



Review

Importance of Endogenous Fibrinolysis in Platelet Thrombus Formation

Ying X. Gue ¹  and Diana A. Gorog ^{1,2,3,*}

¹ Department of Cardiology, East and North Hertfordshire NHS Trust, Hertfordshire SG1 4AB, UK; y.gue@nhs.net

² Department of Postgraduate Medicine, University of Hertfordshire, Hertfordshire AL10 9AB, UK

³ National Heart & Lung Institute, Imperial College, London SW3 6LY, UK

* Correspondence: d.gorog@imperial.ac.uk; Tel.: +44-(0)1-707-247-512

Received: 7 July 2017; Accepted: 21 August 2017; Published: 25 August 2017

Abstract: The processes of thrombosis and coagulation are finely regulated by endogenous fibrinolysis maintaining healthy equilibrium. When the balance is altered in favour of platelet activation and/or coagulation, or if endogenous fibrinolysis becomes less efficient, pathological thrombosis can occur. Arterial thrombosis remains a major cause of morbidity and mortality in the world despite advances in medical therapies. The role endogenous fibrinolysis in the pathogenesis of arterial thrombosis has gained increasing attention in recent years as it presents novel ways to prevent and treat existing diseases. In this review article, we discuss the role of endogenous fibrinolysis in platelet thrombus formation, methods of measurement of fibrinolytic activity, its role in predicting cardiovascular diseases and clinical outcomes and future directions.

Keywords: endogenous; spontaneous; thrombolysis; fibrinolysis; thrombosis; platelets; cardiovascular

1. Background

1.1. Haemostasis

Haemostasis is a complex sequence of biochemical response to injury to allow formation of a blood clot and repair of damaged endothelium. The maintenance of the equilibrium between coagulation and fibrinolysis is vital, as imbalance would lead to abnormal bleeding or increased risk of thrombosis. Thrombosis is pathological clot formation within the blood vessels in the absence of injury. In fact, the recognition of thrombosis has shaped treatment and prevention of vascular events such as acute coronary syndrome (ACS), which is typically caused by arterial thrombosis following the rupture of vulnerable atheroma within the arterial wall [1–3].

1.2. Atherosclerosis and Plaque Rupture

Atherosclerosis is induced by many factors including endothelial dysfunction, elevated low-density lipoprotein (LDL), oxygen free radicals and hypertension. These factors trigger an inflammatory response within the endothelial cells and, with repeated insults, lead to proliferation of the smooth muscle cells and formation of lipid-rich or fibrous plaques which intrude into the lumen and alter blood flow dynamics within [4]. Plaques have varying morphology and propensity to rupture [5]. Rupture of the plaque allows the contents of the plaque and the subendothelium to come into contact with the contents of flowing blood. Emphasis has been placed on thin-cap fibroatheroma (TCFA) as they are considered very vulnerable to rupture and are most causally associated with the occurrence of coronary thrombosis [6].

1.3. Platelet Aggregation and Thrombus Formation

Following plaque rupture, the exposure of tissue factor and collagen creates a pro-thrombotic environment by initiating the coagulation cascade, platelet aggregation and thrombus formation [1]. Von Willebrand factor (vWF), platelet glycoprotein receptors, adenosine diphosphate (ADP), thromboxane A₂ (TxA₂) and thrombin all play important roles in platelet activation and recruitment of other platelets [7]. Thrombin is also responsible for converting fibrinogen into fibrin, which stabilizes the platelet-platelet contacts leading to thrombus formation.

2. Regulators of Endogenous Fibrinolysis

2.1. Endogenous Fibrinolysis

If thrombus can propagate indefinitely, it will lead to complete occlusion of the vessel and loss of blood flow resulting in disastrous tissue damage, as in the case of acute myocardial infarction or cerebrovascular events or acute peripheral vascular occlusion [8]. Endogenous or spontaneous fibrinolysis is the physiological counter-measure against lasting arterial thrombosis. It is divided into two key steps: (1) activation of plasminogen to serine proteinase plasmin by tissue (tPA) and urokinase (uPA) plasminogen activator; and (2) breaking down of fibrin into fibrin degradation products, thereby dissolving the thrombus to allow restoration of blood flow [9].

2.2. Regulation of Fibrinolysis

Hyperfibrinolysis can result in uncontrolled bleeding, as in the case with disseminated intravascular coagulation (DIC) where systemic inflammation causes increased consumption of fibrin and clotting factors. In a healthy individual, fibrinolysis is regulated by inhibiting plasminogen activator or antagonizing plasmin through α -2-antiplasmin. Plasminogen activator inhibitor (PAI) has been found to work on both tPA and uPA [10,11]. Thrombin-activatable fibrinolysis inhibitor (TAFI) is a glycoprotein which works by reducing activation of plasminogen [10]. The structure of the thrombus also impact on its lysis [12,13] and Factor XIII (FXIII) plays an important role [14]. FXIII not only helps in formation of the fibrin network, but also crosslinks α -2-antiplasmin to fibrin, making it more resistant to fibrinolysis [15]. Lipoprotein (a) (Lp(a)) is a subclass of low density lipoprotein which reduces fibrinolysis through competitive inhibition of plasmin [16].

Plasminogen activator inhibitors are the main glycoproteins responsible for inhibiting the actions of tPA and uPA, with PAI-1 being the most relevant as it is produced by platelets and endothelial cells. PAI-2 is produced by the placenta and is only present in detectable amount during pregnancy. Upon stimulation by thrombin, PAI-1 is released from within platelets as a protective mechanism against premature lysis [11]. PAI-1, synthesized in platelets and endothelial cells, binds with tPA and uPA in a 1:1 ratio to form a stable compound which is then cleared by the liver [11,17].

Thrombin-activatable fibrinolysis inhibitor is a fibrinolysis inhibitor that is converted into its active form, TAFIa, during coagulation after thrombin cleavage. It acts by reducing plasminogen binding to fibrin leading to increase in lysis time [10,18,19]. Its generation is dependent on thrombin levels and greatly potentiated by thrombomodulin [19,20].

α -2-antiplasmin is a serine protease inhibitor which is produced in the liver. It inhibits fibrinolysis via three mechanisms: (1) forming a complex with plasmin; (2) inhibits adsorption of plasminogen onto fibrin; and (3) cross-links with FXIIIa to make fibrin more resistant to plasmin [8].

Factor XIII is a clotting factor which is activated by thrombin in the presence of calcium. Its role is in generating cross-links between fibrin strands within fibrin mesh which strengthens it. It also cross-links inhibitors of fibrinolysis, such as α -2-antiplasmin and TAFI, onto fibrin reducing its solubility and making it more resistant to the effects of plasmin [15]. FXIII is present in large quantities within platelets thereby making platelet rich clots more resistant to lysis than whole blood clots [21].

Lipoprotein (a) competitively binds to fibrin as its molecular structure is similar to plasminogen, leading to an anti-fibrinolytic effect [16]. It also increases synthesis of PAI-1 by endothelial cells which

further reduces plasmin levels [16]. Lp(a) is also believed to be involved in atherogenesis and studies have shown links between Lp(a) levels and cardiovascular risk [16,22–24]. A summary of the regulators of fibrinolysis is provided in Table 1.

Table 1. Regulators of fibrinolysis and their role.

Regulators of Fibrinolysis	Role
PAI-1	Inhibits tPA/uPA
TAFI	Decreases binding of plasminogen to fibrin
α -2-antiplasmin	Forms complex with plasmin Increase adsorption of plasminogen on fibrin Crosslinks FXIIIa
FXIII	Generates crosslinks between fibrin strands Crosslinks other inhibitors (TAFI, α -2-antiplasmin) onto fibrin
Lp(a)	Competitively binds to plasminogen Increases synthesis of PAI-1

PAI: Plasminogen activator inhibitor; TAFI: Thrombin-activatable fibrinolysis inhibitor; FXIII: Factor XIII; Lp(a): Lipoprotein (a).

3. Measurement of Endogenous Fibrinolysis

3.1. Clinical Tests

Electrocardiography (ECG) is the gold-standard non-invasive test to evaluate chest pain. Its use has directed classification of ACS and its treatment. The presence of ST-segment elevation on an ECG is usually an indication of acute thrombotic occlusion of the coronary artery. The natural history of ECG changes in ST-segment elevation myocardial infarction (STEMI) begins with hyperacute T-waves, ST-segment elevation, T-wave inversion, ST-segment resolution followed lastly by abnormal Q waves which indicates transmural infarction. Although in almost all patients with STEMI eventually the ST-segment elevation will eventually resolve with time within 12 h, even if reperfusion does not occur, early spontaneous resolution is associated with improved prognosis [25,26]. It is a crude measure of reperfusion and therefore a marker of fibrinolysis (spontaneous or iatrogenic). Serial ECGs can be used in the setting of STEMI to measure the time taken for reperfusion to occur, which is a surrogate measure of fibrinolytic activity. Therefore, patients with delayed ST-segment recovery are considered to be at higher risk and may require further treatment and closer follow up.

