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A molecular docking study of potential inhibitors and repurposed drugs against SARS-CoV-2 main protease enzyme[☆]



Selami Ercan^{*}, Ercan Çınar

Department of Nursing, School of Health Sciences, Batman University, Batman, Turkey

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ABSTRACT

COVID-19 has affected millions of people. Although many drugs are in use to combat disease, there is not any sufficient treatment yet. Having critical role in propagation of the novel coronavirus (SARS-CoV-2) works Main Protease up into a significant drug target. We have performed a molecular docking study to define possible inhibitor candidates against SARS-CoV-2 Main Protease enzyme. Besides docking Remdesivir, Ribavirin, Chloroquine and 28 other antiviral inhibitors (totally 31 inhibitors) to Main Protease enzyme, we have also performed a molecular docking study of 2177 ligands, which are used against Main Protease for the first time by using molecular docking program Autodock4. All ligands were successfully docked into Main Protease enzyme binding site. Among all ligands, EY16 coded ligand which previously used as EBNA1-DNA binding blocker candidate showed the best score for Main Protease with a binding free energy of -10.83 kcal/mol which was also lower than re-docking score of N3 ligand (-10.72 kcal/mol) contained in crystal structure of Main Protease. After analyzing the docking modes and docking scores we have found that our ligands have better binding free energy values than the inhibitors in use of treatment. We believe that further studies such as molecular dynamics or Molecular Mechanic Poisson Boltzmann Surface Area studies can make contribution that is more exhaustive to the docking results.

1. Introduction

Arising COVID-19 outbreak in Hubei province, P.R. China was a milestone for humanity when it started to affect the entire world in a short span of time like a tornado. It has changed our lifestyles, education methods and whatever we got accustomed. Affecting all the world, COVID 19, enabled scientists, companies, and countries to act together on a topic for the first time. While World Health Organization (WHO) declared the COVID-19 as pandemic on March 11, 2020, vaccine and appropriate drugs for treatment of disease remain aliment yet. According to the current data of the WHO [1], there are a total of 120 383 919 infected people and 2 664 386 deaths in the world by the March 18th 2021.

Being a member of *Nidovirales*, *Coronaviridae* includes *Coronaviruses* which can be investigated in four genera; *alphacoronavirus*, *betacoronavirus*, *gammacoronavirus*, and *deltacoronavirus*. From these species, *alpha*- and *betacoronaviruses* known to infect mammalian including human. Until 2019, it was reported that six types of coronavirus (HCoV-NL63, HCoV-229E, HCoV-HKU1, HCoV-OC43, severe acute respiratory syndrome (SARS-CoV), and Middle East respiratory syndrome

(MERS-CoV)) infected humans. Among these viruses, SARS-CoV killed 916 people in 37 countries between 2002 and 2003, MERS which was firstly reported in the Middle East killed 1791 people in 27 countries [2,3]. The severe acute respiratory syndrome (SARS-CoV) outbreak was started in 2003 in China followed by the second outbreak of the Middle East respiratory syndrome (MERS-CoV) in 2012 in Saudi Arabia. Again, the SARS-CoV-2 infection was started in later part of the year 2019 and was named as a novel coronavirus (COVID-19) on January 12, 2020, which attacks the immunity and the immune system, breaks down in the worst condition [4–6]. Among all other six coronaviruses, which are known to infect humans, **SARS-CoV-2** seems to be the most pathogenic kind of these viruses. The **SARS-CoV-2** is a new strain that has emerged due to which they are not familiar to our immune system causing infection in the respiratory tract. WHO has declared that fever, dry cough, and tiredness are the most common symptoms of COVID-19. Other symptoms are nasal congestion, diarrhea, sore throat, runny nose, and pains which are mainly mild and arise gradually. The less common symptoms are aches and pains, sore throat, conjunctivitis, headache, loss of taste or smell, a rash on skin, or discoloration of fingers or toes, as well as the serious symptoms: viz difficulty breathing or shortness of breath

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^{*} Corresponding author.

E-mail address: slmrcn@gmail.com (S. Ercan).

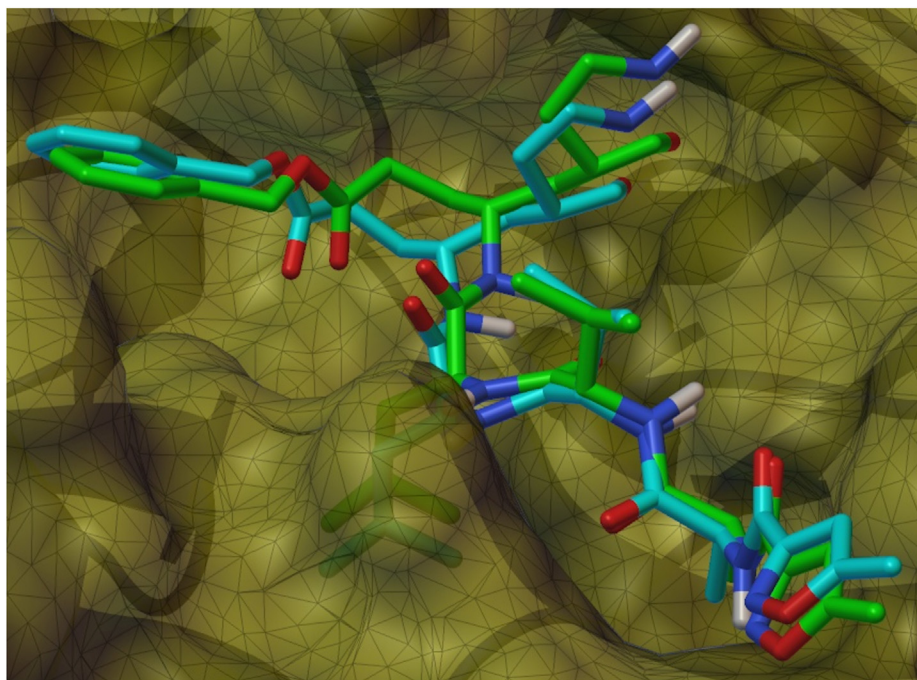


Fig. 1. Binding modes of crystal structure and re-docked structure of N3 ligand in the binding site of M^{pro} enzyme (re-docked structure: green, crystal structure: blue). (For interpretation of the references to color in this figure legend, the reader is referred to the Web version of this article.)

chest pain or pressure and loss of speech or movement. It should also be noted that not all infected people show these indications and thus most people recuperate without any special treatment. About 17% of infected patients have severe illness and breathing difficulties. Disease affect older people and people who have chronic diseases such as diabetes, heart problems, and high blood pressure more seriously [7].

Virus genome encodes two overlapping polyproteins, pp1a and pp1ab. These polyproteins are later subjected to proteolytic processing by viral proteases, main protease (M^{pro}; 3C-like protease) and accessory proteases (papain-like cysteine proteases) to produce functional polypeptides. M^{pro} is a 33.8 kDa enzyme, which also processes itself from polyproteins besides processing all other subunits of virus [8,9]. While M^{pro} cleaves subunits of virus at no less than 11 cleavage sites on 1 ab, this specific cutting makes M^{pro} an attractive drug target since no human proteases have a similar cleavage property. The main title is attributed to enzyme due to its dominant role in the processing replicase polyproteins and gene expression while 3C-like protease referred to enzyme due to similarity of the enzyme with picornavirus 3C proteases [8,10]. SARS-COV-2 main protease (M^{pro}) is one of the promising drug targets in the many possible drug targets of SARS-COV2 such as spike proteins, non-structural proteins (NSPs) and human receptor angiotensin-converting enzyme (ACE2). M^{pro} feature in mediating viral replication and transcription.

Recently, several M^{pro} crystal structures [9,11–13] were published on Protein Data Bank (www.rcsb.org). Jin et al. [9], designed a combination of high-throughput screening and structure-based virtual study over 10 000 compounds including drug candidates in clinical trials and approved drugs against M^{pro} enzyme. They have reported that six of these ligands have IC₅₀ values ranging from 0.67 to 21.4 μM inhibition activity. In another study, Chandel et al. [14], used FDA approved antiviral compounds and a library of active phytochemicals through molecular docking. Durdagi et al. [15], performed a molecular docking study for 7922 compounds from NIH Chemical Genomics Center (NCGC) Pharmaceutical Collection (NPC) database (<https://tripod.nih.gov/npc/>) which then followed by molecular dynamics and MM/GBSA studies for best scored ligands. There are also many antivirals, antimalarials,

anti-parasitic, and antibacterial are in clinical investigations for the treatment of COVID-19[16]. An *in vitro* study of such drugs (ribavirin, penciclovir, nitazoxanide, nafamostat, chloroquine, favipiravir, and Remdesivir) was performed by Wang et al. [17], and they have reported that Remdesivir and Chloroquine can inhibit 2019-nCov effectively.

