Review Article

Ferroptosis Mediated by Lipid Reactive Oxygen Species: A Possible Causal Link of Neuroinflammation to Neurological Disorders

Ying Cheng,¹ Yiting Song,¹ Huan Chen,¹ Qianqian Li,² Yuan Gao,¹ Guanchao Lu,³ and Chengliang Luo,¹

¹Department of Forensic Medicine, School of Basic Medicine and Biological Sciences, Soochow University, Suzhou 215123, China ²School of Forensic Medicine, Wannan Medical College, Wuhu 241002, China ³Department of Neurology, Fuping County Hospital, Weinan 711700, China

Correspondence should be addressed to Yuan Gao; gaoyuansz@suda.edu.cn, Guanchao Lu; luyishisanxian@163.com, and Chengliang Luo; clluo@suda.edu.cn

Received 26 August 2021; Accepted 13 October 2021; Published 23 October 2021

Academic Editor: Francisco Rios

Copyright © 2021 Ying Cheng et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Increasing evidence indicates a possible causal link between neuroinflammation and neurological disorders, including Alzheimer's disease (AD), Parkinson's disease (PD), Huntington's disease (HD), and stroke. A putative mechanism underlying such a link can be explained by ferroptosis. Current studies have shown that disturbances of iron homeostasis, glutamate excitatory toxicity, lipid reactive oxygen species (ROS), and other manifestations related to ferroptosis can be detected in several neurological disorders caused by neuroinflammation. To date, compelling evidence indicates that damage-associated molecular pattern (DAMP) molecules (e.g., ROS) produced in the process of ferroptosis activate glial cells by activating neuroimmune pathways and then produce a series of inflammatory factors which contribute to neurological disorders. Our review article provides a current view of the involvement of ferroptosis or ROS in the pathological process of neuroinflammation, the effects of neuroinflammation mediated by ferroptosis in neurological disorders, a better understanding of the mechanisms underlying ferroptosis participates in neuroinflammation, and the potential treatments for neurological disorders. In addition, further research on the mechanisms of ferroptosis as well as the link between ferroptosis and neuroinflammation will help provide new targets for treatment.

1. Introduction

In 2003, Dolma et al. discovered that camptothecin (CPT) and a new compound erastin could selectively kill tumor cells that expressed RasV12 protein, but their lethal mechanisms are different. Cell death induced by CPT showed changes in nuclear morphology, deoxyribonucleic acid (DNA) fragmentation, and caspase activation, which could be inhibited by caspase inhibitors. However, cell death induced by erastin was different from what had been seen before [1]. Therefore, erastin may have induced a new type of cell death. Later, Yang and Stockwell and Yagoda et al. showed that this mode of cell death could be inhibited by iron-chelating agents, and increased ROS appeared in cells

[2, 3]. Dixon et al. officially named this mode of cell death ferroptosis, an iron-dependent and nonapoptotic form of cell death characterized by intracellular accumulation of ROS. Through electron microscopy, it can be observed that the mitochondria of cells with ferroptosis are significantly shrunken with an increased membrane density [4].

Neuroinflammation is a complex process attributing to the interactions of glial cells and peripheral immune cells in the central nervous system (CNS). The activation of glial cells and peripheral lymphocytes releases inflammatory factors that cause damage to the nervous system. Current studies have shown that disturbances of iron homeostasis, lipid ROS accumulation, and other manifestations related to ferroptosis can be detected in some neurological disorders caused by neuroinflammation [5–10], indicating that ferroptosis may play a tremendous role in the occurrence and development of neurological disorders, including Alzheimer's disease (AD), Parkinson's disease (PD), Huntington's disease (HD), and stroke. Therefore, we give a current view on the relationship between ferroptosis and neuroinflammation in neurological disorders, including the putative pathway for ferroptosis that participates in neuroinflammation and the potential treatments for neurological disorders.

2. The Pathway of Ferroptosis

As a new type of cell death, ferroptosis is iron-dependent and induced by intracellular lipid peroxidation accumulation. The occurrence of ferroptosis is caused by the imbalance between the production and degradation of lipid ROS in cells. Here, we summarize the main mechanisms involved in the process of ferroptosis such as accumulation of lipid ROS, inhibition of system Xc⁻, glutathione peroxidase 4 (GPX4) enzyme activity, and iron metabolism disorder (Figure 1).

2.1. Ferroptosis Mediated by Lipid Reactive Oxygen Species. Apart from the iron-mediated ROS produced by the Fenton reaction, there are plentiful potential sources of ROS in the cell. Among them, one pivotal generator of intracellular ROS is NADPH oxidases (NOX) which is a family of seven transmembrane proteins which transfer electrons across biological membranes, including five monooxidases (NOX1-5) and two dual oxidases (Duox1-2) [11, 12]. When oxygen accepts an electron, the product of the electron transfer reaction is superoxide. The biological function of NOX enzymes is therefore the generation of ROS, including hydroperoxides (H_2O_2) [13], which contributes to Fenton reaction. Members of the NOX family have been shown to promote ferroptotic cancer cell death [14–16]. The NOX inhibitor could prevent erastin-induced ferroptotic death [4], whereas NOX activation sensitizes ferroptosis [15]. Mitochondria are another main source of intracellular oxidant production. Electrons are often lost during the transfer of electron transport chain complexes in oxidative phosphorylation of mitochondria. ROS are generated after electrons binding with molecular oxygen, eventually leading to oxidative stress.

As transmembrane channels, voltage-dependent anion channels (VDACs) transport ions and metabolites [17]. Yagoda et al. found that erastin could act on VDACs. With the help of RNA interference (RNAi) technology, it was found that cells with lower expression of VDAC2 or VDAC3 were tolerant to ferroptosis induced by erastin. Furthermore, erastin can also lead to changes in mitochondrial outer membrane permeability. Consequently, Yagoda et al. believed that the interference of VDACs may conduce to mitochondrial dysfunction and ROS release [3].

