, 16.VI.1974, 2900', C. \& L. O’Brien, G. Marshall (129, ASUCOB; 3, CASENT); Francisco Morazán, 23.8 km SW Talanga, 3.VI.1993, R. Turnbow (1, CMNC); Francisco Morazán, Tamara valley, 5.X.1993, R. Turnbow (1, CMNC).

Remarks. The holotype was collected in flowers of Lantana sp. (Verbenaceae; see Howden, 1961). Adults have been collected between $\sim 880$ and $1,200 \mathrm{~m}$ above sea level. Males remain unknown.

Comments. Isodacrys ellipticum was described from one specimen from Honduras. The holotype, deposited in USNM, was not examined. Identity of material examined was corroborated from specimens identified by Anne Howden and by agreement with the original species description.

## Isodacrys fasciatum Cortés-Hernández, new species

(Figs. 2, 3C, 5E, 9, 16-17)
urn:lsid:zoobank.org:act:881D4538-9CB8-49E5-9641-CCF3973623E1
Diagnosis. Setae lanceolate, completely arched; rostrum with median sulcus short, without foveae at its endings (Fig. 16D); posterior portion of scrobe separated from eye by distance 1.5-2.1 times width of scrobe; antennal scape without scales (Fig. 16B); anterolateral margin of prothorax without postocular vibrissae (Fig. 16B), pronotum covered with white scales medially, with two broad, lateral longitudinal vittae of brown scales (Fig. 16A); profemora 1.2 times wider than metafemora, inner margin of protibiae without teeth, all tarsomeres with appressed scales; and spermatheca comma-shaped with collum not contiguous with corpus (Fig. 17D).

This species is very similar to Isodacrys confusum, nonetheless, in I. fasciatum the banded scale pattern along the elytra is more conspicuous. Furthermore, I. fasciatum does not present scales at the apex of the scape, and the collum of the spermatheca is not contiguous with the corpus like in I. confusum.

Description - Habitus. ㅇ $^{2}$ 2.1-2.9 mm long, $1.0-1.3 \mathrm{~mm}$ wide. Scales polygonal, granulate, mainly overlapping irregularly, from brown to white in coloration. Head and rostrum mainly covered by white scales, with irregular patches of brown scales from vertex to frons. Disc of pronotum covered with white scales medially, with two broad, lateral longitudinal vittae of brown scales (Fig. 16A); sides of prothorax with white scales, often with small, irregular patches of light brown scales. Disc of elytra mainly covered with white scales, interval six at basal third with brown scales, then forming a brown fascia with intervals five and four at second third, apical third with brown fascia from interval six to interval one at summit of apical declivity; epipleura mostly with white scales. Setae lanceolate, completely arched. Rostrum. Dorsolateral margins of rostrum subparallel; dorsal surface flat, becoming slightly depressed near median sulcus; median sulcus deep, short, from posterior margin of epistome to near anterior margin of eyes, without foveae (Fig. 16D). Nasal plate with surface shiny; anterior margin medially indented; posterior margin V-shaped, carinate. Epistome in continuous plane with remainder of rostrum, covered by three rows of shiny, smaller scales (Fig. 16D); anterior margin bearing four epistomal setae on each side; posterior margin limiting with median sulcus. Scrobe deep, bent, reaching ventral surface of rostrum; dorsal margin obtusely angled; anterior portion of scrobe about $1 / 2$ length of posterior portion; posterior portion of scrobe separated from eye by distance 1.5-2.1 times width of scrobe. Antenna setose, without scales (Fig. 16B); scape capitate; funicular antennomere I clavate, wider and longer than remaining antennomeres; funicular antennomere II clavate, longer than remaining funicular antennomeres; funicular antennomeres III-VII moniliform, distal antennomeres becoming broader. Head. Head in lateral view with dorsal outline deflexed at frons towards rostrum; eyes moderate in size, subcircular, prominent, lateral, separated from outline of frons by about $4 / 5$ diameter of eye. Prothorax. Pronotum in dorsal view subcylindrical, as wide as long, lateral outlines sinuate (Fig. 16A); in lateral view dorsal outline slightly sinuate, anterior and posterior constrictions inconspicuous. Anterolateral margin of prothorax without postocular vibrissae (Fig. 16B), with row of scales slightly projected anteriorly. Elytra. 2.0-2.2 times longer than pronotum, at base as wide as base of prothorax. Basal margin of elytra roundly emarginate. Elytra in dorsal view obovate, widest before midlength, five intervals visible at base (Fig. 5E). Elytra in lateral view with dorsal outline gently evenly curved towards summit of apical declivity; apical declivity with upper two thirds straight, lower third slightly oblique. Elytral intervals with no elevations or depressions. Legs. Procoxae narrowly separated, intercoxal process $1 / 6-1 / 4$ width of procoxa; profemora fusiform, subequal in length to metafemora, 1.2 times wider than metafemora; protibiae straight, as long as metatibiae, inner margin without teeth. Dorsal surface of tarsi setose, all tarsomeres with appressed scales. Tarsal claws free. Abdomen. Ventrite III with anterior margin narrowly sulcate, sulcus shallow; sulcus of ventrites IV and V progressively wider and deeper (Fig. 16C); sulci enclosed by lateral margins of ventrites; posterior margin of sulci obliterated, ventrites becoming flat posteriorly. Genitalia. Q̣: Lamina of sternite VIII triangular, apex acute (Fig. 17C); spiculum ventrale 4.0-4.2 times length of lamina. Ovipositor with distal gonocoxites slightly sclerotized laterally, styli represented by single thickened seta; two ventral baculi
close to each other, separated by distance similar to width of one baculus; vagina with two pairs of lightly sclerotized proximal rods (Fig. 17B). Spermatheca comma-shaped; corpus slightly subglobose, narrowing towards cornu; collum produced, subcylindrical, bent basally to ramus, not contiguous with corpus; ramus slightly tumid; cornu strongly curved near corpus, then evenly curved, elongate, reaching apex of ramus, apically gradually narrowed (Fig. 17D).

Type material. Holotype $\%$ : [MEX., Coah., 10 mi./E. Saltillo 7000'/VIII-14-1971 C\&L/O'Brien \& Marshall] (ASUCOB). Paratypes (11 ¢ ¢): [MEX., Coah., $10 \mathrm{mi} . / \mathrm{same}$ data as holotype] (2, ASUCOB); [MEXICO: Coahuila/12.4 mi S Saltillo/4-VII-1985, J. Woolley/G. Zolnerowich 83/023] [TAMU-ENTO/X0727876] (1, TAMUIC); [MEXICO, Dgo., 22 mi ./NE. Durango, 6200'/Aug. 20, 1974 C.W. \& L./O’Brien \& Marshall] (4, ASUCOB; 2, MZFC; 1, CMNC); [MEX. N.L. 10 mi//S. Jcn. 57 \& 60/VI-231971 L\&CW/O’Brien \& Marshall] (1, ASUCOB).

Etymology. Named with the Latin word fascia, referring to the white/brown banded scale pattern visible along the elytra.

Remarks. Adults have been collected between $\sim 1,890$ and $2,130 \mathrm{~m}$ above sea level. Males remain unknown.

## Isodacrys frontalis Cortés-Hernández, new species

(Figs. 2, 3F, 6D, 9, 18-19)
urn:lsid:zoobank.org:act:DB24A34B-7EF1-44AD-8DFD-5823FC083899
Diagnosis. Setae lanceolate, completely arched to incompletely arched; rostrum with inconspicuous longitudinal sulci mesad of dorsolateral margins, at bent of scrobes; posterior portion of scrobe separated from eye by distance at least twice width of scrobe (Fig. 18B); eyes separated from dorsal outline of frons by distance at least equal to diameter of eye; frons strongly prominent (Fig. 18B); anterolateral margin of prothorax without postocular vibrissae; elytra in dorsal view with five intervals visible at base, intervals seven to nine slightly depressed at base, interval five slightly elevated at base and at fasciae; inner margin of protibiae without teeth.

Isodacrys frontalis and I. brevirostre have similar scale patterns but the first never has the rostrum as short as I. brevirostre. Isodacrys frontalis also has the frons very prominent. The shape and disposition of setae are also different: I. frontalis has lanceolate, completely arched setae, while I. brevirostre presents spatulate setae, from semierect to erect on elytra.

Description - Habitus. $\ddagger 2.2-3.0 \mathrm{~mm}$ long, $1.0-1.3 \mathrm{~mm}$ wide. Scales polygonal, granulate, mainly overlapping irregularly, from light brown to brown in coloration. Head and rostrum covered with brown scales, with inconspicuous longitudinal vitta of darker scales from occiput to between eyes; scales surrounding eyes, fovea between eyes and sides of rostrum lighter, with metallic reflections. Disc of pronotum with brown scales, with lateral longitudinal vittae of pale scales continuing along base of elytral intervals five and six; sides of prothorax with brown scales. Elytral interval five at basal two fifths with irregular
spot of pale scales, disc of elytra mostly with brown scales, at apical $4 / 5$ with two fasciae from interval five towards interval one at summit of apical declivity, anteriormost fascia constituted by dark brown scales, posteriormost fascia by pale brown scales (Fig. 18A). Setae lanceolate, completely arched to incompletely arched, mostly inconspicuous on temple and disc of pronotum. Rostrum. Dorsolateral margins of rostrum subparallel, with inconspicuous longitudinal sulci mesad of dorsolateral margins at bent of scrobes; dorsal surface flat, becoming slightly depressed near median sulcus; median sulcus conspicuous or not, both endings marked by foveae (Fig. 18D). Nasal plate with surface shiny, rugulose; anterior margin medially indented; posterior margin V -shaped, carinate. Epistome in continuous plane with remainder of rostrum, slightly depressed, covered by two or three rows of shiny, small scales (Fig. 18D); anterior margin bearing six epistomal setae on each side; posterior margin limiting with anterior fovea of median sulcus (Fig. 18D). Scrobe deep, bent, reaching ventral surface of rostrum; dorsal margin obtusely angled; anterior portion of scrobe about as long as posterior portion; posterior portion of scrobe separated from eye by distance at least twice width of scrobe (Fig. 18B). Antenna setose, without scales; scape capitate; funicular antennomere I clavate, wider and longer than remaining funicular antennomeres; funicular antennomere II clavate, longer than remaining funicular antennomeres; funicular antennomeres III to VII moniliform, distal antennomeres becoming broader (Fig. 18D). Head. Head in lateral view with dorsal outline deflexed at frons towards rostrum, frons strongly prominent (Fig. 18B); eyes moderate in size, suboval, prominent, lateral, separated from outline of frons by distance at least equal to diameter of eye. Prothorax. Pronotum in dorsal view subcylindrical, nearly as wide as long, lateral outlines slightly sinuate; in lateral view dorsal outline almost straight, anterior and posterior constrictions inconspicuous. Anterolateral margin of prothorax without postocular vibrissae or scales projected anteriorly. Elytra. 2.2-2.4 times longer than pronotum, at base as wide as base of prothorax. Basal margin of elytra roundly emarginate. Elytra in dorsal view subelliptical, widest at midlength, five intervals visible at base. Elytra in lateral view with dorsal outline evenly curved towards summit of apical declivity; apical declivity with upper half straight, lower half oblique (Fig. 18B). Elytral interval five slightly elevated at base and at fasciae; intervals seven to nine slightly depressed at base. Legs. Procoxae narrowly separated (Fig. 18C), intercoxal process $1 / 8-1 / 5$ width of procoxa; profemora fusiform, subequal in length and width to metafemora; protibiae straight, as long as metatibiae, inner margin without teeth. Dorsal surface of tarsi setose, first and second tarsomeres with appressed scales or not (probably lost by abrasion). Tarsal claws free, brown at base, becoming black apically. Abdomen. Ventrite III with anterior margin narrowly sulcate, sulcus shallow; sulcus of ventrites IV and V progressively wider and deeper (Fig. 18C); sulci enclosed by lateral margins of ventrites; posterior margin of sulci obliterated, ventrites becoming flat posteriorly. Genitalia. Q̣: Lamina of sternite VIII triangular, apex acute (Fig. 19C); spiculum ventrale 5.5-5.6 times length of lamina. Ovipositor with distal gonocoxites slightly sclerotized laterally, styli represented by single thickened seta; two ventral baculi close to each other, separated by distance similar to width of one baculus; vagina with two pairs of lightly sclerotized proximal rods. Spermatheca fishhook-shaped; corpus wide;
collum produced, conical, bent apically to ramus; ramus broadly cylindrical, short; cornu evenly curved, short, not reaching apex of ramus, apically gradually narrowed (Fig. 19D).

Type material. Holotype $\uparrow$ : [MEX.: OAX.; 20 mi S/Juchatengo, $6000^{\prime} / 29 . V .1971$, S. Peck/Ber207, oak litter] (CMNC). Paratypes (23 ¢): [GUATEMALA: Sacatepéquez/ 5 km SE Antigua/14.53495-90.69367 $\pm 50 \mathrm{~m} / 2,150 \mathrm{~m}$, 10.VI.2009/LLAMA Wa-B-08-1-49] [WORLD/WEEVIL/DATABASE/WWD0041418] (1 CMNC); [GUATEMALA: Sacatepéquez/5 km SE Antigua/ $14.53495-90.69367 \pm 50 \mathrm{~m} / 2,150 \mathrm{~m}$, 10.VI.2009/ LLAMA Wa-B-08-1-49] [WORLD/WEEVIL/DATABASE/WWD0041419] (1, CMNC); [GUATEMALA: Sacatepéquez/5 km SE Antigua/14.53495-90.69367 $\pm 50 \mathrm{~m} / 2,150 \mathrm{~m}$, 10.VI.2009/LLAMA Wa-B-08-1-49] [WORLD/WEEVIL/DATABASE/WWD0041420] (1, CMNC); [GUATEMALA: Sacatepéquez/5 km SE Antigua/14.53510-90.69384 $\pm$ $50 \mathrm{~m} / 2,150 \mathrm{~m}, 10 . \mathrm{VI} .2009 / L L A M A$ Wa-B-08-1-44] [WORLD/WEEVIL/DATABASE/ WWD0041311] [Isodacrys/GUA 1/det. R.S. Anderson, 20] (1, CMNC); [GUATEMALA: Sacatepéquez/5 km SE Antigua/14.53529-90.69404 $\pm 50 \mathrm{~m} / 2,150 \mathrm{~m}, 10$.VI.2009/LLAMA Wa-B-08-1-38] [WORLD/WEEVIL/DATABASE/WWD0041144] (1, CMNC); [GUATEMALA: Sacatepéquez/5 km SE Antigua/14.53504-90.69377 $\pm 50 \mathrm{~m} / 2,150 \mathrm{~m}$, 10.VI.2009/LLAMA Wa-B-08-1-46] [WORLD/WEEVIL/DATABASE/WWD0041358] (1, CMNC); [GUATEMALA: Sacatepéquez/5 km SE Antigua/14.53504-90.69377 $\pm$ $50 \mathrm{~m} / 21,50 \mathrm{~m}, 10 . \mathrm{VI} .2009 / L L A M A$ Wa-B-08-1-46] [WORLD/WEEVIL/DATABASE/ WWD0041359] (1, CMNC); [GUATEMALA: Sacatepéquez/5 km SE Antigua/ 14.53504-90.69377 $\pm 50 \mathrm{~m} / 2,150 \mathrm{~m}, 10 . \mathrm{VI} .2009 / L L A M A$ Wa-B-08-1-46] [WORLD/ WEEVIL/DATABASE/WWD0041360] (1, CMNC); [GUATEMALA: Sacatepéquez/5 km SE Antigua/14.53504-90.69377 $\pm 50 \mathrm{~m} / 2,150 \mathrm{~m}, 10$.VI.2009/LLAMA Wa-B-08-1-46] [WORLD/WEEVIL/DATABASE/WWD0041361] (1, CMNC); [GUATEMALA: Sacatepéquez/5 km SE Antigua/14.53504-90.69377 $\pm 50 \mathrm{~m} / 2,150 \mathrm{~m}$, 10.VI.2009/ LLAMA Wa-B-08-1-46] [WORLD/WEEVIL/DATABASE/WWD0041363] (1, TAMUIC); [GUATEMALA: Sacatepéquez/5 km SE Antigua/14.53507-90.69380 $\pm 50 \mathrm{~m} / 2,150 \mathrm{~m}$, 10.VI.2009/LLAMA Wa-B-08-1-45] [WORLD/WEEVIL/DATABASE/WWD0041326] (1, ASUCOB); [GUATEMALA: SACATEPEQUEZ/Guatemala City, Cerro/Alux, $2,260 \mathrm{~m}$., 11.VI.1991/R. Anderson, wet oak/forest, 91-60] (1, CMNC); [GUATEMALA, 9 mi.W./Cd. Guatemala 6900'/VI-7-1974C.W. \&L./O’Brien \& Marshall] (2, ASUCOB). [MEX.: OAX, same data as holotype] (2, CMNC; 1, MZFC; 1, ICZ); [MEXICO: Oaxaca $3.2 \mathrm{~km} . / \mathrm{S}$. San Jose del Pacifico/2,440 m, 22.VII.1992, J.S./Ashe,oak-pine for. litt.] (1, CMNC);
[MEXICO: Oaxaca, $5.1 \mathrm{~km} . / \mathrm{S}$. Suchixtepec, 2,150 m/24.VII.1992,92-024, R.S./Anderson, oak|alder|pine/for., leaf litter Berlese] (1, CMNC; 1, MZFC; 1, ICZ); [MEXICO: Oaxaca, 23 km./S.W. Valle Nacional, KM76/1,300 m, 26.VII.1992,92-029/R.S. Anderson, cloud forest/ leaf litter Berlese] (1, CMNC).