Due to rapid transmission of the virus and urgency in discovering effective drugs for treatment of COVID-19, we have performed an *in-silico* study by using 2208 compounds (31 ligands approved or in clinical trial and 2177 used first time for SARS-CoV-2). The interactions of best-scored ligands were investigated deeply in this study.

2. Materials and methods

2.1. Receptor structure preparation

Recently reported (www.rcsb.org) [18] crystal structure of **SAR-CoV-2** M^{pro}, 6lu7. pdb [9], was selected for molecular docking. Crystal structure of 6lu7 also contains N3 ligand, which was used for dock validation. The receptor and N3 ligand files were saved separately after removing water molecules from crystal structure.

2.2. Ligand library preparation

Ligand library was generated as follows; previously used for both HIV-1 IN inhibition and 3-Hydroxy-3-methyl-glutaryl-CoA Reductase Enzyme (HMGR) (10 ligands) [19–21] inhibition, 104 ligands used for HMGR (ligands coded as HMXXX) [21] enzyme inhibition, 1637 ligands used for EBV EBNA1-DNA binding blockers (ligands coded as EBXXX, EYXX, and EZXXXX) [22], and 426 ligands used as HIV-1 RT-IN dual inhibitors (ligands coded as HBXX) [23]. Designing procedure of these ligands have been described in the related publications. Since these ligands were optimized formerly, they need not to be optimized any more. Beside these ligands, we have also studied 31 ligands to compare docking scores of ligands which some of them are approved inhibitors and some of them are in clinical trials for treatment of COVID-19[24]. The 3D structures of inhibitors in use of treatment for COVID-19 and

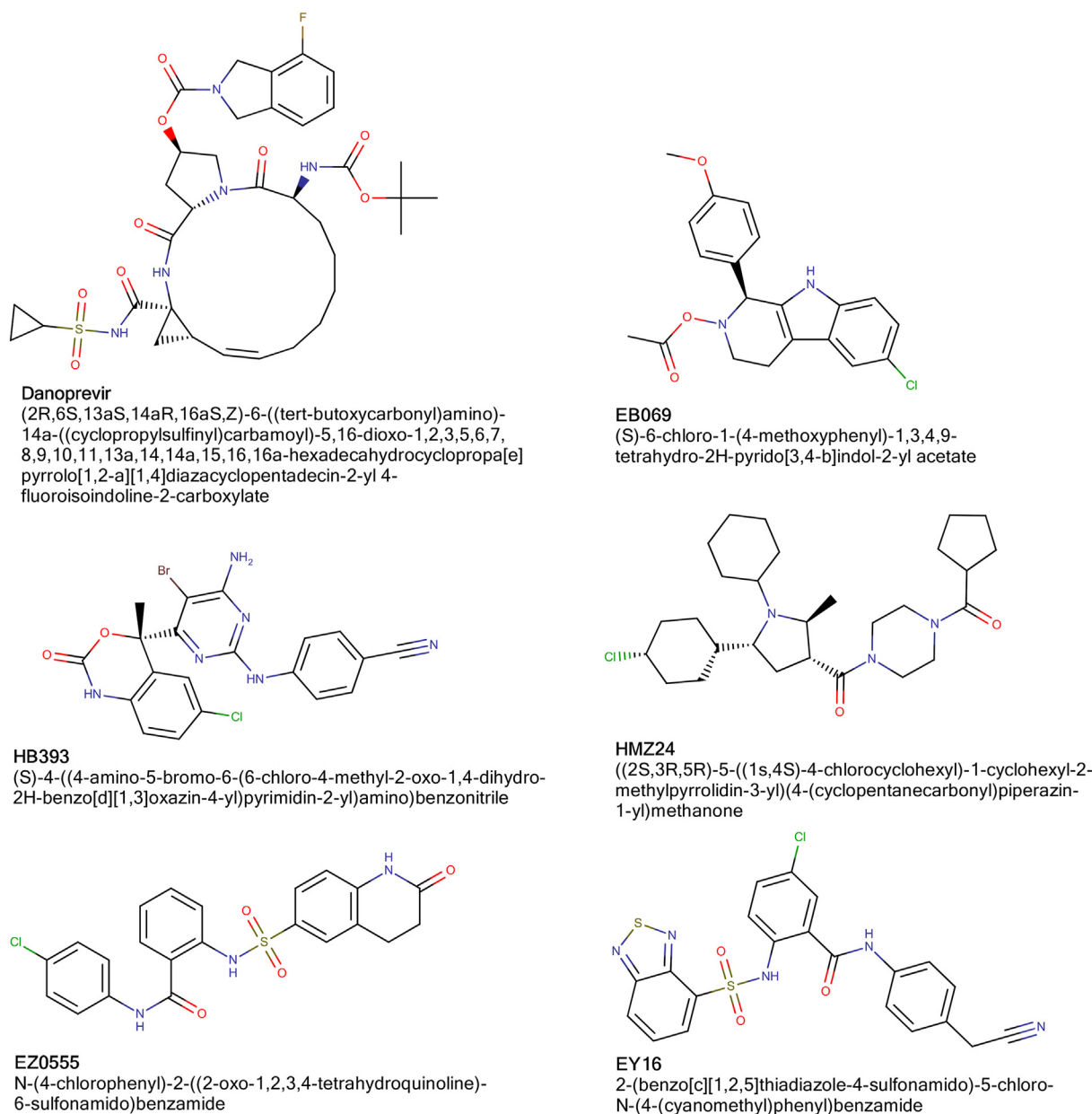


Fig. 2. 2D representations of best-scored ligands in their ligand sets.

which are in clinical trials were downloaded from PubChem [25] and Drugbank [26].

2.3. Molecular docking

While Autodock Vina, Autodock4 with Lamarckian Genetic algorithm (LGA) and Local Search (LS) algorithms were used for validation studies, it has been seen that LS algorithm of Autodock4 showed best docking pose for re-docking of N3 ligand to M^{Pro} enzyme structure (Fig. 1.) Docking site and box dimensions were selected by centering grid box on the N3 ligand, included in crystal structure. Docking score of N3 was -10.72 kcal/mol with a RMSD value of 0.81. Box was placed to -9.73 Å, 11.52 Å, 68.49 through x, y, and z coordinates, respectively, with the dimensions of $36 \times 60 \times 40$ Å. Docking studies were performed using LS algorithm with a number of 300 individuals to result in 250 docking poses for each ligand.

3. Result and discussion

Molecular docking is a key step in structure-based drug design. Allowing flexible ligand docking is also an advantage of the method. In this study, we used Autodock4 docking program to investigate binding modes of a total 2177 ligands in the binding site of M^{Pro} enzyme of SARS-CoV-2. Researchers have been making many efforts to find or develop appropriate drug candidates against possible drug targets of SARS-CoV-2 since arising COVID-19 outbreak by using repurposing drugs or using different ligand sources [27–33]. For the same goal, we have used a different set of ligands in the current study.

Docking studies of 31 inhibitors, which are in usage of COVID-19 treatment or in clinical trials and 2177 ligands, which are subjected to SARS-CoV-2 M^{Pro} enzyme structure for the first time were performed successfully. To validate the accuracy of docking procedure we have

Table 1

Docking scores of 20 best ligands of each sets of ligands.