In addition to NADPH oxidases and mitochondria, extra sources of ROS include plentiful enzymes such as cyclooxygenases (COX), lipoxygenases (LOX), xanthine oxidase (XO), and monoamine oxidase (MAO) which produce ROS because of their enzyme function [18]. Studies have expounded that the polyunsaturated fatty acid (PUFA) chain

of membrane lipids can form lipid ROS by reacting with ROS [4]. PUFAs can undergo enzymatic or nonenzymatic oxidation reactions to form lipid H_2O_2 , which can combine with iron to produce toxic lipid free radicals, causing cell damage. These free radicals can transfer protons adjacent to PUFAs, initiating a new round of lipid oxidation reactions and further transmitting oxidative damage, which directly triggers ferroptosis. Lysophosphatidylcholine acyltransferase 3 (LPCAT3) and acyl-CoA synthetase long-chain family member 4 (ACSL4), two genes related to lipid metabolism, can induce lipid peroxidation. Hence, the suppression of ACSL4 and LPCAT3 may play a fundamental role in inhibiting ferroptosis [19]. Interestingly, Gao et al. found that autophagy may engender ferroptosis by regulating intracellular iron homeostasis and the generation of ROS [20]. Park and Chung also observed that ferroptosis inducer (erastin) enhanced intracellular lipid peroxidation and ROS in wildtype cells, but not in autophagy defective cells, which indicated that autophagy may induce ferroptosis by increasing lipid peroxidation [21]. What is more, Ma's group demonstrated that iron-mediated ROS generation can induce autophagy [22]. This complex cycle continuously induces ferroptosis and the production of lipid ROS, causing cell damage.

2.2. Inducing Ferroptosis by Suppressing System Xc⁻. System Xc⁻, a heterodimer composed of SLC7A11 and SLC3A2 subunits, is an amino acid antiporter widely distributed in the phospholipid bilayer and also well known as an important antioxidant component. System Xc balances excess hydrogen peroxide and peroxides by regulating the synthesis of glutathione (GSH) through the import and export of cysteine and glutamate [23]. Yang and Stockwell found that the decrease of GSH leads to the decrease in the activity of GPXs, which plays a crucial role in inhibiting the generation of lipid ROS [2], and glutathione is also an essential cofactor that contributes as the main substrate of GPX4 [24]. Pharmacologically, several system Xc inhibitors have been reported such as erastin, blocking the absorption of cystine, which induces ROS accumulation and ferroptosis [25]. The mRNA and protein expression levels of SLC7A11 were significantly decreased after the upregulation of P53 gene expression, thus confirming that SLC7A11 is a new target of the P53 gene [26].

2.3. Inducing Ferroptosis by Suppressing GPX4. There are many members of the GPXs family, including GPX1~GPX8. GPX4 converts reduced glutathione (GSH) to oxidized glutathione (GSSH) and reduces cellular lipid hydroperoxides (L-OOH) to corresponding alcohols [27, 28]. Research has already reported that cells with downregulated GPX4 expression were more sensitive to malnutrition, whereas upregulated GPX4 expression inhibited ferroptosis [2], indicating that GPX4 plays an important role in ferroptosis. In addition, current studies founded that an official ferroptosis inducer (RSL3) and some other compounds, such as DPI7 and DPI10, can directly inhibit GPX4 and reduce the antioxidant capacity of cells. Moreover, the mevalonate pathway which targets GPX4 by regulating the maturation of



FIGURE 1: Mechanism pathways of ferroptosis.

selenocysteine tRNA gives rise to ferroptosis [29]. Selenocysteine is one of the amino acids in the active center of GPX4, and the insertion of selenocysteine into GPX4 requires a special transporter—selenocysteine tRNA [30]. Maturation of selenocysteine tRNA depends on the transfer of isopentenyl groups from isopentenyl pyrophosphate (IPP) to the precursor of selenocysteine tRNA by isopentenyl transferase [31]. In consequence, the mevalonate pathway can affect the synthesis of selenocysteine tRNA by downregulating IPP and further interfere with the activity of GPX4, resulting in ferroptosis.

2.4. Ferroptosis Mediated by Iron Metabolism Disorder. Free iron ions in serum are always transported together with transferrin. The Tf-Fe³⁺ complex binds to the transferrin receptor 1 (TfR1) on the cell membrane and enters the cell through endocytosis, forming a stable endosome. Fe³⁺ in the cells was reduced to Fe²⁺ by the six-transmembrane epithelial antigen of the prostate 3 (STEAP3). Fe²⁺ was transported from the nucleus to the unstable cytosol mediated by divalent metal transporter 1 (DMT1). The excessive free iron ions are stored in ferritin or transferred to the outside of the cell by ferroportin (Fpn) to maintain intracellular iron homeostasis [32]. Our recent study expounded that ferritinknockout mice are less resistant to ferroptosis [33]. Excess iron can directly catalyze the production of lipid ROS through the Fenton reaction that brings about the continuous accumulation of intracellular lipid ROS and ferroptosis [34]. In addition, non-transferrin-bound iron (NTBI) can be transferred into the cell by ZIP14 and ZIP8 (SLC39A14 and SLC39A8) [35, 36]. Cascades underpinning imbalance of iron metabolism could be active drivers of neuropathology in major neurological disorders.

3. The Involvement of Ferroptosis in the Pathological Process of Neuroinflammation

Given that ferroptosis has been discovered to promote pathological processes in neurological disorders, many researchers sought to gain insight into mechanisms of ferroptosis that participates in neuroinflammation to neurological disorders. Summarily, DAMP molecules, produced in the process of ferroptosis, activate glial cells by activating neuroimmune pathways. Activated glial cells produce a series of inflammatory factors which contribute to neuronal damage and a series of neurological disorders (Figure 2).

3.1. Activation of NF- κ B Pathway by ROS. In the pathogenesis of ferroptosis, a key point is the deficiency of GPX4, which leads to the accumulation of lipid ROS and further bringing about ferroptosis [29, 37]. GPX4 utilizes GSH as a cofactor to remove lipid peroxidation from the membrane. GPX4 reduces the complex hydroperoxide including phosphatidyl hydroperoxide and cholesterol hydroperoxide to the corresponding alcohol to maintain the redox balance and avoid harmful oxidative stress damage. ROS and



FIGURE 2: Putative pathway for ferroptosis participates in neuroinflammation to neurological disorders. Damage-associated molecular pattern (DAMP) molecules (e.g., ROS, cfDNA, HMGB1, ITs, and PGs) produced in the process of ferroptosis activate glial cells by activating neuroimmune pathways. Activated glial cells produce a series of inflammatory factors which contribute to neuronal damage and a series of neurological disorders, such as Alzheimer's disease, Parkinson's disease, Huntington's disease, and stroke.

