

# Regulation of the Cardiac Sodium/Bicarbonate Cotransporter by Angiotensin II: Potential Contribution to Structural, Ionic and Electrophysiological Myocardial Remodelling

Ernesto Alejandro Aiello\* and Verónica Celeste De Giusti

Centro de Investigaciones Cardiovasculares, Facultad de Ciencias Médicas, Universidad Nacional de La Plata, La Plata, Argentina.

**Abstract:** The sodium/ bicarbonate cotransporter (NBC) is, with the  $\text{Na}^+/\text{H}^+$  exchanger (NHE), an important alkalinizing mechanism that maintains cellular intracellular pH ( $\text{pH}_i$ ). In the heart exists at least three isoforms of NBC, one that promotes the co-influx of 1 molecule of  $\text{Na}^+$  per 1 molecule of  $\text{HCO}_3^-$  (electroneutral isoform; nNBC) and two others that generates the co-influx of 1 molecule of  $\text{Na}^+$  per 2 molecules of  $\text{HCO}_3^-$  (electrogenic isoforms; eNBC). In addition, the eNBC generates an anionic repolarizing current that modulate the cardiac action potential (CAP), adding to such isoforms the relevance to modulate the electrophysiological function of the heart. Angiotensin II (Ang II) is one of the main hormones that regulate cardiac physiology. The alkalinizing mechanisms (NHE and NBC) are stimulated by Ang II, increasing  $\text{pH}_i$  and intracellular  $\text{Na}^+$  concentration, which indirectly, due to the stimulation of the  $\text{Na}^+/\text{Ca}^{2+}$  exchanger (NCX) operating in the reverse form, leads to an increase in the intracellular  $\text{Ca}^{2+}$  concentration. Interestingly, it has been shown that Ang II exhibits an opposite effect on NBC isoforms: it activates the nNBC and inhibits the eNBC. This inhibition generates a CAP prolongation, which could directly increase the intracellular  $\text{Ca}^{2+}$  concentration. The regulation of the intracellular  $\text{Na}^+$  and  $\text{Ca}^{2+}$  concentrations is crucial for the cardiac cellular physiology, but these ions are also involved in the development of cardiac hypertrophy and the damage produced by ischemia-reperfusion, suggesting a potential role of NBC in cardiac diseases.

**Keywords:** Angiotensin II, calcium overload, cardiac arrhythmias, cardiac hypertrophy, electrogenic sodium/bicarbonate cotransporter, electroneutral sodium/bicarbonate cotransporter, sodium overload.

## INTRODUCTION

The fine regulation of intracellular pH ( $\text{pH}_i$ ) is essential for the heart. Fluctuations of  $\text{pH}_i$  occur physiologically in cardiac myocytes, as during changes in heart rate [1,2], but also a major decrease can occur during pathological conditions, such as myocardial ischemia [3,4]. Four sarcolemmal ion transporters regulate  $\text{pH}_i$  homeostasis in order to maintain its value near to 7.2 and prevent the adverse effects of large fluctuations in  $\text{pH}_i$ . Two of these transporters mediate acid-loading, the  $\text{Cl}^-/\text{HCO}_3^-$  exchanger (anion exchanger, AE) and the  $\text{Cl}^-/\text{OH}^-$  exchanger (CHE). On the other hand, two other transporters mediate the acid-extrusion, either exporting  $\text{H}^+$ , the  $\text{Na}^+/\text{H}^+$  antiporter (NHE), or introducing  $\text{HCO}_3^-$  into the cell, the  $\text{Na}^+/\text{HCO}_3^-$  symporter (NBC).

In the present review we will specifically outline the importance of NBC in the maintenance of the cardiomyocytes  $\text{pH}_i$ . Also, due to the relevance of Angiotensin II (Ang II) in heart function, we will review the regulation of NBC by this hormone. Finally, we will recap the knowledge about the impact of NBC on intracellular  $\text{Na}^+$  ( $[\text{Na}^+]_i$ ) and  $\text{Ca}^{2+}$  concentrations ( $[\text{Ca}^{2+}]_i$ ), emphasizing the potential relevance of NBC in structural and electrical cardiac disorders.

\*Address correspondence to this autor at the Centro de Investigaciones Cardiovasculares, Facultad de Ciencias Médicas, UNLP., Calle 60 y 120, 1900, La Plata, Argentina; Tel/Fax: 54-221-483-4833; E-mail: [aaiello@med.unlp.edu.ar](mailto:aaiello@med.unlp.edu.ar)

## ROLE OF THE CARDIAC NBC IN $\text{pH}_i$ AND $[\text{Na}^+]_i$ REGULATION

Although the physiological role of NHE as an alkalinizing mechanism has been well demonstrated [5,6], that of NBC has long been underestimated, sometimes because the investigations were carried out in  $\text{HCO}_3^-$  free-buffered solutions. Another issue is that until few years ago, there was not available a selective NBC inhibitor. Fortunately in 2008 the group of Dr. Vaughan-Jones presented and characterized a novel and selective NBC inhibitor [7], that has been successfully used to demonstrate the importance of total NBC activity in the control of cardiac  $\text{pH}_i$  [8].

At present it is known that NBC is responsible for 40-50% of total acid extrusion in cardiac myocytes [9,10]. Moreover, although at acidic  $\text{pH}_i$  (near to 6.8) the relative importance of NBC is only of 30% against the 70% of NHE [8,11,12], both transporters are equally operative at  $\text{pH}_i$  closed to basal [8,13,14].

Interestingly, it has been demonstrated that NBC increase the  $[\text{Na}^+]_i$ , being responsible for 30% of this increase at  $\text{pH}_i$  6.8 [12]. The increase in  $[\text{Na}^+]_i$  stimulates the reverse mode of  $\text{Na}^+/\text{Ca}^{2+}$  exchanger (NCX), leading to an increase in  $[\text{Ca}^{2+}]_i$  [15-17]. This process is involved in Ang II and Endothelin-1 (ET-1) -induced positive inotropic effects [17,18] and cardiac hypertrophy [19,20]. Furthermore, this phe-

nomenon might be involved in NBC-induced cardiac pathologies [21].

### **ELECTRONEUTRAL (nNBC) AND ELECTROGENIC (eNBC) ISOFORMS OF NBC IN THE HEART**

Cardiac NBC was initially described by Lagadic-Gossmann *et al.* as an electroneutral transporter (nNBC), with a stoichiometry of 1 Na<sup>+</sup>/1 HCO<sub>3</sub><sup>-</sup> [9]. Some years later Dr. Cingolani's group demonstrated that NBC exhibits an electrogenic behavior (eNBC), with a stoichiometry of 1 Na<sup>+</sup>/2 HCO<sub>3</sub><sup>-</sup> [10]. In addition, we have later described and characterized the eNBC current as an anionic bicarbonate and sodium-dependent current which reversed at around -85 mV ( $I_{NBC}$ ) [22,23]. Moreover, the functional diversity of the eNBC in ventricular myocytes from rat, rabbit and guinea pig has been described in detail by Yamamoto *et al.* [24].

The controversy around the cardiac NBC stoichiometry has been resolved and it is now accepted that both mechanisms are present in cardiac cells [13,25]. In the heart exist at least two electrogenic isoforms, called NBCe1 (also named NBC1) and NBCe2 (also named NBC4) which are encoded by the SLC4A4 gene [26], and the SLC4A5 gene [27], respectively, and one electroneutral isoform, named NBCn1 (also named NBC3), that is encoded by the SLC4A7 gene [28].

We have described the influence of eNBC in the configuration of the cardiac action potential (CAP) [27]. Using the patch-clamp technique, we have demonstrated that the change of the extracellular solution from a HEPES- (HCO<sub>3</sub><sup>-</sup>-free solution) to a HCO<sub>3</sub><sup>-</sup>-containing solution hyperpolarized resting membrane potential (RMP) by 3-5 mV and evoked a 25% CAP shortening, both in rat [22] and cat [23] ventricular myocytes. Reciprocally, it has been shown that eNBC increased pH<sub>i</sub> in response to the change in RMP induced by hiperkalemic extracellular solutions [10,24,29] or after increasing the heart rate [30].