Angiography is the use of X-rays and contrast agent to assess blood vessels. In the realm of cardiology, coronary angiography, either by direct fluoroscopy or using computed tomography, is used to assess coronary artery disease. It can identify luminal narrowing within the coronary arteries and flow distal to the disease. Thrombolysis In Myocardial Infarction (TIMI) flow grade is widely used to classify epicardial blood flow [27]. The presence of TIMI 3 flow (normal flow filling distal coronary completely) is associated with improved outcomes compared to lower grades [28]. The restoration of flow on angiography is an indication that fibrinolysis has resulted in the elimination of thrombotic occlusion. Angiography is currently the “gold-standard” measure of the restoration of blood flow in the epicardial coronary arteries in patients with coronary artery disease.

3.2. Laboratory Measurement of Fibrinolysis

Activity of fibrinolytic pathways can be measured through various factorial assays and biomarkers such as tPA [29–31], PAI-1 [32,33], α -2-antiplasmin and plasma- α -2-antiplasmin (PAP) complex [32,34], fibrin degradation products (d-dimer and soluble fibrin) [29,32,34,35], TAFI [32,36] and LP(a) [32,33,37]. Although the measurement of these markers reflects the activity of fibrinolytic pathway, their clinical use and predictive values remains limited [32]. It is very difficult, with so many individual components of the pathway, to build up a complete or comprehensive overview of the efficiency of the fibrinolytic

pathways. Furthermore, the relative contribution of different individual components may vary in different clinical scenarios.

As the structure of the fibrin network regulates mechanical stability and its resistance to fibrinolysis, the assessment of clot structure can provide information about thromboembolic risk and susceptibility to lysis [38]. Fibrin clot permeability is the measure of how tightly packed the fibrin clot is and has been identified to be altered in patients with coronary artery disease (CAD), diabetes and relatives of family with premature CAD [12,39–41]. The method of performing this requires the use of citrated plasma mixed with thrombin to allow clot formation. Volume of buffer flowing through the gel is then added into a formula to calculate the permeation coefficient (K_s) [42]. The main limitation of this assessment is that the range of values is large, thereby making inter-laboratory comparison difficult even with standardisation of measurements [43].

3.3. Clinically Applicable Tests of Fibrinolysis

Thromboelastography or TEG[®] and rotational thromboelastometry or ROTEM[®] are point of care, global test of coagulation status, simultaneously assessing clot development, stabilization, and dissolution. It utilizes a pin suspended by a torsion wire into a cylinder or an optical detector in the case of ROTEM[®], to measure the physical properties of a clot. As blood clot formation occurs around the pin, fibrin strands form between the cylindrical cup and pin. The rotation of the cylindrical cup will be transmitted to the pin whose displacement is then picked up by the torsion wire. This is analysed and presented in graphical form by the machine to allow analysis of different stages of coagulation and fibrinolysis [44]. It is able to provide clot formation time and rate, maximum amplitude (MA) or clot strength and clot lysis time (CLT). It was designed to be used with native blood but modification with different activators and inhibitors have been used [45–47] although the correlation has been poor [48]. Its main use remains in assessing perioperative risks of bleeding, with questionable use with regards to assessment of fibrinolytic status [49]. Its main limitation in assessing fibrinolysis is that it employs a low-flow, static-type situation that more closely resembles venous, rather than arterial thrombosis.

The Global Thrombosis Test (GTT, Thromboquest Ltd., London, UK) is a relatively new automated, point-of-care test that simultaneously assesses platelet reactivity, thrombosis, and fibrinolytic activity, from a native whole blood sample [50]. Blood passing through a plastic conical tube with narrow gaps generates high shear stress. This mimics flow within a narrowed vessel, activates platelets and induces thrombus formation. Reduction of flow, as detected by an optical sensor, is expressed as occlusion time (OT). Blood flow resumes following spontaneous fibrinolysis, and the time taken to do so is expressed as the lysis time (LT). The GTT provides a more comprehensive test to evaluate thrombosis and lysis under physiological conditions that closely resemble arterial thrombosis models. The evaluation of GTT results in association with clinical outcomes has shown promise in various clinical settings for identifying patients at risk of future adverse cardiovascular event [51,52].

3.4. Measuring Lysis and Its Clinical Impact

Cardiovascular disease (CVD) is the leading cause of death globally, accounting for 32% of deaths [53]. There are many interventions in place to modify risk factors for patients at a population level. With the appreciation of the role that thrombosis and fibrinolysis plays in CVD [54], could we equate the measurement of fibrinolysis into defining specific high risk patient groups which could lead to targeted preventative therapy?

Prediction of clinical outcome, or clinical risk of future adverse events, is a vital process of management of patients with pathological conditions. It allows clinicians to decide the need for more aggressive treatments which come with a price of increased likelihood of side effects. Scoring systems have been used in several areas of clinical medicine and normally take into account parameters which can affect clinical outcome. The use of the Global Registry of Acute Coronary Events (GRACE) score to risk stratify patients with ACS and the QRisk2 score used in the UK are examples [55,56]. The use of the GRACE score has allowed hospitals to identify patients who are higher risk for whom in-patient

management and earlier intervention is warranted [57,58]. The QRisk2 score allows the physician to incorporate simple demographics and blood tests to provide patients with their individual risk of a cardiovascular event expressed as a percentage over 10 years [56].

Although ECG is a low cost, low risk and easily available test, its use in screening of asymptomatic, low risk population is negligible [59]. In patients with conventional risk factors for CVD, the use of screening ECG does not add any value to risk management [59]. Early resolution of ST segment elevation on the ECG of patients with ACS that have undergone thrombolysis has been shown to be associated with improved prognosis [25]. In the substudy of HORIZONS-AMI trial, absence of ST-segment resolution post primary percutaneous coronary intervention (PPCI) has been found to be a predictor of major adverse cardiovascular events (MACE) and the need for recurrent target vessel revascularisation [60]. This has also been shown in other trials [61,62]. Lønborg J. et al. [63] evaluated spontaneous ST-segment resolution and identified improved long-term outcome in patients with resolved ST-segment deviation prior to PPCI. Resolution of ST-depression in patients with STEMI by $\geq 50\%$ is associated with more favourable outcomes [64].

With angiography being the standard measure of localised epicardial fibrinolytic activity in patients with STEMI, its usefulness in guiding prognosis is well documented. Early infarct-related artery patency at angiography, an indicator of spontaneous fibrinolysis, is an independent predictor of lower 1 year mortality [65], higher TIMI flow post percutaneous coronary intervention (PCI), more rapid ST-segment resolution and favourable clinical outcome after 90 days [66]. The presence of TIMI 3 flow post PPCI is associated with better outcome [67] and conversely TIMI flow ≤ 2 is associated with adverse events [68].

The use of coronary computed tomography angiography (CCTA) and coronary calcium scoring for detection of coronary artery disease is increasing. There have been various studies which involved the use of CCTA to screen for coronary artery disease. The Subclinical COronary Atherosclerosis Updated With Coronary cT Angiography (SCOUT) study enrolled 1000 asymptomatic individuals of different risk groups and identified plaques in 22% of the patients [69]. However, the prognosis of these patients was good and therefore the risk versus benefit of using screening CCTA remains questionable. A randomised trial looked into screening of high risk diabetics using CCTA (FACTOR-64) did not show evidence to support the use of CCTA screening [70].

Biomarkers, including factorial assays of fibrinolysis pathways have been used with varying degrees of success in identifying high risk cohorts, however their overall usefulness remains controversial and not helpful in individual patients [32]. There has been weak correlation shown between tPA and the development of coronary disease [29,71] however this is not supported other studies [72]. PAI-1 and its relation to cardiovascular events are in similar predicament [72,73]. In predicting outcomes, PAI-1 has been found to be independent predictor of in-hospital and one year mortality [74] and MACE following PCI [33,75]. It has also shown association with higher all cause mortality and ACS [76]. Measurement of TAFI level has had conflicting results with regards to predicting CAD risk with some studies showing a low level being cardioprotective [77,78] whilst others stating the opposite [79,80]. High levels of TAFI levels were shown to predict stent restenosis [81]. A recently published large study found that elevated LP(a) levels were associated with an increased risk of major adverse cardiovascular events (MACE) and CVD especially in diabetic populations [82], whereas previous studies have shown weak correlations [37,83]. However, Lp(a) has shown no associations with mortality in patients with established coronary artery disease [84] or MACE post PCI [33].

