#### **ORIGINAL ARTICLE**



# Comparison of clinical and microbiological diagnoses for older adults with COVID-19 in Wuhan: a retrospective study

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#### **Abstract**

**Background** The potential differences between a clinical diagnosis of coronavirus disease 2019 (COVID-19) (i.e., symptoms without positive virus test) and a microbiological diagnosis (i.e., positive virus test results) of COVID-19 are not known. **Aims** This study explored the differences between the two types of COVID-19 diagnosis among older patients in terms of clinical characteristics and outcomes.

Methods A total of 244 inpatients aged ≥ 60 years with COVID-19 were included in this study, of whom 52 were clinically diagnosed and 192 were microbiologically diagnosed. Clinical and laboratory data on hospital admission and outcomes (discharged or died in hospital) of all patients were retrieved from medical records retrospectively. Patients who met the criteria for clinical diagnosis with negative virus test results were assigned to the clinical diagnosis group, whereas those with positive virus test results were assigned to the microbiological diagnosis group. After univariate analyses, two propensity score analyses [i.e., covariate adjustment using propensity score (CAPS) and propensity score matching (PSM)] were conducted to control bias.

**Results** The clinical and microbiological diagnosis groups demonstrated significant differences in outcomes and in the majority of laboratory findings. After propensity score analyses, many differences between the two groups disappeared and the rate of mortality had no statistically significant difference (P = 0.318 and 0.828 for CAPS and PSM, respectively). **Conclusions** Patients with similar signs, symptoms, and laboratory and imaging findings as confirmed COVID-19 cases may have a similar mortality risk, regardless of the virus test results, and require timely intervention to reduce their mortality.

Keywords Clinical diagnosis · COVID-19 · Microbiological diagnosis · Older patients · SARS-CoV-2

## Introduction

At the end of December 2019, cases of an acute pneumonia of unknown origin were detected in Wuhan, Hubei Province, China [1, 2]. Following the unremitting efforts of clinicians and scientists, the pathogen was isolated and identified as

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severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) [3]. The disease caused by it was officially named as coronavirus disease 2019 (COVID-19) by the World Health Organization (WHO) [2]. Because of the global outbreak and rapidly increasing numbers of patients and deaths, the WHO has declared COVID-19 as a worldwide pandemic and a public health emergency of international concern.

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Recognizing and diagnosing individuals with COVID-19 quickly and providing them with treatment and quarantine in time are becoming a huge challenge worldwide.

As COVID-19 is a respiratory infectious disease caused by an RNA virus, real-time reverse transcriptase polymerase chain reaction (RT-PCR) is regarded as the gold standard method for the diagnosis of COVID-19. However, because of some as yet unknown factors, the detection rate for SARS-CoV-2 using RT-PCR is not high [4, 5]. There have also been several recent reports about false-negative results of nucleic acid tests for COVID-19 [6, 7]. To provide patients with timely treatment and to block viral transmission as soon as possible, the National Health Commission of the People's Republic of China has revised the diagnostic criteria in the fifth version of the guideline for diagnosis and treatment of COVID-19 [8]. A new type of diagnosis, namely clinical diagnosis, was performed in Hubei Province, which is the area with the highest prevalence of COVID-19 in China. Clinical diagnosis of COVID-19 is based mainly on clinical, epidemic, and radiologic features, regardless of the RT-PCR results, whereas microbiological diagnosis is based mainly on SARS-CoV-2 test results. It is still unknown whether there are differences between clinically and microbiologically (i.e. positive RT-PCR results for SARS-CoV-2 RNA) diagnosed patients.

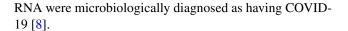
Recent studies have demonstrated that older age was a risk factor for death from COVID-19 [9, 10]. Because of the progressive aging of the population, the diagnosis and treatment for older adults with COVID-19 should be brought to the forefront worldwide. Therefore, this study aimed at exploring the differences between the two types of diagnosis among older patients in terms of clinical characteristics on admission and outcomes (i.e., discharged or died in the hospital) and at contributing toward optimizing the diagnosis and treatment of these patients.