While the roles of NBCe1 and NBCn1 have been well established, the true relevance of the NBCe2 is still unresolved. Although it was initially reported to be present in the heart, the existence of NBCe2 in the cardiomyocytes was recently challenged [31,25]. Moreover the NBCe1 seems to be the only active electrogenic mechanism in normal cat and rat ventricular myocytes [25,32]. We have also recently reported that this NBC isoform is physically and functionally coupled to the carbonic anhydrase in rat ventricular myocytes [33].

A novel contribution to the knowledge about NBC was presented by us last year [25]: We have produced and characterized two different and selective functional antibodies against the extracellular loops of NBCe1, that were called a-L3 and a-L4, which recognized the extracellular loop 3 and loop 4, respectively. The pre-incubation of the myocytes with a-L3 canceled NBCe1 function, which allowed us to demonstrate that this isoform was the main, if not the only, electrogenic active isoform in normal cardiac myocytes. On the other hand, the pre-incubation with a-L4 improved the NBCe1 activity [25]. Inhibitory antibodies of NBCe1 have been previously used to investigate the implication of this isoform in the contractile dysfunction induced by ischemia-

reperfusion [34]. However, a direct activator of NBCe1 has never been used before our work. Nevertheless, besides that NBCe1 seems to be the only active electrogenic isoform in cardiac myocytes under physiological conditions, it still unknown the relevance of each NBC isoform during the development and progress of cardiac pathology. Thus, we proposed these antibodies as new pharmacological tools that will allow us to investigate the participation of NBCe1 in isolation in cardiac pathophysiology.

### **SYSTEMIC AND LOCAL ANG II IN THE HEART**

Ang II is an important hormone that regulates the excitation and contraction in the heart. Ang II is an octapeptide that was classically known to be synthesized from Ang I by the angiotensin-converting enzyme (ACE) present in the endothelial vessels in response to increases levels of Aldosterone (Ald), conforming the endocrine system known as renin-angiotensin-aldosterone-system (RAAS). Moreover, at present it is well recognized that Ang II is produced and secreted in several tissues, including the heart [35]. Dr. Sadoshima's group has shown for the first time that Ang II exerts autocrine and paracrine effects when it is secreted from intracellular vacuoles in response to myocyte stretching, leading to cardiac hypertrophy [36,37]. Furthermore, Dr. Cingolani's group have deeply investigated the presence of this autocrine pathway in the slow force response (SFR) to myocardial stretch, proposing the NHE stimulation as the final effect triggered by the endogenous Ang II action [38,39]. Furthermore, it was shown that >75% of cardiac Ang II was synthesized locally, and that its source was also in situ-synthesized Ang I [40]. In concordance, it has been demonstrated that Ald synthase exist in the myocyte [41], supporting the existence of a local RAAS [42]. On the other hand, it is important to mention that, under pathological conditions, like post-myocardial infarction and in response to pressure and volume overload, increased cardiac Ang II levels [43-45] and upregulation of AT-1 receptors [46] were reported.

It is well-known that Ang II effects involve MAP kinases (MAPK) stimulation and reactive oxygen species (ROS) generation [47-50]. In this regard, it has been demonstrated that low concentration of ROS, instead of being deleterious, acts as intracellular molecules that regulate myocyte physiology [50-52]. Moreover, the exposure of cardiac myocytes to extracellular H<sub>2</sub>O<sub>2</sub> activates the ERK 1/2 kinase and stimulates NHE in a dose and time-dependent manner [53,54].

Interestingly, at present it is accepted that many effects initially thought to be produced directly by Ang II, are really induced by ET-1 [19,55], Ald [56,57] and more recently, after the transactivation of the epidermal growth factor receptor (EGFR) [58-61]. Because of the close relationship between Ang II and the regulation of ion membrane transporters, in the last few years the investigation of Ang II-induced NBC modulation gained increasing interest.

### **REGULATION OF NBC BY ANG II**

It was demonstrated that Ang II stimulates total NBC activity during the recovery of pH<sub>i</sub> after an intracellular

acidosis both in rat [11] and cat [8] adult ventricular myocytes in a ROS- [8] and ERK 1/2- dependent manner [8,11]. Moreover, the recently described phenomenon of “ROS-induce-ROS-release” [62-64] was also involved in Ang II-induced NBC stimulation [8]. In addition, in neonatal rat myocytes, Ang II was reported to stimulate NBC activity in a phosphoinositide-independent mechanism after activation of AT-2 receptors [65].

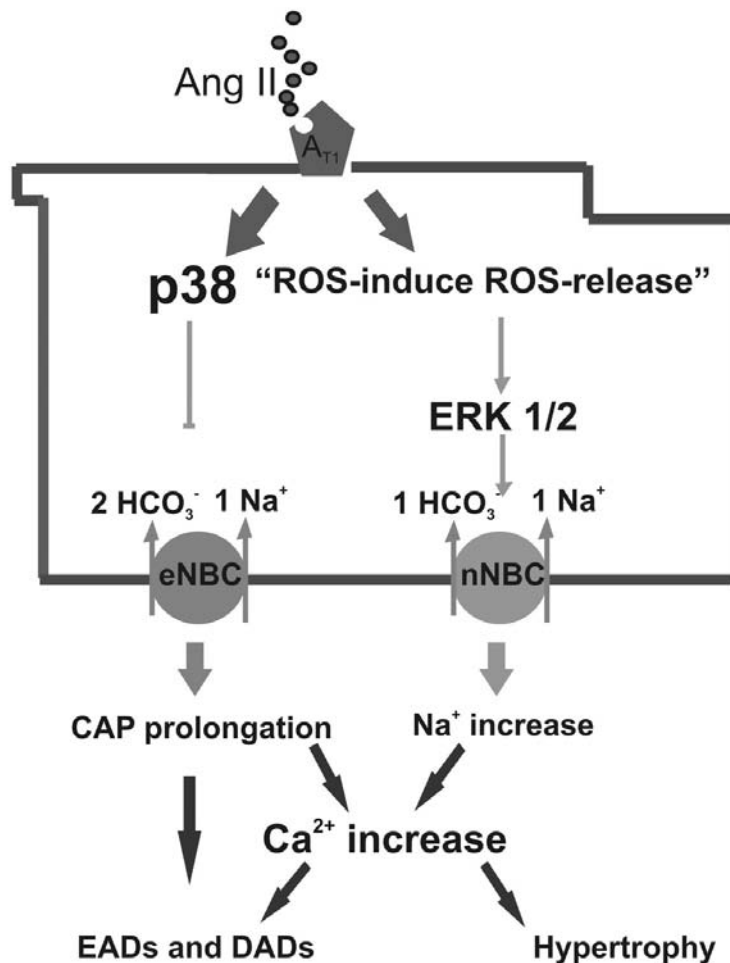
We have recently shown for the first time a differential effect of Ang II on NBC isoforms [29]. We demonstrated that Ang II inhibits eNBC in a p38 kinase-dependent, but ERK 1/2 and ROS-independent manner, whereas activates nNBC via an ERK 1/2 and ROS-dependent mechanism (Fig. 1). Thus, we suggested that Ang II, binding to the AT-1 receptor, activates the nNBC and inhibits the eNBC through parallel pathways [29]. Since previous studies, in which the effect of the hormone on both isoforms was not discriminated, have shown that Ang II stimulates total NBC activity [8,11], it might be possible to speculate that Ang II-induced stimulation of nNBC is able to overrule the inhibition of eNBC [29]. Consistently, when we measured total NBC activity during the recovery from acidosis, we found a stimulatory effect of Ang II which was further enhanced when p38-kinase was blocked, demonstrating that Ang II-induced in-

hibition of eNBC was only partly compensating the excitatory effect of the hormone on nNBC [29]. Furthermore, in the presence of NHE inhibition with HOE642, Ang II also significantly increased resting  $pH_i$ , again likely due to nNBC stimulation overcoming eNBC inhibition [29].

Although we presented the first evidence for the Ang II-induced cardiac eNBC inhibition, this effect was in agreement with several studies that demonstrated a biphasic regulation of NBCe1 by Ang II in renal tubules: low concentrations (picomolar to nanomolar) stimulated NBCe1 activity whereas higher concentrations (nanomolar to micromolar) inhibited it in an arachidonic acid-dependent way [66,67]. Consistently, it has been previously reported that p38 kinase is related with the arachidonic acid pathway and its metabolites in several tissues [68-71]. Further investigations are needed to evaluate the participation of arachidonic acid in Ang II-induced cardiac eNBC regulation. Moreover, it would be interesting to investigate the effect of low concentrations of Ang II in cardiac NBC activity.