EBNA1 ligand libraries				HIV1-RT, IN ligand library				HMGR ligand library	
Mol. ^a	Score	Mol.	Score	Mol.	Score	Mol.	Score	Mol.	Score
EB069	-9,84	EY16	-10,83	EZ0555	-10,46	HB393	-10,51	HMZ24	-10,51
EB071	-9,55	EY06	-9,67	EZ0239	-10,25	HB099	-10,01	HMZ29	-9,89
EB143	-9,48	EY03	-9,66	EZ0172	-9,92	HB110	-10,00	HMZ12	-9,74
EB045	-9,40	EY07	-9,63	EZ0271	-9,81	HB070	-9,82	HMZ34	-9,73
EB047	-9,38	EY04	-9,51	EZ0464	-9,74	HB051	-9,33	HMZ19	-9,72
EB070	-9,27	EY32	-9,30	EZ0593	-9,72	HB104	-9,32	HMZ16	-9,58
EB367	-9,24	EY28	-9,27	EZ0718	-9,68	HB095	-9,31	HMZ01	-9,54
EB183	-9,20	EY17	-9,22	EZ0171	-9,64	HB101	-9,20	HMZ26	-9,27
EB043	-9,17	EY01	-9,18	EZ0401	-9,64	HB061	-9,13	HMZ02	-9,23
EB196	-9,13	EY46	-9,11	EZ0542	-9,60	HB220	-9,05	HMZ09	-9,05
EB054	-9,12	EY12	-9,10	EZ0915	-9,57	HB062	-9,01	HMZ37	-9,02
EB190	-9,12	EY33	-9,08	EZ0763	-9,52	HB102	-8,98	HMZ10	-8,95
EB189	-9,11	EY45	-8,99	EZ0929	-9,51	HB105	-8,96	HMZ18	-8,93
EB354	-9,08	EY11	-8,92	EZ0248	-9,49	HB228	-8,96	HMM11	-8,91
EB004	-9,03	EY47	-8,81	EZ0097	-9,48	HB395	-8,91	HMZ17	-8,85
EB145	-9,03	EY20	-8,73	EZ0314	-9,48	HB219	-8,84	HMM32	-8,84
EB359	-9,02	EY19	-8,59	EZ0363	-9,46	HB229	-8,83	HMZ21	-8,84
EB122	-8,98	EY30	-8,59	EZ0296	-9,43	HB187	-8,80	HMZ25	-8,77
EB032	-8,96	EY29	-8,56	EZ0851	-9,42	HB071	-8,79	HMD04	-8,67
EB116	-8,95	EY14	-8,55	EZ0198	-9,40	HB107	-8,79	HMB04	-8,66

^a Mol. = Molecule.**Table 2**

Docking scores of inhibitors in clinical trials or in use of COVID-19 treatment.

Molecule	Score	Molecule	Score	Molecule	Score
Danoprevir	-8,93	Ritonavir	-7,20	Pirfenidone	-5,86
Nelfinavir	-8,92	Camostat	-6,91	Triazavirin	-5,52
Ebastine	-8,47	Chloroquine	-6,89	Azvodine	-5,38
Baloxavir_marboxil	-8,23	Cobicistat	-6,80	Fingolimod	-5,10
Losartan	-8,14	Darunavir	-6,73	Emtricitabine	-4,94
Methylprednisolone	-8,08	Remdesivir	-6,68	Ribavirin	-4,48
Ivermectin	-8,07	Oseltamivir	-6,41	Tenofovir	-4,35
Ruxolitinib	-8,04	Hydroxychloroquine	-6,37	Favipiravir	-4,21
Lopinavir	-7,90	Dipyridamole	-6,22	Acetylcysteine	-3,74
Umifenovir	-7,65	Tranilast	-6,17		
Thalidomide	-7,26	Azithromycin	-6,15		

Table 3

Average scores of ligands (kcal/mol) according to ligand sets.

Ligand Type	Numbers of Ligands	Average Docking Score
EBNA1 Designed ligand library	409	-7,40
EBNA1 Literature ligand library	58	-8,05
EBNA1 ZINC15 ligand library	1170	-7,77
HIV-1 RT, IN ligand library	426	-6,86
HMGR ligand library	114	-7,88
COVID-19 Inhibitors	31	-6,56
Total	2208	-7,42

performed a re-docking study. According to docking scores, EY16 coded ligand (Fig. 2.), which is previously used as EBNA1 inhibitor *in vitro* [34] and *in silico* [22,34], was the best-scored ligand with a score of -10.83 kcal/mol while re-docking of N3 ligand has a score of -10.72 kcal/mol. Other great white hope ligands are ligands coded as EZ055, EZ0239, HB393, HB099, HB110 and HMZ24 (Fig. 2.). We also want to report that newly studied ligands have better scores than approved inhibitors or inhibitors in clinical trials where their docking scores are in the range of (-3.74)-(-8.92) kcal/mol. While Danoprevir (Fig. 2.) has the best docking score (-8.93 kcal/mol), Acetylcysteine has the lowest docking score (-3.74 kcal/mol) among this set of ligands.

Docking scores of 20 ligands in each set and docking scores of inhibitors in clinical trials or in use are shown in Table 1 and Table 2, respectively. Docking scores of all ligands were added as supplementary material (S.M. Table 1).

As mentioned before, we used different sets of ligands in the study. From the three different sets of ligands formerly used for blocking DNA binding to the EBNA1, EB069, EY16, and EZ0555 were the best binding ligands with the scores of -9.84, -10.83, and -10.46 kcal/mol, respectively. HB393 coded ligand was the best-scored ligand from the set of ligands designed for dual inhibition of HIV-1 RT and IN enzymes with a binding free energy of -10.51 kcal/mol. While HMZ24, a ligand designed for inhibition of HMGR, scored -10.51 kcal/mol, Danoprevir was the best scored ligand through inhibitors in use or in clinical trials for treatment of COVID-19 with a binding free energy of -8.93 kcal/mol.

We have analyzed the interactions of receptor residues with five best-scored ligands of each sets of ligands and two best scored inhibitors, which are in use or in clinical trials for treatment of COVID-19.

We have analyzed average scores of used all ligands to define the best ligand library, which will also contribute to understanding of more appropriate structural properties of ligands. As seen from Table 3, ligands used for EBNA1 those selected from literature [34-40] showed best average docking scores through all ligands with an average score of -8.05 kcal/mol.

Table 4
M^{PRO} residues in interaction with best scored 20 ligands.

Molecule	Score	Interacted M ^{PRO} enzyme residues
EY16	-10.83	Thr26, His41, Met49, Leu141, Ser144, Cys145, His163, His164, Met165, Glu166
HB393	-10.51	His41, Met49, Leu141, Cys145, His164, Met165, Glu166
HMZ24	-10.51	His41, Met49, Ser144, Cys145, Met165, Gln189,
EZ0555	-10.46	Thr24, His41, Met49, Pro52, Cys145, His164
EZ0239	-10.25	Thr26, Leu27, His41, Met49, Gly143, Cys145, His163, Met165, His172
HB099	-10.01	His41, Met49, Cys145, His163, His164, Glu166, Pro168, His172, Arg188, Gln189
HB110	-10.00	His41, Cys44, Met49, Tyr54, Cys145, His163, Met165, Glu166, Pro168, Gln192
EZ0172	-9.92	Cys145, His163, Met165, Arg188, Thr190
HMZ29	-9.89	Leu27, Thr26, His41, Phe140, Gly143, Cys145, Met165
EB069	-9.84	His41, Met49, Gly143, Cys145, His164, Met165, His172
HB070	-9.82	Leu27, His41, Gly143, Cys145, Met165, Glu166, Leu167
EZ0271	-9.81	Met49, Ser144, Cys145, His163, Met165, Glu166
EZ0464	-9.74	His41, Met49, Cys145, His164, Met165, Arg188, Thr190, Gln192
HMZ12	-9.74	His41, Met49, Cys145, Met165, Gln189
HMZ34	-9.73	Glu166, Gly143, Ser144, Cys145, Met165, Pro168
EZ0593	-9.72	His41, His163, Met165, Pro168, His172, Arg188, Gln189, Thr190, Gln192
HMZ19	-9.72	Gly143, Ser144, Cys145, Met165, Glu166, Leu167
EZ0718	-9.68	His41, Met49, Cys145, His164, Met165, Glu166
EY06	-9.67	His41, Asn142, Gly143, Met49, His163, His164, Cys145, His172, Arg188
EY03	-9.66	Leu27, His41, Cys44, Met49, Pro52, Tyr54, Asn142, Ser144, Cys145, Met165, Pro168, Arg188, Gln189

Before analyzing ligand-receptor interactions, we have defined binding site residues of M^{PRO} enzyme. Binding site of M^{PRO} enzyme contains Thr24, Thr25, Thr26, Leu27, His41, Met49, Tyr54, Phe140, Leu141, Asn142, Gly143, Ser144, Cys145, His163, His164, Met165, Glu166, Leu167, Pro168, His172, Asp187, Arg188, Gln189, Thr190, Ala191, and Gln192 residues. The interactions of these residues with ligands' atoms are valuable and are investigated from docking modes of ligands. Ligand N3 interacts with M^{PRO} enzyme residues Phe140, Gly143, His163, His164, Glu166, Gln189, and Thr190 by hydrogen bonds. Other interactions are amide- π stacked interaction with Leu141 and Asn142 residues, alkyl-alkyl interactions with Met49, Met165, and Leu167 residues, π -alkyl interactions with His41, Pro168 and Ala191 residues in crystal structure 6lu7. pdb. The enzyme residues with 20 best-scored ligands and 31 drugs or inhibitors in clinical trial interacted with are summarized in Table 4 and Table 5, respectively.