### **Methods**

## **Definitions**

Clinical diagnosis Patients from epidemic areas who showed clinical (i.e., fever or respiratory symptoms) and laboratory (i.e., normal or decreased white blood cell counts, or decreased lymphocyte counts) features as well as imaging findings of viral pneumonia (i.e., ground-glass opacities, multifocal patchy consolidation, infiltration shadowing and interstitial abnormalities with a peripheral distribution) were clinically diagnosed as having COVID-19 by two experienced respiratory physicians [8].

Microbiological diagnosis: Patients who met the criteria for clinical diagnosis with positive evidence of SARS-CoV-2



# Study design and participants

Our study was based on retrospective observation of inpatients in the Sino-French New City Branch of Tongji Hospital, which was an assigned hospital for the treatment of severe and critical patients with COVID-19 in Wuhan. It has 23 wards (including 2 intensive care units) with 1085 beds. The study's observational end point was either death due to COVID-19 during hospitalization or eligible discharge [8] between January 29, 2020 and March 5, 2020. Patients aged 60 years or older with definite outcomes (i.e., discharged or died in hospital) by March 5, 2020 were included in our study. All patients provided nasopharyngeal or oropharyngeal swab samples upon admission, and the SARS-CoV-2 tests were repeated during the course of the hospitalization. The presence of SARS-CoV-2 in the patients' specimens was detected by RT-PCR using kits produced by DAAN GENE Co., Ltd of Sun Yat-Sen University. Participants who met the criteria for clinical diagnosis with only negative virus test results (including the tests at admission and those repeated during hospitalization) were allocated to the clinical diagnosis group, and those with positive virus test results were allocated to microbiological diagnosis groups.

### **Data collection**

Data on each patient's clinical characteristics, laboratory and imaging findings on admission, and outcomes were collected and reviewed by at least two experienced clinicians through assessment of electronic medical records. A trained team of researchers analyzed the data, including symptoms, signs, laboratory test results, and outcomes. Peripheral oxygen saturation (SpO<sub>2</sub>) was measured in the patient's oxygen-absorbing state.

#### **Statistics**

Comparison of categorical variables (except outcomes) between the two groups was evaluated using the Pearson Chi-square test with continuity correction or Fisher's exact test. The Mann–Whitney U test was used for continuous variables. Propensity score analyses, including covariate adjustment using propensity score (CAPS) and propensity score matching (PSM), were used to control the bias of confounding factors. Age, white blood cell (WBC) count, and lymphocyte (LYM) count were included in the propensity score model (using a logistic regression), which were the most important risk factors for death in the hospital among older patients with COVID-19 [11]. WBC count and LYM count were also the most important laboratory indicators of



clinical diagnosis criteria for COVID-19. Continuous variables were treated as continuous measures in our analysis. Samples with missing values in the three variables above were excluded from the propensity score model. Univariate logistic regression was used to estimate the differences in outcomes between the two groups before propensity score analyses and after PSM. The PSM was performed at a ratio of 1:1 with 0.01 caliper and random sampling without replacement. Only matched samples were included in the PSM analysis. Statistical analyses were performed using SPSS 21.0. Statistical tests were considered significant when two-sided *P* values were less than 0.05.

# **Results**

A total of 244 older patients were included in this study, with 52 patients undergoing clinical diagnosis and 192 patients, microbiological diagnosis. Of the patients, 16 (30.8%) and 107 (55.7%) in the clinical and microbiological diagnosis groups, respectively, were discharged. The median patient age was 68 years for the clinical diagnosis group and 70 years for microbiological diagnosis group (P=0.286; Table 1). Male individuals accounted for 69.2% of patients in the clinical diagnosis group and 50.5% in the microbiological group, a between-group difference that was statistically significant (P = 0.016). The majority of patients in both the clinical and microbiological diagnosis groups had fever (86.5%, 86.5%, P = 0.988) and respiratory symptoms (86.5%, 89.1%, P = 0.612), especially cough (69.2%, 74.5%, P = 0.448). The proportion of patients with gastrointestinal symptoms in the clinical diagnosis group was higher than that in the microbiological group (48.1% vs. 29.2%, P = 0.010). The between-group distribution of the WBC count was statistically significantly different (median value 10.17 vs. 6.07, P < 0.001). The P value of the distribution difference for the LYM count was 0.053. The erythrocyte sedimentation rate (ESR) distribution was similar between the two groups (P = 0.864). Other laboratory findings, such as amino-terminal pro-brain natriuretic peptide (NTproBNP), procalcitonin, high-sensitivity cardiac troponin I (hsTnI), D-dimer, alanine aminotransferase (ALT), aspartate aminotransferase (AST), creatinine, eGFR, high-sensitivity C reactive protein (hsCRP), serum ferritin, and interleukin-6 (IL-6) levels, were significantly different (P < 0.05) between the groups.