Etymology. Named with the Latin word frontalis, referring to its strongly prominent frons.
Remarks. Most adults were collected from leaf litter between $\sim 1,830$ and $2,440 \mathrm{~m}$ above sea level, mainly in various types of oak forests. Males remain unknown.

Comments. Specimens of I. frontalis were first recognized as an undescribed Isodacrys species by R.S. Anderson.

## Isodacrys geminatum Howden, 1961

(Figs. 2, 6A, 6C, 9)
Isodacrys geminatum Howden, 1961: 80-81. Holotype: $\uparrow$, examined; labeled as [Texcoco 7000'/Mexico, Mex./20-VIII-1958] [H.F. Howden/Collector] [HOLOTYPE/Isodacrys/ geminatum/A.T. Howden/No.7384] [CNC/379714]. O’Brien \& Wibmer, 1982: 46 (checklist); Morrone, 1999: 145 (checklist); Cortes-Hernández \& Morrone, 2019: 50.

Diagnosis. Setae of elytra lanceolate, incompletely arched; rostrum with longitudinal sulci mesad of dorsolateral margins of rostrum at bent of scrobes; scrobe right-angled (Fig. 6A), anterior portion as long as posterior portion, posterior portion separated from eye by distance shorter than width of scrobe; anterolateral margin of prothorax without postocular vibrissae (Fig. 6A); elytra in dorsal view obovate, widest before midlength, with five intervals visible at base; prolegs not larger than metalegs (Figs. 6A and 6C), inner margin of protibiae without teeth; and spermatheca $U$-shaped, corpus subcylindrical, collum produced, conical, long, curved, ramus subcylindrical, very short.

Isodacrys geminatum can be separated from I. antrum by the elytral setae incompletely arched, the frons slightly deflexed to the rostrum and elytral intervals seven to nine not concave. In Isodacrys antrum the elytral setae completely arched, dorsal outline of the head continuously curved towards the rostrum, frons not deflexed and elytral intervals seven to nine concave near base. Additionally, Isodacrys geminatum can be easily confused with I. kuchii, I. carlae, I. confusum and I. fasciatum because of the strong superficial resemblance among these species with non-denticulate protibiae. Isodacrys kuchii, I. carlae, I. confusum and I. fasciatum can be separated from I. geminatum by the elytral setae completely arched, the scrobe obtusely angled, anterior portion of the scrobe shorter than posterior portion and posterior portion of the scrobe separated from eye by at least distance equal to width of the scrobe.

Additional material examined. MEXICO: Mexico City, Mpio. Xilotepec, Hwy57, 25.VII.2006, oak forest litter, $2,700 \mathrm{~m}, 20^{\circ} 04^{\prime} .797^{\prime} \mathrm{N}, 99^{\circ} 37.906^{\prime} \mathrm{W}$, R.S. Anderson (1, CMNC); Oaxaca, 22 km NE Oaxaca, 18.VI.1979, 2,600 m, H. \& A. Howden (1, CMNC); Oaxaca, 3 mi. N Suchixtepec, 4.VI.1971, Ber209, oaklitter, $9500^{\prime}$, S. Peck (1, CMNC); Puebla, 6 mi. NE Zacatepec, 27.VI.1975, L.E. Watrous (9, ASUCOB); Querétaro, Mpio. Colón, Cerro Zamorano, 26.VII.2006, oyamel/oak forest litter, 3,100 m, $20^{\circ} 55.967^{\prime} \mathrm{N}$, $100^{\circ} 11.021^{\prime}$ W, R.S. Anderson (1, CMNC); Querétaro, Mpio. Huimilpan, 1 km NW La Beata, 2.XII.2003, sample \#6, leaf litter, $2,500 \mathrm{~m}, 20^{\circ} 28.42^{\prime} \mathrm{N}, 100^{\circ} 14.49^{\prime} \mathrm{W}$, R. Jones (1, UAQE); Tlaxcala, 6.8 km N Taxco, Hwy 119, 10.VII.1992, ex. misc. Mushrooms, $2,820 \mathrm{~m}$, J.S. Ashe \#36 (1, CMNC).

Remarks. Except for one adult collected from an unidentified mushroom, most of the examined specimens were collected from leaf litter. This species has been collected between $\sim 2,100$ and $3,100 \mathrm{~m}$ above sea level. Males remain unknown.

Comments. Isodacrys geminatum was described based on four specimens from Texcoco, Estado de México, Mexico. The holotype, deposited in CNC, was examined. The three paratypes are deposited in CNC. Identity of additional material examined was based on comparisons with the holotype and by agreement with the original species description.

## Isodacrys guatemalenum Sharp, 1911

(Figs. 1, 5D, 9)
Isodacrys guatemalenus Sharp, 1911: 175. Syntypes not examined. Howden, 1993a: 1; Alonso-Zarazaga \& Lyal, 1999: 179.

Isodacrys guatemalensis; Pierce, 1913: 401 (designation as type species of Isodacrys).
Isodacrys guatemalena; Blackwelder, 1947: 799 (checklist).
Isodacrys guatemalenum; Howden, 1961: 83-84 (redescription); O’Brien \& Wibmer, 1982: 46 (checklist); Morrone, 1999: 145 (checklist).

Diagnosis. Setae of elytra parallel-sided, very long, erect (Fig. 5D); epistome with longitudinal carina joined with posterior margin of nasal plate (e.g., Fig. 4C); scrobe with anterior portion as long as posterior portion; anterolateral margin of prothorax without postocular vibrissae; elytra in dorsal view subelliptical, widest at midlength (Fig. 5D), with seven intervals visible at base; profemora wider than metafemora, protibiae longer than metatibiae, inner margin of protibiae with teeth; median lobe stout, shorter than spiculum gastrale, slightly longer than temones, apex in dorsal view acute, manubrium as long as median lobe; and spermatheca u-shaped, corpus subcylindrical, collum and ramus inconspicuous.

Isodacrys guatemalenum is separated from I. ellipticum by its parallel-sided long elytral setae, conspicuously erect, and by the spermatheca with collum and ramus inconspicuous. In I. ellipticum the elytral setae are spatulate, from semierect to erect, and the spermatheca with collum inconspicuous, ramus short.

Material examined. GUATEMALA: Sacatepéquez, Antigua, 16.VIII.1947, 5000', C. \& P. Vaurie (1, CMNC); Sacatepéquez, Santa Lucía Milpas Altas, 24.VI.1993, 1,800 m, B.D. Gill (1, CMNC). HONDURAS: Choluteca, 1 km W San Marcos de Colón, 23.VII.1977, C. \& L. O’Brien, G. Marshall (12, ASUCOB); Comayagua, Comayagua, 18.VII.1977, J.V. Mankins (2, CMNC); Comayagua, 22 km N Comayagua, 18.VII.1977, C. \& L. O'Brien, G. Marshall (80, ASUCOB; 12, CASENT).

Remarks. Some specimens have been collected between $\sim 1,500$ and $1,800 \mathrm{~m}$ above sea level.

Comments. Isodacrys guatemalenum was described based on 15 specimens from Dueñas, Sacatepéquez, Guatemala. The syntypes are distributed in BMNH and USNM and were not examined. Identity of material examined was based on comparison with specimens
identified by Anne Howden and by agreement with the original species description (Sharp, 1911) and redescription (Howden, 1961).

From the males examined, none of them possess postocular vibrissae. Nonetheless, Howden (1961) stated that postocular vibrissae can be poorly developed in males.

## Isodacrys kuchii Cortés-Hernández, new species

(Figs. 1, 7B-7D, 9, 20-21)
urn:lsid:zoobank.org:act:CB805B2D-80BA-4B36-9B6D-90740B222917
Diagnosis. Setae lanceolate, completely arched, more inconspicuous on head; posterior portion of scrobe separated from eye by distance equal to width of scrobe; antennal scape without scales (Fig. 20D); eyes large; anterolateral margin of prothorax without postocular vibrissae (Fig. 20B), pronotum covered with brown scales medially (Fig. 20A), with broad, lateral longitudinal vittae of white scales; profemora 1.2 times wider than metafemora, inner margin of protibiae without teeth, all tarsomeres with appressed scales (Fig. 20B); male terminalia with median lobe shorter than spiculum gastrale and manubrium (Fig. 21B); spermatheca comma-shaped, collum produced, conical, not directed to ramus (Fig. 21E).

Isodacrys kuchii can be confused with I. carlae, I. confusum and I. fasciatum. Isodacrys kuchii can be easily separated from those species by its inconspicuous median sulcus, with both endings marked by foveae, and the posterior portion of the scrobe separated from eye by distance equal to width of the scrobe. In I. carlae, I. confusum and I. fasciatum the median sulcus is conspicuous, usually deep, both endings without fovea, and posterior portion of the scrobe separated from eye by distance greater than width of the scrobe.

Description - Habitus. $O^{\prime \prime} 2.1-2.2 \mathrm{~mm}$ long, $0.8-0.9 \mathrm{~mm}$ wide; $\bigcirc 2.4 \mathrm{~mm}$ long, 1.1 mm wide. Scales polygonal, granulate, mainly overlapping irregularly, from brown to white in coloration. Head from vertex to frons covered by brown and white scales; rostrum and genae covered with white scales. Disc of pronotum covered with brown scales medially (Fig. 20A), with broad, lateral longitudinal vittae composed of white scales; sides of prothorax with white scales, with irregular longitudinal brown vittae between anterior and posterior constrictions of prothorax. Disc of elytra mostly with brown scales, intervals six to three at second third with irregular white fasciae; epipleura mostly with white scales, intervals one to four irregularly spotted with white scales at summit of apical declivity. Setae lanceolate, completely arched, more inconspicuous on head. Rostrum. Dorsolateral margins of rostrum subparallel; dorsal surface flat, becoming slightly depressed near median sulcus; median sulcus inconspicuous, both endings marked by foveae (Fig. 20D). Nasal plate with surface rugulose; anterior margin medially indented; posterior margin V-shaped, carinate. Epistome in continuous plane with remainder of rostrum, covered by three or four rows of shiny, smaller scales (Fig. 20D); anterior margin bearing three or four epistomal setae on each side; posterior margin limiting with anterior fovea of median sulcus. Scrobe deep, bent, reaching ventral surface of rostrum; dorsal margin obtusely angled; anterior portion of scrobe about $2 / 3$ length of posterior portion; posterior portion of scrobe separated from eye by distance equal to width of scrobe. Antenna setose, without
scales (Fig. 20D); scape capitate; funicular I antennomere clavate, wider and longer than remaining funicular antennomeres; funicular antennomere II clavate; funicular antennomeres III to VII moniliform, distal antennomeres becoming broader. Head. Head in lateral view with dorsal outline deflexed at frons towards rostrum; eyes large, subcircular, prominent, lateral, separated from outline of frons by distance about half diameter of eye. Prothorax. Pronotum in dorsal view subcylindrical, 1.1-1.2 times longer than wide, lateral outlines sinuate; in lateral view dorsal outline sinuate, anterior and posterior constrictions conspicuous. Anterolateral margin of prothorax without postocular vibrissae (Fig. 20B), with row of scales slightly projected anteriorly. Elytra. 1.6-1.8 times longer than pronotum, at base as wide as base of prothorax. Basal margin of elytra roundly emarginate. Elytra in dorsal view obovate, widest before midlength, five intervals visible at base. Elytra in lateral view with dorsal outline gently evenly curved towards summit of apical declivity; apical declivity with upper two thirds straight, lower third oblique. Elytral intervals with no elevations or depressions. Legs. Procoxae narrowly separated (Fig. 20C), intercoxal process $1 / 5-1 / 7$ width of procoxa; profemora fusiform, subequal in length to metafemora, 1.2 times wider than metafemora; protibiae straight, as long as metatibiae, inner margin without teeth. Dorsal surface of tarsi setose, all tarsomeres with appressed scales. Tarsal claws free. Abdomen. Ventrite III with anterior margin narrowly sulcate, sulcus shallow; sulcus of ventrites IV and V progressively wider and deeper (Fig. 20C); sulci enclosed by lateral margins of ventrites; posterior margin of sulci obliterated, ventrites becoming flat posteriorly. Genitalia. ơ: hemisternites VIII teardrop-shaped, outer corner not truncate (Fig. 21C); basal plate of sternite IX lightly sclerotized, distally bifurcate, tips of bifurcations sclerotized (Fig. 21C); spiculum gastrale 1.3 times longer than median lobe, distally continuing through basal plate; ring of tegmen without parameres; manubrium 1.6 times longer than median lobe (Fig. 21B); temones as long as median lobe (Fig. 21B); median lobe 4.4 times longer than wide in dorsal view, basal half more curved in lateral view, apical opening elliptical, base of opening prolonged anteriorly, apex acute (Fig. 21A). © : Lamina of sternite VIII triangular, apex acute. Ovipositor with distal gonocoxites slightly sclerotized laterally, styli represented by single thickened seta; two ventral baculi close to each other, separated by distance similar to width of one baculus. Spermatheca comma-shaped; corpus wide, narrowing towards cornu; collum produced, conical, not directed to ramus; ramus slightly tumid; cornu strongly bent near corpus, then straight, slightly sinuate, apically gradually narrowed (Fig. 21E). Sexual dimorphism. Ventrites I and II slightly concave mesally in males, convex in females.

Type material. Holotype ớ: [MEXICO: Puebla/Mpio: Zapotitlán Salinas/1 km N San Juan Raya] [ $18^{\circ} 19^{\prime} 11^{\prime \prime}, 97^{\circ} 36^{\prime} 50^{\prime \prime} / 19-v i i-2011$, A. Obregón/1, 755 m , Matorral] [Isodacrys/geminatum/Howden 1961/Det. Jesús Luna Cozar 2015] (CNIN). Paratypes ( 1 O', $^{\prime} 1$ O): [MEXICO: Puebla/same data as holotype] [ $18^{\circ} 19^{\prime} 11^{\prime \prime}, 97^{\circ} 36^{\prime} 50^{\prime \prime} /$ same data as holotype] [Isodacrys/same data as holotype] ( 1 O, CNIN); [MEXICO: Puebla/6 Mi. SW. Tehacan/July 8, 1973/Schaffner \& Mastro] [TAMU-ENTO/X0725504] [Isodacrys/? geminatum/DET./A. Howden] (1 \&, TAMUIC).

Etymology. This species is named with the word kuchî́ meaning weevil in Popoloca, one of the native languages in the Tehuacán valley, Puebla, where the specimens were collected.

Remarks. The type series was collected in the Tehuacán valley, in areas predominantly covered with xerophytic vegetation. The holotype and one of the paratypes were collected at $1,755 \mathrm{~m}$ above sea level.

Comments. Part of the female genitalia was lost during dissection, that is why some characters of the sternite VIII were not observed, described or coded for the phylogenetic analysis.

Isodacrys mexicanum Sharp, 1911
(Figs. 1, 6E, 9)
Isodacrys mexicanus Sharp, 1911: 176. Syntypes not examined.
Isodacrys mexicana; Blackwelder, 1947: 799 (checklist).
Isodacrys mexicanum; Howden, 1961: 92-93 (redescription); O’Brien \& Wibmer, 1982: 46 (checklist); Morrone, 1999: 145 (checklist); Cortes-Hernández \& Morrone, 2019: 50.

Diagnosis. Setae lanceolate, small, completely arched, mostly inconspicuous on disc of pronotum; scrobe with anterior portion as long as posterior portion, posterior portion separated from eye by distance nearly 1.2 times width of scrobe; eyes large, subcircular, convex; pronotum nearly 1.2 times longer than wide; prothorax strongly sinuate between anterior and posterior constrictions, anterolateral margin of prothorax with one to five vestigial postocular vibrissae; elytra in dorsal view obovate, widest before midlength, with seven intervals visible at base, intervals three to five slightly elevated at base; profemora wider than metafemora, protibiae longer than metatibiae, inner margin of protibiae with teeth; median lobe nearly as long as spiculum gastrale, apex in lateral view bent, apex in dorsal view broadly rounded; spermatheca comma-shaped, corpus slightly subglobose, collum slightly produced, subconical, directed to ramus, ramus broadly subcylindrical, short.

Isodacrys mexicanum can be separated from I. apicale by setae lanceolate, small, completely arched, the pronotum nearly 1.2 times longer than wide, the postocular vibrissae reduced in number and size and apex of the elytra conjointly rounded. In Isodacrys apicale setae of the elytra are spatulate, incompletely arched, erect on declivity, the pronotum as wide as long, more than six postocular vibrissae, some well-developed, and apex of the elytra bisinuate.