Although there are prospects for the use of biomarkers in risk detection of populations at risk, limitations still exist. The limitations stem from the complex nature of fibrinolysis. Although the measurement of different assays may reflect part of the activity, the complex interaction between the structure of the thrombin clot, cells within the arterial blood i.e., neutrophils and effects of drugs added into the mix limits its predictive value [32]. Furthermore, levels of these factorial components may fluctuate over time and in response to multiple clinical factors, so prediction based on an isolated sample has very limited value.

Clot permeability and the assessment of fibrin clot properties are more useful in prediction of venous thromboembolism [14,85,86]. Clots resistant to fibrinolysis have also been identified in patients with end-stage renal disease (ESRD) [87]. In patients with a family history of premature coronary artery disease, fibrin clots are thicker and less permeable [39]. Patients with type 2 diabetes have also been found to have lower permeability clots that are resistant to lysis [88,89]. In a matched case control study, patients with in-stent stenosis were found to have denser clots which were more resistant to lysis [90] and in patients with ischemic strokes, it has been found that less porous clots are associated with neurological deficit [91]. There have been very limited clinical studies exploring the relationship of clot properties with clinical outcome.

TEG[®] and ROTEM[®] have been in use for a long time in the prediction of bleeding and the requirement for blood and blood products in the settings of trauma resuscitation and in operating theatres [92–95]. Although their use in the capacity of hyperfibrinolysis is well described in the literature, with regards of hypofibrinolysis (namely thrombosis risk assessment), it has had less success [96]. High platelet-fibrin clot strength (MA) has been linked to increased ischemic events [97–100], the use of TEG[®] in the detection of patients at CVD risk is extremely limited. A study comparing patients with different levels of angina looked at time to clot and clot strength using TEG[®] and has shown a stepwise increase in clot strength from clinically stable to unstable stages of angina [101]. This might be an area where TEG[®] may provide some insight.

Clinical studies evaluating the Global Thrombosis Test have shown relation between lysis time (LT), a measure of endogenous fibrinolysis, and MACE. A study of 300 patients with ACS revealed that LT >3000 s was identified as the optimal cut-point to predict MACE [51]. Even with adjustments for baseline cardiovascular risk factors, LT remained a statistically significant predictor of recurrent future MACE. In patients undergoing PPCI, pre-PPCI impaired endogenous fibrinolysis is associated with MACE whilst intact fibrinolysis was associated with spontaneous reperfusion, ST resolution and TIMI 3 flow pre-PPCI [102]. There have been clinical studies evaluating the use of the Global Thrombosis Test in the context of cardiovascular risk factors such as smoking and metabolic syndrome [103–105] and they have shown correlation between differences in LT and presence of risk factor. The GTT shows promise in the use of this technique for risk factor detection as well as assessing the impact of particular risk factors on thrombotic status.

The methods of measuring fibrinolysis and their clinical applications are summarised in Table 2.

Table 2. Summary of measurements of fibrinolysis, its clinical application, strengths and limitations.

Methods of Measurement of Fibrinolysis	Clinical Application		Strengths and Limitations
	Detection of High Risk Patients	Prediction of Clinical Outcomes	
Electrocardiography (ECG)	Screening ECG does not add value to risk management. Might lead to unnecessary downstream investigations and invasive tests.	ST change resolution does help predict outcomes in patients with ACS	Low cost, low risk. Easily available.
Angiography	Coronary computed tomography angiography (CCTA) screening has been able to identify coronary plaques in asymptomatic individuals.	Well documented evidence of epicardial blood flow on angiogram and correlation to outcomes	Standard test. Invasive and requires use of ionising radiation.
Factorial Assays	Varying reports of correlation between different assays and development of cardiovascular disease	TAFI and PAI 1 have been found to correlate with clinical outcomes.	Fluctuates over time and in response to other clinical factors

Table 2. Cont.

Methods of Measurement of Fibrinolysis	Clinical Application		Strengths and Limitations
	Detection of High Risk Patients	Prediction of Clinical Outcomes	
Assessment of Clot Structure	Clot structure has been found to be more resistant to lysis in some patients with higher risk.	Limited studies to associate with clinical outcomes	Complex procedure to analyse samples.
Thromboelastography (TEG)	Has shown difference in clot strength (MA) in patients with varying levels of unstable angina	Only clot strength (MA) has been shown to help predict outcome.	Not relevant to arterial thrombosis. Established method in hyperfibrinolytic status, i.e., bleeding. Limited data on hypofibrinolysis. No standard method of using TEG (native or with additives).
The Global Thrombosis Test (GTT)	Clinical studies have shown correlation of impaired lysis with conventional CV risk factors. Impaired lysis is able to detect high risk ACS patients and high risk PPCI patients.	Impaired fibrinolysis shown to be predictive of recurrent adverse cardiovascular events in patients with acute coronary syndrome and renal impairment.	Point of care test. High shear stress which mimics flow within narrowed vessel. Relatively new therefore limited data.

4. Current Situation, Limitations and Future Directions

Cardiovascular disease remains the leading cause of mortality and morbidity in the world despite many prevention and treatment strategies. Currently in the UK, the screening for cardiovascular disease is done thorough risk scores (such as QRisk2) [56] to allow primary care practitioners to start primary prevention treatment for patients to avoid future thrombotic events. Traditional cardiovascular risk factors such as smoking, hypertension and hypercholesterolemia are the targets for primary prevention drugs.

Current treatment for arterial thrombosis relies on antiplatelet agents, such as aspirin, and P2Y₁₂ inhibitors, such as clopidogrel or ticagrelor, to help alleviate the pro-thrombotic environment. However, use of such medications to reduce thrombosis in allcomers puts many patients at unnecessary risk of bleeding [106,107].

Although the use of fibrinolytic therapy is still recommended for STEMI if PPCI cannot be provided within 120 min [108], its use remains limited outside of it. With the understanding of the importance of spontaneous, endogenous fibrinolysis and its role in modulating platelet thrombus formation, medication that impacts favourably on this pathway may be an alternative approach to reperfusion or to regulate thrombus formation. This could be in the form of inhibition of regulators of fibrinolysis (TAFI inhibitors [109,110], PAI-1 inhibitors [111,112] and α -2 plasmin inhibitors [113,114]).

The use of clinical tests such as ECG is incorporated as a baseline assessment of chest pain to allow classification of pathology i.e. STEMI, NSTEMI, unstable angina which is useful in specific treatment pathways to be followed. Angiography with PCI is the main treatment for patients presenting with ACS. These clinical tests will likely remain as the backbone for assessment of CVD.

Laboratory assays of individual components of the fibrinolytic pathway may be used as a research tool in population studies, but plays little role in the clinical setting in individual patients as the use of such factorial assays to determine risk and clinical outcome has been controversial. Although Lp(a)

has shown promise in identifying high risk patients with diabetes [82], there is still work to be done for it to have a place in risk stratification for CVD. The lack of standardised approach to fibrin clot structure assessment and permeability studies limits reproducibility and usefulness [43]. However, the possibility of an automated system might allow this technique to be used more widely [115].

Point-of-care global assays have a more important role within the clinical environment. Thromboelastography, including TEG[®] and ROTEM[®], has become well established in the assessment hyperfibrinolysis, namely bleeding, in emergency and theatre settings but has not been shown to help predict recurrent thrombotic events due to hypofibrinolysis. Standardisation techniques are also required to allow for universal interpretation of results [116]. The Global Thrombosis Test provides a more physiological assessment of both thrombus formation and thrombus lysis in a high shear setting reflective of arterial flow conditions. Studies suggest that it may play a role in identifying patients at high risk of future cardiovascular events and predict outcomes in patients following cardiovascular events.

5. Conclusions

Endogenous fibrinolysis plays an important role in impeding the continuous propagation of a growing platelet thrombus. It is an expanding area of interest in cardiovascular research and provides an alternative viewpoint to the approach of treatment of cardiovascular disease. A number of different ways of assessing the capacity of the fibrinolytic pathways are available, which may aid future cardiovascular risk stratification and the tailoring of medications to favourably alter this.

Acknowledgments: This article is not funded by any external sources.

Author Contributions: Ying X Gue drafted the article and made revisions. Diana A Gorog critically reviewed, edited and approved the final version of the article.

Conflicts of Interest: Diana A Gorog is related by family to a company director in Thromboquest Ltd., but has no financial involvement or equity interest in, and has received no financial assistance, support or grant from the aforementioned company. There are no other contracts, benefits or personal relationships to disclose.