Propensity score analyses were applied to balance the confounding factors between the two groups. Older age, higher WBC count, and lower LYM count were previously identified as three important prognostic factors for COVID-19 [11]. The propensity score was calculated using a logistic regression model based on these three factors. Both the clinical and microbiological diagnosis groups contained one

case each with missing values for WBC count and LYM count, and those cases were excluded from the propensity score model. A total of 92 patients were included in the PSM analysis, with 46 patients from each group (Table 2). The median ages were 69 and 70 years for the matched clinical and microbiological groups, respectively, and the majority of patients in the two matched groups were male (69.5% vs. 56.5%, P = 0.195). After matching, there were no statistical differences in most variables between the two groups, especially the laboratory variables (P > 0.05). The P value of D-dimer was 0.047 [median (IQR) 3.60 (1.22-21.00) vs. 1.34 (0.74–15.13)]. Among the comorbidities, hypertension was still the most common, with the proportion of patients with hypertension at 63% (29/46) and 50% (23/46) in the matched clinical and microbiological diagnosis groups (P = 0.207). However, gastrointestinal symptoms were significantly different between the matched clinical and microbiological groups (P=0.001). The proportion of patients with gastrointestinal symptoms were 47.8% and 15.2%, respectively. Diarrhea was a common symptom, and the distribution of this symptom was also significantly different between the two matched groups (41.3% vs. 13.0%, P = 0.002).

The disease outcomes between the two groups were significantly different (P=0.002) before propensity score analyses (Table 3). However, after controlling for bias by CAPS, the difference in outcomes between the two groups disappeared (P=0.318) (Table 3). In PSM samples, 16 patients were discharged, and 30 patients died in the matched clinical diagnosis group, while 17 were discharged and 29 died in the matched microbiological diagnosis group. The outcomes between the two groups after PSM were similar [P=0.828, OR 1.099, 95% CI (0.469, 2.578)] (Table 3).

#### Discussion

In this retrospective study, we found that there were many differences in clinical features, laboratory findings, and patient outcomes between the two types of COVID-19 diagnoses. Mortality in the clinical diagnosis group was significantly higher than that in the microbiological diagnosis group. Treatments for COVID-19 were based on patients' conditions according to the guidelines and were not related to the type of diagnosis. This revealed that patients with a clinical diagnosis of COVID-19 had more severe disease than those with a microbiological diagnosis. However, after PSM, most of the differences disappeared. Of note, the mortalities of the two groups were no longer significantly different after CAPS or PSM analysis. Gastrointestinal symptoms and consciousness disorders were significantly different before and after matching in the two clinical and microbiological groups.



Table 1 Differences of characteristics between clinically and microbiologically diagnosed cases (total samples) before propensity score analyses