Material examined. MEXICO: Aguascalientes, 24 miles West of Aguascalientes, 5.VI.1956, B. Banta (1, CASENT); Mexico City, Milpa Alta, 26.VII.1980, H. Brailovsky (1, CNIN); Mexico City, Pedregal de San Ángel, Loc. 21, 22.VIII. 1961 (2, CNIN); Mexico City, Zumpango, 18.VII.1981, Peter Jump (1, CMNC; 1, TAMUIC); Michoacán, km 154 car. Irapuato-Guadalajara, 26.VII.1984, M. García (1, CNIN); Morelos, Tepoztlán, 11.X.1968, R. Muñíz V. (1, CMNC; 1, ASUCOB); Morelos, YMCA camp. Tepoztlán,
21.VIII.1958, H.F. Howden (4, CMNC); Oaxaca, 1.1 mi . W El Tule, 87/048, 5400', Woolley, Zolnerowich (2, TAMUIC).

Remarks. Adults have been collected beating trees at elevations of $\sim 1,500 \mathrm{~m}$ above sea level (Howden, 1961).

Comments. Isodacrys mexicanum was described based on five specimens from Guanajuato, Mexico. The syntypes, distributed in BMNH and USNM, were not examined. Identity of material examined was corroborated by comparing with specimens identified by Anne Howden and by agreement with the original species description (Sharp, 1911) and redescription (Howden, 1961).

## Isodacrys obrienorum Cortés-Hernández, new species

(Figs. 9, 22-23)
urn:lsid:zoobank.org:act:9DB9D7AA-F4AB-4424-A810-3DD5BF6ABA56
Diagnosis. Scales mostly contiguous and not overlapping (Fig. 22A); setae long, mostly erect (Fig. 22B); rostrum with short longitudinal sulci mesad of dorsolateral margins at bent of scrobes; epistome with scattered scales, integument exposed (Fig. 22D); scrobe with dorsal margin right-angled, posterior portion separated from eye by distance shorter than width of scrobe; eyes large (Fig. 22B), prominent; inner margin of protibiae without teeth; manubrium longer than median lobe; lamina of female sternite VIII with apex blunt (Fig. 23E).

From its closest relative, I. antrum, I. obrienorum can be recognized by the inner margin of the protibiae smooth, without teeth, scales not overlapping and setae semierect to erect. In I. antrum the protibiae possess teeth along their inner margin, scales overlapping irregularly, and setae completely arched.

Description - Habitus. Ơ' $^{1} 1.9-2.0 \mathrm{~mm}$ long, $0.8-0.9 \mathrm{~mm}$ wide; $\bigcirc 2.0-2.2 \mathrm{~mm}$ long, $0.9-1.1 \mathrm{~mm}$ wide. Scales polygonal, with raised margins, granulate, mostly contiguous and not overlapping, from dark brown to brown in coloration (Fig. 22A). Head and rostrum covered with brown scales. Disc of pronotum with dark brown to brown scales, with lateral broad vittae of brown to pale brown scales continuing with base of elytral intervals six and seven; sides of prothorax with dark brown scales. Elytra with irregular scale pattern. Setae lanceolate, long, slightly wide apically, from incompletely arched to erect, mostly erect (Fig. 22B). Rostrum. Dorsolateral margins of rostrum subparallel, with short longitudinal sulci mesad of dorsolateral margins at bent of scrobes; dorsal surface flat; median sulcus deeply impressed (Fig. 22D), both endings marked by foveae. Nasal plate with surface shiny, rugulose; anterior margin medially indented; posterior margin V-shaped, carinate. Epistome in continuous plane with remainder of rostrum, covered by shiny, small and scattered scales, integument of epistome exposed (Fig. 22D); anterior margin bearing four epistomal setae on each side; posterior margin limiting with anterior fovea of median sulcus. Scrobe deep, bent, reaching ventral surface of rostrum; dorsal margin right-angled; anterior portion of scrobe $2 / 3$ length of posterior portion; posterior portion of scrobe separated from eye by distance shorter than width of scrobe.

Antenna setose, without scales; scape capitate; funicular antennomere I clavate, wider and longer than remaining funicular antennomeres; funicular antennomere II clavate; funicular antennomeres III to VII moniliform, distal antennomeres becoming broader (Fig. 22D). Head. In lateral view with dorsal outline slightly deflexed at frons towards rostrum, frons not prominent (Fig. 22B); eyes large, subcircular (Fig. 22B), prominent, lateral, separated from outline of frons by less than half diameter of eye. Prothorax. Pronotum in dorsal view subcylindrical, as wide as long, lateral outlines sinuate (Fig. 22A); in lateral view dorsal outline sinuate between anterior and posterior constrictions. Anterolateral margin of prothorax with one or two rudimentary, small postocular vibrissae or lacking. Elytra. 1.9-2.1 times longer than pronotum, at base as wide as base of prothorax. Basal margin of elytra roundly emarginate. Elytra in dorsal view obovate, widest before midlength (Fig. 22A); five intervals visible at base. Elytra in lateral view with dorsal outline evenly curved towards summit of apical declivity; apical declivity oblique, subtle (Fig. 22B). Elytral intervals with no elevations or depressions. Legs. Procoxae narrowly separated (Fig. 22C), intercoxal process $1 / 6-1 / 5$ width of procoxa; profemora fusiform, subequal in length and width to metafemora; protibiae straight, as long as metatibiae, inner margin without teeth. Dorsal surface of tarsi setose, without scales. Tarsal claws free. Abdomen. Ventrite III with anterior margin narrowly sulcate; sulcus of ventrites IV and V progressively wider and deeper (Fig. 22C); sulci enclosed by lateral margins of ventrites; posterior margin of sulci obliterated, ventrites becoming flat posteriorly. Genitalia. O': Hemisternites VIII teardrop-shaped, outer corner not truncate (Fig. 23C); basal plate of sternite IX lightly sclerotized, distally bifurcate, tips of bifurcations sclerotized; spiculum gastrale as long as median lobe, distally continuing through basal plate; ring of tegmen without parameres; manubrium 1.5 times longer than median lobe; temones $4 / 5$ length of median lobe; median lobe 8.9 times longer than wide in dorsal view, evenly curved in lateral view (Fig. 23B), apical opening elliptical, apex acute. ㅇ: lamina of sternite VIII triangular, apex blunt; spiculum ventrale 4.3 times length of lamina. Ovipositor with distal gonocoxites slightly sclerotized laterally, styli represented by single thickened seta; two ventral baculi close to each other, separated by distance similar to width of one baculus; vagina with two pairs of lightly sclerotized proximal rods (Fig. 23G). Spermatheca comma-shaped; corpus slightly subglobose, narrowing towards cornu; collum produced, conical, bent apically towards ramus; ramus broadly cylindrical, short; cornu strongly curved near corpus, then gently curved, elongate, reaching apex of ramus, apically gradually narrowed (Fig. 23F). Sexual dimorphism. Ventrites I and II slightly concave mesally in males, convex in females; apex of ventrite V truncate in males, rounded in females.

Type material. Holotype $O^{\prime}$ : [GUATEMALA, $12 \mathrm{mi} . S E . /$ San Cristobal de To-/tonicapan, 9800'/VII-28.1974] [under stones] [C.W. \& L.B. O’Brien/\& G.B.Marshall] (ASUCOB). Paratypes ( 6 ƠƠ, 6 Q Y ): [GUATELAMA: Jalapa/4 km E Mataquescuintla/ 14.52705-90.14671 $\pm 55 \mathrm{~m} / 2,660 \mathrm{~m}, 2 . \mathrm{VI} .2009$,/LLAMA \#Go-B-07-1-04/cloud forest beating vegetation] [WORLD/WEEVIL/DATABASE/WWD0125181] (1 ¢, CMNC); [GUATELAMA: Jalapa/4 km E Mataquescuintla/14.52705-90.14671 $\pm 55 \mathrm{~m} / 2,660 \mathrm{~m}$,
2.VI.2009,/LLAMA \#Go-B-07-1-04/cloud forest beating vegetation] [WORLD/ WEEVIL/DATABASE/WWD0125188] ( $1 \mathrm{o}^{*}, \mathrm{CMNC}$ ); [GUATELAMA: Jalapa/4 km E Mataquescuintla/14.52705-90.14671 $\pm 55 \mathrm{~m} / 2,660 \mathrm{~m}, 2 . \mathrm{VI} .2009$,/LLAMA \#Go-B-07-104/cloud forest beating vegetation] [WORLD/WEEVIL/DATABASE/WWD0125177]

 SanCristobal de/Totonicapan,9300'] [VII-29-1974C.W.\&L./O'Brien \& Marshall] [under stones/and dead/grass clumps] (1 O, ASUCOB); [GUATEMALA: San Marcos/above Sibinal/3,009 m, 15.14366-92.06662/8 Jun 2015, R. Anderson/alder litter, 2015-127] (1 O', CMNC).

Etymology. Named after Charlie and Lois O'Brien, who collected most of the type series, in honor of their many years of world-wide collecting and their huge contributions to weevil knowledge.

Remarks. Adults have been collected under rocks, on dead grass clumps, on Alnus sp. (Betulaceae) litter, and beating vegetation, between 2,660 and 3,009 m above sea level. Some collected in cloud forests.

Comments. It was first recognized as an undescribed Isodacrys species by Charles W. O'Brien.

## Isodacrys okuiltontli Cortés-Hernández, new species

(Figs. 9, 24-25)
urn:lsid:zoobank.org:act:01FC3BDC-D6AC-48D1-BF6A-60CF65BB6626
Diagnosis. Setae short, spatulate, erect (Fig. 24B); scrobe with anterior portion as long as posterior portion, posterior portion of scrobe separated from eye by distance shorter than width of scrobe; eyes large, subcircular; anterolateral margin of prothorax without postocular vibrissae (Fig. 24B); protibiae with 2 or 3 small teeth.

Isodacrys okuiltontli can be readily separated from I. apicale by its short and erect setae, the posterior portion of the scrobe separated from eye by distance shorter than width of the scrobe, postocular vibrissae absent and apical margin of the elytra conjointly rounded. In I. apicale setae are longer, incompletely arched or becoming erect towards the summit of apical declivity, posterior portion of the scrobe separated from eye by distance greater than width of the scrobe, the prothorax with well-developed postocular vibrissae and apical margins of the elytra bisinuate.
 Scales polygonal, granulate, mainly overlapping irregularly, from brown to gray in coloration. Head with three longitudinal brown vittae at vertex, converging at frons; genae and rostrum mainly covered with gray scales. Longitudinal brown vittae of head continuing along disc of pronotum, wider, the lateral ones irregular; sides of prothorax irregularly scattered with brown and white scales. Disc of elytra with apparent irregular scale color pattern, epipleura mostly with white scales. Setae short, spatulate, erect
(Fig. 24B); mostly inconspicuous on temple. Rostrum. Dorsolateral margins of rostrum subparallel; dorsal surface flat; median sulcus conspicuous or not, anterior end marked by fovea (Fig. 24D). Nasal plate with surface shiny, rugulose; anterior margin medially indented; posterior margin V-shaped, carinate. Epistome in continuous plane with remainder of rostrum, covered by three rows of shiny, smaller scales (Fig. 24D); anterior margin bearing three epistomal setae on each side; posterior margin limiting with anterior fovea of median sulcus. Scrobe deep, bent, reaching ventral surface of rostrum; dorsal margin obtusely angled; anterior portion of scrobe as long as posterior portion; posterior portion of scrobe separated from eye by distance shorter than width of scrobe. Antenna setose, without scales; scape capitate; funicular antennomere I clavate, wider and longer than remaining funicular antennomeres; funicular antennomere II clavate, longer than remaining funicular antennomeres; funicular antennomeres III to VII moniliform, distal antennomeres becoming broader. Head. In lateral view with dorsal outline gently deflexed at frons towards rostrum, frons not prominent; eyes large, subcircular (Fig. 24B), prominent, lateral, separated from outline of frons by near $1 / 2$ diameter of eye. Prothorax. Pronotum in dorsal view subcylindrical, 1.1-1.2 times longer than wide, lateral outlines sinuate; in lateral view dorsal outline sinuate, anterior and posterior constrictions conspicuous. Anterolateral margin of prothorax without postocular vibrissae or scales projected anteriorly (Fig. 24B). Elytra. 1.6-1.7 times longer than pronotum, at base as wide as base of prothorax. Basal margin of elytra roundly emarginate. Elytra in dorsal view obovate, widest before midlength (Fig. 24A), seven intervals visible at base. Elytra in lateral view with dorsal outline evenly curved towards summit of apical declivity; apical declivity oblique (Fig. 24B). Elytral intervals three and five inconspicuously elevated before midlength. Legs. Procoxae narrowly separated, intercoxal process $1 / 6-1 / 5$ width of procoxa; profemora fusiform, subequal in length and width to metafemora; protibiae straight, as long as metatibiae, inner margin sinuate, with two or three small teeth. Dorsal surface of tarsi setose, first and second tarsomere with appressed scales. Tarsal claws free. Abdomen. Ventrites III, IV and V with anterior margin narrowly sulcate, progressively wider (Fig. 24C); sulci enclosed by lateral margins of ventrites; posterior margin of sulci obliterated, ventrites becoming flat posteriorly. Genitalia. ơ: Hemisternites VIII teardrop-shaped, outer corner not truncate (Fig. 25C); basal plate of sternite IX lightly sclerotized, distally bifurcate, tips of bifurcations sclerotized (Fig. 25C); spiculum gastrale $\sim 1.5$ times longer than median lobe, distally continuing through basal plate; median lobe $\sim 7$ times longer than wide in dorsal view, basal half more curved in lateral view, apical opening elliptical, base of opening prolonged anteriorly, apex acute (Fig. 25A). O: Lamina of sternite VIII triangular, apex blunt (Fig. 25E); spiculum ventrale 5.3 times length of lamina. Ovipositor with distal gonocoxites slightly sclerotized laterally, styli represented by single thickened seta; two ventral baculi close to each other, separated by distance similar to width of one baculus; vagina with two pairs of lightly sclerotized proximal rods (Fig. 25G). Spermatheca u-shaped; corpus cylindrical; collum produced, cylindrical, curved; ramus cylindrical, shorter than collum; cornu strongly curved near corpus, then evenly curved, elongate, reaching apex of collum, apically gradually narrowed (Fig. 25F).

Type material. Holotype ơ: [MEXICO, Oax., Hwy131/5 mi. N. Telixtlahuaca/6200' 31Aug. 1982 CW. \&/L. O’Brien \& G. Wibmer] (ASUCOB). Paratypes ( 5 ¢ 母)):
[MEXICO, Oax, 19.5/km. NW. Huitzo, Hwy./190, 2,260 m, 9.VII./1992 C.L. Bellamy] (1 \&, ASUCOB); [MEXICO, Oax., Hwy131/same data as holotype] (1 \&, ASUCOB; 1 \&, MZFC; 1 \& , CMNC); [MEXICO, Oax., Hwy131/same data as holotype] [Isodacrys/ minutus/Sharp/det. C. W. O’Brien, 1999] (1 \&, ASUCOB).

Etymology. Following the Náhuatl roots of the municipalities where the specimens were collected, this species is named with the Náhuatl word for weevil, okuiltontli.

Remarks. Adults have been collected between $\sim 1,890$ and $2,260 \mathrm{~m}$ above sea level.
Comments. All specimens except one have the scales of the elytral disc abraded at different degrees. The tegmen and some parts of the median lobe were not described in detail because they were damaged during dissection. Apparently with no sexual dimorphism.

## Isodacrys orizabae Sharp, 1911

(Fig. 9)
Isodacrys orizabae Sharp, 1911: 176. Syntypes not examined. Blackwelder, 1947: 799 (checklist); Howden, 1961: 85-86 (redescription); O’Brien \& Wibmer, 1982: 46 (checklist); Morrone, 1999: 145 (checklist); Cortes-Hernández \& Morrone, 2019: 50.

Diagnosis. Setae of elytra spatulate, from incompletely arched to erect; scrobe with anterior portion as long as posterior portion, posterior portion separated from eye by distance nearly 1.2 times width of scrobe; eyes moderate in size, slightly convex, not prominent; anterolateral margin of prothorax with one or two vestigial postocular vibrissae, when present; elytra in dorsal view variable, obovate, widest before midlength in females, subelliptical, widest at midlength in males; profemora wider than metafemora, protibiae longer than metatibiae, inner margin of protibiae with teeth; last abdominal segment tumid in females; median lobe shorter than spiculum gastrale, apex in dorsal view acute, manubrium shorter than median lobe; spermatheca u-shaped, corpus subcylindrical, slightly wide, collum inconspicuous, ramus broadly subcylindrical, short.

Isodacrys orizabae can be easily confused with I. ellipticum and I. guatemalenum, but it can be easily separated by setae from incompletely arched to erect, the epistome without longitudinal carina, eyes slightly convex, not prominent and last abdominal segment tumid in females. In I. ellipticum and I. guatemalenum setae are longer, from semierect to erect, the epistome with longitudinal carina joined with posterior margin of nasal plate, eyes convex, prominent, and last abdominal segment flat in both sexes.