oxidative stress are common features associated with inflammation [38]. ROS can activate the nuclear factor kappa B (NF- κ B) transcription factor system, which is a key player in inflammation-related cancers. The core components of the NF- κ B pathway are mainly the inhibitory I κ B protein, the IkB kinase (IKK) complex, and the transcription factor

NF- κ B itself. External stimuli trigger IKK, initiating the phosphorylation, ubiquitination, and degradation of IkB protein. The released dimer of NF-kB is further activated by various posttranslational modifications and is transferred to the nucleus to bind to specific DNA sequences, promoting the transcription of target genes. Studies have shown that

ROS, especially H₂O₂, can activate IKK in certain cell types [39]. ROS promotes IKKy/NEMO dimerization which is the subunit of IKK by promoting the formation of disulfide bonds between Cys54 and Cys347, resulting in the activation of IKK and the initiation of the NF-kB pathway. Additionally, ROS regulation of the upstream NF-*k*B activation pathway is also reflected in selective I κ B α phosphorylation [40]. $I\kappa B\alpha$ is usually phosphorylated at Ser32 and Ser36, but H_2O_2 causes ubiquitination and degradation of IkBa by influencing the phosphorylation of $I\kappa B\alpha$ at Tyr42 or other tyrosine residues. The released dimer of NF- κ B migrates to the nucleus, where it binds to the κB site on the DNA sequence and promotes the transcription of target genes. Therefore, GPX4 plays a significant role in the elimination of peroxides, reducing the accumulation of ROS and inhibiting the activation of NF- κ B. NF- κ B is a nuclear transcription factor closely related to inflammation [41], which is expressed in almost all cell tissues, such as microglia, neurons, and astrocytes. It regulates multiple functional target genes and participates in a variety of biological processes such as immune response, inflammatory response, apoptosis, and tumorigenesis. Microglia are one of the most important immune cells in the CNS and play a vital role in both normal brain development and neuroinflammatory diseases. The NF- κ B signaling pathway of the glial also plays a critical role in inflammatory responses in the nervous system. Chronic activation of NF- κ B can induce transcription of various neuroinflammatory genes [42] and promote the secretion of inflammatory cytokines, such as TNF- α , IL-6, IL- 1β , nitric oxide, adhesion molecules, and neurotoxins, inducing neuronal apoptosis. Accumulated ROS during the ferroptosis process in the brain leads to the activation of the NF- κ B pathway in glial cells and the occurrence of neuroinflammation.

3.2. Neuroinflammatory Damage Caused by Lipid Peroxidation. In the complex mechanism of ferroptosis, another hallmark lipid peroxidation must be mentioned. Researches have shown that lipids not only form lipid peroxides to interfere with the structures of membrane and functions of membrane proteins but also induce intracellular signal transduction and stimulate other pathways leading to cell death [4, 43]. Many studies have illustrated that lipid may play a vital role in inflammation, immunity, metabolism, and various biofilms. Excessive ROS such as H₂O₂ and hydroxyl radical (·OH), as well as reactive nitrogen species (RNS) such as NO. and ONOO-, may give rise to oxidative stress [44], oxidizing biological macromolecules such as lipids, DNA, or proteins. The ·OH is the most dynamic and dangerous reactive oxygen radical, which is formed by H_2O_2 in the presence of metal ions. Hydroxyl radicals play a key role in lipid peroxidation, which oxidizes lipids containing carbon-carbon double bonds, especially PUFAs. In PUFAs, the C=C units are separated by a single-bonded C atom, and the hydrogen atoms (diallyl hydrogens) attached to these C atoms (bis-allylic hydrogens) are vulnerable to hydroxyl radical. Therefore, PUFA such as arachidonic acid (AA) is highly oxidized fatty acid. The peroxides of PUFAs and their final production-reactive aldehydes like 4-

hydroxynonaldehyde (4-HNE)-lead to carbonylation of proteins, which is associated with atherosclerosis, neurodegenerative diseases, and other diseases [45]. Oxidation of PUFAs can be carried out by nonenzymatic or enzymatic means. Lipoxygenases are ferric enzymes that catalyze the oxygenation of PUFAs to produce fatty acid hydroperoxides. According to the research, the cascade reaction of arachidonic acid involves enzymes related to lipid peroxidation metabolism [46]. The oxidation cascade of AA first releases the polyunsaturated fatty acid AA from the N-2 position of the membrane phospholipid by phosphatase A2 (PLA2). AA produces bioactive eicosanoids (prostaglandins, leukotrienes, and epoxy-fatty acids) using enzymes related to lipid peroxidase (COX, LOX, and cytochrome P450 enzymes). Under physiological conditions, COX-2 plays a role in regulating cerebral vascular and synaptic plasticity in the nervous system. Under pathological conditions, the metabolism of AA mediated by COX to produce prostaglandin was also enhanced in a rat model of focal cerebral ischemia [47]. Prostaglandins, a class of carotenoids, are reported to be a key factor in ischemic and excitotoxic nerve injury. Overexpression of COX-2 during lipid peroxidation mediates neurotoxicity through prostaglandin E2 (PGE2). If the toxic stimulation persists, it may also lead to secondary neuroinflammation damage of the CNS. Experimental studies support that the application of COX-2 inhibitors reduces the level of proinflammatory cytokines such as IL-1 β and TNF- α [48, 49]. Drug suppression or gene knockout of 15-LOX also leads to reduced production of IL-1 and TNF- α [50, 51].

3.3. DAMPs Inducing Neuroinflammation. Iron disturbance is a basic element of ferroptosis. Excess iron from hemoglobin and the periphery can be transported through a series of ion channels and transporters, including divalent metal transporter 1 (DMT1), transferrin receptor 1 (TfR1), and ferroportin (Fpn). Fe²⁺ directly catalyzes the generation of lipid ROS through the Fenton reaction, resulting in the continuous accumulation of intracellular lipid ROS and causing ferroptosis. It was founded that excess Fe²⁺ transported from the microglia resulted in iron metabolism disorder [34]. In the CNS, glial cells not only continuously monitor the dynamic balance of the internal environment but also act as immune cells, resisting endogenous and exogenous pathogenic stimuli [52]. Immune cells recognize pathogens primarily by pattern recognition receptors (PRRs), which include pathogen-associated molecular patterns (PAMPs) and damage-associated molecular patterns (DAMPs). The main form of PRRs is the Toll-like receptor (TLR) family, which is not only abundant in immune cells but also expressed in CNS glial cells. The danger signal DAMP is an endogenous molecule encoded directly by endogenous genes of the host. It can be released immediately under certain conditions of tissue damage, without the need for protein synthesis. DAMPs from ferroptosis cells and proinflammatory cytokines provide prefeedback signals that reinforce the programmed death of more cells [53]. Severe cycles of death can lead to barrier dysfunction, with inflammation spreading throughout the body, which may