Ligands coded as EB069, EY16, and EZ0555 were previously used for preventing DNA binding to EBV EBNA1 protein [22]. EY16 gave the best docking score through all ligands used in this study. As mentioned before, this ligand was studied formerly as *in vitro* and *in silico* for binding EBNA1 antigen. EB069 was designed by using some ligands as template which explained in one of our study [22] while EZ0555 (ZINC12583256) downloaded from ZINC14 database [41]. EB069 forms three hydrogen bonds with Gly143, Cys145, and His164 residues of receptor (Fig. 3.). Other interactions seen between EB069 and receptor residues are π -sulfur interaction with the residue Met49, alkyl-alkyl interactions with residues Cys145 and Met49, π -alkyl interactions with His41, Cys145, and His172, and a halogen interaction of ligands Cl atom with Met49 as given in Fig. 3. Ligand EY16 forms four hydrogen bonds with residues Thr26, Leu141, His164, and Glu166. π -sulfur interaction with Met165, π - π T shaped interactions with His163, π - π stacked interaction with His41, alkyl-alkyl interaction with Met49 and π -alkyl interaction with Met49 and Cys145 residues are other interactions seen between EY16 and receptor residues. There is also an unfavorable H-H interaction between

Table 5
M^{PRO} residues in interaction with drugs or inhibitors in clinical trials.

Molecule	Score	Interactions
Danoprevir	-8.93	His41, Asn142, Met165, Glu166, Thr190
Nelfinavir	-8.92	Met49, Cys145, His163, Met165, Glu166, Leu167, Pro168, Thr190, Gln192
Ebastine	-8.47	Leu27, Met49, His41, Cys145, Glu166
Baloxavir_marboxil	-8.23	Thr26, Phe140, Leu141, Gly143, Ser144, Cys145, His163, Met165, Glu166
Losartan	-8.14	Met49, Leu141, Ser144, Cys145, Glu166, Met165, Gln189
Methylprednisolone	-8.08	Cys145, Met165, Glu166, Thr190, Gln192
Ivermectin	-8.07	Thr26, His41, Met49, Gly143, Met165, Thr190, Ala191
Ruxolitinib	-8.04	His41, Met49, Cys145, Met165, Leu167, Pro168, Gln189, Thr190, Gln192
Lopinavir	-7.90	Met49, Gly143, Cys145, His164, Met165, Leu167, Pro168
Umifenovir	-7.65	His41, Met49, His164, Met165, Glu166
Thalidomide	-7.26	His41, Met165, Glu166, Thr190, Gln192
Ritonavir	-7.20	His41, Met49, Gly143, Cys145, Met165, Pro168
Camostat	-6.91	Phe140, Asn142, Gly143, Cys145, His163, Glu166
Chloroquine	-6.89	His41, Met49, Phe140, Cys145, His163, His164, Met165, Glu166, His172
Cobicistat	-6.80	Thr26, Ser144, Cys145, Met165, Glu166, Gln189
Darunavir	-6.73	His41, Met49, Cys145, His163, Met165, Glu166, Pro168, Gln192
Remdesivir	-6.68	Leu27, His41, Met49, Leu141, Ser144, Cys145, His164, Met165, Glu166, Thr190
Oseltamivir	-6.41	His41, Met165, Pro168, Arg188, Thr190, Gln192
Hydroxychloroquine	-6.37	Gly143, Met165, His163, Pro168, Gln189
Dipyridamole	-6.22	Met49, Cys145, Met165, Glu166
Tranilast	-6.17	Leu27, His41, Ser144, Cys145, Met165
Azithromycin	-6.15	Met49, Asn142, Cys145, His164, Met165, Glu166, Pro168, Gln189, Ala191
Pirfenidone	-5.86	His41, Met49, Met165, Gln189
Triazavirin	-5.52	Leu141, Asn142, Gly143, Cys145, Ser144
Azvadine	-5.38	Gly143, Ser144, His163, Glu166, Gln189
Fingolimod	-5.10	His41, Asn142, Cys145, Glu166
Emtricitabine	-4.94	Phe140, Asn142, Gly143, Cys145, His163, Glu166
Ribavirin	-4.48	Leu141, Asn142, Ser144, His163, Glu166
Tenofovir	-4.35	Gly143, Cys145, Glu166
Favipiravir	-4.21	Phe140, Leu141, Asn142, Ser144, Glu166
Acetylcysteine	-3.74	Leu141, Gly143, Ser144, Cys145, His164

Ser144:HN and Y16:H15 atoms (Fig. 4.). Ligand EZ0555 forms hydrogen bonds with Thr24, Cys145 and His164 residues, π -sulfur interaction with His41 and Cys145 residues, alkyl-alkyl interactions with Met49 and Pro52 residues and π -alkyl interaction with Met49 residue (Fig. 5.).

HB393 is a ligand designed for dual inhibition of HIV-1 RT and IN enzymes. Ligand showed a binding free energy of -10.51 kcal/mol for inhibition of M^{PRO} enzyme. Ligand HB393 creates three hydrogen bonds with Leu141, His164, and Glu166 residues. HB393 also in interaction with Met49, Cys145 by forming π -sulfur interactions, with His41 by forming π - π stacked interaction and with His41 and Met165 residues by forming π -alkyl interactions (Fig. 6.).

HMZ24 is the best scored ligand in the HMGR ligand set with a binding free energy of -10.51 kcal/mol. Interactions those ligand forms are; hydrogen bonds with Ser144 and Cys145 residues, alkyl-alkyl interactions with Met49, Cys145, and Met165, π - σ interaction with His41 and π -alkyl interaction with His41 residue (Fig. 7.).

Danoprevir has the lowest binding free energy through inhibitors, which are in use for treatment of COVID-19 or in clinical trials with a binding score of -8.93 kcal/mol. Danoprevir forms two hydrogen bonds with Asn142 and Glu166 residues of enzyme. Ligand also construct π -sulfur, alkyl-alkyl and a halogen interaction with His41, Met165, and Thr190 residues, respectively (Fig. 8.).

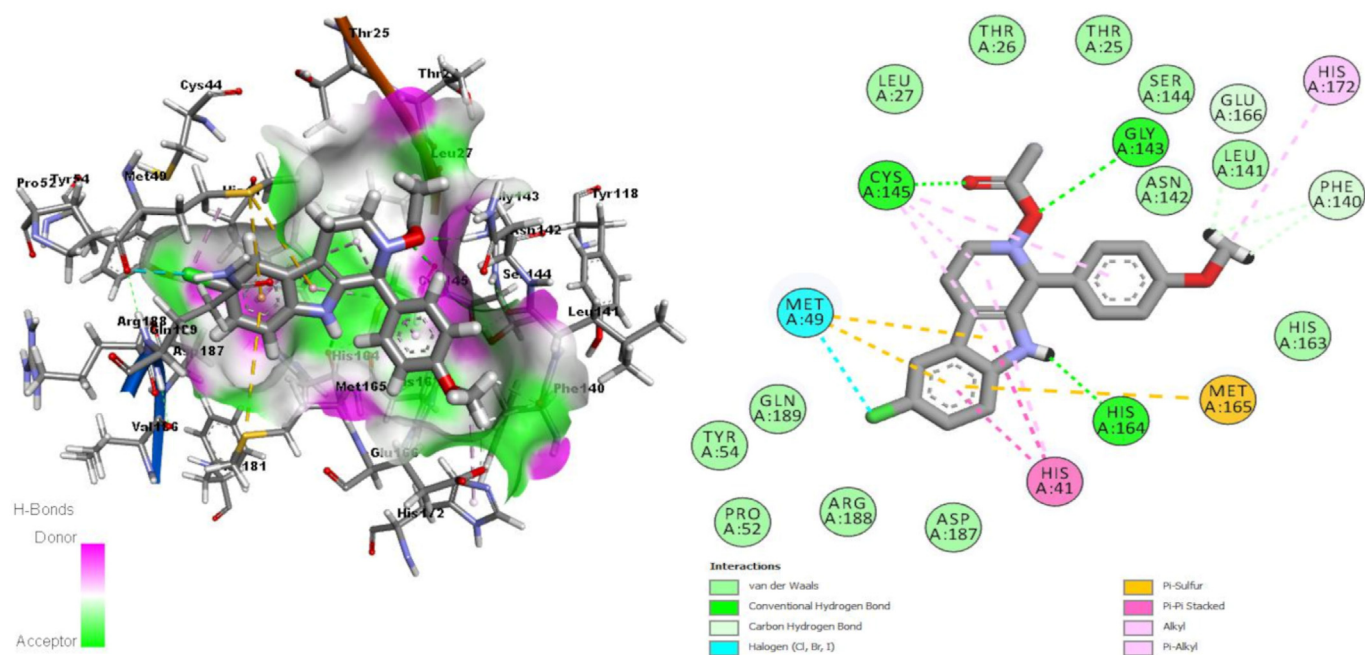


Fig. 3. Interactions of M^{pro} binding site residues with EB069 (3D (right) and 2D (left)).