Abbreviations

ACS	Acute Coronary Syndrome
ADP	Adenosine diphosphate
CAD	Coronary artery disease
CCTA	Coronary computed tomography angiography
CVD	Cardiovascular disease
CLT	Clot lysis time
DIC	Disseminated intravascular coagulation
ECG	Electrocardiogram
ESRD	End-stage renal disease
FXIII	Factor XIII
GRACE	Global Registry of Acute Coronary Event
GTT	Global thrombosis test
Ks	Permeation coefficient/fibrin clot permeability
LDL	Low density lipoprotein
Lp(a)	Lipoprotein (a)
LT	Lysis time
MA	Maximum amplitude
MACE	Major adverse cardiovascular event
OT	Occlusion time
PAI	Plasmin activator inhibitor
PAP	Plasma- α 2-antiplasmin
PCI	Percutaneous coronary intervention
PPCI	Primary percutaneous coronary intervention
ROTEM	Rotational thromboelastometry

STEMI	ST-segment elevation myocardial infarction
TAFI	Thrombin-activatable fibrinolysis inhibitor
TCFA	Thin-cap fibroatheroma
TEG	Thromboelastography
TIMI	Thrombolysis in myocardial infarction
tPA	Tissue plasminogen activator
TxA2	Thromboxane A2
uPA	Urokinase plasminogen activator
vWF	Von Willebrand Factor

References

- Reininger, A.J.; Bernlochner, I.; Penz, S.M.; Ravanat, C.; Smethurst, P.; Farndale, R.W.; Gachet, C.; Brandl, R.; Siess, W. A 2-Step mechanism of arterial thrombus formation induced by human atherosclerotic plaques. *J. Am. Coll. Cardiol.* **2010**, *55*, 1147–1158. [[CrossRef](#)] [[PubMed](#)]
- Fuster, V.; Moreno, P.R.; Fayad, Z.A.; Corti, R.; Badimon, J.J. Atherothrombosis and high-risk plaque Part I: Evolving concepts. *J. Am. Coll. Cardiol.* **2005**, *46*, 937–954. [[CrossRef](#)] [[PubMed](#)]
- Bonaca, M.P.; Steg, P.G.; Feldman, L.J.; Canales, J.F.; Ferguson, J.J.; Wallentin, L.; Califf, R.M.; Harrington, R.A.; Giugliano, R.P. Antithrombotics in acute coronary syndromes. *J. Am. Coll. Cardiol.* **2009**, *54*, 969–984. [[CrossRef](#)] [[PubMed](#)]
- Epstein, F.H.; Ross, R. Atherosclerosis? An inflammatory disease. *N. Engl. J. Med.* **1999**, *340*, 115–126. [[CrossRef](#)]
- Virmani, R.; Kolodgie, F.D.; Burke, A.P.; Farb, A.; Schwartz, S.M. Lessons from sudden coronary death: A comprehensive morphological classification scheme for atherosclerotic lesions. *Arterioscler. Thromb. Vasc. Biol.* **2000**, *20*, 1262–1275. [[CrossRef](#)] [[PubMed](#)]
- Finn, A.V.; Nakano, M.; Narula, J.; Kolodgie, F.D.; Virmani, R. Concept of vulnerable/unstable plaque. *Arterioscler. Thromb. Vasc. Biol.* **2010**, *30*, 1282–1292. [[CrossRef](#)] [[PubMed](#)]
- Yun, S.-H.; Sim, E.-H.; Goh, R.-Y.; Park, J.-I.; Han, J.-Y. Platelet activation: The mechanisms and potential biomarkers. *Biomed. Res. Int.* **2016**, *2016*, 1–5. [[CrossRef](#)] [[PubMed](#)]
- Carpenter, S.L.; Mathew, P. α 2-Antiplasmin and its deficiency: Fibrinolysis out of balance. *Haemophilia* **2008**, *14*, 1250–1254. [[CrossRef](#)] [[PubMed](#)]
- Longstaff, C.; Kolev, K. Basic mechanisms and regulation of fibrinolysis. *J. Thromb. Haemost.* **2015**, *13*, 98–105. [[CrossRef](#)] [[PubMed](#)]
- Rijken, D.C.; Lijnen, H.R. New insights into the molecular mechanisms of the fibrinolytic system. *J. Thromb. Haemost.* **2009**, *7*, 4–13. [[CrossRef](#)] [[PubMed](#)]
- Zhu, Y.; Carmeliet, P.; Fay, W.P. Plasminogen activator inhibitor-1 is a major determinant of arterial thrombolysis resistance. *Circulation* **1999**, *99*, 3050–3055. [[CrossRef](#)] [[PubMed](#)]
- Undas, A.; Ariens, R.A.S. Fibrin clot structure and function. *Arterioscler. Thromb. Vasc. Biol.* **2011**, *31*, e88–e99. [[CrossRef](#)] [[PubMed](#)]
- Gabriel, D.; Muga, K.; Boothroyd, E. The effect of fibrin structure on fibrinolysis. *J. Biol. Chem.* **1992**, *267*, 24259–24263. [[PubMed](#)]
- Undas, A. Fibrin clot properties and their modulation in thrombotic disorders. *Thromb. Haemost.* **2014**, *112*, 32–42. [[CrossRef](#)] [[PubMed](#)]
- Mitchell, J.L.; Lionikiene, A.S.; Fraser, S.R.; Whyte, C.S.; Booth, N.A.; Mutch, N.J. Functional factor XIII-A is exposed on the stimulated platelet surface. *Blood* **2014**, *124*, 3982–3990. [[CrossRef](#)] [[PubMed](#)]
- Deb, A.; Caplice, N.M. Lipoprotein(a): New insights into mechanisms of atherogenesis and thrombosis. *Clin. Cardiol.* **2004**, *27*, 258–264. [[CrossRef](#)] [[PubMed](#)]
- Epstein, F.H.; Kohler, H.P.; Grant, P.J. Plasminogen-activator inhibitor Type 1 and coronary artery disease. *N. Engl. J. Med.* **2000**, *342*, 1792–1801. [[CrossRef](#)]
- Wang, W.; Boffa, M.B.; Bajzar, L.; Walker, J.B.; Nesheim, M.E. A study of the mechanism of inhibition of fibrinolysis by activated thrombin-activatable fibrinolysis inhibitor. *J. Biol. Chem.* **1998**, *273*, 27176–27181. [[CrossRef](#)] [[PubMed](#)]