#### INVOLVEMENT OF NBC IN CARDIAC PATHOLOGY

Although little is yet known about the implication of NBC in cardiac pathologies, in the last years there was a



**Fig. (1).** Schematic diagram showing the proposed Ang II- induced opposite effects on NBC isoforms and the possible implications in cardiac pathologies, as hypertrophy and arrhythmias. p38: p38 kinase; ERK 1/2: ERK kinase.

considerable increase in the knowledge about this issue, which suggested the involvement of NBC in several heart diseases, such as myocardial ischemia [34,72-77], infarction [78] and cardiac hypertrophy [31,79]. Interestingly, Ang II is also involved in these pathologies.

#### a) Ischemic Disease and Myocardial Infarction

Myocardial infarction is a major cause of death and disability worldwide. It is mainly developed during an unstable period of a coronary atherosclerosis disease. The term myocardial infarction reflects the existence of cardiomyocyte death caused by prolonged ischemia, which is the result of a perfusion imbalance between supply and demand.

Previous studies have demonstrated that cardiac NBC is activated during ischemia-reperfusion [72-77]. Moreover, Khandoudi and coauthors [34] have demonstrated that selective inhibition of NBCe1 during reperfusion after ischemia significantly improved contractile recovery, indicating that this transporter contributes to the characteristic intracellular  $\text{Na}^+$  and  $\text{Ca}^{2+}$  overload produced by this pathology. These authors also showed that NBCe1 is over-expressed in human heart failure [34].

Experimentally, it was demonstrated that local myocardial infarction (MI) leads to an increase in both, mRNA level and NBC protein expression and, as a consequence, NBC activity was enhanced [78]. Chronic treatment with blockers of the Ang II signaling, either with ACE inhibitor or AT-1 receptor antagonists, effectively reduced mRNA and protein NBC upregulation and transport activity [78], demonstrating a close relationship between NBC, Ang II and myocardial infarction.

#### b) Cardiac Hypertrophy

Cardiac hypertrophy is a response of the heart to a variety of extrinsic and intrinsic stimuli, some of which finally lead to a maladaptative state. On the basis of the external stimuli and ongoing molecular changes, cardiac hypertrophy is divided in two types; the physiological cardiac hypertrophy, mostly seen in athlete's heart and the pathological cardiac hypertrophy induced by mechanical stress, due to pressure overload or volume overload. In the physiological cardiac hypertrophy the increase of the cardiac muscle reduces ventricular wall stress and compensates for the increased hemodynamic demand, improving heart contractility. It is generally associated with increased cardiac mass without collagen deposition. Opposite, the pathological cardiac hypertrophy is characterized by the fibrosis and induction of fetal gene expression, which lead to a reduction in cardiac output and enhancement of the risk of sudden death, arrhythmia and heart failure [80,81].

Yamamoto *et al.* have demonstrated that NBCe1 and NBCn1 were over-expressed in ventricular myocytes isolated from hypertrophied rat hearts subjected to non-ischemic pressure overload [31]. Moreover, these changes are prevented by Losartan [31]. When these authors evaluated the function of both isoforms in the hypertrophied hearts, they could not find a clear upregulation of NBCe1 [31]. Consistently, we have recently shown preliminary data

suggesting that, although NBCe1 is also over-expressed in hypertrophied hearts of spontaneous hypertensive rats (SHR), its activity is impaired [79]. It is possible that Ang II induced the NBCe1 internalization, explaining the discordance between the protein expression and the transport activity. In agreement, Ang II-induced NBCe1 internalization was described in *Xenopus* oocytes transfected with this NBC isoform [82]. Nevertheless, it is important to mention that it could not be determined yet if the changes on NBC were the cause or the consequence of the development of cardiac hypertrophy. Additional studies are required to fully resolve this important issue.

#### ROLE OF NBC-INDUCED $[\text{Na}^+]_i$ AND $[\text{Ca}^{2+}]_i$ OVERLOAD: POTENTIAL IMPLICATIONS IN CARDIAC HYPERTROPHY

It is well-known that increased  $[\text{Ca}^{2+}]_i$  activates hypertrophic pathways, such as the one of calcineurin [83,84].  $\text{Ca}^{2+}$  regulation is closely linked to  $[\text{Na}^+]_i$  because one of the routes for  $\text{Ca}^{2+}$  influx into the myocytes is via the reverse mode of NCX. When  $[\text{Na}^+]_i$  increases, NCX is shifted to less forward mode activity ( $\text{Ca}^{2+}$ - efflux) and/or to reverse operation mode, leading to  $[\text{Ca}^{2+}]_i$  overload [85-87].

In animal models of hypertrophy, as well as in human heart failure, it has been demonstrated an increase in  $[\text{Na}^+]_i$  and  $[\text{Ca}^{2+}]_i$  [88-90]. Furthermore, it was shown that chronic inhibition of NHE, which attenuates the  $[\text{Na}^+]_i$  overload, prevented or reverted cardiac hypertrophy [91-94]. On the other hand, the over-expression of NHE induced cardiac hypertrophy [95].

As it was demonstrated that NBC is responsible for 30% of  $\text{Na}^+$  influx into the myocyte at pH<sub>i</sub> 6.8 [12], it may be also important in the development of cardiac hypertrophy. In this regard and as commented above, it has been shown that nNBC function is up-regulated in cardiac hypertrophy [31], while eNBC transport seems to be impaired [79]. Taking into account the stoichiometry of both NBC isoforms, which could lead to the consideration of eNBC as a " $\text{Na}^+$ - sparing" bicarbonate transporter, it is feasible to anticipate that this remodeling in NBC isoforms function in the hypertrophied hearts would lead to more deleterious effects on  $[\text{Na}^+]_i$  and  $[\text{Ca}^{2+}]_i$  overload.

#### ROLE OF NBC- INDUCED $[\text{Na}^+]_i$ AND $[\text{Ca}^{2+}]_i$ OVERLOAD: POTENTIAL IMPLICATION IN DELAYED AFTER DEPOLARIZATIONS (DADs)

It has been shown that either the inhibition of the  $\text{Na}^+/\text{K}^+$  ATPase [96,97] or the NHE stimulation [98] generate  $[\text{Na}^+]_i$  overload and cardiac arrhythmias. The proposed mechanism is the following:  $[\text{Na}^+]_i$  overload reduces  $\text{Ca}^{2+}$  extrusion and/or increases  $\text{Ca}^{2+}$  influx through the NCX. The increase in  $[\text{Ca}^{2+}]_i$  enhance the sarcoplasmic reticulum (SR) calcium load, exceeding the ryanodine receptor channel (RyR) threshold necessary to be opened and finally leading to spontaneous diastolic calcium release. The transient increase in cytosolic  $\text{Ca}^{2+}$  (waves) activates an inward (depolarizing) current ( $I_{ti}$ ), mediated by the forward mode of NCX [99,100].  $I_{ti}$  is responsible for the generation of DADs which, when are sufficiently large to achieve the threshold, generate spontaneous CAP, leading to triggered activity [101].

As NBC activity promotes the increase in  $[\text{Na}^+]_i$  [12], it is also possible to speculate that Ang II and ROS-induced NBC stimulation [8] might be implicated in DADs generation (Fig. 1). According to this, it was demonstrated that Ang II induces DADs in a ROS-dependent manner [102].

However, it is important to note that increased SR  $\text{Ca}^{2+}$  load is not sufficient to promote diastolic spontaneous SR  $\text{Ca}^{2+}$  release [97], and that also a functional or structural alteration in RyR is needed to induce DADs [103-107]. In addition, it was recently demonstrated that ROS can directly oxidise the RyR, making it leaky [108-111]. Interestingly, the inverse conclusion seems to be also true: just an impaired RyR is not sufficient to induce  $\text{Ca}^{2+}$  release, since a parallel increase in SR  $\text{Ca}^{2+}$  load is also required [112].

### ROLE OF eNBC INHIBITION-INDUCED CAP PROLONGATION: POTENTIAL IMPLICATION IN EARLY AFTER DEPOLARIZATIONS (EADs)

Classically, Ang II is known to modulate the properties of ion channels leading to CAP prolongation [113-116]. It has been reported that Ang II both inhibits repolarizing currents as  $I_{K1}$ ,  $I_{Kr}$  and  $I_{to}$  [114,116-118] and stimulates depolarizing currents as  $I_{CaL}$  [119,120]. Moreover, we have recently demonstrated that Ang II abrogated the eNBC-induced CAP shortening, likely due to the inhibition of the repolarizing current generated by the transporter [29].