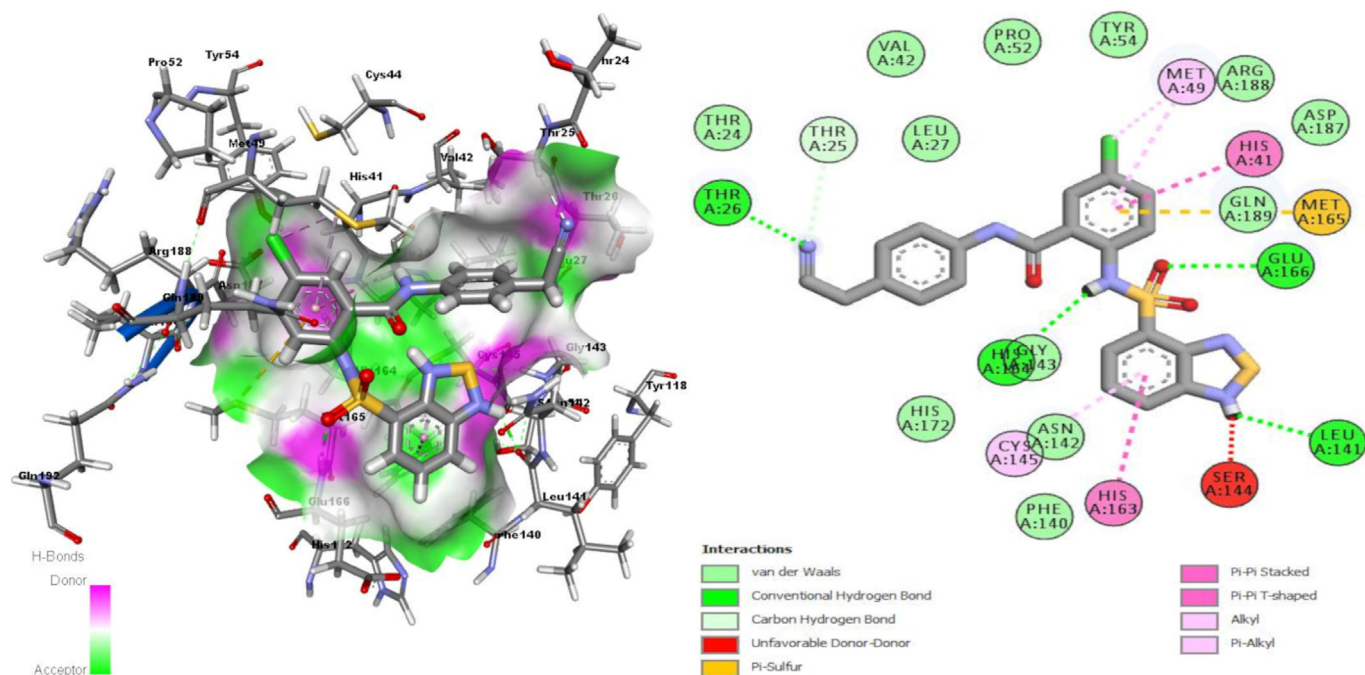


Fig. 4. Interactions of M^{pro} binding site residues with EY16 (3D (right) and 2D (left)).

Nelfinavir was the second-best scored ligand through COVID-19 inhibitors with a binding free energy of -8.92 kcal/mol. Nelfinavir forms three hydrogen bonds with Gln192, Glu166, and Thr190 residues of M^{pro}. Alkyl-alkyl interactions with Met49, Pro168, π -sulfur interaction with Met165, and π -alkyl interactions with His41, Met49, and Pro168 are the other interactions observed between Nelfinavir and enzyme residues (Fig. 9).

Among the binding site residues, His41, Met49, Gly143, Cys145, and Met165 are the most common residues, which interact with ligands' atoms. While Met49 interacts with ligands' atoms only by hydrophobic interactions other residues are also seen in hydrogen bond interactions with ligands' atoms. The importance of these residues' interactions was also discussed by Yoshino and coworkers [42]. According to binding free energy scores, we can state that the study bids fair different structural

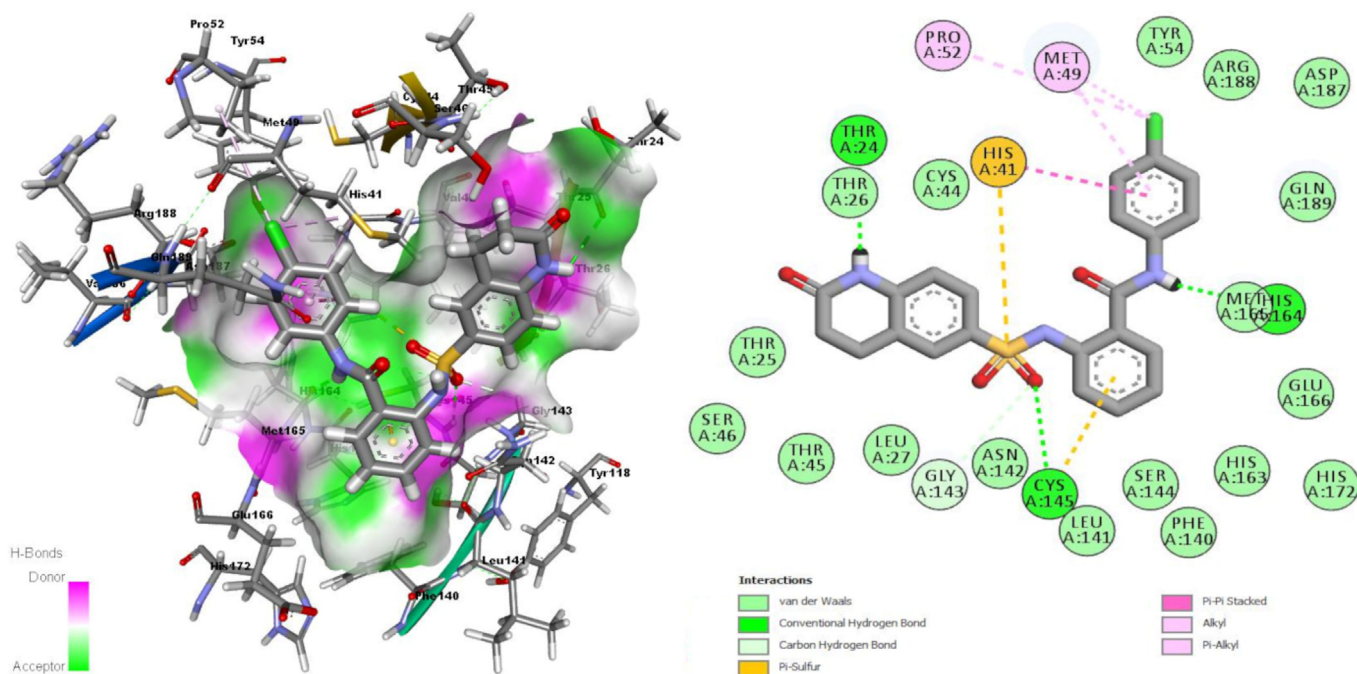


Fig. 5. Interactions of M^{Pro} binding site residues with EZ0555 (3D (right) and 2D (left)).