19. Renucci, J.F.; Grimaux, M.; Morange, P.E.; Gouvernet, J.; Gourmelin, Y.; Alessi, M.C. Thrombin-activatable fibrinolysis inhibitor antigen levels and cardiovascular risk factors. *Arterioscler. Thromb. Vasc. Biol.* **2000**, *20*, 2156–2161.
20. Kokame, K.; Zheng, X.; Sadler, J.E. Activation of thrombin-activable fibrinolysis inhibitor requires epidermal growth factor-like domain 3 of thrombomodulin and is inhibited competitively by protein C. *J. Biol. Chem.* **1998**, *273*, 12135–12139. [[CrossRef](#)] [[PubMed](#)]
21. Jang, I.K.; Gold, H.K.; Ziskind, A.A.; Fallon, J.T.; Holt, R.E.; Leinbach, R.C.; May, J.W.; Collen, D. Differential sensitivity of erythrocyte-rich and platelet-rich arterial thrombi to lysis with recombinant tissue-type plasminogen activator. A possible explanation for resistance to coronary thrombolysis. *Circulation* **1989**, *79*, 920–928. [[CrossRef](#)] [[PubMed](#)]
22. Maranhão, R.C.; Carvalho, P.O.; Strunz, C.C.; Pileggi, F. Lipoprotein(a): Structure, pathophysiology and clinical implications. *Arq. Bras. Cardiol.* **2014**, *103*, 76–84. [[CrossRef](#)] [[PubMed](#)]
23. Corsetti, J.P.; Ryan, D.; Rainwater, D.L.; Moss, A.J.; Zareba, W.; Block, R.C.; Sparks, C.E. Lp(a) and risk of recurrent cardiac events in obese postinfarction patients. *Obesity* **2008**, *16*, 2717–2722. [[CrossRef](#)] [[PubMed](#)]
24. Kim, J.W.; Seo, H.S.; Suh, S.Y.; Choi, C.U.; Kim, E.J.; Rha, S.-W.; Park, C.G.; Oh, D.J. Relationship between Lipoprotein(a) and spontaneous recanalization of infarct-related arteries in the early phase of acute myocardial infarction. *Clin. Cardiol.* **2008**, *31*, 211–216. [[CrossRef](#)] [[PubMed](#)]
25. Zeymer, U.; Schröder, K.; Wegscheider, K.; Senges, J.; Neuhaus, K.-L.; Schröder, R. ST resolution in a single electrocardiographic lead: A simple and accurate predictor of cardiac mortality in patients with fibrinolytic therapy for acute ST-elevation myocardial infarction. *Am. Heart J.* **2005**, *149*, 91–97. [[CrossRef](#)] [[PubMed](#)]
26. Piérard, L.A. ST elevation after myocardial infarction: What does it mean? *Heart* **2007**, *93*, 1329–1330. [[CrossRef](#)] [[PubMed](#)]
27. The TIMI Study Group, 1. The Thrombolysis in Myocardial Infarction (TIMI) trial. *N. Engl. J. Med.* **1985**, *312*, 932–936.
28. Anderson, J.L.; Karagounis, L.A.; Califf, R.M.; Candela, R.; Abbottsmith, C.; Ellis, S.; Sigmon, K.; Kereiakes, D.; George, B.; Stack, R.; et al. Metaanalysis of five reported studies on the relation of early coronary patency grades with mortality and outcomes after acute myocardial infarction. *Am. J. Cardiol.* **1996**, *78*, 1–8. [[CrossRef](#)]
29. May, M.; Lawlor, D.A.; Patel, R.; Rumley, A.; Lowe, G.; Ebrahim, S. Associations of von Willebrand factor, fibrin D-dimer and tissue plasminogen activator with incident coronary heart disease: British Women's heart and health cohort study. *Eur. J. Cardiovasc. Prev. Rehabil.* **2007**, *14*, 638–645. [[CrossRef](#)] [[PubMed](#)]
30. Kinlay, S.; Schwartz, G.G.; Olsson, A.G.; Rifai, N.; Bao, W.; Libby, P.; Ganz, P.; Myocardial Ischemia Reduction with Aggressive Cholesterol Lowering (MIRACL) study investigators. Endogenous tissue plasminogen activator and risk of recurrent cardiac events after an acute coronary syndrome in the MIRACL study. *Atherosclerosis* **2009**, *206*, 551–555. [[CrossRef](#)] [[PubMed](#)]
31. Lee, C.W.; Ahn, J.-M.; Park, D.-W.; Kim, Y.-H.; Hong, M.-K.; Song, J.-K.; Kim, J.-J.; Park, S.-W.; Chi, H.-S.; Park, S.-J. Tissue plasminogen activator on admission is an important predictor of 30-day mortality in patients with acute myocardial infarction undergoing primary angioplasty. *Atherosclerosis* **2008**, *196*, 327–332. [[CrossRef](#)] [[PubMed](#)]
32. Gorog, D.A. Prognostic value of plasma fibrinolysis activation markers in cardiovascular disease. *J. Am. Coll. Cardiol.* **2010**, *55*, 2701–2709. [[CrossRef](#)] [[PubMed](#)]
33. Marcucci, R.; Brogi, D.; Sofi, F.; Giglioli, C.; Valente, S.; Liotta, A.A.; Lenti, M.; Gori, A.M.; Prisco, D.; Abbate, R.; et al. PAI-1 and homocysteine, but not lipoprotein(a) and thrombophilic polymorphisms, are independently associated with the occurrence of major adverse cardiac events after successful coronary stenting. *Heart* **2005**, *92*, 377–381. [[CrossRef](#)] [[PubMed](#)]
34. Morange, P.E.; Bickel, C.; Nicaud, V.; Schnabel, R.; Rupprecht, H.J.; Peetz, D.; Lackner, K.J.; Cambien, F.; Blankenberg, S.; Tiret, L. Haemostatic factors and the risk of cardiovascular death in patients with coronary artery disease: The AtheroGene study. *Arterioscler. Thromb. Vasc. Biol.* **2006**, *26*, 2793–2799. [[CrossRef](#)] [[PubMed](#)]
35. Alehagen, U.; Dahlström, U.; Lindahl, T.L. Elevated D-dimer level is an independent risk factor for cardiovascular death in out-patients with symptoms compatible with heart failure. *Thromb. Haemost.* **2004**, *92*, 1250–1258. [[CrossRef](#)] [[PubMed](#)]

36. Tregouet, D.A.; Schnabel, R.; Alessi, M.C.; Godefroy, T.; Declerck, P.J.; Nicaud, V.; Munzel, T.; Bickel, C.; Rupprecht, H.J.; Lubos, E.; et al. Activated thrombin activatable fibrinolysis inhibitor levels are associated with the risk of cardiovascular death in patients with coronary artery disease: The Athero Gene study. *J. Thromb. Haemost.* **2009**, *7*, 49–57. [[CrossRef](#)] [[PubMed](#)]
37. Erqou, S.; Kaptoge, S.; Perry, P.L.; Di Angelantonio, E.; Thompson, A.; White, I.R.; Marcovina, S.M.; Collins, R.; Thompson, S.G.; Danesh, J. Lipoprotein(a) concentration and the risk of coronary heart disease, stroke, and nonvascular mortality. *JAMA* **2009**, *302*, 412–423. [[CrossRef](#)] [[PubMed](#)]
38. Blombäck, B.; Okada, M. Fibrin gel structure and clotting time. *Thromb. Res.* **1982**, *25*, 51–70. [[CrossRef](#)]
39. Mills, J.D.; Ariëns, R.A.S.; Mansfield, M.W.; Grant, P.J. Altered fibrin clot structure in the healthy relatives of patients with premature coronary artery disease. *Circulation* **2002**, *106*, 1938–1942. [[CrossRef](#)] [[PubMed](#)]
40. Undas, A.; Szuldrzynski, K.; Stepien, E.; Zalewski, J.; Godlewski, J.; Tracz, W.; Pasowicz, M.; Zmudka, K. Reduced clot permeability and susceptibility to lysis in patients with acute coronary syndrome: Effects of inflammation and oxidative stress. *Atherosclerosis* **2008**, *196*, 551–557. [[CrossRef](#)] [[PubMed](#)]
41. Zalewski, J.; Undas, A.; Godlewski, J.; Stepien, E.; Zmudka, K. No-reflow phenomenon after acute myocardial infarction is associated with reduced clot permeability and susceptibility to lysis. *Arterioscler. Thromb. Vasc. Biol.* **2007**, *27*, 2258–2265. [[CrossRef](#)] [[PubMed](#)]
42. Siudut, J.; Grela, M.; Wypasek, E.; Plens, K.; Undas, A. Reduced plasma fibrin clot permeability and susceptibility to lysis are associated with increased risk of postthrombotic syndrome. *J. Thromb. Haemost.* **2016**, *14*, 784–793. [[CrossRef](#)] [[PubMed](#)]
43. Pieters, M.; Undas, A.; Marchi, R.; De Maat, M.P.M.; Weisel, J.W.; Ariens, R.A.S. An international study on the standardization of fibrin clot permeability measurement: Methodological considerations and implications for healthy control values. *J. Thromb. Haemost.* **2012**, *10*, 2179–2181. [[CrossRef](#)] [[PubMed](#)]
44. Thakur, M.; Ahmed, A.B. A review of thromboelastography. *Int. J. Perioper. Ultrasound Appl. Technol.* **2012**, *11*, 25–29. [[CrossRef](#)]
45. Zambruni, A.; Thalheimer, U.; Leandro, G.; Perry, D.; Burroughs, A.K. Thromboelastography with citrated blood: Comparability with native blood, stability of citrate storage and effect of repeated sampling. *Blood Coagul. Fibrinolysis* **2004**, *15*, 103–107. [[CrossRef](#)] [[PubMed](#)]
46. Chen, A.; Teruya, J. Global hemostasis testing thromboelastography: Old technology, new applications. *Clin. Lab. Med.* **2009**, *29*, 391–407. [[CrossRef](#)] [[PubMed](#)]
47. Young, G.; Zhang, R.; Miller, R.; Yassin, D.; Nugent, D.J. Comparison of kaolin and tissue factor activated thromboelastography in haemophilia. *Haemophilia* **2009**, *16*, 518–524. [[CrossRef](#)] [[PubMed](#)]
48. Thalheimer, U.; Triantos, C.K.; Samonakis, D.N.; Zambruni, A.; Senzolo, M.; Leandro, G.; Patch, D.; Burroughs, A.K. A comparison of kaolin-activated versus nonkaolin-activated thromboelastography in native and citrated blood. *Blood Coagul. Fibrinolysis* **2008**, *19*, 495–501. [[CrossRef](#)] [[PubMed](#)]
49. Okafor, O.N.; Gorog, D.A. Endogenous fibrinolysis: An important mediator of thrombus formation and cardiovascular risk. *J. Am. Coll. Cardiol.* **2015**, *65*, 1683–1699. [[CrossRef](#)] [[PubMed](#)]
50. Yamamoto, J.; Inoue, N.; Otsui, K.; Ishii, H.; Gorog, D.A. Global Thrombosis Test (GTT) can detect major determinants of haemostasis including platelet reactivity, endogenous fibrinolytic and thrombin generating potential. *Thromb. Res.* **2014**, *133*, 919–926. [[CrossRef](#)] [[PubMed](#)]
51. Saraf, S.; Christopoulos, C.; Salha, I.B.; Stott, D.J.; Gorog, D.A. Impaired endogenous thrombolysis in acute coronary syndrome patients predicts cardiovascular death and nonfatal myocardial infarction. *J. Am. Coll. Cardiol.* **2010**, *55*, 2107–2115. [[CrossRef](#)] [[PubMed](#)]
52. Farag, M.; Srinivasan, M.; Wellsted, D.; Sullivan, K.; Gorog, D.A. Assessment of endogenous thrombolysis predicts cardiovascular risk in patient with ST-elevation myocardial infarction. *Heart* **2016**, *102*, A69–A70. [[CrossRef](#)]
53. Roth, G.A.; Huffman, M.D.; Moran, A.E.; Feigin, V.; Mensah, G.A.; Naghavi, M.; Murray, C.J.L. Global and regional patterns in cardiovascular mortality from 1990 to 2013. *Circulation* **2015**, *132*, 1667–1678. [[CrossRef](#)] [[PubMed](#)]
54. Barrabés, J.A.; Galian, L. Endogenous thrombolysis. A hidden player in acute coronary syndromes? *J. Am. Coll. Cardiol.* **2010**, *55*, 2116–2117. [[CrossRef](#)] [[PubMed](#)]
55. Granger, C.B.; Goldberg, R.J.; Dabbous, O.; Pieper, K.S.; Eagle, K.A.; Cannon, C.P.; van de Werf, F.; Avezum, A.; Goodman, S.G.; Flather, M.D.; et al. Predictors of hospital mortality in the global registry of acute coronary events. *Arch. Intern. Med.* **2003**, *163*, 2345. [[CrossRef](#)] [[PubMed](#)]