Material examined. MEXICO: Oaxaca, 1.8 km S El Tule, 16.VII.1987, dry Acacia scrub, 1,524 m, R.S. Anderson (1, CMNC); Oaxaca, 20 km E Mitla, 3.VIII.1986, 1,800 m, H. \& A. Howden (2, CMNC); Oaxaca, Hwy 175, 10 km NE Oaxaca, 16.VI.1979, 1,800 m, H. \& A. Howden (1, CMNC); Oaxaca, 5 mi. SW Oaxaca, 4.VIII.1983, Acacia thorn scrub, 4700', R. . Anderson, W. Maddison (9, CMNC); Oaxaca, 13 mi. SE Tlalcolula, 2.VI.1974, 6100', C. \& L. O’Brien, G. Marshall (4, ASUCOB); Puebla, 4.4 mi. SW Acatepec, 26.VII.1974,

Clark, Murray, Ashe, Schaffner (5, CMNC; 6, TAMUIC); Puebla, 6 km S Acatepec, 19.VIII.1982, Clark, Cave (1, ASUCOB); Veracruz, VIII.1942, Koebele (2, CMNC); Veracruz, Hwy 150, 31 km W Orizaba, 15.VIII.1975, beating Pine, $1,880 \mathrm{~m}$, B. García G. (1, ASUCOB).

Remarks. Adults have been collected between $\sim 1,400$ and $1,880 \mathrm{~m}$ above sea level, some specimens were collected on Acacia sp. (Fabaceae). One specimen was collected beating Pinus sp. (Pinaceae).

Comments. Isodacrys orizabae was described based on nine specimens from Orizaba, Veracruz, Mexico. The syntypes are at BMNH and USNM and were not examined. Identity of material examined was corroborated by comparing with specimens identified by Anne Howden and by agreement with the original species description (Sharp, 1911) and redescription (Howden, 1961).

Isodacrys ovipennis (Schaeffer, 1908)
(Figs. 5F and 9)
Pandeletejus ovipennis Schaeffer, 1908: 215. Lectotype (Howden, 1961): $\uparrow$, not examined.
Pandeleteius ovipennis; Pierce, 1909: 359.
Isodacrys ovipennis; Pierce, 1913: 401 (new combination); Burke, 1959: 39 (actually Isodacrys burkei); Howden, 1961: 87-88 (redescription and lectotype designation); O’Brien \& Wibmer, 1982: 46 (checklist); Howden, 1993a: 2 (checklist); Morrone, 1999: 145 (checklist); Anderson, 2002: 780; Cortes-Hernández \& Morrone, 2019: 50.

Diagnosis. Setae lanceolate, small, from completely to incompletely arched, mostly inconspicuous; epistome with longitudinal carina joined with posterior margin of nasal plate; scrobe with anterior portion as long as posterior portion, posterior portion separated from eye by distance nearly 1.2 times width of scrobe; antennal scape and funicular antennomere I covered with scales dorsally; eyes large, slightly convex, not prominent; anterolateral margin of prothorax with three to six postocular vibrissae, mostly vestigial; elytra in dorsal view ovate, widest after midlength, intervals two and ten, and intervals five and six tumescent at their apical junctions (Fig. 5F), intervals three to five slightly elevated at base; profemora wider than metafemora, protibiae longer than metatibiae, inner margin of protibiae with teeth; median lobe shorter than spiculum gastrale, longer than temones, apex in dorsal view acute, manubrium as long as median lobe; spermatheca u-shaped; corpus subcylindrical, collum inconspicuous, ramus broadly subcylindrical, short.

Isodacrys ovipennis can be easily separated from I. burkei by anterolateral margin of the prothorax with three to six postocular vibrissae. Isodacrys burkei does not presents postocular vibrissae, instead a tooth extending anteriorly towards the eyes.

Material examined. MEXICO: Tamaulipas, Hwy101 11 mi. SW Cd. Victoria, 22.VII.1982, $4000^{\prime}$, C. \& L. O’Brien, G. Wibmer (37, ASUCOB); Tamaulipas, 2 mi. NE San Fernando,

Hwy 101, 31.VII.1970, C.W. O’Brien (1, ASUCOB); Tamaulipas, 4 mi. S San Fernando, Hwy 101, 5.V.1983, C. \& L. O’Brien, G. Marshall (2, ASUCOB). UNITED STATES OF AMERICA: Texas, Cameron Co. Laguna Atascosa Nat. Wild. Ref. rd. to Laguna Madre, 20. X.1989, sweeping Palafoxia texana var. ambigua, R.S. Anderson, E. Riley (13, CMNC); Texas, Cameron Co. Laguna Atascosa Wdlf. Ref, 20.X.1989, E.G. Riley (5, ASUCOB); Texas, Encino, 13.III.1966, Modiola caroliniana, M.F. Schuster (1, CMNC); Texas, Kenedy Co. Los Olmos Creek at \#77, 2 mi. S Riviera, 20.X.1989, sweeping Palafoxia texana var. ambigua, R.S. Anderson, E. Riley (13, CMNC; 1, CASENT); Texas, Kenedy Co. 10 mi. N Norias, 10.X.1989, D.J. Heffern (1, CMNC); Texas, Edinburg, 3.VI.1962, W.L. Sterling (1, TAMUIC); Texas, Kingsville, Kleberg Co., 25.VII.1951, H.R. Burke (1, TAMUIC); Texas, 10 miles NW La Gloria, 6.XI.1989, W.F. Chamberlain (1, TAMUIC); Texas, Kleberg Co., Jct. Los Olmos Creek \& Hwy 77, 19.X.1990, T. Carlow, E. Riley (2, TAMUIC); Texas, Palmitto Hill, 10 mi . W Boca Chica, 13.X.1985, R. Turnbow (1, CMNC).

Remarks. Adults have been collected on Palafoxia texana var. ambigua (Asteraceae) and Modiola caroliniana (Malvaceae). A big series from Tamaulipas was collected by the O'Briens at $\sim 1,200 \mathrm{~m}$ above sea level.

Comments. Isodacrys ovipennis was described based on 11 specimens (Howden, 1961) from Brownsville, Texas, United States of America. The lectotype is in USNM and was not examined. Paralectotypes are also at USNM. Although Howden (1961) stated that the type series was deposited in USNM, there is an additional syntype from databased material in Brigham Young University Arthropod Museum (BYUC; Gries, Gilbert \& Franz, 2014, available at https://scan-bugs.org/portal/collections/individual/index.php?occid=37487711).

## Isodacrys schwarzi Champion, 1911

(Figs. 3D and 9)
Isodacrys schwarzi Champion, 1911: 341. Syntypes not examined. Blackwelder, 1947: 799 (checklist); Howden, 1961: 81-82 (redescription); O’Brien \& Wibmer, 1982: 46 (checklist); Morrone, 1999: 145 (checklist); Cortes-Hernández \& Morrone, 2019: 50.

Diagnosis. Antennal scape covered with scales at apex dorsally (Fig. 3D); anterolateral margin of prothorax with tuft of postocular vibrissae; basal margin of elytra angularly emarginate, with sutural interval at summit of declivity strongly tumid, apical declivity strongly concave (Fig. 3D); and inner margin of protibiae without teeth.

Isodacrys schwarzi can be easily separated by its well-developed postocular vibrissae, which in I. buchanani are reduced to one or two small postocular vibrissae.

Comments. Isodacrys schwarzi was described based on two specimens from Monterrey, Nuevo León, Mexico. One female syntype is deposited at USNM and the other probably at BMNH. None of them were examined. No additional material was examined. Information regarding the deposition and sex of the remaining syntype is unknown.

Diagnosis is based on habitus photographs and on redescription by Howden (1961). See Champion (1911) and Howden (1961) to see the full description.


Figure 26 Isodillex plumosum new genus and new species. (A) Dorsal habitus; (B) lateral habitus; (C) ventral habitus; (D) head in anterior view.

Full-size DOI: 10.7717/peerj.10191/fig-26

## Isodillex Cortés-Hernández, new genus

(Figs. 1, 26-27)
Type species: Isodillex plumosum urn:lsid:zoobank.org:act:2AD213BD-E6DE-4146-A578-426D26C2002E

Diagnosis. Dorsal scales contiguous, not overlapping, polygonal; dorsolateral margins of rostrum subparallel, nasal plate with anterior margin medially indented, posterior margin of nasal plate carinate, epistome in continuous plane with remainder of rostrum, scales of different shape and shinier than remainder of rostrum, posterior margin of epistome indistinct; scrobe obtusely angled, reaching ventral surface of rostrum; anterior portion of scrobe about as long as posterior portion; posterior portion of scrobe separated from eye by distance shorter than width of scrobe; head deflexed, directed ventrally; mandibles covered with setae, without scales, prementum incompletely concealing maxillae; antennal scape capitate, abruptly thickened at apical region, funicular antennomere I longer than funicular antennomere II; pronotum in dorsal view subcylindrical, nearly as wide as long; anterolateral margin of prothorax without postocular vibrissae; intercoxal process of prosternum continuous, separating procoxae; humeri absent, elytra in dorsal view with seven intervals visible at base, basal margin of elytra roundly emarginate; prolegs larger than meso- and metalegs, tarsal claws free;


Figure 27 Isodillex plumosum new genus and new species, genitalia. (A) Median lobe in dorsal view; (B) median lobe in lateral view; (C) male hemisternites VIII and sternite IX in dorsal view; (D) ovipositor in dorsal view; (E) spermatheca; (F) ovipositor in lateral view; (G) female sternite VIII in dorsal view.

> Full-size DOI: 10.7717/peerj.10191/fig-27
anterior margin of ventrites III, IV and V flat, not sulcate; apex of lamina of female sternite VIII blunt; manubrium shorter than median lobe.

Isodillex can be separated from Isodacrys by combination of the following characters: dorsal scales contiguous, not overlapping, anterolateral margin of the prothorax without postocular vibrissae, prolegs larger than metalegs, anterior margin of ventrites III, IV and V flat, not sulcate and male genitalia with manubrium shorter than median lobe. In Isodacrys the dorsal scales are overlapping non-linearly (except I. obrienorum), anterolateral margin of the prothorax with postocular vibrissae or not, the prolegs from slightly larger than the metalegs to equal in size, anterior margin of ventrites III, IV and V sulcate and male genitalia with manubrium as long as median lobe or longer (except I. orizabae).

Description - Habitus. Males $2.8-3.0 \mathrm{~mm}$ long, $1.0-1.2 \mathrm{~mm}$ wide; females $2.4-3.2 \mathrm{~mm}$ long, $1.0-1.4 \mathrm{~mm}$ wide. Scales polygonal, contiguous, not overlapping. Rostrum. Dorsolateral margins of rostrum sub-parallel; dorsal surface flat. Epistome in continuous plane with remainder of rostrum. Nasal plate with anterior margin mesally indented;
posterior margin carinate. Scrobe deep, bent, reaching ventral surface of rostrum; anterior portion of scrobe about as long as posterior portion; posterior portion of scrobe separated from eye by distance shorter than width of scrobe. Funicular antennomere I clavate, large, wider and longer than remaining funicular antennomeres. Head. In lateral view with dorsal outline smoothly continuing with rostrum. Prothorax. Pronotum in dorsal view subcylindrical, nearly as wide as long, lateral outlines sinuate; base of pronotum slightly emarginate medially. Anterolateral margin of prothorax without postocular vibrissae. Elytra. 2.2-2.6 times longer than pronotum. Basal margin of elytra emarginate. Humeri absent. Elytra in dorsal view subelliptical, widest at midlength, seven intervals visible at base. Legs. Procoxae narrowly separated; prolegs larger than meso- and metalegs; profemora fusiform, subequal in length to metafemora, one fifth wider than metafemora; protibiae straight, one fifth longer than metatibiae, inner margin with teeth. Tarsal claws free. Abdomen. Anterior margins of ventrites III, IV and V flat, not sulcate. Genitalia. © ${ }^{\text {: }}$ Hemisternites VIII teardrop-shaped, outer corner not truncate; basal plate of sternite IX lightly sclerotized, distally bifurcate, tips of bifurcations sclerotized; spiculum gastrale as long as median lobe, distally continuing through basal plate; ring of tegmen without parameres; manubrium shorter than median lobe. ©: Lamina of sternite VIII triangular, apex blunt. Ovipositor with distal gonocoxites slightly sclerotized laterally, styli represented by single thickened seta; two ventral baculi close to each other, separated by distance similar to width of one baculus; vagina with two pairs of lightly sclerotized proximal rods.

Etymology. From the combination of Isodacrys and the latin adjective ilex, which means false/fraudulent. Isodillex therefore means "false Isodacrys" based on their external resemblance. Gender neuter.

## New combination

## Isodillex minutum (Sharp, 1911), new combination

(Fig. 2)
Isodacrys minutus Sharp, 1911: 176-177. Holotype or lectotype not designated; syntypes not examined.

Isodacrys minuta; Blackwelder, 1947: 799 (checklist).
Isodacrys minutum; Howden, 1961: 84 (redescription); O’Brien \& Wibmer, 1982: 46 (checklist); Morrone, 1999: 145 (checklist); Cortes-Hernández \& Morrone, 2019: 50.

Diagnosis. Setae of elytra lanceolate, completely arched at base, becoming erect towards declivity; frons convex, rounded; anterolateral margin of prothorax without modified scales, without postocular vibrissae; elytra in lateral view with dorsal outline evenly arcuate from base to apex; interval six slightly depressed at base; spermatheca Y-shaped, collum subcylindrical, short, ramus broadly subcylindrical, longer than collum, cornu curved near corpus, wide, short, not reaching apex of ramus.

Isodillex minutum can be readily separated from I. plumosum by its convex frons, anterolateral margin of the prothorax without modified scales and the elytra in lateral view with dorsal outline evenly arcuate from base towards apex. In Isodillex plumosum the frons is not prominent, anterolateral margin of the prothorax with plumose scales and elytra with apical declivity oblique.

Material examined. "MEXICO: MICHOACÁN, 2 mi. S Carapan, 6.VII.1985, Woolley, Zolnerowich" ( 5 specimens, TAMUIC; 3 specimens, CMNC), "MEXICO: MICHOACÁN, 18.5 km E Morelia, 28.VII.1988, Pine/oak/Acacia, 2,220 m, R. S. Anderson" ( 1 specimen, CMNC), "MEXICO: MICHOACÁN, 17 mi. W Pátzcuaro, 14.VIII.1982, 7700', C. \& L. O'Brien, G. Wibmer" ( 1 specimen, ASUCOB).

Remarks. Adults have been collected between $\sim 850$ and $2,440 \mathrm{~m}$ above sea level, some of them sweeping lupine plants (Fabaceae; see Howden, 1961). Males unknown.

Comments. Isodillex minutum was described based on four specimens from Omilteme, Guerrero, Mexico. One syntype is deposited at BMNH and deposition of remaining syntypes remains unknown. Syntypes not examined. Identity of the material examined was based on specimens identified by Anne Howden and by agreement with the original species description.

## Isodillex plumosum Cortés-Hernández, new species

(Figs. 1, 26-27)
urn:lsid:zoobank.org:act:955038E3-B522-4A22-9149-896ED0092C6B
Diagnosis. Setae of elytra spatulate, completely arched at base, becoming erect towards apical declivity (Fig. 26B); frons not prominent (Fig. 26B); anterolateral margin of prothorax with plumose scales; elytra with apical declivity oblique, intervals with no elevations or depressions; median lobe with basal half more curved in lateral view, apical opening subelliptical, apex in dorsal view rounded; spermatheca u-shaped, corpus cylindrical, collum and ramus inconspicuous.

Isodillex plumosum is easily separated from I. minutum by anterolateral margin of the prothorax with plumose scales, whereas in I. minutum the plumose scales are absent.