Characteristics	Clinical <sup>a</sup>	Microbiological <sup>a</sup>	P value
Demographics			
Age (years)	68 (64-75)	70 (65–76)	0.286
Gender			0.016
Male	36 (69.2)	97 (50.5)	
Female	16 (30.8)	95 (49.5)	
Vital signs			
SpO <sub>2</sub> (%)	95 (85-98)	96 (88–98)	0.711
Pulse rate (bpm)	89 (80-102)	89 (81-102)	0.858
Respiratory rate (bpm)	23 (20-30)	20 (20-24)	0.025
Consciousness disorders	16 (30.8)	16 (8.3)	< 0.001
Symptoms			
Fever	45 (86.5)	166 (86.5)	0.988
Temperature-highest (°C)	38.5 (38.0-39.0)	38.4 (37.8-39.0)	0.261
Respiratory symptoms	45 (86.5)	171 (89.1)	0.612
Cough	36 (69.2)	143 (74.5)	0.448
Dyspnea	35 (67.3)	122 (63.5)	0.615
Gastrointestinal symptoms	25 (48.1)	56 (29.2)	0.010
Diarrhea	21 (40.4)	51 (26.6)	0.053
Abdominal pain	4 (7.7)	6 (3.1)	0.227
Histories			
Hypertension	32 (61.5)	106 (55.5)	0.436
Diabetes	8 (15.4)	43 (22.5)	0.263
Coronary heart disease	10 (19.2)	25 (13.1)	0.263
Respiratory diseases	4 (7.7)	20 (10.5)	0.552
Laboratory findings			
WBC count ( $\times 10^9$ /L)	10.17 (6.20-12.94)	6.07 (4.73-9.34)	< 0.001
LYM count ( $\times 10^9$ /L)	0.60 (0.44-0.96)	0.73 (0.51-1.12)	0.053
NT-proBNP ( $\times 10^2$ pg/mL)	8.00 (1.75-36.14)	3.22 (1.49-8.24)	0.008
Procalcitonin (ng/mL)	0.31 (0.10-1.31)	0.10 (0.04-0.32)	< 0.001
hsTnI (pg/mL)	29.3 (7.4-425.0)	11.4 (4.7–30.5)	0.001
D-dimer (µg/mL FEU)	4.47 (1.26-21.00)	1.34 (0.59-3.26)	< 0.001
ALT (U/L)	28.0 (19.0-46.0)	23.0 (15.0-38.0)	0.047
AST (U/L)	45.0 (29.0-65.0)	33.0 (24.0-52.0)	0.008
Creatinine (µmol/L)	87.0 (72.0-118.0)	76.0 (57.0-94.0)	0.009
eGFR (mL/min/1.73 m <sup>2</sup> )	71.8 (48.9-89.0)	80.3 (60.9-92.5)	0.037
hsCRP (mg/L)	103.2 (47.5-169.3)	65.0 (27.4-114.1)	0.001
ESR (mm/H)	36.0 (20.0-63.0)	37.0 (20.0-59.0)	0.864
Serum ferritin ( $\times 10^2 \mu\text{g/L}$ )	13.50 (6.88-24.10)	7.73 (4.56–16.46)	0.010
Interleukin-6 (pg/mL)	58.01 (31.07-131.95)	30.37 (5.61-77.65)	0.005

WBC white blood cells, LYM lymphocyte, NT-proBNP amino-terminal pro-brain natriuretic peptide, hsTnI high-sensitivity cardiac troponin I, ALT alanine aminotransferase, AST aspartate aminotransferase, eGFR estimated glomerular filtration rate, hsCRP high-sensitivity C-reactive protein, ESR erythrocyte sedimentation rate

<sup>a</sup>Data presented as median (IQR) and n (%) for continuous and categorical variables, respectively, unless otherwise indicated

A false-negative result of SARS-CoV-2 detection may delay the diagnosis of COVID-19. Since the outbreak of COVID-19, detection of etiology has played a decisive role in the diagnosis of infected people. Scientists from China have obtained and uploaded the full-length genomic

sequence of SARS-CoV-2 [12], and several nucleic acid detection kits based on this sequence have been developed. However, differences in sensitivity and specificity have been found on clinical use of different testing kits. For instance, the incidence of false-negative virus test results was high