56. Hippisley-Cox, J.; Coupland, C.; Vinogradova, Y.; Robson, J.; Minhas, R.; Sheikh, A.; Brindle, P. Predicting cardiovascular risk in England and Wales: Prospective derivation and validation of QRISK2. *BMJ* **2008**, *336*, 1475–1482. [[CrossRef](#)] [[PubMed](#)]
57. Yan, A.T.; Yan, R.T.; Tan, M.; Eagle, K.A.; Granger, C.B.; Dabbous, O.H.; Fitchett, D.; Grima, E.; Langer, A.; Goodman, S.G.; et al. In-hospital revascularization and one-year outcome of acute coronary syndrome patients stratified by the GRACE risk score. *Am. J. Cardiol.* **2005**, *96*, 913–916. [[CrossRef](#)] [[PubMed](#)]
58. Huang, W.; FitzGerald, G.; Goldberg, R.J.; Gore, J.; McManus, R.H.; Awad, H.; Waring, M.E.; Allison, J.; Saczynski, J.S.; Kiefe, C.I.; et al. Performance of the GRACE risk Score 2.0 simplified algorithm for predicting 1-year death after hospitalization for an acute coronary syndrome in a contemporary multiracial cohort. *Am. J. Cardiol.* **2016**, *118*, 1105–1110. [[CrossRef](#)] [[PubMed](#)]
59. Moyer, V.A.; Greenland, P.; Alpert, J.; Beller, G.; Benjamin, E.; Budoff, M.; Fayad, Z. Screening for coronary heart disease with electrocardiography: U.S. preventive services task force recommendation statement. *Ann. Intern. Med.* **2012**, *122*, 2748–2764. [[CrossRef](#)] [[PubMed](#)]
60. Farkouh, M.E.; Reiffel, J.; Dressler, O.; Nikolsky, E.; Parise, H.; Cristea, E.; Baran, D.A.; Dizon, J.; Merab, J.P.; Lansky, A.J.; et al. Relationship between ST-segment recovery and clinical outcomes after primary percutaneous coronary intervention. *Circ. Cardiovasc. Interv.* **2013**, *6*, 216–223. [[CrossRef](#)] [[PubMed](#)]
61. McLaughlin, M.G.; Stone, G.W.; Aymong, E.; Gardner, G.; Mehran, R.; Lansky, A.J.; Grines, C.L.; Tchong, J.E.; Cox, D.A.; Stuckey, T.; et al. Prognostic utility of comparative methods for assessment of ST-segment resolution after primary angioplasty for acute myocardial infarction. *J. Am. Coll. Cardiol.* **2004**, *44*, 1215–1223. [[CrossRef](#)] [[PubMed](#)]
62. De Luca, G.; Suryapranata, H.; Ottervanger, J.P.; Hoorntje, J.C.A.; Gosselink, A.T.M.; Dambrink, J.-H.; De Boer, M.-J.; van't Hof, A.W.J. Postprocedural single-lead ST-segment deviation and long-term mortality in patients with ST-segment elevation myocardial infarction treated by primary angioplasty. *Heart* **2008**, *94*, 44–47. [[CrossRef](#)] [[PubMed](#)]
63. Lønborg, J.; Kelbæk, H.; Holmvang, L.; Helqvist, S.; Vejstrup, N.; Jørgensen, E.; Saunamäki, K.; Dridi, N.P.; Kløvgaard, L.; Kaltoft, A.; et al. Comparison of outcome of patients with ST-segment elevation myocardial infarction and complete versus incomplete ST-resolution before primary percutaneous coronary intervention. *Am. J. Cardiol.* **2016**, *117*, 1735–1740. [[CrossRef](#)] [[PubMed](#)]
64. Tjandrawidjaja, M.C.; Fu, Y.; Westerhout, C.M.; White, H.D.; Todaro, T.G.; van de Werf, F.; Mahaffey, K.W.; Wagner, G.S.; Granger, C.B.; Armstrong, P.W. Resolution of ST-segment depression: A new prognostic marker in ST-segment elevation myocardial infarction. *Eur. Heart J.* **2010**, *31*, 573–581. [[CrossRef](#)] [[PubMed](#)]
65. Rakowski, T.; Dudek, D.; Dziewierz, A.; Yu, J.; Witzensbichler, B.; Guagliumi, G.; Kornowski, R.; Hartmann, F.; Lansky, A.J.; Brener, S.J.; et al. Impact of infarct-related artery patency before primary PCI on outcome in patients with ST-segment elevation myocardial infarction: The HORIZONS-AMI trial. *EuroIntervention* **2013**, *8*, 1307–1314. [[CrossRef](#)] [[PubMed](#)]
66. Zeymer, U.; Huber, K.; Fu, Y.; Ross, A.; Granger, C.; Goldstein, P.; van de Werf, F.; Armstrong, P.; ASSENT-4 PCI Investigators. Impact of TIMI 3 patency before primary percutaneous coronary intervention for ST-elevation myocardial infarction on clinical outcome: Results from the ASSENT-4 PCI study. *Eur. Heart J. Acute Cardiovasc. Care* **2012**, *1*, 136–142. [[CrossRef](#)] [[PubMed](#)]
67. Kammler, J.; Kypta, A.; Hofmann, R.; Kerschner, K.; Grund, M.; Sihorsch, K.; Steinwender, C.; Lambert, T.; Helml, W.; Leisch, F. TIMI 3 flow after primary angioplasty is an important predictor for outcome in patients with acute myocardial infarction. *Clin. Res. Cardiol.* **2009**, *98*, 165–170. [[CrossRef](#)] [[PubMed](#)]
68. Mehta, R.H.; Harjai, K.J.; Cox, D.; Stone, G.W.; Brodie, B.; Boura, J.; O'Neill, W.; Grines, C.L.; Primary Angioplasty in Myocardial Infarction (PAMI) investigators. Clinical and angiographic correlates and outcomes of suboptimal coronary flow inpatients with acute myocardial infarction undergoing primary percutaneous coronary intervention. *J. Am. Coll. Cardiol.* **2003**, *42*, 1739–1746. [[CrossRef](#)] [[PubMed](#)]
69. Choi, E.-K.; Choi, S.I.; Rivera, J.J.; Nasir, K.; Chang, S.-A.; Chun, E.J.; Kim, H.-K.; Choi, D.-J.; Blumenthal, R.S.; Chang, H.-J. Coronary computed tomography angiography as a screening tool for the detection of occult coronary artery disease in asymptomatic individuals. *J. Am. Coll. Cardiol.* **2008**, *52*, 357–365. [[CrossRef](#)] [[PubMed](#)]
70. Muhlestein, J.B.; Lappé, D.L.; Lima, J.A.C.; Rosen, B.D.; May, H.T.; Knight, S.; Bluemke, D.A.; Towner, S.R.; Le, V.; Bair, T.L.; et al. Effect of Screening for coronary artery disease using CT angiography on mortality and cardiac events in high-risk patients with diabetes. *JAMA* **2014**, *312*, 2234. [[CrossRef](#)] [[PubMed](#)]