Description - Habitus. ớ 2.8-3.0 mm long, $1.0-1.2 \mathrm{~mm}$ wide; $\uparrow+2.7-3.2 \mathrm{~mm}$ long, $1.1-1.4 \mathrm{~mm}$ wide. Scales polygonal, with raised margins, granulate, contiguous, not overlapping, from brown to pale brown in coloration (Fig. 26A). Head at vertex inconspicuous longitudinal brown vitta, frons mainly with brown scales; genae and rostrum covered pale brown scales. Disc of pronotum with three longitudinal vittae of brown scales; sides of prothorax with pale brown scales. Disc of elytra and epipleura mostly brown scales, with irregular patches of pale brown scales; intervals nine and ten mostly covered with pale brown scales. Setae lanceolate on head and pronotum, most completely arched to incompletely arched on head, inconspicuous on temple; completely arched in pronotum. Setae of elytra spatulate, completely arched at base, becoming erect at apical declivity (Fig. 26B). Rostrum. Dorsolateral margins of rostrum sub-parallel; dorsal surface
flat; median sulcus conspicuous, posterior end marked by fovea between eyes (Fig. 26D). Epistome in continuous plane with remainder of rostrum, covered by three rows of shiny, small scales (Fig. 26D); anterior margin bearing five epistomal setae on each side; posterior margin limiting with median sulcus. Nasal plate with surface shiny, rugulose; anterior margin mesally indented; posterior margin $V$-shaped, carinate. Scrobe deep, bent, reaching ventral surface of rostrum; dorsal margin obtusely angled; anterior portion of scrobe about as long as posterior portion; posterior portion of scrobe separated from eye by distance shorter than width of scrobe. Antenna setose, without scales (Fig. 26A); scape capitate; funicular antennomere I clavate, wider and longer than remaining funicular antennomeres; funicular antennomere II clavate, longer than remaining funicular antennomeres; funicular antennomeres III to VII moniliform, distal antennomeres becoming broader. Head. In lateral view with dorsal outline smoothly continuing with rostrum, frons not prominent (Fig. 26B); eyes moderate in size, sub-oval, prominent, lateral, separated from outline of frons by distance shorter than half diameter of eye. Prothorax. Pronotum in dorsal view subcylindrical, nearly as wide as long, lateral outlines sinuate; in lateral view dorsal outline sinuate, anterior and posterior constrictions conspicuous; base of pronotum slightly emarginate medially (Fig. 26A). Anterolateral margin of prothorax without postocular vibrissae, with set of plumose scales projected anteriorly. Elytra. 2.2-2.4 times longer than pronotum, at base 1.1-1.2 times wider than base of prothorax. Basal margin of elytra emarginate. Elytra in dorsal view subelliptical, widest at midlength, seven intervals visible at base (Fig. 26A). Elytra in lateral view with dorsal outline nearly straight, becoming slightly convex towards summit of apical declivity; apical declivity oblique (Fig. 26B). Elytral intervals with no elevations or depressions. Legs. Procoxae narrowly separated (Fig. 26C), intercoxal process 1/6-1/4 width of procoxa; profemora fusiform, subequal in length to metafemora, 1.2 times wider than metafemora; protibiae straight, 1.2 times longer than metatibiae, inner margin with four to six teeth. Dorsal surface of tarsi setose, first and second tarsomeres with appressed scales or not (probably lost by abrasion). Tarsal claws free. Abdomen. Anterior margin of ventrites III, IV and V flat, not sulcate (Fig. 26C). Genitalia. Ó: Hemisternites VIII teardrop-shaped, outer corner not truncate (Fig. 27C); basal plate of sternite IX lightly sclerotized, distally bifurcate, tips of bifurcations sclerotized; spiculum gastrale as long as median lobe, distally continuing through basal plate; ring of tegmen without parameres; manubrium 0.7 times length of median lobe; median lobe 9.5 times longer than wide in dorsal view, basal half more curved in lateral view, apex in lateral view slightly bent (Fig. 27B), apical opening subelliptical, apex rounded. ©: Lamina of sternite VIII triangular, apex blunt, bifurcate (Fig. 27G); spiculum ventrale 4.4-4.8 times length of lamina. Ovipositor with distal gonocoxites slightly sclerotized laterally, styli represented by single thickened seta; two ventral baculi close to each other, separated by distance similar to width of one baculus; vagina with two pairs of lightly sclerotized proximal rods (Fig. 27F). Spermatheca u-shaped, corpus cylindrical; collum and ramus inconspicuous; cornu strongly curved near corpus, distal $2 / 3$ bent, elongate, reaching apex of ramus, apex abruptly narrowed (Fig. 27E). Sexual dimorphism. Ventrites I and II slightly flattened mesally in males, convex in females; apex of ventrite V emarginate in males, rounded in females.

Type material. Holotype (ơ): [61 mi. W. Fresnillo/Zacatecas, MEX/8000 ft. VI-25-54/R. H. Brewer] [III\#3] [Isodacrys/n.sp. \#2/AH ‘62] [CASENT/8424336]. Paratypes (3 Ơơ and 11 ¢ Q): [61 mi. W. Fresnillo/same data as holotype] [III\#3] [Cal Acad.] [CASENT/ 8424322] (1 Ơ CASENT); [61 mi. W. Fresnillo/same data as holotype] [III\#3] [CASENT/8424323] ( 1 ơ CASENT); [61 mi. W. Fresnillo/same data as holotype] [III\#3] [CASENT/8424331] (1 Ơ CASENT); [61 mi. W. Fresnillo/same data as holotype] [III\#3] [Cal Acad.] [Isodacrys/n.sp.\#2] [CASENT/8424321] (1 ¢ CASENT); [61 mi. W. Fresnillo/same data as holotype] [III\#3] [CASENT/8424324] ( 1 ¢ CASENT); [61 mi. W. Fresnillo/same data as holotype] [III\#3] [CASENT/8424325] (1 ¢ CASENT); [61 mi. W. Fresnillo/same data as holotype] [III\#3] [CASENT/8424326] ( 1 ¢ CASENT); [61 mi. W. Fresnillo/same data as holotype] [III\#3] [CASENT/8424327] (1 ¢ CASENT); [61 mi. W. Fresnillo/ same data as holotype] [III\#3] [CASENT/8424328] (1 O CASENT); [61 mi.
W. Fresnillo/same data as holotype] [III\#3] [CASENT/8424329] (1 ¢ CASENT); [61 mi. W. Fresnillo/same data as holotype] [III\#3] [CASENT/8424330] (1 \& CASENT); [61 mi. W. Fresnillo/same data as holotype] [III\#3] [CASENT/8424332] (1 ¢ CASENT); [61 mi. W. Fresnillo/same data as holotype] [III\#3] [CASENT/8424333] (1 q CASENT); [61 mi. W. Fresnillo/same data as holotype] [III\#3] [CASENT/8424334] (1 \& CASENT); [61 mi. W. Fresnillo/same data as holotype] [III\#3] [CASENT/8424335] (1 ¢ CASENT).

Etymology. From the latin word plumosum, referring to the plumose setae present at the anterolateral margin of prothorax.

Comments. Length of temones relative to median lobe is not described because temones were incomplete on the specimens dissected.

## Key to identify Isodacrys species

1 Elytra with sutural interval at summit of declivity strongly tumid, apical declivity strongly concave (Fig. 3D); inner margin of protibiae without teeth; antennal scape covered with scales only at apex dorsally (Fig. 3D)
$1^{\prime}$ Elytra with sutural interval at summit of declivity not or only slightly tumid, apical declivity straight (Fig. 6A); inner margin of protibiae variable; antennal scape with scales or not 3

2 Anterolateral margin of prothorax with one or two postocular vibrissae
$\qquad$ I. antrum new species $3^{\prime}$ Elytra with intervals seven to nine at most slightly depressed; lamina of female sternite VIII triangular (Fig. 13C) 4

4 Elytra in dorsal view with seven visible elytral intervals at base (Fig. 5F); inner margin of protibiae with teeth or small denticles; anterolateral margin of prothorax with postocular vibrissae usually present (Fig. 3B)
$4^{\prime}$ Elytra in dorsal view with five visible elytral intervals at base (Fig. 5E); inner margin of protibiae without teeth; prothorax with postocular vibrissae usually absent 13 5 Antennal scape covered with scales at dorsal apex (Fig. 3D); elytra in dorsal view ovate, widest after midlength (Fig. 5F), apical junction of intervals two and ten, five and six swollen (Fig. 5F)
$5^{\prime}$ Antennal scape without scales (Fig. 5A); elytra in dorsal view elliptical (Fig. 5D) or obovate, never widest after midlength, apical junction of intervals two and ten, and intervals five and six not swollen 7
6 Anterolateral margin of prothorax with conspicuous tooth, without postocular vibrissae; anterior portion of scrobe shorter than posterior portion I. burkei $6^{\prime}$ Anterolateral margin of prothorax without tooth, with three to six postocular vibrissae; anterior portion of scrobe as long as posterior portion (Fig. 4E) .................... I. ovipennis 7 Prolegs larger than metalegs (Figs. 5D, F), promefoma wider than metafemora, protibiae longer than metatibiae .. 8
$7^{\prime}$ Prolegs subequal in size to metalegs (Fig. 6A) 11

8 Anterolateral margins of prothorax without postocular vibrissae (Fig. 10B); posterior margin of scrobe separate from eye by distance equal to width of scrobe; setae of elytra long, from semierect to erect (Fig. 5D)
$8^{\prime}$ Anterolateral margins of prothorax with postocular vibrissae (Fig. 3B); posterior margin of scrobe separate from eye by distance at least one fifth longer than width of scrobe (Fig. 4E); setae of elytra moderate in length, curvature of setae various 10
9 Setae of elytra very long, subparallel, erect (Fig. 5D); spermatheca with collum and ramus inconspicuous (Fig. 27E) $\qquad$ I. guatemalenum
$9^{\prime}$ Setae of elytra spatulate, from semierect to erect; spermatheca with collum inconspicuous, ramus very short, broadly cylindrical I. ellipticum 10 Setae of elytra spatulate, from incompletely arched to erect; eyes moderate in size, slightly convex, not prominent; prothorax in dorsal view with subtle constrictions, lateral margins slightly sinuate $\qquad$ I. orizabae $10^{\prime}$ Setae of elytra lanceolate, small, completely arched; eyes large, subcircular, conspicuously convex (Fig. 4B); prothorax in dorsal view with lateral margins strongly sinuate between anterior and posterior constrictions I. mexicanum 11 Setae of elytra lanceolate, completely arched; scrobe right-angled (Fig. 6A), anterior portion shorter than posterior portion; sulci of ventrites III to V with posterior margin sharp, conspicuously delimiting sulci I. crispum
$11^{\prime}$ Setae of elytra spatulate, from incompletely arched to erect; scrobe obtusely angled (Fig. 4E), anterior portion as long as posterior portion; sulci of ventrites III to V with posterior margin effaced, ventrites becoming flat posteriorly (Fig. 6F)
12 Anterolateral margin of prothorax with more than six postocular vibrissae, some well-developed (Fig. 4E); apical margins of elytra bisinuate (Fig. 6B); posterior portion of scrobe separated from eye by distance 1.2 times width of scrobe (Fig. 4E) ........ I. apicale $12^{\prime}$ Anterolateral margin of prothorax without postocular vibrissae (Fig. 10B); apical margins of elytra straight, conjointly rounded; posterior portion of scrobe separated from eye by distance shorter than width of scrobe (Fig. 24B) ............. I. okuiltontli new species

13 Scales of dorsal surface of body mostly contiguous and not overlapping (Fig. 22A); setae mostly erect (Fig. 22B); epistome with scattered scales, integument exposed (Fig. 22D) $\qquad$ I. obrienorum new species
$13^{\prime}$ Scales of dorsal surface of body variously overlapping non-linearly (Fig. 5E); setae usually not erect, at most erect at apical declivity of elytra; epistome with scattered scales but integument not substantially exposed (Fig. 24D) 14
14 Rostrum with longitudinal sulci mesad of dorsolateral margins at bent of scrobes (Fig. 4D); scrobe with anterior portion as long as posterior portion; profemora as wide as metafemora 15
$14^{\prime}$ Rostrum without longitudinal sulci mesad of dorsolateral margins at bent of scrobes; scrobe with anterior portion shorter than posterior portion (Fig. 12B); profemora slightly wider than metafemora
15 Scrobe right-angled (Fig. 6A); posterior portion separated from eye by distance shorter than width of scrobe; elytra obovate, widest before midlength, without depressions
$15^{\prime}$ Scrobe obtusely angled (Fig. 4E); posterior portion separated from eye at least by distance equal to width of scrobe; elytra subelliptical, widest at midlength (Fig. 5D), intervals seven to nine slightly depressed at base
16 Frons not prominent (Fig. 5C); posterior portion of scrobe separated from eye by distance equal to width of scrobe; elytral setae spatulate, semierect, becoming erect at apex of elytra I. brevirostre
$16^{\prime}$ Frons very prominent (Fig. 18B); posterior portion of scrobe separated from eye by distance at least twice width of scrobe (Fig. 18B); elytral setae lanceolate, completely arched to incompletely arched
I. frontalis new species

17 Antennal scape covered with scales at apex dorsally (Fig. 3D); all tarsomeres with appressed scales (Fig. 14B); pronotum covered with white scales medially; spermatheca with collum produced, bent basally towards ramus, contiguous with corpus (Fig. 15F) $\qquad$ I. confusum new species
$17^{\prime}$ Antennal scape without scales (Fig. 5A), if present reduced to 1 or 2 scales; all tarsomeres with appressed scales or not; scale pattern of pronotum various; spermatheca with collum produced but not contiguous with corpus (Fig. 17E) 18
18 Pronotum covered with white scales medially (Fig. 16A); spermatheca comma-shaped, collum produced, subcylindrical, bent basally towards ramus, not contiguous with corpus $\qquad$ I. fasciatum new species
$18^{\prime}$ Pronotum covered with brown scales medially (Fig. 12A); shape of spermatheca various, collum not bent basally towards ramus 19
19 Posterior portion of scrobe separated from eye by distance equal to width of scrobe (Fig. 20B); spermatheca comma-shaped, collum produced, conical I. kuchii new species
$19^{\prime}$ Posterior portion of scrobe separated from eye by distance 1.5-1.6 times width of scrobe (Fig. 12B); spermatheca peanut-shaped (Fig. 13D) $\qquad$ I. carlae new species

## DISCUSSION

## Generic relationships amongst Isodacrys-related Tanymecini

The present analysis has several implications on the relationships among the New World Tanymecini. The monophyly of Tanymecini has not been tested under a comprehensive approach. Nonetheless, current studies suggest that they may be polyphyletic (Gillet et al., 2018), with historical mistakes in classification and interpretation related to the presence of postocular vibrissae at the anterolateral margin of the prothorax in the tribe that consequently ended up in taxonomic amendments excluding several lineages from the tribe (Morrone, 1998; Franz, 2012; Marvaldi et al., 2018; Del Río \& Lanteri, 2019b). Nine of the 16 genera of Tanymecini distributed in the New World were included in the analysis, but further discussion of the natural history of the tribe is out of the scope of this work. Further analyses should be carried out for tanymecine lineages that remain poorly understood (Cortes-Hernández \& Morrone, 2019). As previously stated, traditional characters for grouping New World tanymecines are highly homoplastic and therefore should be taken with caution. This does not mean that they are not phylogenetically informative, instead they likely represent recent transformations that can contribute to the understanding of smaller clades within Tanymecini.

The Tanymecini clade is well supported by eight synapomorphies (epistome distinct 7:1, scrobe in lateral view strongly curved, directed to ventral surface of rostrum 15:1, rostrum in lateral view directed ventrally 19:1, antennal scape capitate, abruptly thickened at apical region 26:1, funicular antennomere I longer than funicular antennomere II 27:2, pronotum in dorsal subglobose 28:1, pronotum in dorsal view with constriction of margins 30:1 and prothorax in lateral view oblique, dorsally produced anteriorly $31: 1$ ) and one homoplastic character state (presence of postocular vibrissae 32:0). The basal split between Minyomerus and the remaining Tanymecini can be related with the psammophilic condition of the genus (i.e., adapted to live in sandy environments; Jansen \& Franz, 2015).

Neither Pandeleteinus nor Pandeleteius were recovered as monophyletic. Pandeleteinus presented a high degree of overlap of several character states with both West Indian and continental lineages. In the case of Pandeleteius, which contains approximately 161 described species, the present taxon sampling is far from representing a comprehensive extent to go deep on insights about its naturalness.

The clade aff. Pandeleteinus sp.-Isodrusus guajavus is the sister group of the Paradacrys ensiformis- I. antrum clade, supported by one apomorphy (anterior margin of nasal plate slightly indented 5:1) and two homoplastic character states (dorsal scales contiguous, not overlapping 1:0 and manubrium of tegmen shorter than median lobe 72:0, both with different character states for Isodacrys clade). The clade aff. Pandeleteinus sp.-Isodrusus guajavus is supported by two homoplastic character states regarding the shape of the scrobe and its distance to eye (chars. 15 and 18). It has a basal division splitting aff. Pandeleteinus sp. from the clade aff. Isodrusus sp.-Isodrusus guajavus. The undescribed species from Colombia fits under the current Isodrusus circumscription (CortésHernández \& Anderson, 2019), here supported by three character states (tarsal claws connate $60: 1$, sulcus in ventrites III, IV and V along its entire width 62:1 and posterior
margin of sulcus in ventrites III, IV and V present, sharply delimiting sulcus 63:0); although homoplastic, they could represent true synapomorphies under a different scope.

The West Indian Paradacrys ensiformis-Scalaventer subtropicus clade is the sister group to the Isodillex-Isodacrys clade based on one synapomorphy (seven elytral intervals visible in dorsal view 40:0, although state 1 , which is five elytral intervals visible in dorsal view, is present in Isodacrys kuchii-I. obrienorum clade) and one homoplastic character state (postocular vibrissae at anterolateral margin of prothorax absent 32:0). Within the Paradacrys ensiformis-S. subtropicus clade, supported by two homoplastic characters (presence of longitudinal carina on the epistome 10:1 and posterior margin of scrobe ending before ventral surface of rostrum 16:0, with reversal in the clade S. cyrillaeS. subtropicus), each genus is recovered as monophyletic in the following sequence: Paradacrys (Paululusus (Scalaventer)).

There are four homoplastic character states supporting the relationship between Isodacrys and Isodillex (anterior portion of scrobe as long as posterior portion 17:1, elytra in dorsal view subelliptical 38:3, elytral humeri absent 39:1 and basal margin of elytra roundly emarginate $42: 1$ ). Some of these homoplastic character states (chars. 38, 39 and 42), all related to the condition of the elytra, as noted by Lanteri \& Del Río (2017), likely evolve convergently in relation with the loss of metathoracic wings and if not accompanied by other synapomorphies should not be considered evidence of a close phylogenetic relationship. Although apterism can be homoplastic in certain lineages like island-inhabiting entimines (Howden, 1970; Franz, 2012; Paululusus as here presented), in continental clades like Isodacrys and Minyomerus (see Jansen \& Franz, 2015: 117) it remained constant. This can be related to both historical and ecological constraints. Based on the information in hand on adult morphology, a close relationship between Isodacrys and Isodillex clade is proposed but additional sources of information should be added to strengthen this hypothesis.