eventually bring about extensive multiorgan failure. DAMPs, hemoglobin, erythrocyte, and iron are produced during ferroptosis [54], which can be recognized by PRRs, namely, TLRs in glial cells. Both microglia and astrocytes are activated and secrete inflammatory cytokines such as IFN- α , IFN- β , IL-1 β , IL-6, IL-10, IL-12, IL-18, and TNF- α , as well as ROS and chemokines, participating in neuroinflammation caused by an infection in the nervous system [55-57]. In the TLR family, the TLR3 signal induces the strongest proinflammatory polarization response, which is characterized by the secretion of high levels of IL-12, TNF- α , IL-6, CXCL10, IL-10, and IFN- β [58]. In acute infectious or noninfectious CNS diseases caused by ferroptosis of nerve cells, activated TLR signals in glial cells are involved in neuronal injury under the action of endogenous and exogenous pathogens or the release of products from cell damage. In AD, amyloid, S100 protein, and high mobility group box-1 (HMGB1) are secondary key factors following primary neuritis, further exacerbating the production of proinflammatory cytokines. HMGB1 belongs to the Ia DAMP category, which can be perceived by PRRs on several cells. Class Ia DAMPs signal danger to surrounding PRR-carrying cells, such as phagocytes, triggers sterile inflammation [59].

3.4. Ferroptosis Mediated by P53 Gene Inducing Neuroinflammation. P53 gene, as a tumor suppressor gene that mediates the inhibition, senescence, and apoptosis of cell cycle, plays an important role in tumorigenesis and development. Recently, acetylated defective P53 mutants have been found to promote ferroptosis [60]. Jiang et al. discovered that the activity of H1299 cells with P53 gene silenced did not change when they were treated with ROS [61]. However, treated with ROS after the P53 gene was activated, 90% of the H1299 cells died, indicating that the activation of P53 reduced the antioxidant capacity of these cells. Studies have demonstrated that the P53 gene mainly affects the synthesis of GSH by inhibiting system Xc⁻ [32]. In addition, P53 can promote ferroptosis by enhancing the expression of spermine N1-acetyltransferase 1 (SAT1), which acetylates spermine and is an important ratelimiting enzyme in polyamine catabolism. The abnormal metabolism of polyamine and aberrant SAT1 expression is associated with a variety of pathological conditions, including neurological disorders and cancer [62, 63]. The activity of SAT1 will increase in a variety of stress reactions, including oxidative stress and inflammatory stimuli. The activation of SAT1 may lead to lipid peroxidation and ferroptosis induced by ROS, resulting in neuroinflammatory damage. Previous studies have also observed that overexpression of SAT1 leads to rapid depletion of spermine in cells, ultimately giving rise to significant growth inhibition and mitochondrial apoptosis [64]. The induction of SAT1 was related to the expression level of 15-LOX, and pharmacological inhibition of 15-LOX by PD146176 attenuated SAT1-mediated ferroptosis [65]. Moreover, 15-LOX, together with COX-2 which is overexpressed during ferroptosis, is also involved in the metabolic process of arachidonic acid, producing prostaglandins, peroxides, ROS, and other substances to induce inflammatory injury of the nervous system [66].

4. The Effects of Neuroinflammation Mediated by Ferroptosis in Neurological Disorders

Neuroinflammation of the CNS is an extremely complex process. In the initial stage, glial cells act as immune cells, performing defensive reactions in response to traumatic stimuli. Mediated by ATP, microglia are activated to produce proinflammatory cytokines [67, 68]. Local damage of the CNS may affect other brain regions far away from the damage site through Ca²⁺ waves [69]. Lipid ROS dysregulation can also destroy physiological calcium signals [70]. During the progressive stage of neuroinflammation, the inflammatory microenvironment may induce glial cells to transform into different phenotypes and regulate inflammation through cell feedback and cell-to-cell communication mechanisms, resulting in the proinflammatory and antiinflammatory reactions of neuroinflammation. Besides, ferroptotic neurons themselves also could release proinflammatory DAMPs, which then triggered the innate immune system in brain tissues [54]. In the prognostic stage of neuroinflammation, glial cells are subjected to endogenous changes such as persistent stimulation, aging, or gene defects under severe pathological conditions, which will lead to chronic neuroinflammation of the CNS and degeneration of neurons. Chronic neuroinflammation is one of the most important mechanisms in the pathogenesis of neurodegenerative diseases. Physiologically, iron, as an important redox metal, produces ATP in the cellular metabolism of the CNS, which supplies energy for cell metabolism in physiological processes. However, when iron metabolism is disrupted, the nerve system is vulnerable to damage caused by excess iron and oxidative stress. Recently, studies supported that iron metabolism disorders, glutamate excitatory toxicity, lipid ROS accumulation, and other characteristics related to ferroptosis can be detected in some neurological disorders. Accumulated evidence demonstrates that ferroptosis participates in the neuroinflammatory pathological process. Ferroptosis inhibitors have also been found to improve the prognosis of neurological disorders. Therefore, further investigation into the mechanism of ferroptosis involved in the neuroinflammatory process will be valuable to the treatment and improvement of neurological disorders. The effects of neuroinflammation mediated by ferroptosis in several characteristic neurological disorders are described below.