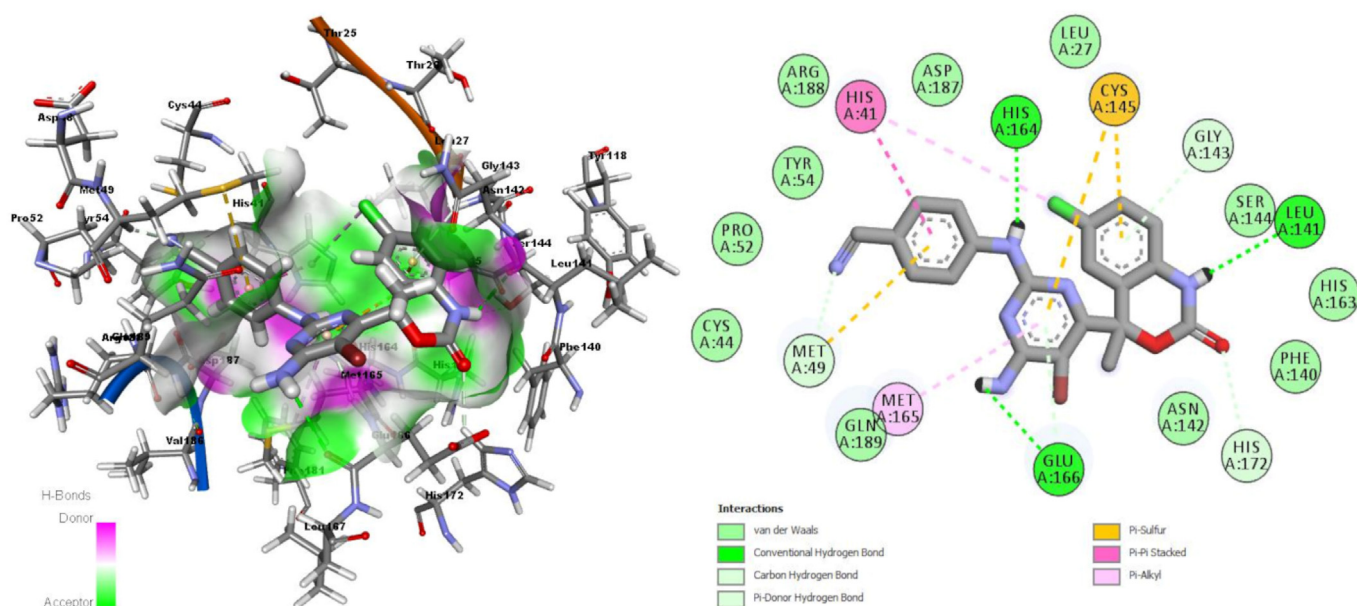


Fig. 6. Interactions of M^{Pro} binding site residues with HB393 (3D (right) and 2D (left)).

inhibitor samples, which need to be forwarded to *in vitro* studies.

We should mention that drugs such as Hydroxychloroquine, Remdesivir, Favipiravir and Ribavirin are currently in use of COVID 19 treatment and show high activity against SARS-CoV-2. Nguyen and coworkers [43] have performed a computational study on binding modes of Remdesivir to the RNA-Dependent RNA Polymerase and Main Protease. They have used Autodock Vina for docking studies where Remdesivir gave the binding energy of -7.9 kcal/mol for Mpro.

In another study, Naik et al. [44], used LGA algorithm of Autodock4 and they have obtained the docking binding energies of -8.2 kcal/mol,

-6.2 kcal/mol, -6.1 kcal/mol, -5.7 kcal/mol, -5.0 kcal/mol for Remdesivir, Ribavirin, Oseltamivir, Chloroquine, and Favipiravir, respectively. Obtaining similar results with study of Naik et al. also confirm our results since we experienced from docking tries that LGA algorithm of Autodock4 gives 2–3 kcal/mol lower scores than LS algorithm of Autodock4.

In a study performed by Chakraborti coworkers, some drugs were repurposed against main protease of SARS-CoV-2. They used Autodock Vina for docking studies of drugs and they have acquired the docking binding free energies of -8.8 kcal/mol and -8.0 kcal/mol for Danoprevir

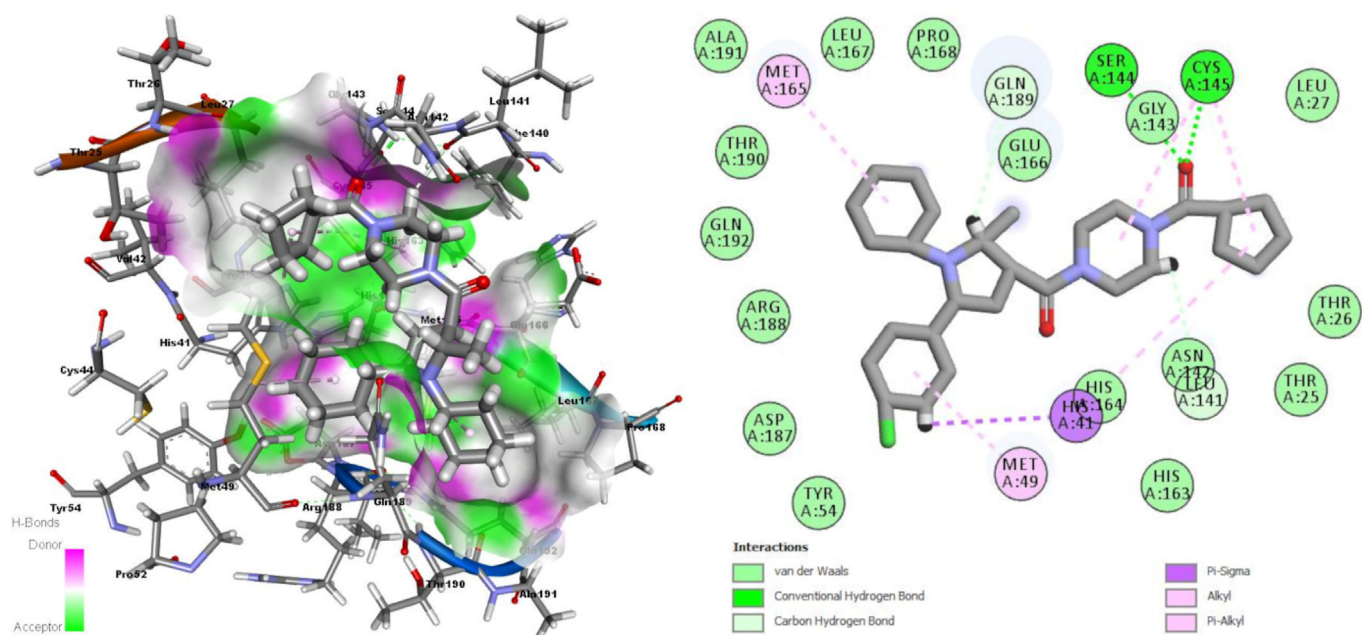


Fig. 7. Interactions of M^{Pro} binding site residues with HMZ24 (3D (right) and 2D (left)).

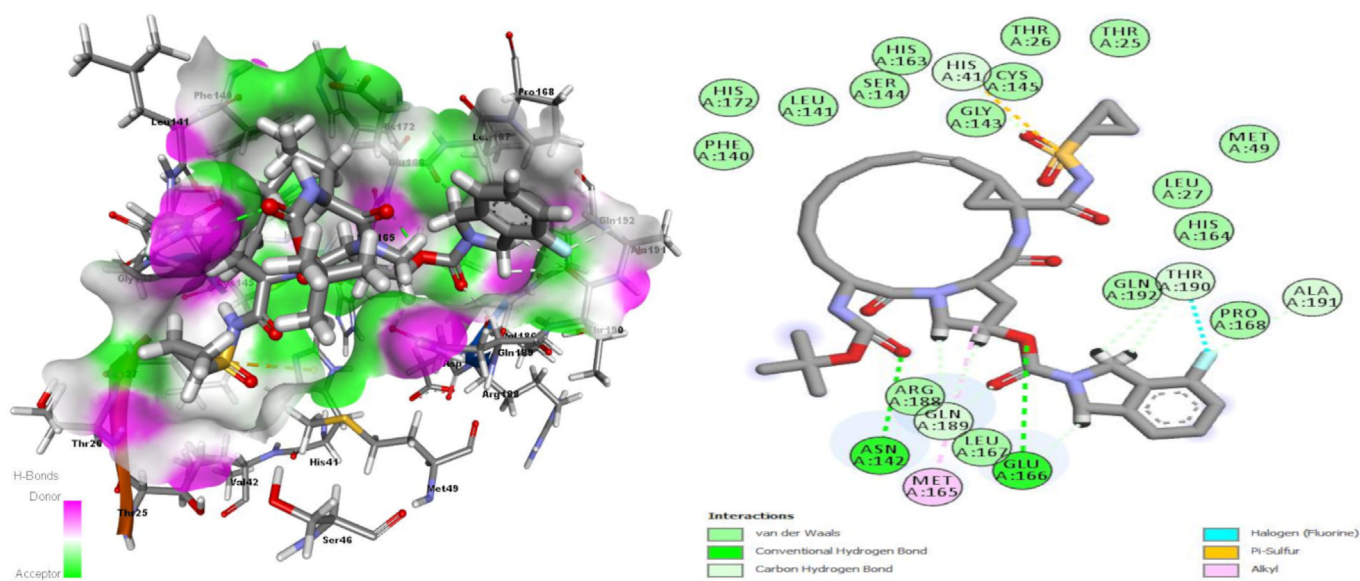


Fig. 8. Interactions of M^{Pro} binding site residues with Danoprevir (3D (right) and 2D (left)).

and Remdesivir, respectively which also support the better binding result of Danoprevir than Remdesivir of our study. In another docking study [45], Chloroquine and Hydroxychloroquine have been placed to different Mpro crystal structures with Autodock4 and while Chloroquine had a binding score of -4.3 kcal/mol and Hydroxychloroquine had a binding score of -4.8 kcal/mol for 6LU7 crystal Mpro structure which also have been used in our study.