71. Gram, J.; Bladbjerg, E.-M.; Moller, L.; Sjol, A.; Jespersen, J. Tissue-type plasminogen activator and C-reactive protein in acute coronary heart disease. A nested case-control study. *J. Intern. Med.* **2000**, *247*, 205–212. [[CrossRef](#)] [[PubMed](#)]
72. Smith, A.; Patterson, C.; Yarnell, J.; Rumley, A.; Ben-Shlomo, Y.; Lowe, G. Which hemostatic markers add to the predictive value of conventional risk factors for coronary heart disease and ischemic stroke? *Circulation* **2005**, *112*, 3080–3087. [[CrossRef](#)] [[PubMed](#)]
73. Wang, T.J.; Gona, P.; Larson, M.G.; Tofler, G.H.; Levy, D.; Newton-Cheh, C.; Jacques, P.F.; Rifai, N.; Selhub, J.; Robins, S.J.; et al. Multiple biomarkers for the prediction of first major cardiovascular events and death. *N. Engl. J. Med.* **2006**, *355*, 2631–2639. [[CrossRef](#)] [[PubMed](#)]
74. Akkus, M.N.; Polat, G.; Yurtdas, M.; Akcay, B.; Ercetin, N.; Cicek, D.; Doven, O.; Sucu, N. Admission levels of C-reactive protein and plasminogen activator Inhibitor-1 in patients with acute myocardial infarction with and without cardiogenic shock or heart failure on admission. *Int. Heart J.* **2009**, *50*, 33–45. [[CrossRef](#)] [[PubMed](#)]
75. Golukhova, E.Z.; Grigorian, M.V.; Ryabinina, M.N.; Bulaeva, N.I.; Fortmann, S.; Serebruany, V.L. Independent predictors of major adverse events following coronary stenting over 28 months of follow-up. *Cardiology* **2015**, *132*, 176–181. [[CrossRef](#)] [[PubMed](#)]
76. Battes, L.C.; Akkerhuis, K.M.; Cheng, J.M.; Garcia-Garcia, H.M.; Oemrawsingh, R.M.; de Boer, S.P.M.; Regar, E.; van Geuns, R.-J.; Serruys, P.W.; Boersma, E.; et al. Circulating acute phase proteins in relation to extent and composition of coronary atherosclerosis and cardiovascular outcome: Results from the ATHEROREMO-IVUS study. *Int. J. Cardiol.* **2014**, *177*, 847–853. [[CrossRef](#)] [[PubMed](#)]
77. De Bruijne, E.L.E.; Gils, A.; Guimaraes, A.H.C.; Dippel, D.W.J.; Deckers, J.W.; van Den Meiracker, A.H.; Poldermans, D.; Rijken, D.C.; Declercq, P.J.; de Maat, M.P.M.; et al. The role of thrombin activatable fibrinolysis inhibitor in arterial thrombosis at a young age: The ATTAC study. *J. Thromb. Haemost.* **2009**, *7*, 919–927. [[CrossRef](#)] [[PubMed](#)]
78. Schroeder, V.; Chatterjee, T.; Mehta, H.; Windecker, S.; Pham, T.; Devantay, N.; Meier, B.; Kohler, H.P. Thrombin activatable fibrinolysis inhibitor (TAFI) levels in patients with coronary artery disease investigated by angiography. *Thromb. Haemost.* **2002**, *88*, 1020–1025. [[PubMed](#)]
79. Meltzer, M.E.; Doggen, C.J.M.; de Groot, P.G.; Meijers, J.C.M.; Rosendaal, F.R.; Lisman, T. Low thrombin activatable fibrinolysis inhibitor activity levels are associated with an increased risk of a first myocardial infarction in men. *Haematologica* **2009**, *94*, 811–818. [[CrossRef](#)] [[PubMed](#)]
80. Juhan-Vague, I.; Morange, P.E.; Aubert, H.; Henry, M.; Aillaud, M.F.; Alessi, M.C.; Samnegård, A.; Hawe, E.; Yudkin, J.; Margaglione, M.; et al. Plasma thrombin-activatable fibrinolysis inhibitor antigen concentration and genotype in relation to myocardial infarction in the north and south of Europe. *Arterioscler. Thromb. Vasc. Biol.* **2002**, *22*, 867–873. [[CrossRef](#)] [[PubMed](#)]
81. Lau, H.K.; Segev, A.; Hegele, R.A.; Sparkes, J.D.; Teitel, J.M.; Chisholm, R.J.; Strauss, B.H. Thrombin-activatable fibrinolysis inhibitor (TAFI): A novel predictor of angiographic coronary restenosis. *Thromb. Haemost.* **2003**, *90*, 1187–1191. [[CrossRef](#)] [[PubMed](#)]
82. Waldeyer, C.; Makarova, N.; Zeller, T.; Schnabel, R.B.; Brunner, F.J.; Jrgensen, T.; Linneberg, A.; Niiranen, T.; Salomaa, V.; Jousilahti, P.; et al. Lipoprotein(a) and the risk of cardiovascular disease in the European population: Results from the BiomarCaRE consortium. *Eur. Heart J.* **2017**. [[CrossRef](#)] [[PubMed](#)]
83. Pineda, J.; Marin, F.; Marco, P.; Roldan, V.; Valencia, J.; Ruiz-Nodar, J.M.; Sogorb, F.; Lip, G.Y.H. Premature coronary artery disease in young (age < 45) subjects: Interactions of lipid profile, thrombophilic and haemostatic markers. *Int. J. Cardiol.* **2009**, *136*, 222–225. [[CrossRef](#)] [[PubMed](#)]
84. Zewinger, S.; Kleber, M.E.; Tragante, V.; McCubrey, R.O.; Schmidt, A.F.; Direk, K.; Laufs, U.; Werner, C.; Koenig, W.; Rothenbacher, D.; et al. Relations between lipoprotein(a) concentrations, LPA genetic variants, and the risk of mortality in patients with established coronary heart disease: A molecular and genetic association study. *Lancet Diabetes Endocrinol.* **2017**. [[CrossRef](#)]
85. Undas, A.; Zawilska, K.; Ciesla-Dul, M.; Lehmann-Kopydlowska, A.; Skubiszak, A.; Ciepluch, K.; Tracz, W. Altered fibrin clot structure/function in patients with idiopathic venous thromboembolism and in their relatives. *Blood* **2009**, *114*, 4272–4278. [[CrossRef](#)] [[PubMed](#)]
86. Zabczyk, M.; Plens, K.; Wojtowicz, W.; Undas, A. Prothrombotic fibrin clot phenotype is associated with recurrent pulmonary embolism after discontinuation of anticoagulant therapy. *Arterioscler. Thromb. Vasc. Biol.* **2016**. [[CrossRef](#)]