In this regard, it has been shown that CAP prolongation enhances the occurrence of EADs, due to the recovery from the inactivation and the reactivation of voltage-dependent L-type  $\text{Ca}^{2+}$  channels [102,121,122] and the impairment of sodium current [102,122]. In addition, Ang II was shown to increase the occurrence of EADs in a ROS and CaMKII-

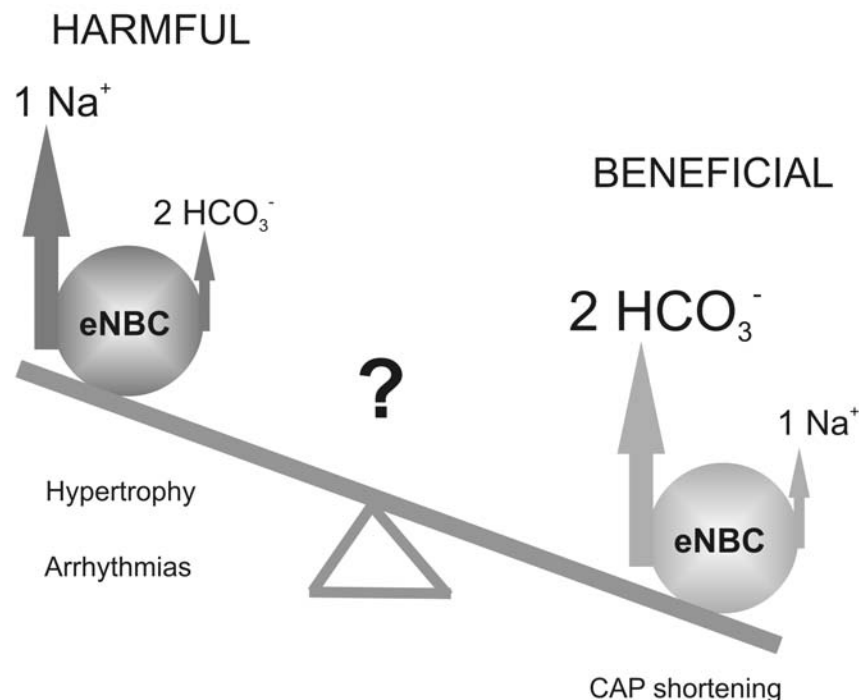
dependent manner [102]. Furthermore, it was shown that chronic inhibition of NHE could reverse the ionic (sodium overload and disturbance in calcium management) and electrical (CAP prolongation) cellular remodeling during heart failure, and reduce arrhythmic events [98].

The relationship between NBC and EADs is currently unknown, but keeping in mind the relevance of eNBC for CAP configuration [29], and the regulation of  $[\text{Na}^+]_i$  by total NBC activity [31], it might be also possible that Ang II-induced eNBC inhibition and nNBC stimulation participate in the generation of EAD, secondary to CAP prolongation and  $[\text{Na}^+]_i$  overload, respectively (Fig. 1).

Importantly, there is a close relationship between CAP prolongation and hypertrophy. Prolongation of CAP is consistently observed in several experimental models of cardiac hypertrophy and failure [123]. It is also known that this can lead to QT prolongation in the electrocardiogram, that in turn promotes arrhythmogenic events [98,124]. Interestingly, Lebeche *et al.* have reported that CAP prolongation promotes an increase in  $[\text{Ca}^{2+}]_i$ , which activate a hypertrophic signaling pathway, that might be a cause and not a consequence of cardiac hypertrophy [125].

### eNBC ACTIVITY: BENEFICIAL OR DETRIMENTAL?

The double participation of eNBC in cardiac physiology makes it difficult to call as a “beneficial” or “detrimental” mechanism (Fig. 2). Since eNBC is a cellular “ $\text{Na}^+$ -loading” mechanism, it might contribute to  $[\text{Na}^+]_i$  and  $[\text{Ca}^{2+}]_i$  overload and arrhythmias generation [21,73]. In concordance, Khandoudi *et al.* reported that blockade of rat cardiac eNBC during reperfusion results in cardioprotection [34]. Thus, we can speculate that the Ang II-induced eNBC inhibition



**Fig. (2).** Schematic representation of eNBC activity showing the balance between its beneficial repolarizing current (*right*) and the consequences of its  $\text{Na}^+$ -loading effect (*left*) in cardiac pathologies.

would carry beneficial effects under this pathological state. Similar paradigmatic speculations about the effect of Ang II on eNBC could be made with cardiac hypertrophy and heart failure, which are cardiac diseases associated to elevated Ang II concentration [40,126].

However, in cardiac hypertrophy or heart failure, the potential attenuation of  $[Na^+]_i$  overload that might be produced by eNBC inhibition [31] could be overruled by the deleterious effects that might be carried by the prolongation of CAP also induced by the blockade of eNBC [29]. Moreover, Camili3n de Hurtado *et al.* have demonstrated that a rate-dependent decrease in  $pH_i$ , probably due to the increased anaerobic glycolysis, was markedly reduced in the presence of  $CO_2$ /bicarbonate in comparison to a free-bicarbonate solution [30]. In this work, the authors proposed that eNBC activity, which leads to  $HCO_3^-$  influx, substantially increases the cell ability to recover from enhanced proton production [30].

## FINAL CONCLUSION

The purpose of this review was to focus the attention on the cardiac NBC and specially consider its regulation by Ang II and the implications of this modulation, either in physiology or in the development of cardiac diseases. Classically, the NBC is known as an alkalinizing mechanism. However, it is important to keep in mind that this is not its only function, but it also controls  $[Na^+]_i$ , and indirectly  $[Ca^{2+}]_i$  through the NCX activity and SR behavior. Moreover, eNBC modulates the shape and the duration of the CAP, adding to this isoform the important role of contributing to cellular electrophysiology.

We consider of significant relevance the fact that a hormone as Ang II, which has a central role in cardiac pathophysiology, regulates NBC activity. Moreover, this peptide exerts an opposite effect on each NBC isoform due to the activation of two different and parallel pathways. The inhibitory effect that Ang II exerts on eNBC, via the activation of the p38-kinase, seems to be more relevant for CAP duration than for  $pH_i$  regulation. On the opposite, the stimulatory effect of this hormone on nNBC, dependent on ROS production and ERK 1/2- activation, overrule the negative effect on eNBC, leading to an increase in  $pH_i$ ,  $[Na^+]_i$ , and  $[Ca^{2+}]_i$  that could be important to explain, at least in part, the hypertrophic effects of Ang II signaling.

The knowledge of the singular regulation of each NBC isoform should be the base for following investigations. The use of specific inhibitors of the ERK 1/2 or p38- kinases pathways and the employment of functional antibodies as new pharmacological tools, will allow the study of the differential implication of eNBC and nNBC in cardiac pathologies. As an example of clinical relevance, it is feasible to suggest that the stimulatory antibody (a-L4) against eNBC, which would induce CAP shortening, could be useful to investigate the potential protective effect of eNBC activation during the development of cardiac hypertrophy or the damage during reperfusion after ischemia. Nevertheless, besides the fact that the amount of pharmacological tools has been growing up, we are still in debt, and the more precise knowledge about development of cardiac diseases that we can elu-

cidate, the more close we will be to find the specific treatment for them.

## CONFLICT OF INTEREST

The authors confirm that this article content has no conflicts of interest.

## ACKNOWLEDGEMENTS

Declared none.