## Delimitation of Isodacrys and phylogenetic insights

The present analysis represents a first approximation of the relationships within Isodacrys and is suited to address the research questions regarding the monophyly of Isodacrys and its relationships with other Tanymecini. None of the character states used to diagnose Isodacrys (Howden, 1961, p. 76) resulted as a synapomorphy when analyzed in a phylogenetic context.

Although the length of the tegmen relative to the median lobe (char. 72:1,2) represents an apomorphy for Isodacrys in the trees obtained with apparently two unique character states shared among its species, there is a reversal in I. orizabae.

Although there is apparently a high degree of incongruence, translated in many clades supported by one or several homoplastic characters, they still represent relevant grouping information for Isodacrys species. As one of the main research questions was to elucidate the generic position of the genus within the American Tanymecini (as discussed in the previous section), an extensive taxon sampling of putatively related genera was conducted. This inclusive approach implied the possibility of putative convergent evolution of non-immediate sister lineages, as supported by our results. A more exclusive
approach could transform some of the homoplastic character states into synapomorphies but would also weaken/modify the displayed phylogenetic relationships and the question would not be even tested.
Howden $(1961,1969,1970)$ highlighted the relevance of genitalia within Tanymecini lineages. According to our results and preliminary data matrices including some of the traits discussed by Howden (e.g., length/width ratio of aedeagus, curvature of aedeagus in lateral view), these characters did not show clear grouping information in the taxa included and exhibited high degree of overlap among New World Tanymecini. Some characters related to the spermatheca have been considered in previous works (Lanteri \& Del Río, 2008; Girón \& Franz, 2012; Franz, 2012; Jansen \& Franz, 2015; Lanteri \& Del Río, 2017). Efforts to include characters related to this structure were conducted in preliminary data matrices but the structures resulted quite variable interspecifically within several genera of Tanymecini, including Isodacrys, and also convergently similar in species of non-sister genera. If parthenogenesis takes place in Isodacrys, it could be a possible explanation for the strong variation of the spermatheca. Dybas (1966) stated that a spermatheca is a non-functional structure in completely parthenogenetic species. Such structures may become vestigial in time, or, at least, more variable because of less stringent selection. Additional to this, depending on how the specimens are prepared for examination (temperature, type of liquid), the spermathecal shape can visibly change during preparation. Based on this and on the high degree of incongruence depicted by the spermathecal characters, we decided to exclude them from the final analysis. Future analyses should consider intraspecific variation when studying reproductive characters in entimine weevils where parthenogenesis has been corroborated or suspicious. Further approaches remain to be explored (e.g., morphometry) in order to evaluate the phylogenetic significance of these characters for parthenogenetic species.

From the 20 species here included in Isodacrys males remain unknown for eight of them: Isodacrys antrum, I. brevirostre, I. carlae, I. crispum, I. ellipticum, I. fasciatum, I. frontalis and I. geminatum (see also comments on Isodacrys schwarzi). Based on the lack of males throughout different localities where specimens have been collected (see material examined for further information) we hypothesize that these species might be parthenogenetic (Howden, 1961). According to Lanteri \& Normark (1995), this mode of reproduction is the most plausible explanation when males are absent in different populations of the same species in naupactine weevils. Further collecting will be needed as well as cytological and genetic studies to corroborate parthenogenesis in Isodacrys species and other genera of Tanymecini where parthenogenesis is suspicious (Jansen \& Franz, 2015; Cortés-Hernández \& Anderson, 2019). Additionally, parthenogenesis can be supported by its correlation with flightlessness given the potential difficulties of flightless insects to locate mates (Roff, 1990; Lanteri \& Normark, 1995). Under this scenario, no energy is invested in mating and wing-development. The loss of metathoracic wings has been also correlated with greater fecundity and environmental stability where dispersal does not impact in long-term survival of populations (Roff, 1990; Wagner \& Liebherr, 1992), as may be the case for most Isodacrys species (parthenogenetic or not), which are found in mountains and tropical montane forests, considered stable habitats (Roff, 1990;

Wagner \& Liebherr, 1992). Parthenogenesis can increase fecundity too, because all the eggs can produce females and are not "wasted" in producing males (Dybas, 1966). In stable environments, low genetic variability of parthenogenetic species may result in greater success of progeny (Jansen \& Franz, 2015).

If parthenogenesis exists in Isodacrys, and assuming that all species where males remain unknown are parthenogenetic, accordingly to the MPT's (Fig. 2) parthenogenesis evolved at least five times within Isodacrys: I. ellipticum, I. crispum clade, I. apicale, I. kuchii clade and $I$. brevirostre clade. Parallel evolution of parthenogenesis has been also discussed for naupactine weevils (Lanteri \& Normark, 1995; Normark, 1996) and for Minyomerus (Jansen \& Franz, 2015). This can be reflected on the interspecific variation of the genitalia (e.g., shape of spermatheca, length/width of median lobe) among closely related Isodacrys species and the consistent information exhibited by the genitalia-related characters in major clades within the genus (Fig. 2, see optimizations in characters 65, 70, 71 and 72 for Isodacrys). Although genitalia characters are traditionally associated as species-grouping characters (Vaurie, 1976; Del Río \& Lanteri, 2019a), it is not necessarily the pattern when alternative modes of reproduction take place. Other sources of information should be explored to better support the interspecific relationships among Isodacrys species.

According to the most-parsimonious trees, a remarkable tendency to certain traits within Isodacrys can be noticed: Isodacrys kuchii-I. antrum clade (Fig. 2) is represented by species with only five elytral intervals (char. 40:1); within this clade there is a tendency to lose the postocular vibrissae at anterolateral margin of the prothorax and loss of teeth along inner margin of the protibiae (chars. 32:0 and 54:0), reduction in size of the prolegs relative to the metalegs (chars. 52:0, although with reversal to state 1 in I. kuchii clade, and 53:0) and the manubrium longer than the median lobe (char. 72:2). These trends to reduction and loss seem to be an important division among Isodacrys species.
Developmental studies should be carried out in order to obtain further information regarding these characters and their putative implications for the natural history of lineages within Isodacrys.

We believe in the potential existence of more species of Isodacrys unknown to science throughout Mexico and Central America. The compiled information from museum specimens indicates that adults of five species were found in leaflitter, a niche that has largely been unexplored for Tanymecini.

## Taxonomic amendments

We propose the following nomenclatural changes: (i) The genus Isodillex is here proposed for grouping Isodillex minutum (Sharp) new combination and Isodillex plumosum new species and (ii) Isodacrys is re-defined as a genus containing 20 species, eight here described as new.

## CONCLUSIONS

The phylogenetic analysis based on adult morphology recovers the genus Isodacrys as a monophyletic entity composed by 20 described species, eight of them herein described.

Additionally, Isodillex new genus, which is sister to Isodacrys, is erected to accommodate one new species and species previously considered as Isodacrys. Finally, the West-Indian Paradacrys-Scalaventer clade is sister to the Isodillex-Isodacrys clade. Future directions to enhance our understanding of the natural history of these weevil lineages can be field observations, breeding experiments of potentially parthenogenetic species and biogeographic analyses regarding its relationships with West Indian tanymecines. Additional sources of information with potential phylogenetic signal (e.g., molecular data) will be necessary to strengthen/test the current species-level relationships inferred within Isodacrys as well as its relationships with other Tanymecini lineages.

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## ADDITIONAL INFORMATION AND DECLARATIONS

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## Competing Interests

Juan J. Morrone is an Academic Editor for PeerJ.

## Author Contributions

- Kevin A. Cortés-Hernández conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.
- Juan J. Morrone conceived and designed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.


## Data Availability

The following information was supplied regarding data availability:
Specimens examined
Acronyms of the collections:
MZFC, Museo de Zoología "Alfonso L. Herrera", Facultad de Ciencias, Universidad Nacional Autónoma de México, Mexico City, Mexico.

IEXA, Colección Entomológica del Instituto de Ecología A.C., Xalapa, Veracruz, Mexico. CNIN, Colección Nacional de Insectos, Instituto de Biología, Universidad Nacional Autónoma de México, Mexico City, Mexico.

UAQE, Colección de Insectos de la Licenciatura en Biología, Facultad de Ciencias
Naturales de la Universidad Autónoma de Querétaro, Campus Juriquilla, Querétaro, Mexico.
ICZ, Insect Collection of Zacapa, Instituto de Investigaciones del Centro Universitario de Zacapa, Universidad de San Carlos de Guatemala, Zacapa, Guatemala.

CMNC, Canadian Museum of Nature Collection, Ottawa, Ontario, Canada.
TAMUIC, Texas A\&M University Insect Collection, Department of Entomology, Texas A\&M University, College Station, Texas, United States of America.

CAS, California Academy of Sciences, Department of Entomology, San Francisco, California, United States of America.

ASUCOB, Arizona State University Charles O'Brien Collection, Tempe, Arizona, United States of America.

CNC, Canadian National Collection of Insects, Arachnids and Nematodes, Agriculture and Agri-Food Canada, Ottawa, Canada.

Isodacrys antrum
Holotype $\uparrow$ : [MEXICO: Tamaulipas/El Cielo, 3 km SE Alta Cima/23.03440-99.186967 $\pm 20 \mathrm{~m} / 860 \mathrm{~m}, 24$. VIII.2009/L.D. Sáenz LSD-370/cloud forest, nest under rock] [WORLD/ WEEVIL/DATABASE/WWD0133374] [Isodrusus/or/Isodacrys/det. R.S. Anderson, 20] (CMNC). Paratypes ( 5 O O ): [GUAT.: BAJA VERAPAZ/8.6 km.W. Chilascó, 1,560 m./24. V.1991, R. Anderson/oak|pine|liquidambar/forest litter, 91-18] (2, CMNC). [MEXICO: Chiapas/Mpio: Tapalapa/12 Km N Coapilla/12-III-94, R. Jones] [Isodacrys/Det. R.S. Anderson, 1995] (1, CMNC); [MEX.: CHIAPAS, 3 km./W. San Cristobal, 2,000 m./18. IX.1991, R. Anderson/oak|pine forest, 91-108] (1, MZFC); [MEXICO: Tamaulipas/same data as holotype] [WORLD/WEEVIL/DATABASE/WWD0133381] (1, CMNC).

Isodacrys apicale
Holotype: ©̛, examined; labeled as [151|35/Choix, SIN./13-VIII-35] [HOLOTYPE/ Isodacrys/apicale/A.T. Howden/No.7385] [CNC/379712]. MEXICO: Sinaloa, same data as holotype (1 paratype, CMNC); Sonora, 1 mi. W Álamos, 16.VII.1964, H. R. Burke, J. Apperson (1, TAMUIC).

## Isodacrys brevirostre

Holotype: $\uparrow$, examined; labeled as [Cuernavaca 6 Mi.,/N., 7500' Morelos/15-VIII-1954/J. G. Chillcott] [HOLOTYPE/Isodacrys/brevirostre/A.T.Howden/No. 7383] [CNC/379713].

MEXICO: Estado de México, 6 km al SE de Atlautla, 27.IX.2017, Redeo en vegetación, B. Pino-Encino, 2450, $19^{\circ} 00^{\prime} 46^{\prime \prime} \mathrm{N}, 98^{\circ} 45^{\prime} 26^{\prime \prime}$ W, K. Cortés, L. Delgado (1, IEXA); Estado de México, La Mirasol, 7 km SW Santiago de Tianguistengo, 2.XI.1973, 2, 800 m , C.W. O'Brien (36, ASUCOB; 3, CASENT); Estado de México, Ocoyoacac, 28.X.1973, on misc. flowers, C.W. O’Brien (4, CASENT; 89, ASUCOB); Estado de México, Bejucos, Mex. Temascaltepec, 3.VII.1933, H.E. Hinton, R.L. Usinger (3 paratypes, CASENT); Estado de México, Real de Arriba, VII.1932, H. Hinton (1 paratype, TAMUIC); Estado de México, Hwy 190, 11 mi. W Río Frío, 5.VI.1983, 9800', C. \& L. O’Brien, G. Marshall (3, ASUCOB); Estado de México, Hwy15, 28 mi. W. Toluca, 6.VIII.1982, 9100', L. O’Brien, G. Wibmer (38, ASUCOB); Guanajuato, 3.6 mi. NE Guanajuato, 5.VII.1985, Woolley, Zolnerowich (1, TAMUIC); Guerrero, 6 km NE Ayotoxtla (Tlatlauquitepec Rd), 24.X.1973, A.N. García A. (2, ASUCOB); Guerrero, 9 km SE Tlatlauquitepec, 24.X.1974, A.N. García A. (1, ASUCOB); Hidalgo, Hwy105 16 mi. N Metzquititlán, 30.VII.1982, $7200^{\prime}$, L. O'Brien, G. Wibmer (23, ASUCOB); Hidalgo, Hwy105 Mineral Real del Monte, 14.VI.1983, 9400', C. \& L. O’Brien, G. Marshall (1, ASUCOB); Hidalgo, Hwy105, 3 mi. S Zacualtipan, 3.VIII.1982, 6959', C. \& L. O’Brien, G. Wibmer (2, ASUCOB); Mexico City, Cañada Contreras, 5.X.1982, s/composit, 2, 550 m, K. Luna (10, MZFC); Mexico City, Road from Mexico City P.N. Cumbres de Ajusco, 4.IX.1982, 2, 750 m, C. \& L. O’Brien, G. Wibmer (17, ASUCOB); Mexico City, Delegación Tlalpan, Tlalpuente, 4.XI.1993,
L. Torres-Miller (1, CMNC); Mexico City, El Pedregal, 4.IX.1982, 2,340 m, C. \& L. O’Brien, G. Wibmer (1, ASUCOB); Mexico City, Unión Perif. Ajusco, 4.IX.1982, 2,340 m, K. Luna (1, MZFC); Mexico City, 7 mi . N Valle de Bravo, 7.VIII.1982, 8300', C. \& L. O’Brien, G. Wibmer (1, ASUCOB); Michoacán, 2 mi. S Carapan, 6.VII.1986, H. \& A. Howden (1, CMNC); Michoacán, 2 mi. S Carapan, 6.VII.1985, Woolley, Zolnerowich (1, TAMUIC); Michoacán, 28.5 mi. S Nueva Italia, 9.VII.1985, Woolley, Zolnerowich (1, TAMUIC); Michoacán, 17 mi. W Pátzcuaro, 4.VIII.1982, 7700', C. \& L. O’Brien, G. Wibmer (6, ASUCOB); Morelos, 8 km N Cuernavaca, Hwy 95, 5.IX.1982, C. \& L. O’Brien, G. Wibmer (5, ASUCOB); Morelos, Santa María Ahuacatitlán, 6.XI.1980, F. Aguirre (1, CNIN); Morelos, Tetela del Volcán, 23.VIII.1984, V. Butze (1, CNIN); Oaxaca, Hwy 175, 27 km NE Oaxaca, 29.VIII.1982, 8500', C. \& L. O’Brien, G. Wibmer (1, ASUCOB); Oaxaca, Hwy 175, 66 km NE Oaxaca, 29.VIII.1982, 8500', C. \& L. O’Brien, G. Wibmer (1, ASUCOB); Oaxaca, Carr. Oaxaca-Tuxtepex, bosque de pino-encino, en hojarasca, $2,296 \mathrm{~m}, 17^{\circ} 20^{\prime} 57.8^{\prime \prime} \mathrm{N}, 96^{\circ} 30^{\prime} 57.5^{\prime \prime} \mathrm{W}, 9 . V I .2018, \mathrm{M}$. Barrios (6, MZFC); Oaxaca, 2 mi. north San José Pacífico, 16.VII.1974, Clark, Murray, Ashe, Schaffner (4, CMNC); Oaxaca, 2 miles north San José Pacífico, 17.VII.1974, Clark, Murray, Ashe, Schaffner (2, TAMUIC); Oaxaca, 8 km S Suchixtepec, Río Molino, 19.VI.1979, 2,200 m, H. \& A. Howden (30, CMNC; 2, ASUCOB); Oaxaca 8 km S Suchixtepec, Río Molino, 6.VIII.1986, H. \& A. Howden (1, CMNC); Oaxaca, 10 km S Suchixtepec, 24.VII.1992, Roadside beating, $2,000 \mathrm{~m}$, R.S. Anderson (24, CMNC); Oaxaca, 1 mi. S Suchixtepec Hwy 175, 2.VI.1983, 9200', C. \& L. O'Brien, G. Marshall (2, ASUCOB); Puebla, 67 mi. E Puebla, 26.VI.1971, 7800', C.W. \& L. O'Brien, Marshall (1, CASENT); Puebla, 7 mi. N Zacapoaxtla, 12.VI.1983, 5000', C. \& L. O’Brien, G. Marshall (1, ASUCOB); Puebla, 6 mi. NE Zacatepec, 27.VI.1975, sifting leaf litter, D.S. Chanddler (1, ASUCOB); Puebla, 6 mi. NE Zacatepec, 27.VI.1975, L.E. Watrous (1, ASUCOB); Puebla, Zacatlán km 2 desv. a Piedras Encimadas, 6.VII.2006, $2,449 \mathrm{~m}, 20^{\circ} 01^{\prime} 07^{\prime \prime} \mathrm{N}, 98^{\circ} 04^{\prime} 43^{\prime \prime} \mathrm{W}$, L. Cervantes, D. Brzoska (1, CNIN); Veracruz, Hwy 140, 17 mi. NW Jalapa, 20.VIII.1982, 8000', C. \& L. O'Brien, G. Wibmer (1, ASUCOB). Isodacrys buchanani
MEXICO: Puebla, 6 mi. SE Tehuacán, 7.VII.1973, Mastro, Schaffner (2, TAMUIC; 1, CMNC); Tamaulipas, 4 mi. S San Fernando, Hwy 101, 5.V.1983, C. \& L. O’Brien, G. Marshall (1, ASUCOB). UNITED STATES OF AMERICA: Texas, Duval Co., 1 mi . W San Diego, 10.IV.1973, W.E. Clark (1, TAMUIC); Texas, 15 mi. SW Jct FR 3073 y Hwy 16, 10. IV.1987, B.F. \& J.L. Carr (1, CMNC; 2, CNC); Texas, Jim Hogg Co., 5 mi. W Hebbronville, 3.XI.1990, T. Carlow (3, TAMUIC); Texas, 37 mi. N Laredo, Webb Co., 15.X.1970, C.W. O'Brien (53, ASUCOB; 6, CMNC).