87. Undas, A.; Kolarz, M.; Kope, G.; Tracz, W. Altered fibrin clot properties in patients on long-term haemodialysis: Relation to cardiovascular mortality. *Nephrol. Dial. Transplant.* **2008**, *23*, 2010–2015. [[CrossRef](#)] [[PubMed](#)]
88. Konieczynska, M.; Fil, K.; Bazanek, M.; Undas, A. Prolonged duration of type 2 diabetes is associated with increased thrombin generation, prothrombotic fibrin clot phenotype and impaired fibrinolysis. *Thromb. Haemost.* **2013**, *111*, 685–693. [[CrossRef](#)] [[PubMed](#)]
89. Alzahrani, S.H.; Hess, K.; Price, J.F.; Strachan, M.; Baxter, P.D.; Cubbon, R.; Phoenix, F.; Gamlen, T.; Arins, R.A.S.; Grant, P.J.; et al. Gender-specific alterations in fibrin structure function in type 2 diabetes: Associations with cardiometabolic and vascular markers. *J. Clin. Endocrinol. Metab.* **2012**, *97*, 2282–2287. [[CrossRef](#)] [[PubMed](#)]
90. Undas, A.; Zalewski, J.; Krochin, M.; Siudak, Z.; Sadowski, M.; Pregowski, J.; Dudek, D.; Janion, M.; Witkowski, A.; Zmudka, K. Altered plasma fibrin clot properties are associated with in-stent thrombosis. *Arterioscler. Thromb. Vasc. Biol.* **2010**, *30*, 276–282. [[CrossRef](#)] [[PubMed](#)]
91. Undas, A.; Slowik, A.; Wolkow, P.; Szczudlik, A.; Tracz, W. Fibrin clot properties in acute ischemic stroke: Relation to neurological deficit. *Thromb. Res.* **2010**, *125*, 357–361. [[CrossRef](#)] [[PubMed](#)]
92. Davenport, R.; Khan, S. Management of major trauma haemorrhage: Treatment priorities and controversies. *Br. J. Haematol.* **2011**, *155*, 537–548. [[CrossRef](#)] [[PubMed](#)]
93. MacIvor, D.; Rebel, A.; Hassan, Z.-U. How do we integrate thromboelastography with perioperative transfusion management? *Transfusion* **2013**, *53*, 1386–1392. [[CrossRef](#)] [[PubMed](#)]
94. Shore-Lesserson, L.; Manspeizer, H.E.; DePerio, M.; Francis, S.; Vela-Cantos, F.; Ergin, M.A. Thromboelastography-guided transfusion algorithm reduces transfusions in complex cardiac surgery. *Anesth. Analg.* **1999**, *88*, 312–319. [[CrossRef](#)] [[PubMed](#)]
95. Whiting, D.; Dinardo, J.A. TEG and ROTEM: Technology and clinical applications. *Am. J. Hematol.* **2014**, *89*, 228–232. [[CrossRef](#)] [[PubMed](#)]
96. Pepperell, D.; Morel-Kopp, M.-C.; Ward, C. Clinical application of fibrinolytic assays. In *Fibrinolysis and Thrombolysis*; InTech: Rijeka, Croatia, 2014.
97. Jeong, Y.-H.; Bliden, K.P.; Shuldiner, A.R.; Tantry, U.S.; Gurbel, P.A. Thrombin-induced platelet-fibrin clot strength: Relation to high on-clopidogrel platelet reactivity, genotype, and post-percutaneous coronary intervention outcomes. *Thromb. Haemost.* **2013**, *111*, 713–724. [[CrossRef](#)] [[PubMed](#)]
98. Gurbel, P.A.; Bliden, K.P.; Navickas, I.A.; Mahla, E.; Dichiaro, J.; Suarez, T.A.; Antonino, M.J.; Tantry, U.S.; Cohen, E. Adenosine diphosphate-induced platelet-fibrin clot strength: A new thrombelastographic indicator of long-term poststenting ischemic events. *Am. Heart J.* **2010**, *160*, 346–354. [[CrossRef](#)] [[PubMed](#)]
99. Gurbel, P.A.; Bliden, K.P.; Guyer, K.; Cho, P.W.; Zaman, K.A.; Kreutz, R.P.; Bassi, A.K.; Tantry, U.S. Platelet reactivity in patients and recurrent events post-stenting. *J. Am. Coll. Cardiol.* **2005**, *46*, 1820–1826. [[CrossRef](#)] [[PubMed](#)]
100. Gurbel, P.A.; Bliden, K.P.; Kreutz, R.P.; Dichiaro, J.; Antonino, M.J.; Tantry, U.S. The link between heightened thrombogenicity and inflammation: Pre-procedure characterization of the patient at high risk for recurrent events after stenting. *Platelets* **2009**, *20*, 97–104. [[CrossRef](#)] [[PubMed](#)]
101. Tantry, U.S.; Bliden, K.P.; Suarez, T.A.; Kreutz, R.P.; Dichiaro, J.; Gurbel, P.A. Hypercoagulability, platelet function, inflammation and coronary artery disease acuity: Results of the Thrombotic Risk Progression (TRIP) study. *Platelets* **2010**, *21*, 360–367. [[CrossRef](#)] [[PubMed](#)]
102. Christopoulos, C.; Farag, M.; Sullivan, K.; Wellsted, D.; Gorog, D.A. Impaired thrombolytic status predicts adverse cardiac events in patients undergoing primary percutaneous coronary intervention. *Thromb. Haemost.* **2017**, *117*, 457–470. [[CrossRef](#)] [[PubMed](#)]
103. Suehiro, A.; Wakabayashi, I.; Yamashita, T.; Yamamoto, J. Attenuation of spontaneous thrombolytic activity measured by the global thrombosis test in male habitual smokers. *J. Thromb. Thrombolysis* **2014**, *37*, 414–418. [[CrossRef](#)] [[PubMed](#)]
104. Ikarugi, H.; Yamashita, T.; Aoki, R.; Ishii, H.; Kanki, K.; Yamamoto, J. Impaired spontaneous thrombolytic activity in elderly and in habitual smokers, as measured by a new global thrombosis test. *Blood Coagul. Fibrinolysis* **2003**, *14*, 781–784. [[CrossRef](#)] [[PubMed](#)]
105. Suehiro, A.; Wakabayashi, I.; Uchida, K.; Yamashita, T.; Yamamoto, J.; Trevisan, M. Impaired spontaneous thrombolytic activity measured by global thrombosis test in males with metabolic syndrome. *Thromb. Res.* **2012**, *129*, 499–501. [[CrossRef](#)] [[PubMed](#)]

106. Mega, J.L.; Braunwald, E.; Wiviott, S.D.; Bassand, J.-P.; Bhatt, D.L.; Bode, C.; Burton, P.; Cohen, M.; Cook-Bruns, N.; Fox, K.A.A.; et al. Rivaroxaban in patients with a recent acute coronary syndrome. *N. Engl. J. Med.* **2012**, *366*, 9–19. [[CrossRef](#)] [[PubMed](#)]
107. Sørensen, R.; Gislason, G. Triple antithrombotic therapy: Risky but sometimes necessary. *Rev. Esp. Cardiol. (Engl. Ed.)* **2014**, *67*, 171–175. [[CrossRef](#)]
108. National Institute for Health and Care Excellence. Myocardial Infarction with ST-Segment Elevation: Acute Management. Available online: <https://www.nice.org.uk/guidance/cg167> (accessed on 22 August 2017).
109. Zhou, X.; Weeks, S.D.; Ameloot, P.; Callewaert, N.; Strelkov, S.V.; Declerck, P.J. Elucidation of the molecular mechanisms of two nanobodies that inhibit thrombin-activatable fibrinolysis inhibitor activation and activated thrombin-activatable fibrinolysis inhibitor activity. *J. Thromb. Haemost.* **2016**, *14*, 1629–1638. [[CrossRef](#)] [[PubMed](#)]
110. Plug, T.; Marquart, J.A.; Marx, P.F.; Meijers, J.C.M. Selective modulation of thrombin-activatable fibrinolysis inhibitor (TAFI) activation by thrombin or the thrombin-thrombomodulin complex using TAFI-derived peptides. *J. Thromb. Haemost.* **2015**, *13*, 2093–2101. [[CrossRef](#)] [[PubMed](#)]
111. Rupin, A.; Gaertner, R.; Mennecier, P.; Richard, I.; Benoist, A.; De Nanteuil, G.; Verbeuren, T.J. S35225 is a direct inhibitor of Plasminogen Activator Inhibitor type-1 activity in the blood. *Thromb. Res.* **2008**, *122*, 265–270. [[CrossRef](#)] [[PubMed](#)]
112. Wyseure, T.; Rubio, M.; Denorme, F.; Martinez de Lizarrondo, S.; Peeters, M.; Gils, A.; De Meyer, S.F.; Vivien, D.; Declerck, P.J. Innovative thrombolytic strategy using a heterodimer diabody against TAFI and PAI-1 in mouse models of thrombosis and stroke. *Blood* **2015**, *125*, 1325–1332. [[CrossRef](#)] [[PubMed](#)]
113. Sakata, Y.; Eguchi, Y.; Mimuro, J.; Matsuda, M.; Sumi, Y. Clot lysis induced by a monoclonal antibody against α -2-plasmin inhibitor. *Blood* **1989**, *74*, 2692–2697. [[PubMed](#)]
114. Reed, G.L.; Matsueda, G.R.; Haber, E. Inhibition of clot-bound alpha 2-antiplasmin enhances in vivo thrombolysis. *Circulation* **1990**, *82*, 164–168. [[CrossRef](#)] [[PubMed](#)]
115. Ząbczyk, M.; Piłat, A.; Awsiuk, M.; Undas, A. An automated method for fibrin clot permeability assessment. *Blood Coagul. Fibrinolysis* **2015**, *26*, 104–109. [[CrossRef](#)] [[PubMed](#)]
116. Quarterman, C.; Shaw, M.; Johnson, I.; Agarwal, S. Intra- and inter-centre standardisation of thromboelastography (TEG[®]). *Anaesthesia* **2014**, *69*, 883–890. [[CrossRef](#)] [[PubMed](#)]



© 2017 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).