## REFERENCES

- [1] Bountra C, Kaila K, Vaughan-Jones RD. Effect of repetitive activity upon intracellular pH, sodium and contraction in sheep cardiac Purkinje fibres. *The Journal of physiology* 1988; 398: 341-60.
- [2] Elliott AC, Smith GL, Allen DG. The metabolic consequences of an increase in the frequency of stimulation in isolated ferret hearts. *The Journal of physiology* 1994; 474(1): 147-59.
- [3] Steenbergen C, Deleew G, Rich T, Williamson JR. Effects of acidosis and ischemia on contractility and intracellular pH of rat heart. *Circulation research* 1977; 41(6): 849-58.
- [4] Garlick PB, Radda GK, Seeley PJ. Studies of acidosis in the ischaemic heart by phosphorus nuclear magnetic resonance. *The Biochemical journal* 1979; 184(3): 547-54.
- [5] Cha CY, Oka C, Earm YE, Wakabayashi S, Noma A. A model of  $Na^+/H^+$  exchanger and its central role in regulation of pH and  $Na^+$  in cardiac myocytes. *Biophysical journal* 2009; 97(10): 2674-83.
- [6] Vaughan-Jones RD, Swietach P. Pushing and pulling the cardiac sodium/hydrogen exchanger. *Circulation research* 2008; 103(8): 773-5.
- [7] Ch'en FF, Villafuerte FC, Swietach P, Cobden PM, Vaughan-Jones RD. S0859, an N-cyanosulphonamide inhibitor of sodium-bicarbonate cotransport in the heart. *British journal of pharmacology* 2008; 153(5): 972-82.
- [8] De Giusti VC, Garcarena CD, Aiello EA. Role of reactive oxygen species (ROS) in angiotensin II-induced stimulation of the cardiac  $Na^+/HCO_3^-$  cotransport. *Journal of molecular and cellular cardiology* 2009; 47(5): 716-22.
- [9] Lagadic-Gossmann D, Buckler KJ, Vaughan-Jones RD. Role of bicarbonate in pH recovery from intracellular acidosis in the guinea-pig ventricular myocyte. *The Journal of physiology* 1992; 458: 361-84.
- [10] Camilion de Hurtado MC, Perez NG, Cingolani HE. An electrogenic sodium-bicarbonate cotransport in the regulation of myocardial intracellular pH. *Journal of molecular and cellular cardiology* 1995; 27(1): 231-42.
- [11] Baetz D, Haworth RS, Avkiran M, Feuvray D. The ERK pathway regulates  $Na^+-HCO_3^-$  cotransport activity in adult rat cardiomyocytes. *Am J Physiol Heart Circ Physiol* 2002; 283(5): H2102-9.
- [12] Vaughan-Jones RD, Villafuerte FC, Swietach P, Yamamoto T, Rossini A, Spitzer KW. pH-Regulated  $Na^+$  influx into the mammalian ventricular myocyte: the relative role of  $Na^+-H^+$  exchange and  $Na^+-HCO_3^-$  Co-transport. *Journal of cardiovascular electrophysiology* 2006; 17 Suppl 1: S134-S40.
- [13] Vaughan-Jones RD, Spitzer KW, Swietach P. Intracellular pH regulation in heart. *Journal of molecular and cellular cardiology* 2009; 46(3): 318-31.
- [14] Le Prigent K, Lagadic-Gossmann D, Mongodin E, Feuvray D.  $HCO_3^-$ -dependent alkalinizing transporter in adult rat ventricular myocytes: characterization and modulation. *The American journal of physiology* 1997; 273(6 Pt 2): H2596-603.
- [15] Rothstein EC, Byron KL, Reed RE, Fliegel L, Lucchesi PA.  $H_2O_2$ -induced  $Ca^{2+}$  overload in NRVM involves ERK1/2 MAP kinases: role for an NHE-1-dependent pathway. *Am J Physiol Heart Circ Physiol* 2002; 283(2): H598-605.
- [16] Bril A. [Ion transporters and cardiovascular diseases: pH control or modulation of intracellular calcium concentration]. *Annales de cardiologie et d'angiologie* 2003; 52(1): 41-51.
- [17] Aiello EA, Villa-Abrille MC, Dulce RA, Cingolani HE, Perez NG. Endothelin-1 stimulates the  $Na^+/Ca^{2+}$  exchanger reverse mode through intracellular  $Na^+$  ( $Na^+$ )-dependent and  $Na^+$ -independent pathways. *Hypertension* 2005; 45(2): 288-93.