4.1. Alzheimer's Disease (AD). Among the classical properties of AD, the amyloid cascade theory is an important study to explain the pathogenesis of AD. Amyloid β -protein (A β) accumulation is also considered to be an initial event in the pathogenesis of AD. Another feature of AD is intracellular neurofibrillary tangles (NFT) due to hyperphosphorylation of Tau protein. Under physiological conditions, tau protein stabilizes microtubules and regulates axon transport. Pathological changes will make tau separate from microtubules, resulting in synaptic loss and neuronal dysfunction. What is more, the neurocognitive decline is associated with synaptic reduction and neurotransmitter oxidation, which could

be attributed to oxidative stress, mainly the increase of ROS and intracellular and extracellular hydrogen peroxide [71, 72]. There has been early evidence of an association between iron accumulation in the cerebral cortex and the development of AD [73, 74]. Svobodová et al. proved that free iron and ferritin accumulated after amyloid plaque formation in the cerebral cortex of β -amyloid precursor protein (APP)/presenilin 1 (PS1) transgenic mouse model [75]; iron deposition is involved in the formation of $A\beta$ plaque and the misfolding process of NFT. Inhibition of lipid peroxidation and ferroptosis mediated by ACSL4 and ACSL4 special protein 1 (SP1) mimicked the beneficial effects of aldehyde dehydrogenase (ALDH2) overexpression in APP/PS1mutant mice [76]. APP is a type 1 transmembrane protein; the proteolytic cleavage of which forms the A β is associated with the pathogenesis of AD. In the presence of free iron, $A\beta$ plaques are effectively involved in ROS production, leading to increased lipid peroxidation, protein oxidation, and DNA damage [77]. Proteolysis cleavage in APP occurs through enzyme complexes including α -secretase or β -secretase and y-secretase. Tsatsanis et al. showed that APP promotes the iron efflux of neurons by stabilizing the expression of ferroportin on the cell surface [78]. The proteolytic cleavage of APP under the action of β -secretase leads to the accumulation of iron in the neuron cells, resulting in the generation of A β . Hambright et al. found in a mouse model with a condition deletion in neurons of the forebrain of GPX4 that GPX4-deficient mice have significant learning and memory dysfunction, which is related to markers of ferroptosis, such as increased lipid peroxidation and neuroinflammation [79]. Furthermore, treatment with liproxstatin-1, a ferroptosis inhibitor, ameliorated neuronal death and memory impairment induced by $A\beta$ [80]. In addition, enzymes involved in lipid peroxidation, such as COX-2, LOX, and cytochrome P450 enzymes, metabolize AA into substances such as prostaglandins, leukotrienes, and epoxide fatty acids. COX-2 mediates neurotoxicity through PGE2 produced from AA, which is an important member of DAMPs. If the toxic stimulation persists, it may also conduce secondary neuroinflammation damage in the CNS. More and more evidence indicates that the amyloid cascade theory alone may not be convinced to explain the pathogenesis of AD. Therefore, researchers attempt to explore the relationship with other pathological mechanisms. The increased levels of inflammatory markers in the brain of AD patients and the discovery of AD risk genes related to innate immune function suggest that neuroinflammation may play a significant role in the pathogenesis of AD [81-85]. A hallmark of neuroinflammation is the production of proinflammatory cytokines, and the innate immune cells involved in the process of neuroinflammation are mainly microglia and astrocytes. Capillary endothelial cells and permeable blood cells also participate in neuroinflammation, especially when the blood-brain barrier (BBB) is subjected to biochemical or mechanical damage [86, 87]. Astrocytes respond to pathological injury through reactive gliosis, and inflammatory injury induces A1 astrocyte phenotype through the NF- κ B pathway, expressing inflammatory mediators. Microglia recognize pathogens, cell debris, or abnor-

mal proteins through TLR and induce microglia cell response [82], internalizing and degrading pathogenic substances through pinocytosis. The process usually disappears once the pathogen or irritant is eliminated. Nonetheless, the function of microglia in the brain of the elderly is impaired. In the case of severe pathological conditions or continuous stimulation by pathogens, the glia is always in the activated state and secreting inflammatory factors, leading to neuroinflammatory damage [88]. Furthermore, crosstalk between glial cells and neurons can also be observed in AD. A β activates NF- κ B in astrocytes, resulting in an increased release of complement C3, which acts on the C3a receptors of microglia and neurons, and the secretion of IL-1 α in activated microglia induces the production of A1 astrocytes. In the inflammatory environment of AD, glial crosstalk may form positive feedback, leading to nervous system disorders and self-amplifying inflammatory responses. Evidence from genome-wide association studies also supports the initiating role of neuroinflammation in AD, and mutations in microglia or innate immune genes are associated with an increased incidence of AD in the population [84]. The influence of neuroinflammation on AD has also been confirmed in clinical imaging studies, and the activation of microglia in AD patients is negatively correlated with the structural integrity or functional activity of the brain [89]. Preclinical studies correspondingly support clinical imaging observations that microglia activated by A β produce neurotoxic cytokines that lead to neuronal dysfunction [90]. Several current studies have proved that ferroptosis and neuroinflammation play a fundamental role in the pathogenesis and progression of AD, and glial cells are the core of neuroinflammation. Regulating the occurrence of lipid peroxidation and ferroptosis in neurons has important therapeutic significance for neurodegenerative diseases. Meanwhile, targeting glial cells as a breakthrough for drug treatment of AD also has a certain prospect.

4.2. Parkinson's Disease (PD). The typical pathological feature of Parkinson's disease is the degeneration of dopaminergic neurons in the substantia nigra, which is linked with the systematic and progressive accumulation of iron in the substantia nigra, leading to the consumption of dopamine in the striatum. Meanwhile, Lewy bodies also appear in the cells, with α -synuclein aggregation as the main component. Studies have shown that the accumulated α -synuclein is accountable for the generation of ROS followed by lipid peroxidation in an iron-dependent manner, leading to cell death [70]. The increase of iron in the substantia nigra and globus pallidus in PD patients has also been observed by magnetic resonance imaging (MRI), which is closely related to the duration of disease, neurodegeneration, and the severity of sports injuries [6, 91]. The study indicated that treatment with deferiprone, an iron chelator, substantially reduced unstable iron and biological damage in oxidatively stressed cells, which improve motor function. In addition, studies have shown that the related mechanisms of neuroinflammation, such as activation of microglia, the proliferation of astrocytes, and other peripheral immune cells, may contribute to the impairment of dopaminergic neurons in the