## Isodacrys burkei

UNITED STATES OF AMERICA: Texas, Brazos Co., 8.V.1956, H. R. Burke (1 paratype, TAMUIC); Texas, Brazos Co., 12.V.1960, H. R. Burke (1 paratype, TAMUIC); Texas, Brazos Co. Bastrop St. Pk., 24-27.V.1963, M. Kaulbars (2, CMNC); Texas, Brazos Co. Bastrop St. Pk., 5.VI.1989, Sweeping composites, R.S. Anderson (6, CMNC); Texas, Brazos Co., College Station, Lick Creek Park, 12.VI.1993, R. Jones (2, UAQE); Texas, College Station, Brazos Co., 15.IV.1963, H.R. Burke (20, CMNC); Texas, College Station, 3.V.1931, swept from weeds, H.J. Reinhard (1 paratype, TAMUIC); Texas, Brazos Co., on Hwy30 7.6
miles east jct. Hwy. 6, 7.V.1981, Sweeping Coreopsis sp., S. J. Merritt (1, TAMUIC); Texas, Brazos Co., Lick Ck. Pk., ca. 3 mi. S College Station, 18.X.1987, R.S. Anderson (4, CASENT; 50, CMNC); Texas, Brazos Co. Lick Creek Park ca. 5 mi. S College Station, 16.IV.1989, Sweeping flowers, R.S. Anderson (2, CMNC); Texas, Lee Co., 29.V.1959, S.D. \& R.H. Burke (1 paratype, TAMUIC); Texas, 0.5 mile west of Minter Springs, Brazos Co., 20.IV.1970, V.V. Board (1, TAMUIC); Texas, Leon Co. 9 km N Flynn, 28.V.1995, Quercus incanus, H. \& A. Howden (5, CMNC); Texas, Milano, Milam Co., 28.IV.1939, \#7315, On tomato plants, very numerous, eating stems particularly, causing plants to fall over (1 paratype, TAMUIC).

Isodacrys carlae
Holotype $\mathrm{Q}:$ : $77000^{\prime} \mathrm{nr}$. Jame,/33 mi. S.E. Saltillo,/Coah. Mex. VII.18'63/A. Howden Acacia] [Isodacrys/n.sp.] (CMNC). Paratypes (45 ¢ O ): [7000' nr. Jame,/33 mi. S.E. Saltillo,/Coah. Mex. VII.18'63/A. Howden Acacia] (1, CMNC); [7500', nr. Jame,/33 mi.S.E. Saltillo,/Coah. Mex. VII.18'63/A. T. Howden] (6, CMNC; 3, MZFC); [7500' nr. Jame,/ 33 mi S.E. Saltillo,/Coah. Mex. VII.25'63/A. T. Howden] (7, CMNC; 1, UAQE; 1 IEXA); [20 mi. S.E. Saltillo,/6000', Rt. 015, Coah./Mex., VI.20-21, 1971/H. F. Howden]; [MEXICO: Coahuila/12.4 mi S Saltillo/4-VII-1985, J. Woolley/G. Zolnerowich 85/023] [TAMUENTO/X0725914] (1, TAMUIC); [MEXICO: Coahuila/12.4 mi S Saltillo/4-VII-1985, J. Woolley/G. Zolnerowich 85/023] [TAMU-ENTO/X0725888] (1, TAMUIC); [MEXICO, Coah., Hwy./57, 15 mi. SE. Saltillo,/7000' 12Sept. 1982 C.\&/L. O'Brien \& G. Wibmer] (1, ASUCOB; 1, MZFC); [MEX., Coah., 10 mi./E. Saltillo 7000'/VIII-14-1971 C\&L/O’Brien \& Marshall] (1, ASUCOB]; [MEXICO, Hidalgo,/Municipio Metztitlán/1 km W. San Cristobal, $20.635^{\circ} \mathrm{N}, 98.846^{\circ} \mathrm{W}, / 15-\mathrm{VIII}-2017$, S.M. Clark] (1, ASUCOB); [MEXICO: Nuevo Leon/road to Galeana, 1/km. N jct. rte. 58/17 July 1988/R. Turnbow] (1, CMNC); [MEXICO: Nuevo Leon/9 mi. west Iturbide/July 3, 1974/Clark, Murray,/Ashe, Schaffner] [TAMU-ENTO/X0725413] (1, TAMUIC); [MEXICO: Nuevo Leon/9 mi. west Iturbide/ July 3, 1974/Clark, Murray,/Ashe, Schaffner] [TAMU-ENTO/X0724919] (1, TAMUIC); [MEXICO: Nuevo Leon/9 mi. west Iturbide/July 3, 1974/Clark, Murray,/Ashe, Schaffner] [TAMU-ENTO/X0725718] (1, IEXA); [MEXICO: Nuevo Leon/9 mi. west Iturbide/July 3, 1974/Clark, Murray,/Ashe, Schaffner] [TAMU-ENTO/X0726871] (1, TAMUIC);
[MEXICO: Nuevo Leon/9 mi. west Iturbide/July 3, 1974/Clark, Murray,/Ashe, Schaffner] [TAMU-ENTO/X0730294] (1, TAMUIC); [MEXICO: Nuevo Leon/9 mi. west Iturbide/ July 3, 1974/Clark, Murray,/Ashe, Schaffner] [TAMU-ENTO/X0726000] (1, UAQE); [MEX. N.L. $40 \mathrm{mi} . / \mathrm{NW}$ Jcn. Hwy 57\&60/VI-22-1971 C.W./O'Brien \& Marshall] (1, ASUCOB; 1, MZFC); [MEXICO, N.L., Hwy./58, 32 mi. W. Linares,/6000' 11Sept. 1982 C.\&/L. O’Brien \& G. Wibmer] (3, ASUCOB); [MEXICO, N.L., 11 km./NE. S. Antonio Peña/Nevada, 2,000 m. Aug16/1977A.N.Garcia A.] [beating pine and/miscellaneous] (1, ASUCOB); [MEX., N.L., $29 \mathrm{mi} . / \mathrm{SW}$. Linares 5700'/VIII-15-1971 C\&L/O'Brien \& Marshall] (1, ASUCOB); [MEXICO: San Luis Potosi/4 km. W Guadalcazar/18 July 1988/ R. Turnbow] [Isodacrys/geminatum Howd./det. R.S. Anderson 90] (1, CMNC); [MEXICO: Tamaulipas/19.6 mi. sw. Jaumave/18-VII-1973/Gaumer and Clark] [TAMU-ENTO/X0725801] [Isodacrys/sp./DET./A. Howden] (1, TAMUIC); [MEXICO:

Tamaulipas/19.6 mi. sw. Jaumave/18-VII-1973/Gaumer and Clark] [TAMU-ENTO/ X0727602] (1, TAMUIC); [MEXICO: Tamaulipas/19.6 mi. sw. Jaumave/18-VII-1973/ Gaumer and Clark] [TAMU-ENTO/X0725553] (1, TAMUIC); [MEXICO, Tam., Hwy./ 101, 28 mi. NE. Tula/4500' 22July 1982 CW\&/L. O'Brien \& G. Wibmer] (1, ASUCOB).

Isodacrys confusum
Holotype ơ: [MEXICO: Tamaulipas/16.3 mi. ne. Jaumave/18-VII.1973/Gaumer and Clark] [taken on/Mimosa/monancistra] [TAMU-ENTO/X0729035] (TAMUIC); paratypes ( 8 ƠƠ', 28 ¢ ¢ ) : [MEXICO, Tam., Hwy 101/18 mi. SW. Cd. Victoria, $/ 4300^{\prime}$ 22July1982 C.W.\&/L. O’Brien \& G. Wibmer] ( 1 O', 1 \&, ASUCOB; 1 O', MZFC); [MEXICO, Tam., Hwy 101/24 mi. SW. Cd. Victoria,/2150' 23July1982 C.W.\&/L. O’Brien \& G. Wibmer] ( 1 Ơ, 2 Q, ASUCOB); [MEXICO, Tam., Hwy 101/22 mi. SW. Cd. Victoria,/ 3000' 22July1982 C.W.\&/L. O’Brien \& G. Wibmer] ( 1 ơ, 1 \&, ASUCOB); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/ X0726618] (1 \&, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/ same data as holotype] [TAMU-ENTO/X0727115] (1 \&, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/ X0725169] (1 \&, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/ same data as holotype] [TAMU-ENTO/X0724605] ( 1 O , TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/ X0725410] (1 \& , TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/ same data as holotype] [TAMU-ENTO/X0726220] (1 \&, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/ X0726502] (1 \&, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/ same data as holotype] [TAMU-ENTO/X0727010] ( 1 ơ, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/ X0726280] (1 \&, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/ same data as holotype] [TAMU-ENTO/X0725437] (1 \&, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/ X0725446] (1 \&, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/ same data as holotype] [TAMU-ENTO/X0725407] (1 \&, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/ X0725141] (1 \&, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/ same data as holotype] [TAMU-ENTO/X0725168] (1 $\uparrow$, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/ X0725412] (1 \&, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/ same data as holotype] [TAMU-ENTO/X0726973] (1 \&, IEXA); [MEXICO: Tamaulipas/ same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0725112] (1 \&, IEXA); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0725393] (1 \& , UAQE); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0729284] (1 \&, UAQE); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0728579] (1 Ơ, TAMUIC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [Isodacrys/sp./DET./A. Howden]
[TAMU-ENTO/X0728911] (1 \& , MZFC); [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [TAMU-ENTO/X0728902] [ơ] (1 Ơ, MZFC); [MEXICO: Tamaulipas/same data as holotype] [TAMU-ENTO/X0726515]; [MEXICO: Tamaulipas/same data as holotype] [taken on/same data as holotype] [Isodacrys/ geminatum/Howden/det. R.S. Anderson 1981] (1 O, CMNC); [MEXICO: Tamaulipas/ same data as holotype] [taken on/same data as holotype] ( $10^{\prime \prime}, 3 申 \subset$, CMNC).

Isodacrys crispum
MEXICO: Durango, 25 mi . W Durango, 10.VI.1964, Oak, H. Howden (5, CMNC); Durango, 25 mi. W Durango, 29.VI.1964, H. Howden (30, CMNC; 2 ASUCOB); Durango, 25 mi. W Durango, 3.VII.1964, H. \& A. Howden (3, CNC); Durango, 28 mi. W Durango, 9.VI.1967, Quercus clivicola F. consanguinea, C.H. Maen (1, CMNC); Durango, 25 mi . W Durango, 18.VIII.1974, on Quercus, 7800', C.W. \& L. O'Brien, Marshall (2, UAQE); Durango, 25 mi. W Durango, 18.VIII.1974, on pine/on Quercus, 7800', C.W. \& L. O'Brien, Marshall (34, ASUCOB); Durango, 26.5 mi . SW Durango, 21.VII.1982, General sweeping, Fred. G. Andrews (9, CMNC); Zacatecas, 27 mi NW Fresnillo, Hwy 45, 17.VIII.1974, on Quercus, 7800 ', C.W. \& L. O'Brien, Marshall (32, ASUCOB).

Isodacrys ellipticum
GUATEMALA: Zacapa, San Lorenzo Quarry Road, 3-7 km N Sta. Cruz, CA9, 17.VII.2008, Dry forest, C. \& L. O'Brien, F. Skillman (1, ASUCOB). HONDURAS: Comayagua, Comayagua, 2.VI.1978, Gary V. Manley (1, CMNC); Cortés, 5 km N Cofradía, 5.VIII.1977, C. \& L. O’Brien, G. Marshall" (41, ASUCOB); Francisco Morazán, Distrito Central, 5 mi. S Tegucigalpa, 9.VI.1974, night, 4000', C. \& L. O'Brien, G. Marshall (3, CASENT); Francisco Morazán, 17 mi. NE Talanga, 16.VI.1974, 2900', C. \& L. O'Brien, G. Marshall (129, ASUCOB; 3, CASENT); Francisco Morazán, 23.8 km SW Talanga, 3.VI.1993, R. Turnbow (1, CMNC); Francisco Morazán, Tamara valley, 5.X.1993, R. Turnbow (1, CMNC).

## Isodacrys fasciatum

Holotype $\uparrow$ : [MEX., Coah., 10 mi./E. Saltillo 7000'/VIII-14-1971 C\&L/O’Brien \& Marshall] (ASUCOB). Paratypes ( 11 ¢ ¢ ) : [MEX., Coah., $10 \mathrm{mi} . / \mathrm{same}$ data as holotype] (2, ASUCOB); [MEXICO: Coahuila/12.4 mi S Saltillo/4-VII-1985, J. Woolley/G.
Zolnerowich 83/023] [TAMU-ENTO/X0727876] (1, TAMUIC); [MEXICO, Dgo., 22 mi // NE. Durango, 6200'/Aug. 20, 1974 C.W.\&L./O'Brien \& Marshall] (7, ASUCOB); [MEX. N. L. $10 \mathrm{mi} / / \mathrm{S}$. Jcn. 57 \& 60/VI-23-1971 L\&CW/O’Brien \& Marshall] (1, ASUCOB). Isodacrys frontalis
Holotype $\uparrow:$ [MEX.: OAX.; $20 \mathrm{mi} \mathrm{S} /$ Juchatengo, $6000^{\prime} / 29 . V .1971$, S. Peck/Ber207, oak litter] (CMNC). Paratypes (23 ¢): [GUATEMALA: Sacatepéquez/5 km SE Antigua/ 14.53495-90.69367 $\pm 50 \mathrm{~m} / 2,150 \mathrm{~m}$, 10.VI.2009/LLAMA Wa-B-08-1-49] [WORLD/ WEEVIL/DATABASE/WWD0041418] (1 CMNC); [GUATEMALA: Sacatepéquez/5 km SE Antigua/14.53495-90.69367 $\pm 50 \mathrm{~m} / 2,150 \mathrm{~m}$, 10.VI.2009/LLAMA Wa-B-08-1-49] [WORLD/WEEVIL/DATABASE/WWD0041419] (1, CMNC); [GUATEMALA: Sacatepéquez/5 km SE Antigua/14.53495-90.69367 $\pm 50 \mathrm{~m} / 2,150 \mathrm{~m}, 10$. VI.2009/LLAMA Wa-B-08-1-49] [WORLD/WEEVIL/DATABASE/WWD0041420] (1, CMNC); [GUATEMALA: Sacatepéquez/5 km SE Antigua/14.53510-90.69384 $\pm 50 \mathrm{~m} / 2,150 \mathrm{~m}$, 10.VI.2009/LLAMA Wa-B-08-1-44] [WORLD/WEEVIL/DATABASE/WWD0041311]
[Isodacrys/GUA 1/det. R.S. Anderson, 20] (1, CMNC); [GUATEMALA: Sacatepéquez/ 5 km SE Antigua/ $14.53529-90.69404 \pm 50 \mathrm{~m} / 2,150 \mathrm{~m}$, 10.VI.2009/LLAMA Wa-B-08-138] [WORLD/WEEVIL/DATABASE/WWD0041144] (1, CMNC); [GUATEMALA: Sacatepéquez/5 km SE Antigua/14.53504-90.69377 $\pm 50 \mathrm{~m} / 2,150 \mathrm{~m}, 10$. VI.2009/LLAMA Wa-B-08-1-46] [WORLD/WEEVIL/DATABASE/WWD0041358] (1, CMNC); [GUATEMALA: Sacatepéquez/5 km SE Antigua/14.53504-90.69377 $\pm 50 \mathrm{~m} / 2,150 \mathrm{~m}$, 10.VI.2009/LLAMA Wa-B-08-1-46] [WORLD/WEEVIL/DATABASE/WWD0041359] (1, CMNC); [GUATEMALA: Sacatepéquez/5 km SE Antigua/14.53504-90.69377 $\pm 50 \mathrm{~m} /$ 2,150 m, 10.VI.2009/LLAMA Wa-B-08-1-46] [WORLD/WEEVIL/DATABASE/ WWD0041360] (1, CMNC); [GUATEMALA: Sacatepéquez/5 km SE Antigua/ 14.53504-90.69377 $\pm 50 \mathrm{~m} / 2,150 \mathrm{~m}, 10$. VI.2009/LLAMA Wa-B-08-1-46] [WORLD/ WEEVIL/DATABASE/WWD0041361] (1, CMNC); [GUATEMALA: Sacatepéquez/5 km SE Antigua/14.53504-90.69377 $\pm 50 \mathrm{~m} / 2,150 \mathrm{~m}, 10 . \mathrm{VI} .2009 / L L A M A ~ W a-B-08-1-46]$ [WORLD/WEEVIL/DATABASE/WWD0041363] (1, CMNC); [GUATEMALA: Sacatepéquez/5 km SE Antigua/14.53507-90.69380 $\pm 50 \mathrm{~m} / 2,150 \mathrm{~m}, 10 . \mathrm{VI} .2009 /$ LLAMA Wa-B-08-1-45] [WORLD/WEEVIL/DATABASE/WWD0041326] (1, CMNC); [GUATEMALA: SACATEPEQUEZ/Guatemala City, Cerro/Alux, 2,260 m., 11.VI.1991/ R. Anderson, wet oak/forest, 91-60] (1, CMNC); [GUATEMALA, 9 mi.W./Cd. Guatemala 6900'/VI-7-1974C.W. \&L./O’Brien \&Marshall] (2, ASUCOB). [MEX.: OAX, same data as holotype] (4, CMNC); [MEXICO: Oaxaca $3.2 \mathrm{~km} . / \mathrm{S}$. San Jose del Pacifico/2,440 m, 22.VII.1992, J.S./Ashe,oak-pine for. litt.] (1, CMNC); [MEXICO: Oaxaca, $5.1 \mathrm{~km} . / \mathrm{S}$. Suchixtepec, 2,150 m/24.VII.1992,92-024,R.S./Anderson, oak|alder|pine/for., leaf litter Berlese] (3, CMNC); [MEXICO: Oaxaca, $23 \mathrm{~km} . / \mathrm{S} . \mathrm{W}$. Valle Nacional, KM76/1,300 m, 26.VII.1992,92-029/R.S. Anderson, cloud forest/leaf litter Berlese] (1, CMNC).