- [18] De Giusti VC, Correa MV, Villa-Abrille MC, Beltrano C, Yeves AM, de Cingolani GE, *et al.* The positive inotropic effect of endothelin-1 is mediated by mitochondrial reactive oxygen species. *Life sciences* 2008; 83(7-8): 264-71.
- [19] Cingolani HE, Perez NG, Aiello EA, Ennis IL, Garcarena CD, Villa-Abrille MC, *et al.* Early signals after stretch leading to cardiac hypertrophy. Key role of NHE-1. *Front Biosci* 2008; 13: 7096-114.
- [20] Dulce RA, Hurtado C, Ennis IL, Garcarena CD, Alvarez MC, Caldiz C, *et al.* Endothelin-1 induced hypertrophic effect in neonatal rat cardiomyocytes: involvement of  $\text{Na}^+/\text{H}^+$  and  $\text{Na}^+/\text{Ca}^{2+}$  exchangers. *Journal of molecular and cellular cardiology* 2006; 41(5): 807-15.
- [21] Baartscheer A, van Borren MM. Sodium ion transporters as new therapeutic targets in heart failure. *Cardiovascular & hematological agents in medicinal chemistry* 2008; 6(4): 229-36.
- [22] Aiello EA, Petroff MG, Mattiuzzi AR, Cingolani HE. Evidence for an electrogenic  $\text{Na}^+/\text{HCO}_3^-$  symport in rat cardiac myocytes. *The Journal of physiology* 1998; 512 ( Pt 1): 137-48.
- [23] Villa-Abrille MC, Petroff MG, Aiello EA. The electrogenic  $\text{Na}^+/\text{HCO}_3^-$  cotransport modulates resting membrane potential and action potential duration in cat ventricular myocytes. *The Journal of physiology* 2007; 578(Pt 3): 819-29.
- [24] Yamamoto T, Swietach P, Rossini A, Loh SH, Vaughan-Jones RD, Spitzer KW. Functional diversity of electrogenic  $\text{Na}^+/\text{HCO}_3^-$  cotransport in ventricular myocytes from rat, rabbit and guinea pig. *The Journal of physiology* 2005; 562(Pt 2): 455-75.
- [25] De Giusti VC, Orłowski A, Villa-Abrille MC, de Cingolani GE, Casey JR, Alvarez BV, *et al.* Antibodies against the cardiac sodium/bicarbonate co-transporter (NBCe1) as pharmacological tools. *British journal of pharmacology* 2011; 164(8): 1976-89.
- [26] Choi I, Romero MF, Khandoudi N, Bril A, Boron WF. Cloning and characterization of a human electrogenic  $\text{Na}^+/\text{HCO}_3^-$  cotransporter isoform (hhNBC). *The American journal of physiology* 1999; 276(3 Pt 1): C576-84.
- [27] Sassani P, Pushkin A, Gross E, Gomer A, Abuladze N, Dukkupati R, *et al.* Functional characterization of NBC4: a new electrogenic sodium-bicarbonate cotransporter. *Am J Physiol Cell Physiol* 2002; 282(2): C408-16.
- [28] Choi I, Aalkjaer C, Boulpaep EL, Boron WF. An electroneutral sodium/bicarbonate cotransporter NBCn1 and associated sodium channel. *Nature* 2000; 405(6786): 571-5.
- [29] De Giusti VC, Orłowski A, Aiello EA. Angiotensin II inhibits the electrogenic  $\text{Na}^+/\text{HCO}_3^-$  cotransport of cat cardiac myocytes. *Journal of molecular and cellular cardiology* 2010; 49(5): 812-8.
- [30] Camilion de Hurtado MC, Alvarez BV, Perez NG, Cingolani HE. Role of an electrogenic  $\text{Na}^+/\text{HCO}_3^-$  cotransport in determining myocardial pH<sub>i</sub> after an increase in heart rate. *Circulation research* 1996; 79(4): 698-704.
- [31] Yamamoto T, Shirayama T, Sakatani T, Takahashi T, Tanaka H, Takamatsu T, *et al.* Enhanced activity of ventricular  $\text{Na}^+/\text{HCO}_3^-$  cotransport in pressure overload hypertrophy. *Am J Physiol Heart Circ Physiol* 2007; 293(2): H1254-64.
- [32] De Giusti VC, Aiello EA. Generación de anticuerpos inhibitorios de la función del cotransportador sodio/bicarbonato cardíaco. Una posible futura herramienta terapéutica. *Rev Fed Arg Cardiol* 2011; 40(1): 32-40.
- [33] Orłowski A, De Giusti VC, Morgan PE, Aiello EA, Alvarez BV. Binding of carbonic anhydrase IX to extracellular loop 4 of the NBCe1  $\text{Na}^+/\text{HCO}_3^-$  cotransporter enhances NBCe1-mediated  $\text{HCO}_3^-$  influx in the rat heart. *Am J Physiol Cell Physiol* 2012; 303(1): C69-80.
- [34] Khandoudi N, Albadine J, Robert P, Krief S, Berrebi-Bertrand I, Martin X, *et al.* Inhibition of the cardiac electrogenic sodium bicarbonate cotransporter reduces ischemic injury. *Cardiovascular research* 2001; 52(3): 387-96.
- [35] Husain A KA, Sung SS, Urata H, Bumpus FM. Human Heart Chymase. In: Lindpaintner K GDE, editor. *The Cardiac Renin-Angiotensin System*. Armonk, New York: Futura Publishing 1994; p. 309-32.
- [36] Sadoshima J, Xu Y, Slayter HS, Izumo S. Autocrine release of angiotensin II mediates stretch-induced hypertrophy of cardiac myocytes *in vitro*. *Cell* 1993; 75(5): 977-84.
- [37] Sadoshima J, Izumo S. Autocrine secretion of angiotensin II mediates stretch-induced hypertrophy of cardiac myocytes *in vitro*. *Contributions to nephrology* 1996; 118: 214-21.
- [38] Cingolani HE, Perez NG, Camilion de Hurtado MC. An autocrine/paracrine mechanism triggered by myocardial stretch induces changes in contractility. *News Physiol Sci* 2001; 16: 88-91.
- [39] Cingolani HE, Perez NG, Pieske B, von Lewinski D, Camilion de Hurtado MC. Stretch-elicited  $\text{Na}^+/\text{H}^+$  exchanger activation: the autocrine/paracrine loop and its mechanical counterpart. *Cardiovascular research* 2003; 57(4): 953-60.
- [40] De Mello WC, Danser AH. Angiotensin II and the heart : on the intracrine renin-angiotensin system. *Hypertension* 2000; 35(6): 1183-8.
- [41] Silvestre JS, Robert V, Heymes C, Aupetit-Faisant B, Mouas C, Moalic JM, *et al.* Myocardial production of aldosterone and corticosterone in the rat. *Physiological regulation*. *The Journal of biological chemistry* 1998; 273(9): 4883-91.
- [42] Varagic J, Frohlich ED. Local cardiac renin-angiotensin system: hypertension and cardiac failure. *Journal of molecular and cellular cardiology* 2002; 34(11): 1435-42.
- [43] Leenen FH, Skarda V, Yuan B, White R. Changes in cardiac ANG II postmyocardial infarction in rats: effects of nephrectomy and ACE inhibitors. *The American journal of physiology* 1999; 276(1 Pt 2): H317-25.
- [44] Ruzicka M, Skarda V, Leenen FH. Effects of ACE inhibitors on circulating versus cardiac angiotensin II in volume overload-induced cardiac hypertrophy in rats. *Circulation* 1995; 92(12): 3568-73.
- [45] Yoneda M, Sanada H, Yatabe J, Midorikawa S, Hashimoto S, Sasaki M, *et al.* Differential effects of angiotensin II type-1 receptor antisense oligonucleotides on renal function in spontaneously hypertensive rats. *Hypertension* 2005; 46(1): 58-65.
- [46] Cardinale JP, Sriramula S, Pariaut R, Guggilam A, Mariappan N, Elks CM, *et al.* HDAC inhibition attenuates inflammatory, hypertrophic, and hypertensive responses in spontaneously hypertensive rats. *Hypertension* 2010; 56(3): 437-44.
- [47] Zhang GX, Lu XM, Kimura S, Nishiyama A. Role of mitochondria in angiotensin II-induced reactive oxygen species and mitogen-activated protein kinase activation. *Cardiovascular research* 2007; 76(2): 204-12.
- [48] Mehta PK, Griendling KK. Angiotensin II cell signaling: physiological and pathological effects in the cardiovascular system. *Am J Physiol Cell Physiol* 2007; 292(1): C82-97.
- [49] Seshiah PN, Weber DS, Rocic P, Valppu L, Taniyama Y, Griendling KK. Angiotensin II stimulation of NAD(P)H oxidase activity: upstream mediators. *Circulation research* 2002; 91(5): 406-13.
- [50] Caldiz CI, Garcarena CD, Dulce RA, Novaretto LP, Yeves AM, Ennis IL, *et al.* Mitochondrial reactive oxygen species activate the slow force response to stretch in feline myocardium. *The Journal of physiology* 2007; 584(Pt 3): 895-905.
- [51] Bartosz G. Reactive oxygen species: destroyers or messengers? *Biochemical pharmacology* 2009; 77(8): 1303-15.
- [52] Cingolani HE, Villa-Abrille MC, Cornelli M, Nolly A, Ennis IL, Garcarena C, *et al.* The positive inotropic effect of angiotensin II: role of endothelin-1 and reactive oxygen species. *Hypertension* 2006; 47(4): 727-34.
- [53] Snabaitis AK, Hearse DJ, Avkiran M. Regulation of sarcolemmal  $\text{Na}^+/\text{H}^+$  exchange by hydrogen peroxide in adult rat ventricular myocytes. *Cardiovascular research* 2002; 53(2): 470-80.
- [54] Sabri A, Byron KL, Samarel AM, Bell J, Lucchesi PA. Hydrogen peroxide activates mitogen-activated protein kinases and  $\text{Na}^+/\text{H}^+$  exchange in neonatal rat cardiac myocytes. *Circulation research* 1998; 82(10): 1053-62.
- [55] Perez NG, Villa-Abrille MC, Aiello EA, Dulce RA, Cingolani HE, Camilion de Hurtado MC. A low dose of angiotensin II increases inotropism through activation of reverse  $\text{Na}^+/\text{Ca}^{2+}$  exchange by endothelin release. *Cardiovascular research* 2003; 60(3): 589-97.
- [56] Lemarie CA, Paradis P, Schiffrin EL. New insights on signaling cascades induced by cross-talk between angiotensin II and aldosterone. *Journal of molecular medicine (Berlin, Germany)* 2008; 86(6): 673-8.
- [57] Xiao F, Puddefoot JR, Barker S, Vinson GP. Mechanism for aldosterone potentiation of angiotensin II-stimulated rat arterial smooth muscle cell proliferation. *Hypertension* 2004; 44(3): 340-5.
- [58] Freeman EJ, Sheakley ML, Clements RJ. Angiotensin II-dependent growth of vascular smooth muscle cells requires transactivation of the epidermal growth factor receptor via a cytosolic phospholipase A(2)-mediated release of arachidonic acid. *Archives of biochemistry and biophysics* 2010; 498(1): 50-6.