Analyzing docking scores showed that newly studied two ligands have same docking score as Danoprevir, the best scored ligands through drugs in usage, while 157 ligands have a lower score than Danoprevir. This is also an indication of the studied ligands could have better

inhibition effect on M^{Pro} enzyme than drugs in usage of COVID-19 treatment. According to binding free energy scores, we can state that the study promise hope about introducing different structural inhibitor samples as inhibitor candidates for inhibition of SARS-CoV-2 M^{Pro} enzyme.

4. Conclusion

To bring forth new type of ligands to the treatment efforts of COVID-19, we have performed a molecular docking study by using totally 2208 ligands. Our ligand library consists of ligands which are

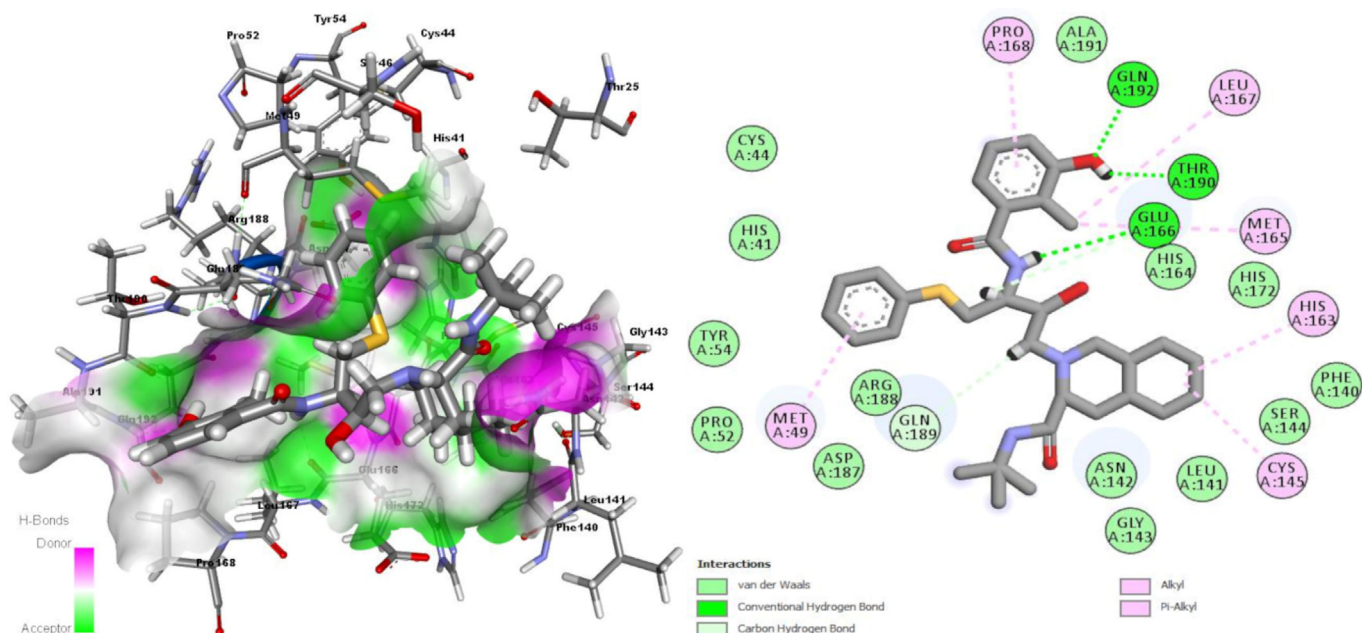


Fig. 9. Interactions of M^{pro} binding site residues with Nelfinavir (3D (right) and 2D (left)).

previously used for different targets as explained in ligand library preparation section.

Ligand coded EY16 was the best scored ligand through all ligands with a binding free energy of -10.83 kcal/mol. Other promising ligands are EZ055, EZ0239, HB393, HB099, HB110 and HMZ24. Although we have observed valuable results, we know that the study should be forward to molecular dynamics and MM/PB(GB)SA studies which will detail the accuracy of the molecular docking results.

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Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.jics.2021.100041>.

References

- [1] Who. WHO coronavirus disease (COVID-19) dashboard. WHO. Accessed 03.18.2021, <https://covid19.who.int/table>; 2021.
- [2] Fung TS, Liu DX. Human coronavirus: host-pathogen interaction. *Annu. Rev. Microbiol.* Sep 8 2019;73:529–57. <https://doi.org/10.1146/annurev-micro-020518-115759>.
- [3] Liu Z, Xiao X, Wei X, et al. Composition and divergence of coronavirus spike proteins and host ACE2 receptors predict potential intermediate hosts of SARS-CoV-2. *J. Med. Virol.* Feb 26 2020. <https://doi.org/10.1002/jmv.25726>.
- [4] Lu H, Stratton CW, Tang YW. Outbreak of pneumonia of unknown etiology in Wuhan, China: the mystery and the miracle. *J Med Virol.* Apr 2020;92(4):401–2. <https://doi.org/10.1002/jmv.25678>.
- [5] Chen Y, Liu Q, Guo D. Emerging coronaviruses: genome structure, replication, and pathogenesis. *J Med Virol.* Oct 2020;92(10):2249. <https://doi.org/10.1002/jmv.26234>.
- [6] Bogoch II, Watts A, Thomas-Bachli A, Huber C, Kraemer MUG, Khan K. Pneumonia of unknown aetiology in Wuhan, China: potential for international spread via commercial air travel. *J. Trav. Med.* Mar 13 2020;27(2). <https://doi.org/10.1093/jtm/taaa008>.
- [7] Who. Q&A on coronaviruses (COVID-19). WHO. Accessed 28.09.2020, <https://www.who.int/news-room/q-a-detail/q-a-coronaviruses>; 2020.
- [8] Yang H, Bartlam M, Rao Z. Drug design targeting the main protease, the Achilles' heel of coronaviruses. *Curr. Pharmaceut. Des.* 2006;12(35):4573–90. <https://doi.org/10.2174/138161206779010369>.
- [9] Jin Z, Du X, Xu Y, Deng Y, Liu M, Zhao Y, Zhang B, Li X, Zhang L, Peng C, Duan Y, Yu J, Wang L, Yang K, Liu F, Jiang R, Yang X, You T, Liu X, Yang X, Bai F, Liu H, Liu X, Guddat L, Xu W, Xiao G, Qin C, Shi Z, Jiang H, Rao Z, Yang H. Structure of Mpro from COVID-19 virus and discovery of its inhibitors. *Biorxiv* 2020. <https://doi.org/10.2210/pdb6lu7/pdb>.
- [10] Ziebuhr J, Snijder EJ, Gorbalenya AE. Virus-encoded proteinases and proteolytic processing in the Nidovirales. *J Gen Virol.* Apr 2000;81(Pt 4):853–79. <https://doi.org/10.1099/0022-1317-81-4-853>.
- [11] Fearon D, Powell AJ, Douangamath A, Owen CD, Wild C, Krojter T, Lukacik P, Strain-Damerell CM, Walsh MA. von Delft, PanDDA analysis of COVID-19 main protease against the DSI-poised Fragment Library. 2020. <https://doi.org/10.2210/pdb5r84/pdb>. To be published.
- [12] Zhang L, Lin D, Sun X, et al. Crystal structure of SARS-CoV-2 main protease provides a basis for design of improved alpha-ketoamide inhibitors. *Science* Mar 20 2020. <https://doi.org/10.1126/science.abb3405>.
- [13] Mesecar AD. A Taxonomically-Driven Approach to Development of Potent, Broad-Spectrum Inhibitors of Coronavirus Main Protease Including SARS-CoV-2 (COVID-19). 2020. <https://doi.org/10.2210/pdb6w63/pdb>.
- [14] Chandel Vr S, Rathi B, Kumar D. In silico identification of potent COVID-19 main protease inhibitors from FDA approved antiviral compounds and active phytochemicals through molecular docking: a drug repurposing approach. *Preprints* 2020. <https://doi.org/10.20944/preprints202003.0349.v1>.
- [15] Durdagi S, Aksoydan Busecan, Dogan Berna, Sahin Kader, Shahraki Aida. Screening of clinically approved and investigation drugs as potential inhibitors of COVID-19 main protease: a virtual drug repurposing study. *ChemRxiv Preprint* 2020. <https://doi.org/10.26434/chemrxiv.12032712.v1>.
- [16] Harrison C. Coronavirus puts drug repurposing on the fast track. *Nat Biotechnol.* Apr 2020;38(4):379–81. <https://doi.org/10.1038/d41587-020-00003-1>.
- [17] Wang M, Cao R, Zhang L, et al. Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro. *Cell Res.* Mar 2020;30(3):269–71. <https://doi.org/10.1038/s41422-020-0282-0>.
- [18] Berman HM, Westbrook J, Feng Z, et al. The protein Data Bank. *Nucleic Acids Res.* Jan 1 2000;28(1):235–42. <https://doi.org/10.1093/nar/28.1.235>.
- [19] Ercan S, Pirincioglu N. Computational design of a full-length model of HIV-1 integrase: modeling of new inhibitors and comparison of their calculated binding energies with those previously studied. *J. Mol. Model.* 2013;19(10):4349–68. <https://doi.org/10.1007/s00894-013-1943-4>. 2013/10/01.
- [20] Ercan S. Docking and molecular dynamics calculations of some previously studied and newly designed ligands to catalytic core domain of HIV-1 integrase and an