**Table 2** Characteristics of clinically and microbiologically diagnosed cases after propensity score matching

Characteristics	Clinical <sup>a</sup>	Microbiological <sup>a</sup>	P value	
Demographics				
Age (years)	69 (64-78)	70 (64–76)	0.944	
Gender				
Male	32 (69.6)	26 (56.5)	0.195	
Female	14 (30.4)	20 (43.5)		
Vital signs				
SpO <sub>2</sub> (%)	95 (85-97)	94 (82-97)	0.484	
Pulse rate (bpm)	89 (79-101)	90 (81-111)	0.761	
Respiratory rate (bpm)	22 (20-30)	21 (20-28)	0.547	
Consciousness disorders	13 (28.3)	4 (8.7)	0.016	
Symptoms				
Fever	40 (87.0)	38 (82.6)	0.562	
Temperature-highest (°C)	38.5 (38.0-39.0)	38.5 (37.8-38.9)		
Respiratory symptoms	40 (87.0)	40 (87.0)	1.000	
Cough	33 (71.7)	28 (60.9)	0.270	
Dyspnea	30 (65.2)	31 (67.4)	0.825	
Gastrointestinal symptoms	22 (47.8)	7 (15.2)	0.001	
Diarrhea	19 (41.3)	6 (13.0)	0.002	
Abdominal pain	3 (6.5)	0 (0.0)	0.242	
Histories				
Hypertension	29 (63.0)	23 (50.0)	0.207	
Diabetes	7 (15.2)	9 (19.6)	0.582	
Coronary heart disease	8 (17.4)	8 (17.4)	1.000	
Respiratory diseases	4 (8.7)	8 (17.4)	0.216	
Laboratory findings				
WBC count ( $\times 10^9$ /L)	9.13 (6.10-12.26)	9.33 (5.89-11.72)	0.941	
LYM count ( $\times 10^9/L$ )	0.64 (0.43-0.98)	0.66 (0.34-1.10)	0.947	
NT-proBNP ( $\times 10^2$ pg/mL)	7.90 (1.72-35.76)	6.01 (2.03-13.75)	0.799	
Procalcitonin (ng/mL)	0.28 (0.09-1.18)	0.18 (0.07-0.68)	0.217	
hsTnI (pg/mL)	35.2 (8.1-425.0)	21.0 (6.6-63.7)	0.199	
D-dimer (µg/mL FEU)	3.60 (1.22-21.00)	1.34 (0.74-15.13)	0.047	
ALT (U/L)	28.0 (19.0-41.0)	28.0 (17.0-41.0)	0.623	
AST (U/L)	44.0 (29.0-58.0)	36.5 (26.0-56.0)	0.269	
Creatinine (µmol/L)	86.5 (72.0-117.0)	87.0 (65.0-103.0)	0.555	
eGFR (mL/min/1.73 m <sup>2</sup> )	71.8 (50.0-85.0)	77.6 (51.3–91.2)	0.458	
hsCRP (mg/L)	103.3 (51.1-169.3)	71.0 (36.7–12.6)	0.034	
ESR (mm/H)	36.0 (20.0-63.0)	41.0 (18.0-60.0)	0.983	
Serum ferritin ( $\times 10^2 \mu\text{g/L}$ )	12.38 (6.64-22.47)	10.72 (5.14-19.44)	0.379	
Interleukin-6 (pg/mL)	55.07 (26.99-103.00)	31.99 (9.99-113.40)	0.168	

WBC white blood cells, LYM lymphocyte, NT-proBNP amino-terminal pro-brain natriuretic peptide, hsTnI high-sensitivity cardiac troponin I, ALT alanine aminotransferase, AST aspartate aminotransferase, eGFR estimated glomerular filtration rate, hsCRP high-sensitivity C-reactive protein, ESR erythrocyte sedimentation rate

<sup>a</sup>Data presented as median (IQR) and n (%) for continuous and categorical variables, respectively, unless otherwise indicated

[13]. As a result, some patients might be misdiagnosed when the positive result of virus test is a necessary criterion for diagnosis of COVID-19. Several explanations could account for the high false-negative rate. Currently, nasal and pharyngeal swabs are the main methods to obtain samples for nucleic acid testing for SARS-CoV-2. However, a recent study revealed that nasal and pharyngeal swabs demonstrated a poor positive rate, while the bronchoalveolar lavage fluid exhibited a high detection rate [14]. As COVID-19 is a virus that can infect the upper and lower respiratory



**Table 3** The outcomes of clinically and microbiologically diagnosed cases before and after propensity score analyses

Grouping	Outcomes		P value	OR	95% CI	
	Total	Discharged	Dead			
Before propensity score analysis				0.002	2.832	1.473–5.448
Clinical	52	16	36			
Microbiological	192	107	85			
CAPS				0.318	1.490	0.681-3.262
Clinical	51 <sup>a</sup>	16	35 <sup>a</sup>			
Microbiological	191 <sup>b</sup>	107	84 <sup>b</sup>			
PSM				0.828	1.099	0.469-2.578
Clinical	46	16	30			
Microbiological	46	17	29			

OR odds ratio, CI confidence interval of OR, CAPS covariate adjustment using propensity score, PSM propensity score matching