- [59] Shah BH, Catt KJ. A central role of EGF receptor transactivation in angiotensin II-induced cardiac hypertrophy. *Trends in pharmacological sciences* 2003; 24(5): 239-44.
- [60] Zhai P, Galeotti J, Liu J, *et al.* An angiotensin II type 1 receptor mutant lacking epidermal growth factor receptor transactivation does not induce angiotensin II-mediated cardiac hypertrophy. *Circulation research* 2006; 99(5): 528-36.
- [61] Villa-Abrille MC, Caldiz CI, Ennis IL, *et al.* The Anrep effect requires transactivation of the epidermal growth factor receptor. *The Journal of physiology* 2010; 588(Pt 9): 1579-90.
- [62] Zorov DB, Filburn CR, Klotz LO, Zweier JL, Sollott SJ. Reactive oxygen species (ROS)-induced ROS release: a new phenomenon accompanying induction of the mitochondrial permeability transition in cardiac myocytes. *The Journal of experimental medicine* 2000; 192(7): 1001-14.
- [63] Zorov DB, Juhaszova M, Sollott SJ. Mitochondrial ROS-induced ROS release: an update and review. *Biochimica et biophysica acta* 2006; 1757(5-6): 509-17.
- [64] Kimura S, Zhang GX, Nishiyama A, *et al.* Role of NAD(P)H oxidase- and mitochondria-derived reactive oxygen species in cardioprotection of ischemic reperfusion injury by angiotensin II. *Hypertension* 2005; 45(5): 860-6.
- [65] Kohout TA, Rogers TB. Angiotensin II activates the  $\text{Na}^+/\text{HCO}_3^-$  symport through a phosphoinositide-independent mechanism in cardiac cells. *The Journal of biological chemistry* 1995; 270(35): 20432-8.
- [66] Horita S, Zheng Y, Hara C, *et al.* Biphasic regulation of  $\text{Na}^+/\text{HCO}_3^-$  cotransporter by angiotensin II type 1A receptor. *Hypertension* 2002; 40(5): 707-12.
- [67] Li Y, Yamada H, Kita Y, Suzuki M, Endo Y, Horita S, *et al.* Arachidonic acid metabolites inhibit the stimulatory effect of angiotensin II in renal proximal tubules. *Hypertens Res* 2008; 31(12): 2155-64.
- [68] Nito C, Kamada H, Endo H, Niizuma K, Myer DJ, Chan PH. Role of the p38 mitogen-activated protein kinase/cytosolic phospholipase A2 signaling pathway in blood-brain barrier disruption after focal cerebral ischemia and reperfusion. *J Cereb Blood Flow Metab* 2008; 28(10): 1686-96.
- [69] Kalyankrishna S, Malik KU. Norepinephrine-induced stimulation of p38 mitogen-activated protein kinase is mediated by arachidonic acid metabolites generated by activation of cytosolic phospholipase A(2) in vascular smooth muscle cells. *The Journal of pharmacology and experimental therapeutics* 2003; 304(2): 761-72.
- [70] Husain S, Abdel-Latif AA. Endothelin-1 activates p38 mitogen-activated protein kinase and cytosolic phospholipase A2 in cat iris sphincter smooth muscle cells. *The Biochemical journal* 1999; 342 (Pt 1): 87-96.
- [71] Upmacis RK, Deeb RS, Resnick MJ, *et al.* Involvement of the mitogen-activated protein kinase cascade in peroxynitrite-mediated arachidonic acid release in vascular smooth muscle cells. *Am J Physiol Cell Physiol* 2004; 286(6): C1271-80.
- [72] Bauza G, Le Moyec L, Eugene M. pH regulation during ischaemia-reperfusion of isolated rat hearts, and metabolic effects of 2,3-butanedione monoxime. *Journal of molecular and cellular cardiology* 1995; 27(8): 1703-13.
- [73] Doggrel SA, Hancox JC. Is timing everything? Therapeutic potential of modulators of cardiac  $\text{Na}^+$  transporters. *Expert opinion on investigational drugs* 2003; 12(7): 1123-42.
- [74] Schafer C, Ladilov YV, Siegmund B, Piper HM. Importance of bicarbonate transport for protection of cardiomyocytes against reoxygenation injury. *Am J Physiol Heart Circ Physiol* 2000; 278(5): H1457-63.
- [75] Skolnick RL, Litwin SE, Barry WH, Spitzer KW. Effect of ANG II on  $\text{pH}_i$ ,  $[\text{Ca}^{2+}]_i$ , and contraction in rabbit ventricular myocytes from infarcted hearts. *The American journal of physiology* 1998; 275(5 Pt 2): H1788-97.
- [76] Ten Hove M, Nederhoff MG, Van Echteld CJ. Relative contributions of  $\text{Na}^+/\text{H}^+$  exchange and  $\text{Na}^+/\text{HCO}_3^-$  cotransport to ischemic  $\text{Na}^+$  overload in isolated rat hearts. *Am J Physiol Heart Circ Physiol* 2005; 288(1): H287-92.
- [77] van Borren MM, Baartscheer A, Wilders R, Ravestloot JH. NHE-1 and NBC during pseudo-ischemia/reperfusion in rabbit ventricular myocytes. *Journal of molecular and cellular cardiology* 2004; 37(2): 567-77.
- [78] Sandmann S, Yu M, Kaschina E, *et al.* Differential effects of angiotensin AT1 and AT2 receptors on the expression, translation and function of the  $\text{Na}^+/\text{H}^+$  exchanger and  $\text{Na}^+/\text{HCO}_3^-$  symporter in the rat heart after myocardial infarction. *Journal of the American College of Cardiology* 2001; 37(8): 2154-65.
- [79] Orłowski A, De Giusti VC, Aiello EA. Dysfunctional Electrogenic Sodium/Bicarbonate Cotransport in Cardiomyocytes of Spontaneously Hypertensive Rats. *Hypertension* 2011; 58:e114.
- [80] Sala V, Gallo S, Leo C, Gatti S, Gelb BD, Crepaldi T. Signaling to cardiac hypertrophy: insights from human and mouse RASopathies. *Molecular medicine* 2012.
- [81] Maulik SK, Kumar S. Oxidative stress and cardiac hypertrophy: a review. *Toxicology mechanisms and methods* 2012; 22(5): 359-66.
- [82] Perry C, Le H, Grichtchenko, II. ANG II and calmodulin/CaMKII regulate surface expression and functional activity of NBCe1 via separate means. *American journal of physiology* 2007; 293(1): F68-77.
- [83] Ennis IL, Garcarena CD, Escudero EM, *et al.* Normalization of the calcineurin pathway underlies the regression of hypertensive hypertrophy induced by  $\text{Na}^+/\text{H}^+$  exchanger-1 (NHE-1) inhibition. *Canadian journal of physiology and pharmacology* 2007; 85(3-4): 301-10.
- [84] Guo J, Gan XT, Haist JV, Rajapurohitam V, Zeidan A, Faruq NS, *et al.* Ginseng inhibits cardiomyocyte hypertrophy and heart failure via NHE-1 inhibition and attenuation of calcineurin activation. *Circ Heart Fail* 2011; 4(1): 79-88.
- [85] Alvarez BV, Perez NG, Ennis IL, Camilion de Hurtado MC, Cingolani HE. Mechanisms underlying the increase in force and  $\text{Ca}^{2+}$  transient that follow stretch of cardiac muscle: a possible explanation of the Anrep effect. *Circulation research* 1999; 85(8): 716-22.
- [86] Kentish JC, Wrzosek A. Changes in force and cytosolic  $\text{Ca}^{2+}$  concentration after length changes in isolated rat ventricular trabeculae. *The Journal of physiology* 1998; 506 (Pt 2): 431-44.
- [87] Cingolani HE, Ennis IL. Sodium-hydrogen exchanger, cardiac overload, and myocardial hypertrophy. *Circulation* 2007; 115(9): 1090-100.
- [88] Verdonck F, Volders PG, Vos MA, Sipido KR. Increased  $\text{Na}^+$  concentration and altered Na/K pump activity in hypertrophied canine ventricular cells. *Cardiovascular research* 2003; 57(4): 1035-43.
- [89] Despa S, Islam MA, Weber CR, Pogwizd SM, Bers DM. Intracellular  $\text{Na}^+$  concentration is elevated in heart failure but Na/K pump function is unchanged. *Circulation* 2002; 105(21): 2543-8.
- [90] Gray RP, McIntyre H, Sheridan DS, Fry CH. Intracellular sodium and contractile function in hypertrophied human and guinea-pig myocardium. *Pflugers Arch* 2001; 442(1): 117-23.
- [91] Camilion de Hurtado MC, Portiansky EL, Perez NG, Rebolledo OR, Cingolani HE. Regression of cardiomyocyte hypertrophy in SHR following chronic inhibition of the  $\text{Na}^+/\text{H}^+$  exchanger. *Cardiovascular research* 2002; 53(4): 862-8.
- [92] Engelhardt S, Hein L, Keller U, Klambt K, Lohse MJ. Inhibition of  $\text{Na}^+/\text{H}^+$  exchange prevents hypertrophy, fibrosis, and heart failure in beta(1)-adrenergic receptor transgenic mice. *Circulation research* 2002; 90(7): 814-9.
- [93] Kusumoto K, Haist JV, Karmazyn M.  $\text{Na}^{(+)}/\text{H}^{(+)}$  exchange inhibition reduces hypertrophy and heart failure after myocardial infarction in rats. *Am J Physiol Heart Circ Physiol* 2001; 280(2): H738-45.
- [94] Ennis IL, Escudero EM, Console GM, Camihort G, Dumm CG, Seidler RW, *et al.* Regression of isoproterenol-induced cardiac hypertrophy by  $\text{Na}^+/\text{H}^+$  exchanger inhibition. *Hypertension* 2003; 41(6): 1324-9.
- [95] Nakamura TY, Iwata Y, Arai Y, Komamura K, Wakabayashi S. Activation of  $\text{Na}^+/\text{H}^+$  exchanger 1 is sufficient to generate  $\text{Ca}^{2+}$  signals that induce cardiac hypertrophy and heart failure. *Circulation research* 2008; 103(8): 891-9.
- [96] Sedej S, Heinzl FR, Walther S, *et al.*  $\text{Na}^+$ -dependent SR  $\text{Ca}^{2+}$  overload induces arrhythmic events in mouse cardiomyocytes with a human CPVT mutation. *Cardiovascular research* 2010; 87(1): 50-9.
- [97] Gonano LA, Sepulveda M, Rico Y, *et al.* CaMKII Mediates Digitalis-Induced Arrhythmias. *Circ Arrhythm Electrophysiol* 2011; 4: 947-57.
- [98] Baartscheer A, Hardziyenka M, Schumacher CA, *et al.* Chronic inhibition of the  $\text{Na}^+/\text{H}^+$  - exchanger causes regression of hypertrophy, heart failure, and ionic and electrophysiological remodeling. *British journal of pharmacology* 2008; 154(6): 1266-75.