Isodacrys geminatum
MEXICO: Mexico City, Mpio. Xilotepec, Hwy57, 25.VII.2006, oak forest litter, 2,700 m, $20^{\circ} 04^{\prime} .797^{\prime} \mathrm{N}, 99^{\circ} 37.906^{\prime} \mathrm{W}$, R.S. Anderson (1, CMNC); Oaxaca, 22 km NE Oaxaca, 18.VI.1979, 2,600 m, H. \& A. Howden (1, CMNC); Oaxaca, 3 mi. N Suchixtepec, 4.VI.1971, Ber209, oaklitter, $9500^{\prime}$, S. Peck (1, CMNC); Puebla, 6 mi. NE Zacatepec, 27.VI.1975, L.E. Watrous (9, ASUCOB); Querétaro, Mpio. Colón, Cerro Zamorano, 26. VII.2006, oyamel/oak forest litter, $3,100 \mathrm{~m}, 20^{\circ} 55.967^{\prime} \mathrm{N}, 100^{\circ} 11.021^{\prime} \mathrm{W}$, R.S. Anderson (1, CMNC); Querétaro, Mpio. Huimilpan, 1 km NW La Beata, 2.XII.2003, sample \#6, leaf litter, $2,500 \mathrm{~m}, 20^{\circ} 28.42^{\prime} \mathrm{N}, 100^{\circ} 14.49^{\prime} \mathrm{W}$, R. Jones (1, UAQE); Tlaxcala, 6.8 km N Taxco, Hwy 119, 10.VII.1992, ex. misc. Mushrooms, 2,820 m, J.S. Ashe \#36 (1, CMNC).

Isodacrys guatemalenum
GUATEMALA: Sacatepéquez, Antigua, 16.VIII.1947, 5000', C. \& P. Vaurie" (1, CMNC); Sacatepéquez, Santa Lucía Milpas Altas, 24.VI.1993, 1,800 m, B.D. Gill (1, CMNC). HONDURAS: Choluteca, 1 km W San Marcos de Colón, 23.VII.1977, C. \& L. O'Brien, G. Marshall (12, ASUCOB); Comayagua, Comayagua, 18.VII.1977, J.V. Mankins (2, CMNC); Comayagua, 22 km N Comayagua, 18.VII.1977, C. \& L. O’Brien, G. Marshall (80, ASUCOB; 12, CASENT).

## Isodacrys kuchii

Holotype ơ: [MEXICO: Puebla/Mpio: Zapotitlán Salinas/ 1 km N San Juan Raya] [ $18^{\circ} 19^{\prime} 11^{\prime \prime}, 97^{\circ} 36^{\prime} 50^{\prime \prime} / 19$-vii-2011, A. Obregón/1,755 m, Matorral] [Isodacrys/ geminatum/Howden 1961/Det. Jesús Luna Cozar 2015] (UAQE). Paratypes (1 Ơ, 1 Q): [MEXICO: Puebla/same data as holotype] [ $18^{\circ} 19^{\prime} 11^{\prime \prime}, 97^{\circ} 36^{\prime} 50^{\prime \prime} /$ same data as holotype] [Isodacrys/same data as holotype] (1 O", UAQE); [MEXICO: Puebla/6 Mi. SW. Tehacan/ July 8, 1973/Schaffner \& Mastro] [TAMU-ENTO/X0725504] [Isodacrys/?geminatum/ DET./A. Howden] (1 \&, TAMUIC).

## Isodacrys mexicanum

MEXICO: Aguascalientes, 24 miles West of Aguascalientes, 5.VI.1956, B. Banta (1, CASENT); Mexico City, Milpa Alta, 26.VII.1980, H. Brailovsky (1, CNIN); Mexico City, Pedregal de San Ángel, Loc. 21, 22.VIII. 1961 (2, CNIN); Mexico City, Zumpango, 18.VII.1981, Peter Jump (1, CMNC; 1, TAMUIC); Michoacán, km 154 car. IrapuatoGuadalajara, 26.VII.1984, M. García" (1, CNIN); Morelos, Tepoztlán, 11.X.1968, R. Muñíz V. (1, CMNC; 1, ASUCOB); Morelos, YMCA camp. Tepoztlán, 21.VIII.1958, H.F. Howden (4, CMNC); Oaxaca, 1.1 mi . W El Tule, 87/048, 5400', Woolley, Zolnerowich (2, TAMUIC).
Isodacrys obrienorum
Holotype ơ: [GUATEMALA, $12 \mathrm{mi} . S E . /$ San Cristobal de To-/tonicapan, $9800^{\prime} /$ VII-28.1974] [under stones] [C.W. \& L.B. O'Brien/\& G.B. Marshall] (ASUCOB). Paratypes ( 6 ƠƠ, 6 O Q P): [GUATELAMA: Jalapa/4 km E Mataquescuintla/14.52705-90.14671 $\pm 55 \mathrm{~m} /$ 2,660 m, 2.VI.2009, /LLAMA \#Go-B-07-1-04/cloud forest beating vegetation] [WORLD/ WEEVIL/DATABASE/WWD0125181] (1 \&, CMNC); [GUATELAMA: Jalapa/4 km E Mataquescuintla/14.52705-90.14671 $\pm 55 \mathrm{~m} / 2,660 \mathrm{~m}, 2$. VI.2009,/LLAMA \#Go-B-07-1-04/ cloud forest beating vegetation] [WORLD/WEEVIL/DATABASE/WWD0125188] ( 1 ơ, CMNC); [GUATELAMA: Jalapa/4 km E Mataquescuintla/14.52705-90.14671 $\pm 55 \mathrm{~m} /$ $2,660 \mathrm{~m}, 2 . \mathrm{VI} .2009, / L L A M A$ \#Go-B-07-1-04/cloud forest beating vegetation] [WORLD/ WEEVIL/DATABASE/WWD0125177] ( $1 \mathrm{o}^{\prime}, \mathrm{CMNC}$ ); [GUATEMALA, 12 mi .SE./San Cristobal deTo-/tonicapan, same data as holotype] ( 3 ƠƠ, $^{\prime} 4 \bigcirc \bigcirc$, ASUCOB); [GUATEMALA, 10 mi.NW./SanCristobal de/Totonicapan, 9300'] [VII-29-1974C.W.\&L./ O'Brien \& Marshall] [under stones/and dead/grass clumps] (1 \&, ASUCOB); [GUATEMALA: San Marcos/above Sibinal/3,009 m, 15.14366-92.06662/8 Jun 2015, R. Anderson/alder litter, 2015-127] ( 1 O", CMNC).

Isodacrys okuiltontli
Holotype ơ: [MEXICO, Oax., Hwy131/5 mi. N. Telixtlahuaca/6200' 31Aug. 1982 CW. \&/L. O’Brien \& G. Wibmer] (ASUCOB). Paratypes ( 5 ¢ $\uparrow$ ): [MEXICO, Oax, 19.5/km. NW. Huitzo, Hwy./190, 2,260 m, 9.VII./1992 C.L. Bellamy] (1 © , ASUCOB); [MEXICO, Oax., Hwy131/same data as holotype] ( 3 \& $\uparrow$, ASUCOB); [MEXICO, Oax., Hwy131/same data as holotype] [Isodacrys/minutus/Sharp/det. C. W. O’Brien, 1999] (1 \&, ASUCOB).

Isodacrys orizabae
MEXICO: Oaxaca, 1.8 km S El Tule, 16.VII.1987, dry Acacia scrub, 1,524 m, R.S. Anderson (1, CMNC); Oaxaca, 20 km E Mitla, 3.VIII.1986, $1,800 \mathrm{~m}, \mathrm{H} . \&$ A. Howden (2, CMNC); Oaxaca, Hwy 175, 10 km NE Oaxaca, 16.VI.1979, 1,800 m, H. \& A. Howden
（1，CMNC）；Oaxaca， 5 mi．SW Oaxaca，4．VIII．1983，Acacia thorn scrub，4700＇， R．S．Anderson，W．Maddison（9，CMNC）；Oaxaca， 13 mi ．SE Tlalcolula，2．VI．1974，6100＇ C．\＆L．O’Brien，G．Marshall（4，ASUCOB）；Puebla， 4.4 mi．SW Acatepec，26．VII．1974， Clark，Murray，Ashe，Schaffner（5，CMNC；6，TAMUIC）；Puebla， 6 km S Acatepec， 19．VIII．1982，Clark，Cave（1，ASUCOB）；Veracruz，VIII．1942，Koebele（2，CMNC）； Veracruz，Hwy 150， 31 km W Orizaba，15．VIII．1975，beating Pine，1，880 m，B．García G． （1，ASUCOB）．

## Isodacrys ovipennis

MEXICO：Tamaulipas，Hwy101 11 mi．SW Cd．Victoria，22．VII．1982，4000＇，C．\＆ L．O’Brien，G．Wibmer（37，ASUCOB）；Tamaulipas， 2 mi．NE San Fernando，Hwy 101， 31．VII．1970，C．W．O’Brien（1，ASUCOB）；Tamaulipas， 4 mi．S San Fernando，Hwy 101， 5．V．1983，C．\＆L．O＇Brien，G．Marshall（2，ASUCOB）．UNITED STATES OF AMERICA： Texas，Cameron Co．Laguna Atascosa Nat．Wild．Ref．rd．to Laguna Madre，20．X．1989， sweeping Palafoxia texana var．ambigua，R．S．Anderson，E．Riley（13，CMNC）；Texas， Cameron Co．Laguna Atascosa Wdlf．Ref，20．X．1989，E．G．Riley（5，ASUCOB）；Texas， Encino，13．III．1966，Modiola caroliniana，M．F．Schuster（1，CMNC）；Texas，Kenedy Co． Los Olmos Creek at \＃77， 2 mi．S Riviera，20．X．1989，sweeping Palafoxia texana var． ambigua，R．S．Anderson，E．Riley（13，CMNC；1，CASENT）；Texas，Kenedy Co． 10 mi. N Norias，10．X．1989，D．J．Heffern（1，CMNC）；Texas，Edinburg，3．VI．1962，W．L．Sterling （1，TAMUIC）；Texas，Kingsville，Kleberg Co．，25．VII．1951，H．R．Burke（1，TAMUIC）； Texas， 10 miles NW La Gloria，6．XI．1989，W．F．Chamberlain（1，TAMUIC）；Texas，Kleberg Co．，Jct．Los Olmos Creek \＆Hwy 77，19．X．1990，T．Carlow，E．Riley（2，TAMUIC）；Texas， Palmitto Hill， 10 mi ．W Boca Chica，13．X．1985，R．Turnbow（1，CMNC）．
Isodillex minutum
MEXICO：MICHOACÁN， 2 mi．S Carapan，6．VII．1985，Woolley，Zolnerowich＂ （ 5 specimens，TAMUIC； 3 specimens，CMNC），＂MEXICO：MICHOACÁN， 18.5 km E Morelia，28．VII．1988，Pine／oak／Acacia，2，220 m，R．S．Anderson＂（1 specimen，CMNC）， ＂MEXICO：MICHOACÁN， 17 mi ．W Pátzcuaro，14．VIII．1982，7700＇，C．\＆L．O＇Brien， G．Wibmer＂（1 specimen，ASUCOB）．

## Isodillex plumosum

Holotype（ $\mathrm{O}^{r}$ ）：［61 mi．W．Fresnillo／Zacatecas，MEX／8000 ft．VI－25－54／R．H．Brewer］ ［III\＃3］［Isodacrys／n．sp．\＃2／AH ‘62］［CASENT／8424336］．Paratypes（3 ƠƠ and 11 O O ¢）：［61 mi．W．Fresnillo／same data as holotype］［III\＃3］［Cal Acad．］［CASENT／8424322］（ 1 O＂ CASENT）；［61 mi．W．Fresnillo／same data as holotype］［III\＃3］［CASENT／8424323］（1 ơ CASENT）；［61 mi．W．Fresnillo／same data as holotype］［III\＃3］［CASENT／8424331］（1 ơ CASENT）；［61 mi．W．Fresnillo／same data as holotype］［III\＃3］［Cal Acad．］［Isodacrys／ n．sp．\＃2］［CASENT／8424321］（1 O CASENT）；［61 mi．W．Fresnillo／same data as holotype］ ［III\＃3］［CASENT／8424324］（ 1 甲 CASENT）；［61 mi．W．Fresnillo／same data as holotype］ ［III\＃3］［CASENT／8424325］（1 \＆CASENT）；［61 mi．W．Fresnillo／same data as holotype］ ［III\＃3］［CASENT／8424326］（ 1 ¢ CASENT）；［61 mi．W．Fresnillo／same data as holotype］ ［III\＃3］［CASENT／8424327］（ 1 甲 CASENT）；［61 mi．W．Fresnillo／same data as holotype］ ［III\＃3］［CASENT／8424328］（1 ¢ CASENT）；［61 mi．W．Fresnillo／same data as holotype］ ［III\＃3］［CASENT／8424329］（1 \＆CASENT）；［61 mi．W．Fresnillo／same data as holotype］
[III\#3] [CASENT/8424330] (1 ¢ CASENT); [61 mi. W. Fresnillo/same data as holotype] [III\#3] [CASENT/8424332] (1 Q CASENT); [61 mi. W. Fresnillo/same data as holotype] [III\#3] [CASENT/8424333] (1 Q CASENT); [61 mi. W. Fresnillo/same data as holotype] [III\#3] [CASENT/8424334] (1 Q CASENT); [61 mi. W. Fresnillo/same data as holotype] [III\#3] [CASENT/8424335] (1 ¢ CASENT).

## New Species Registration

The following information was supplied regarding the registration of a newly described species:

Publication LSID: urn:lsid:zoobank.org:pub:F5D5C6F7-3FB4-4D1F-91F3-0D6380C8FF59 Isodacrys antrum LSID: urn:lsid:zoobank.org:act:FBE7F34A-9C93-4BDC-94233D3A915B926A.

Isodacrys carlae LSID: urn:lsid:zoobank.org:act:146A1489-52AE-477D-A30486136B5880BA.

Isodacrys confusum LSID: urn:lsid:zoobank.org:act:3EDF5293-38FD-4217-8730164074806BAA.

Isodacrys fasciatum LSID: urn:lsid:zoobank.org:act:881D4538-9CB8-49E5-9641CCF3973623E1.

Isodacrys frontalis LSID: urn:lsid:zoobank.org:act:DB24A34B-7EF1-44AD-8DFD5823FC083899.

Isodacrys kuchii LSID: urn:lsid:zoobank.org:act:CB805B2D-80BA-4B36-9B6D90740 B 222917.

Isodacrys obrienorum LSID: urn:lsid:zoobank.org:act:9DB9D7AA-F4AB-4424-A8103DD5BF6ABA56.

Isodacrys okuiltontli LSID: urn:lsid:zoobank.org:act:01FC3BDC-D6AC-48D1-BF6A60CF65BB6626.

Isodillex LSID: urn:lsid:zoobank.org:act:2AD213BD-E6DE-4146-A578-426D26C2002E. Isodillex plumosum LSID: urn:lsid:zoobank.org:act:955038E3-B522-4A22-9149896ED0092C6B.

## Supplemental Information

Supplemental information for this article can be found online at http://dx.doi.org/10.7717/ peerj.10191\#supplemental-information.

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