substantia nigra. Some cellular and molecular responses related to neuroinflammation may induce harmful events such as oxidative stress and apoptosis mediated by cytokine receptors, eventually leading to dopamine cell death [92]. In 1988, McGeer et al. found activated microglia in the substantia nigra of the nervous system in an autopsy study of patients with Parkinson's disease [93]. Bertrand and colleagues reported that there is mild microglial activation in the locus blue in patients with Parkinson's disease [94]. These are important neuropathological features of Parkinson's disease. Activated microglia produce voluminous oxygen-derived and nitrogen-derived products, among which NO and O²⁻ can combine to form a highly reactive nitrogen species peroxynitrite (ONOO-), which can cause oxidative damage to various proteins. Another important neuropathological feature of the substantia nigra in Parkinson's disease is astrocyte response. Astrocytes may protect the nervous system by eliminating reactive oxygen species in the nervous system through GPX4 or secreting glial cellderived neurotrophic factor (GDNF). The deficiency or decreased activity of GPX4 may cause ROS accumulation, lipid peroxidation, or activation of the NF-kB pathway to produce inflammatory mediators, resulting in inflammatory damage to the nervous system. There may be few astrocytes around vulnerable neurons in patients with PD, and a limited astrocyte environment may be a susceptibility factor for the disease [95]. Mogi et al. showed increased concentrations of TNF- α , TGF- α , TGF- β 1, and IL-1 β in the striatum of patients with Parkinson's disease. TNF- α , IL-1 β , and IFN- γ were also detected in the patient's substantia nigra [96]. In addition, dopaminergic neurons express receptors for these cytokines, which combine with the cytokines to cause neurotoxicity. These inflammatory cytokines, such as TNF- α , IL-1 β , and IFN- γ , can induce the expression of nitric oxide synthase (iNOS) or COX-2. LOX and COX are important metabolic enzymes in lipid peroxidation, resulting in the metabolism of arachidonic acid into prostaglandin, leukotriene, and other substances. COX-2 mediates neurotoxicity through PGE2, resulting in inflammatory damage to the nervous system. Concentrations of other enzymes involved in neuroinflammation mediated by oxidative stress are also increased in PD. Biofluid studies also support the role of neuroinflammatory processes in Parkinson's disease, with elevated levels of IL-2, IL-6, and TNF- α in the serum of patients with PD. Regardless of the original nerve conduction in PD, prevention or inhibition of ferroptosis in the nervous system, as well as the inflammatory immune response of glial cells, is important to prevent the progression of the disease.

4.3. Huntington's Disease (HD). A study demonstrated that a larger concentration of iron in neurons was observed in a mouse HD model compared to the wild-type model, suggesting that iron accumulation may be involved in neurode-generation in HD patients. When excessive Fe^{2+} is generated intracellularly, the production of lipid ROS can be directly catalyzed by the Fenton reaction, resulting in the continuous accumulation of lipid ROS and eventually causing cell ferroptosis [34]. Hence, the overexpression of antioxidant

genes in HD can be explained as an adaptive response to ROS imbalance. Ferritin is identified as an iron storage protein with antioxidant activity. Previous research reported the accumulation of ferritin in microglia of R6/2 mice appeared to be a protective mechanism to counteract iron accumulation [9]. Mitochondria are the main source of ROS in cells. During the transfer of electron transport chain complexes in oxidative phosphorylation, electrons are often lost, and ROS are generated after electron binding with molecular oxygen, ultimately, leading to oxidative stress. Furthermore, mutated Huntington's protein (mHTT) in HD is expressed in both neurons and glial cells, which can bring about the production of free radicals in the CNS, eventually triggering oxidative stress [97]. The accumulation of reactive microglia and astrocytes can be observed in the brain of HD patients. Upregulated expression of IL-6, IL-8, and tumor necrosis factor (TNF) in cerebrospinal fluid indicates immune activation. Under the condition of the injury, astrocytes magnify the inflammatory process, remove cell debris, and repair the damaged tissue. Astrocytes that express mHTT were found to be functionally deficient, with low expression of glutamate transporters and inhibited glutamate uptake, resulting in striatal neurons being affected by extracellular glutamate and exhibiting the neuronal glutamate excitability associated with ferroptosis observed in HD. In addition, astrocytes expressing mHTT have been shown to reduce the availability of the inflammatory C-C chemokine ligand 5 (CCL5) to neurons [98]. On the one hand, the mHTT reduced the transcription of CCL5 in astrocytes in an NF- κ B-dependent manner. On the other hand, it prevents astrocytes from secreting the CCL5 protein; the loss of which may adversely affect neuronal function [99, 100]. The death of neurons and protein aggregates caused by various oxidative stresses, known as DAMPs, can be recognized by PRRs, namely, Toll-like receptors (TLRs) in glial cells, resulting in activation of both microglia and astrocytes and the secretion of inflammatory cytokines, which may cause aseptic encephalitis in HD. Neuroinflammation is an important component of the pathogenesis of HD, and the expression of mutant Huntington protein alters specific processes of glial response, such as cell signaling, cytokine release, and migration. Changes in microglia and astrocyte function may result in a chronic pathogenic inflammatory response that leads to the death of other neurons, leading to more neuroinflammation.

4.4. Stroke. Stroke has an extremely high incidence, mortality, and disability, which can be classified into two major categories: hemorrhagic and ischemic stroke according to the original cause of occurrence [101]. Ischemic stroke is caused by the stenosis or occlusion of carotid and vertebral arteries and insufficient blood supply to the brain, resulting in the depletion of oxygen and nutrients, causing a cascade reaction leading to oxidative stress and mitochondrial damage [102]. Researches have indicated that ferroptosis is strongly associated with ischemic stroke. In 2014, Ahmad et al. found that in a mouse model of ischemic stroke, the content of GSH significantly decreased while the content of lipid ROS increased. The imbalance between ROS and antioxidant