- [99] Rizzi N, Liu N, Napolitano C, *et al.* Unexpected structural and functional consequences of the R33Q homozygous mutation in cardiac calsequestrin: a complex arrhythmogenic cascade in a knock in mouse model. *Circulation research* 2008; 103(3): 298-306.
- [100] Bers DM, Pogwizd SM, Schlotthauer K. Upregulated Na/Ca exchange is involved in both contractile dysfunction and arrhythmogenesis in heart failure. *Basic research in cardiology* 2002; 97 Suppl 1: I36-42.
- [101] Liu N, Ruan Y, Denegri M, *et al.* Calmodulin kinase II inhibition prevents arrhythmias in RyR2<sup>(R4496C+/+)</sup> mice with catecholaminergic polymorphic ventricular tachycardia. *Journal of molecular and cellular cardiology* 2011; 50(1): 214-22.
- [102] Zhao Z, Fefelova N, Shanmugam M, Bishara P, Babu GJ, Xie LH. Angiotensin II induces afterdepolarizations via reactive oxygen species and calmodulin kinase II signaling. *Journal of molecular and cellular cardiology* 2011; 50(1): 128-36.
- [103] Liu N, Rizzi N, Boveri L, Priori SG. Ryanodine receptor and calsequestrin in arrhythmogenesis: what we have learnt from genetic diseases and transgenic mice. *Journal of molecular and cellular cardiology* 2009; 46(2): 149-59.
- [104] Venetucci LA, Trafford AW, O'Neill SC, Eisner DA. The sarcoplasmic reticulum and arrhythmogenic calcium release. *Cardiovascular research* 2008; 77(2): 285-92.
- [105] Yano M. Ryanodine receptor as a new therapeutic target of heart failure and lethal arrhythmia. *Circ J* 2008; 72(4): 509-14.
- [106] Tateishi H, Yano M, Mochizuki M, *et al.* Defective domain-domain interactions within the ryanodine receptor as a critical cause of diastolic Ca<sup>2+</sup> leak in failing hearts. *Cardiovascular research* 2009; 81(3): 536-45.
- [107] Said M, Becerra R, Valverde CA, *et al.* Calcium-calmodulin dependent protein kinase II (CaMKII): A main signal responsible for early reperfusion arrhythmias. *Journal of molecular and cellular cardiology* 2011; 51(6): 936-44.
- [108] Mochizuki M, Yano M, Oda T, *et al.* Scavenging free radicals by low-dose carvedilol prevents redox-dependent Ca<sup>2+</sup> leak via stabilization of ryanodine receptor in heart failure. *Journal of the American College of Cardiology* 2007; 49(16): 1722-32.
- [109] Terentyev D, Gyorke I, Belevych AE, *et al.* Redox modification of ryanodine receptors contributes to sarcoplasmic reticulum Ca<sup>2+</sup> leak in chronic heart failure. *Circulation research* 2008; 103(12): 1466-72.
- [110] Gonzalez DR, Treuer AV, Castellanos J, Dulce RA, Hare JM. Impaired S-nitrosylation of the ryanodine receptor caused by xanthine oxidase activity contributes to calcium leak in heart failure. *The Journal of biological chemistry* 2010; 285(37): 28938-45.
- [111] Ho HT, Stevens SC, Terentyeva R, Carnes CA, Terentyev D, Gyorke S. Arrhythmogenic adverse effects of cardiac glycosides are mediated by redox modification of ryanodine receptors. *The Journal of physiology* 2011; 589(Pt 19): 4697-708.
- [112] Venetucci LA, Trafford AW, Eisner DA. Increasing ryanodine receptor open probability alone does not produce arrhythmogenic calcium waves: threshold sarcoplasmic reticulum calcium content is required. *Circulation research* 2007; 100(1): 105-11.
- [113] Vila Petroff MG, Aiello EA, Palomeque J, Salas MA, Mattiazzi A. Subcellular mechanisms of the positive inotropic effect of angiotensin II in cat myocardium. *The Journal of physiology* 2000; 529 Pt 1: 189-203.
- [114] Domenighetti AA, Boixel C, Cefai D, Abriel H, Pedrazzini T. Chronic angiotensin II stimulation in the heart produces an acquired long QT syndrome associated with IK1 potassium current downregulation. *Journal of molecular and cellular cardiology* 2007; 42(1): 63-70.
- [115] Salas MA, Vila-Petroff MG, Palomeque J, Aiello EA, Mattiazzi A. Positive inotropic and negative lusitropic effect of angiotensin II: intracellular mechanisms and second messengers. *Journal of molecular and cellular cardiology* 2001; 33(11): 1957-71.
- [116] Rivard K, Paradis P, Nemer M, Fiset C. Cardiac-specific overexpression of the human type 1 angiotensin II receptor causes delayed repolarization. *Cardiovascular research* 2008; 78(1): 53-62.
- [117] Wang YH, Shi CX, Dong F, Sheng JW, Xu YF. Inhibition of the rapid component of the delayed rectifier potassium current in ventricular myocytes by angiotensin II via the AT1 receptor. *British journal of pharmacology* 2008; 154(2): 429-39.
- [118] Zhou C, Ziegler C, Birder LA, Stewart AF, Levitan ES. Angiotensin II and stretch activate NADPH oxidase to destabilize cardiac Kv4.3 channel mRNA. *Circulation research* 2006; 98(8): 1040-7.
- [119] Aiello EA, Cingolani HE. Angiotensin II stimulates cardiac L-type Ca<sup>2+</sup> current by a Ca<sup>2+</sup>- and protein kinase C-dependent mechanism. *Am J Physiol Heart Circ Physiol* 2001; 280(4): H1528-36.
- [120] Ichiyanagi O, Ishii K, Endoh M. Angiotensin II increases L-type Ca<sup>2+</sup> current in gramicidin D-perforated adult rabbit ventricular myocytes: comparison with conventional patch-clamp method. *Pflügers Arch* 2002; 444(1-2): 107-16.
- [121] Nuss HB, Kaab S, Kass DA, Tomaselli GF, Marban E. Cellular basis of ventricular arrhythmias and abnormal automaticity in heart failure. *The American journal of physiology* 1999; 277(1 Pt 2): H80-91.
- [122] Xie LH, Chen F, Karagueuzian HS, Weiss JN. Oxidative-stress-induced afterdepolarizations and calmodulin kinase II signaling. *Circulation research* 2009; 104(1): 79-86.
- [123] Carmeliet E. Action potential duration, rate of stimulation, and intracellular sodium. *Journal of cardiovascular electrophysiology* 2006; 17 Suppl 1: S2-S7.
- [124] Fischer R, Dechend R, Gapelyuk A, *et al.* Angiotensin II-induced sudden arrhythmic death and electrical remodeling. *Am J Physiol Heart Circ Physiol* 2007; 293(2): H1242-53.
- [125] Lebeche D, Kaprielian R, Hajjar R. Modulation of action potential duration on myocyte hypertrophic pathways. *Journal of molecular and cellular cardiology* 2006; 40(5): 725-35.
- [126] Palomeque J, Delbridge L, Petroff MV. Angiotensin II: a regulator of cardiomyocyte function and survival. *Front Biosci* 2009; 14: 5118-33.