<sup>a</sup>One case of clinical diagnosis group was excluded from CAPS analysis for containing missing data in WBC count and LYM count

<sup>b</sup>One case of microbiological diagnosis group was excluded from CAPS analysis for containing missing data in WBC count and LYM count

tract simultaneously, there may be differences in viral load between samples from different parts of the respiratory tract, which could in turn cause a disparity in the detection rate. The collection of nasal and pharyngeal swabs may cause patients to be slightly uncomfortable. Furthermore, collection of such samples usually requires a certain amount of cooperation from patients. It is difficult to collect nasal and pharyngeal swabs from some patients, such as those who face difficulties in communication, like older patients (e.g., the participants in our study), which will influence the detection rate of the virus. In addition, as mentioned above, testing kit performance is also associated with the microbiological test outcome.

There was a certain proportion of patients who did not have respiratory symptoms but had gastrointestinal symptoms [10, 15]. In our study, the presence of gastrointestinal symptoms was statistically different between the clinical and microbiological groups before and after PSM, and the proportion of patients with gastrointestinal symptoms was greater in the clinical diagnosis group. Our previous study results showed that gastrointestinal symptoms were not a risk factor for death [11], but some studies observed that gastrointestinal symptoms were related to worse outcomes [16]. Gastrointestinal symptoms may mislead or delay the diagnosis of COVID-19, especially in patients mainly presenting with gastrointestinal symptoms [17]. The positivity rate for virus tests of feces samples in patients with gastrointestinal symptoms was much higher than the average [13]. The viral load of the respiratory tract may not be high enough to be detectable in these patients, which may also explain the negative results of their nasopharyngeal swabs. This suggests that for patients with nasopharyngeal swab-negative results,

attention should be paid to their gastrointestinal symptoms, and anal swab virus detection should be considered, when possible.

Before propensity score analyses, the clinical and laboratory features of patients in the clinical diagnosis group were more severe than those of patients in the microbiological group on admission. In particular, consciousness disorders, and increased WBC count, procalcitonin and D-dimer levels are associated with worse outcomes [11]. After 1:1 propensity score matching based on age, WBC, and LYM count, which were identified as important prognostic factors in our previous work [11] and also reported by other recent studies [9, 15], many differences in the clinical features and laboratory findings disappeared. WBC and LYM counts were also the most important indicators in clinical diagnosis criteria in the 5th version of the guideline for diagnosis and treatment of COVID-19. At the same time, the two matched groups demonstrated similar mortality. This means that patients with similar signs, symptoms, and laboratory findings may have similar outcomes, regardless of the results of RT-PCR for viral RNA detection. Therefore, the higher mortality in the clinical diagnosis group before PSM may mainly be because of the more severe disease state than that of patients in the microbiological group on admission.

Although the current study is limited by the small sample size after PSM, it is the first to investigate the differences in clinical and microbiological diagnoses of COVID-19. The negative result achieved from PSM was confirmed by CAPS, thus proving its authenticity.

For clinicians treating COVID-19 in epidemic areas where the supply of virus detection kits is insufficient or the performance of kits is poor, older patients with clinical



features of COVID-19, especially gastrointestinal symptoms such as diarrhea, should be paid more attention when making medical decisions. Even if the virus tests are negative, older adults eligible for clinical diagnosis of COVID-19 may require timely intervention to reduce mortality.

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**Availability of data and materials** All data generated or analyzed during this study are included in this published article.

# **Compliance with ethical standards**

Conflict of interest The authors declare that they have no conflicts of interest.

**Ethics approval** This study followed the Helsinki Declaration of the World Medical Association. The study was approved by the Ethics Committee of Tongji Hospital of Tongji Medical College of Huazhong University of Science and Technology (TJ-IRB20200505).

**Statement of human and animal rights** The present study followed the ethical standards for Humans and animal rights.

**Informed consent** The requirement for written informed consent was waived by the Ethics Committee of Tongji Hospital of Tongji Medical College of Huazhong University of Science and Technology.

Consent for publication The publication was approved and the requirement for written informed consent was waived by the Ethics Committee of Tongji Hospital of Tongji Medical College of Huazhong University of Science and Technology.

Code availability Not applicable.

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