defense results in oxidative stress, lipid peroxidation, and eventually neuronal damage [103]. Tuo et al. also reported in their research that ferroptosis caused neuronal degeneration after ischemic stroke, and the use of ferroptosis inhibitors such as ferrostatin-1 or liproxstatin-1 exerted the neuroprotection in a mouse ischemic model [104]. Studies have shown that selenium ions can enhance the expression of GPX4 and inhibit ferroptosis by activating transcription factors TFAP2c and Sp1 [105]. The addition of GPX4 in a mouse model of ischemic stroke exerted an effect on the improvement of behavioral defects to some extent. After ischemic stroke, microglia are activated and change morphologically and phenotypically. As the innate immune cells in the CNS, microglia play a key role in the removal of foreign bodies, cell debris, and the production of cytokines. Studies have revealed that the release of cytokines can destroy the blood-brain barrier [102], leading to the aggregation of various inflammatory cells such as macrophages, natural killer cells, and glial cells to the ischemic site to produce inflammatory responses. Interestingly, activated microglia play a dual role in ischemic stroke. On the one hand, microglia cause inflammatory damage to nerve tissue by secreting proinflammatory cytokines, and on the other hand, they also secrete anti-inflammatory factors to play a neuroprotective role [106, 107]. Pharmacological reduction of 15-LOX expression exerted to neuroprotection and improved behavioral recovery in a stroke model [108]. Jin et al. demonstrated that the consumption of microglia by PLX3397, a dual colony-stimulating factor-1 inhibitor, could aggravate cerebral infarction and neurological defects, and the failure of microglia could also exaggerate the expression of inflammatory mediators in the nervous system [109]. Different subsets of microglia may play diverse roles after ischemic stroke in the brain, striking a balance between proinflammatory and anti-inflammatory. Astrocytes, like microglia, also play essential immune functions in the CNS. Under normal physiological conditions, astrocytes take glutamate from extracellular sources. Nonetheless, in severe pathological conditions, such as ischemic stroke, the expression of glutamate transporter EAAT2 in astrocytes is defective, which prevents astrocytes from absorbing excessive extracellular glutamate, giving rise to impaired glutamate excitability of neurons. After ischemic stroke, astrocytes are stimulated by cytokines from neurons and glial cells to generate IL-1 β , IL-6, and TNF- α , which stimulate and exacerbate the inflammatory response.

5. Conclusion and Expectation

There are a series of complex biochemical reactions during ferroptosis pathology including lipid ROS accumulation, glutamate transport inhibition, and iron metabolism disorder, which promote neuroinflammation and oxidative stress response and then aggravate the CNS damage. Activated microglia, reactive astrocytes, invading T cells, and overproduction of inflammatory mediators constitute the neuroinflammatory response. In the initial stage, acute inflammatory responses that happen in CNS trauma may play a protective role, limiting the severity of the injury and enhancing neuronal repair. If the acute inflammation does not subside sufficiently, it will turn into chronic inflammation and is difficult to control. In this case, glial cells tend to aggravate oxidative stress on neurons. Although neuroinflammation is not necessary during the initial stage of neurological disorders, a persistent inflammatory response can cause the exacerbation of diseases. Voluminous studies have confirmed that ferroptosis is closely related to AD, PD, HD, stroke, and other neurological disorders. Although the characteristics and pathogenesis of ferroptosis, as well as the pathological mechanism of ferroptosis's involvement in neuroinflammation, have been initially understood, there are still many problems to be solved in this field. For instance, what are the similarities and differences in the molecular mechanisms of ferroptosis in the context of different neurological disorders? The use of inhibitors of ferroptosis in experimental animal models can improve the severity of the diseases, but whether drugs targeting ferroptosis can play a role in the clinical treatment of chronic neuroinflammatory diseases still needs a lot of clinical verification. Further researches on ferroptosis will help us to deeply comprehend the pathogenesis of neurological disorders and provide a theoretical basis and transformation strategies for the clinical prevention and treatment of neurological disorders.

Conflicts of Interest

The authors declare that there are no conflicts of interest regarding the publication of this paper.

Authors' Contributions

Ying Cheng and Yiting Song contributed equally to this work.

Acknowledgments

This study was financially supported by the National Natural Science Foundation of China (No. 81971163 and No. 82101972), the Suzhou Municipal Science and Technology Bureau (SYS2019027), a project funded by the Priority Academic Program Development of Jiangsu Higher Education Institutions (PAPD), the outstanding young backbone teacher of the "Blue Project" in Jiangsu Province (to Chengliang Luo), the National Training Program of Innovation and Entrepreneurship for Undergraduate (202110285046Z), and the Key Project of Science Foundation of Wannan Medical College (WK2020Z05).

References

- S. Dolma, S. L. Lessnick, W. C. Hahn, and B. R. Stockwell, "Identification of genotype-selective antitumor agents using synthetic lethal chemical screening in engineered human tumor cells," *Cancer Cell*, vol. 3, no. 3, pp. 285–296, 2003.
- [2] W. S. Yang and B. R. Stockwell, "Synthetic lethal screening identifies compounds activating iron-dependent, nonapoptotic cell death in oncogenic-RAS-harboring cancer cells," *Chemistry & Biology*, vol. 15, no. 3, pp. 234–245, 2008.

- [3] N. Yagoda, M. von Rechenberg, E. Zaganjor et al., "RAS-RAF-MEK-dependent oxidative cell death involving voltage-dependent anion channels," *Nature*, vol. 447, no. 7146, pp. 865–869, 2007.
- [4] S. J. Dixon, K. M. Lemberg, M. R. Lamprecht et al., "Ferroptosis: an iron-dependent form of nonapoptotic cell death," *Cell*, vol. 149, no. 5, pp. 1060–1072, 2012.
- [5] Y. Mi, X. Gao, H. Xu, Y. Cui, Y. Zhang, and X. Gou, "The emerging roles of ferroptosis in Huntington's disease," *NeuroMolecular Medicine*, vol. 21, no. 2, pp. 110–119, 2019.
- [6] M. Rossi, H. Ruottinen, S. Soimakallio, I. Elovaara, and P. Dastidar, "Clinical MRI for iron detection in Parkinson's disease," *Clinical Imaging*, vol. 37, no. 4, pp. 631–636, 2013.
- [7] J. Wu, J.-J. Yang, Y. Cao et al., "Iron overload contributes to general anaesthesia-induced neurotoxicity and cognitive deficits," *Journal of Neuroinflammation*, vol. 17, no. 1, p. 110, 2020.
- [8] S. Ayton, Y. Wang, I. Diouf et al., "Brain iron is associated with accelerated cognitive decline in people with Alzheimer pathology," *Molecular Psychiatry*, vol. 25, no. 11, pp. 2932– 2941, 2020.
- [9] D. A. Simmons, M. Casale, B. Alcon, N. Pham, N. Narayan, and G. Lynch, "Ferritin accumulation in dystrophic microglia is an early event in the development of Huntington's disease," *Glia*, vol. 55, no. 10, pp. 1074–1084, 2007.
- [10] J.-X. Ren, X. Sun, X.-L. Yan, Z. N. Guo, and Y. Yang, "Ferroptosis in neurological diseases," *Frontiers in Cellular Neuroscience*, vol. 14, p. 218, 2020.
- [11] K. Wingler, J. J. R. Hermans, P. Schiffers, A. L. Moens, M. Paul, and H. H. H. W. Schmidt, "NOX1, 2, 4, 5: counting out oxidative stress," *British Journal of Pharmacology*, vol. 164, no. 3, pp. 866–883, 2011.
- [12] R.-N. Zhang, Q. Pan, R.-D. Zheng et al., "Genome-wide analysis of DNA methylation in human peripheral leukocytes identifies potential biomarkers of nonalcoholic fatty liver disease," *International Journal of Molecular Medicine*, vol. 42, no. 1, pp. 443–452, 2018.
- [13] K. Bedard and K.-H. Krause, "The NOX family of ROSgenerating NADPH oxidases: physiology and pathophysiology," *Physiological Reviews*, vol. 87, no. 1, pp. 245–313, 2007.
- [14] Y. Xie, S. Zhu, X. Song et al., "The Tumor Suppressor p53 Limits Ferroptosis by Blocking DPP4 Activity," *Cell Reports*, vol. 20, no. 7, pp. 1692–1704, 2017.
- [15] W.-H. Yang, Z. Huang, J. Wu, C. K. Ding, S. K. Murphy, and J. T. Chi, "A TAZ-ANGPTL4-NOX2 axis regulates ferroptotic cell death and chemoresistance in epithelial ovarian cancer," *Molecular Cancer Research*, vol. 18, no. 1, pp. 79–90, 2020.
- [16] W.-H. Yang, C.-K. C. Ding, T. Sun et al., "The Hippo pathway effector TAZ regulates ferroptosis in renal cell carcinoma," *Cell Reports*, vol. 28, no. 10, pp. 2501–2508.e4, 2019.
- [17] M. Skonieczna, A. Cieslar-Pobuda, Y. Saenko et al., "The impact of DIDS-induced inhibition of voltage-dependent anion channels (VDAC) on cellular response of lymphoblastoid cells to ionizing radiation," *Medicinal Chemistry*, vol. 13, no. 5, pp. 477–483, 2017.
- [18] T. Finkel, "Signal transduction by reactive oxygen species," *The Journal of Cell Biology*, vol. 194, no. 1, pp. 7–15, 2011.
- [19] Y. Xie, W. Hou, X. Song et al., "Ferroptosis: process and function," *Cell Death and Differentiation*, vol. 23, no. 3, pp. 369– 379, 2016.