- investigation to effects of conformational changes of protein on docking results. *JOTCSA* 2017;4(1): 270-243.
- [21] Ercan S, Çınar E, Özyayın C, Efe Ertürk N, Çakmak R. Inhibitor design for 3-hydroxy-3-methyl-glutaryl-CoA reductase enzyme; molecular docking and determination of molecular and electronic properties of ligands by density functional theory method. *J. Heterocycl. Chem.* 2020;57(7):2875–88. <https://doi.org/10.1002/jhet.3996>. 2020/07/01.
- [22] Ercan S, Şenses Y. Design and molecular docking studies of new inhibitor candidates for EBNA1 DNA binding site: a computational study. *Mol. Simulat.* 2020; 46(4):332–9. <https://doi.org/10.1080/08927022.2019.1709638>. 2020/03/03.
- [23] Ercan S, Senyigit B, Senses Y. Dual inhibitor design for HIV-1 reverse transcriptase and integrase enzymes: a molecular docking study. *J. Biomol. Struct. Dyn.* Feb 2020; 38(2):573–80. <https://doi.org/10.1080/07391102.2019.1700166>.
- [24] Ghddi. Selected Drug Information of Current Ongoing Clinical Studies on COVID-19. The Global Health Drug Discovery Institute; 2020. Accessed 25.04.2020, <https://ghddi-aialab.github.io/Targeting2019-nCoV/clinical>.
- [25] Kim S, Chen J, Cheng T, et al. PubChem 2019 update: improved access to chemical data. *Nucleic Acids Res.* Jan 8 2019;47(D1):D1102–9. <https://doi.org/10.1093/nar/gky1033>.
- [26] Wishart DS, Feunang YD, Guo AC, et al. DrugBank 5.0: a major update to the DrugBank database for 2018. *Nucleic Acids Res.* Jan 4 2018;46(D1):D1074–82. <https://doi.org/10.1093/nar/gkx1037>.
- [27] Marinho EM, Batista de Andrade Neto J, Silva J, et al. Virtual screening based on molecular docking of possible inhibitors of Covid-19 main protease. *Microb. Pathog.* Jun 30 2020;148:104365. <https://doi.org/10.1016/j.micpath.2020.104365>.
- [28] Sepay N, Sekar A, Halder UC, Alarifi A, Afzal M. Anti-COVID-19 terpenoid from marine sources: a docking, ADMET and molecular dynamics study. *J. Mol. Struct.* Oct 2020;10:129433. <https://doi.org/10.1016/j.molstruc.2020.129433>.
- [29] Vardhan S, Sahoo SK. In silico ADMET and molecular docking study on searching potential inhibitors from limonoids and triterpenoids for COVID-19. *Comput. Biol. Med.* Sep 2020;124:103936. <https://doi.org/10.1016/j.combiomed.2020.103936>.
- [30] Yu JW, Wang L, Bao LD. Exploring the active compounds of traditional Mongolian medicine in intervention of novel coronavirus (COVID-19) based on molecular docking method. *J. Funct. Foods* Aug 2020;71:104016. <https://doi.org/10.1016/j.jff.2020.104016>.
- [31] Mu C, Sheng Y, Wang Q, Amin A, Li X, Xie Y. Potential compound from herbal food of rhizoma polygonati for treatment of COVID-19 analyzed by network pharmacology and molecular docking technology. *J. Funct. Foods* Aug 14 2020: 104149. <https://doi.org/10.1016/j.jff.2020.104149>.
- [32] Abo-Zeid Y, Ismail NSM, McLean GR, Hamdy NM. A molecular docking study repurposes FDA approved iron oxide nanoparticles to treat and control COVID-19 infection. *Eur. J. Pharmaceut. Sci.* Oct 1 2020;153:105465. <https://doi.org/10.1016/j.ejps.2020.105465>.
- [33] Kumar Y, Singh H, Patel CN. In silico prediction of potential inhibitors for the Main protease of SARS-CoV-2 using molecular docking and dynamics simulation based drug-repurposing. *J. Infect. Public Heal.* Sep 2020;13(9):1210–23. <https://doi.org/10.1016/j.jiph.2020.06.016>.
- [34] Li N, Thompson S, Schultz DC, et al. Discovery of selective inhibitors against EBNA1 via high throughput in silico virtual screening. *PLoS One* Apr 12 2010;5(4):e10126. <https://doi.org/10.1371/journal.pone.0010126>.
- [35] Akihisa T, Hamasaki Y, Tokuda H, Ukiya M, Kimura Y, Nishino H. Microbial transformation of isosteviol and inhibitory effects on Epstein-Barr virus activation of the transformation products. *Journal of Natural Products.* Mar 2004;67(3): 407–10. <https://doi.org/10.1021/np030393q>.
- [36] Gianti E, Messick TE, Lieberman PM, Zauhar RJ. Computational analysis of EBNA1 "druggability" suggests novel insights for Epstein-Barr virus inhibitor design. *J. Comput. Aid. Mol. Des.* Apr 2016;30(4):285–303. <https://doi.org/10.1007/s10822-016-9899-y>.
- [37] Hu L, Zhang Y, Zhu H, et al. Filicinic acid based meroterpenoids with anti-epstein-barr virus activities from *Hypericum japonicum*. *Org. Lett.* May 6 2016;18(9): 2272–5. <https://doi.org/10.1021/acs.orglett.6b00906>.
- [38] Tikhmyanova N, Schultz DC, Lee T, Salvino JM, Lieberman PM. Identification of a new class of small molecules that efficiently reactivate latent Epstein-Barr Virus. *ACS Chem Biol.* Mar 21 2014;9(3):785–95. <https://doi.org/10.1021/cb4006326>.
- [39] Ukiya M, Akihisa T, Tokuda H, et al. Sunpollenol and five other rearranged 3,4-seco-tirucallane-type triterpenoids from sunflower pollen and their inhibitory effects on Epstein-Barr virus activation. *Journal of Natural Products.* Nov 2003; 66(11):1476–9. <https://doi.org/10.1021/np030276v>.
- [40] Wu T, Wang Q, Jiang C, et al. Neo-clerodane diterpenoids from *Scutellaria barbata* with activity against Epstein-Barr virus lytic replication. *J. Nat. Prod.* Mar 27 2015; 78(3):500–9. <https://doi.org/10.1021/np500988m>.
- [41] Sterling T, Irwin JJ. Zinc 15 – ligand discovery for everyone. *J. Chem. Inf. Model.* 2015;55(11):2324–37. <https://doi.org/10.1021/acs.jcim.5b00559>. 2015/11/23.
- [42] Yoshino R, Yasuo N, Sekijima M. Identification of key interactions between SARS-CoV-2 main protease and inhibitor drug candidates. *Sci. Rep.* Jul 27 2020;10(1): 12493. <https://doi.org/10.1038/s41598-020-69337-9>.
- [43] Nguyen HL, Thai NQ, Truong DT, Li MS. Remdesivir strongly binds to both RNA-dependent RNA Polymerase and main protease of SARS-CoV-2: evidence from molecular simulations. *J. Phys. Chem. B* Dec 17 2020;124(50):11337–48. <https://doi.org/10.1021/acs.jpcc.0c07312>.
- [44] Naik VR, Munikumar M, Ramakrishna U, et al. Remdesivir (GS-5734) as a therapeutic option of 2019-nCoV main protease -in silico approach. *J. Biomol. Struct. Dynam.* Jun 20 2020. <https://doi.org/10.1080/07391102.2020.1781694>.
- [45] Nimgampalle M, Devanathan V, Saxena A. Screening of Chloroquine, Hydroxychloroquine and its derivatives for their binding affinity to multiple SARS-CoV-2 protein drug targets. *J. Biomol. Struct. Dynam.* Jun 23 2020. <https://doi.org/10.1080/07391102.2020.1782265>.