- [20] M. Gao, P. Monian, Q. Pan, W. Zhang, J. Xiang, and X. Jiang, "Ferroptosis is an autophagic cell death process," *Cell Research*, vol. 26, no. 9, pp. 1021–1032, 2016.
- [21] E. Park and S. W. Chung, "ROS-mediated autophagy increases intracellular iron levels and ferroptosis by ferritin and transferrin receptor regulation," *Cell Death & Disease*, vol. 10, no. 11, p. 822, 2019.
- [22] S. Ma, R. F. Dielschneider, E. S. Henson et al., "Ferroptosis and autophagy induced cell death occur independently after siramesine and lapatinib treatment in breast cancer cells," *PLoS One*, vol. 12, no. 8, article e0182921, 2017.
- [23] C.-H. Lin, P.-P. Lin, C.-Y. Lin et al., "Decreased mRNA expression for the two subunits of system x_c⁻, *SLC3A2* and *SLC7A11*, in WBC in patients with schizophrenia: Evidence in support of the hypo- glutamatergic hypothesis of schizophrenia," *Journal of Psychiatric Research*, vol. 72, pp. 58–63, 2016.
- [24] M. Maiorino, M. Conrad, and F. Ursini, "GPx4, lipid peroxidation, and cell death: discoveries, rediscoveries, and open issues," *Antioxidants & Redox Signaling*, vol. 29, no. 1, pp. 61–74, 2018.
- [25] Y. Zhang, H. Tan, J. D. Daniels et al., "Imidazole ketone erastin induces ferroptosis and slows tumor growth in a mouse lymphoma model," *Cell Chemical Biology*, vol. 26, no. 5, pp. 623–633.e9, 2019.
- [26] L. Jiang, N. Kon, T. Li et al., "Ferroptosis as a p53-mediated activity during tumour suppression," *Nature*, vol. 520, no. 7545, pp. 57–62, 2015.
- [27] A. Tarangelo, L. Magtanong, K. T. Bieging-Rolett et al., "P53 suppresses metabolic stress-induced ferroptosis in cancer cells," *Cell Reports*, vol. 22, no. 3, pp. 569–575, 2018.
- [28] J. P. Thomas, P. G. Geiger, M. Maiorino, F. Ursini, and A. W. Girotti, "Enzymatic reduction of phospholipid and cholesterol hydroperoxides in artificial bilayers and lipoproteins," *Biochimica et Biophysica Acta*, vol. 1045, no. 3, pp. 252–260, 1990.
- [29] W. S. Yang, R. SriRamaratnam, M. E. Welsch et al., "Regulation of ferroptotic cancer cell death by GPX4," *Cell*, vol. 156, no. 1–2, pp. 317–331, 2014.
- [30] "Characterization of mammalian selenoproteomes -PubMed," August 2021, https://pubmed.ncbi.nlm.nih.gov/ 12775843/.
- [31] G. J. Warner, M. J. Berry, M. E. Moustafa, B. A. Carlson, D. L. Hatfield, and J. R. Faust, "Inhibition of Selenoprotein Synthesis by Selenocysteine tRNA^{[Ser]Sec} Lacking Isopentenyladenosine," *The Journal of Biological Chemistry*, vol. 275, no. 36, pp. 28110–28119, 2000.
- [32] D. J. R. Lane, A. M. Merlot, M. L.-H. Huang et al., "Cellular iron uptake, trafficking and metabolism: key molecules and mechanisms and their roles in disease," *Biochimica et Biophysica Acta (BBA) - Molecular Cell Research*, vol. 1853, no. 5, pp. 1130–1144, 2015.
- [33] T. Rui, H. Wang, Q. Li et al., "Deletion of ferritin H in neurons counteracts the protective effect of melatonin against traumatic brain injury-induced ferroptosis," *Journal of Pineal Research*, vol. 70, no. 2, article e12704, 2021.
- [34] A. R. Bogdan, M. Miyazawa, K. Hashimoto, and Y. Tsuji, "Regulators of iron homeostasis: new players in metabolism, cell death, and disease," *Trends in Biochemical Sciences*, vol. 41, no. 3, pp. 274–286, 2016.
- [35] Y. Yu, L. Jiang, H. Wang et al., "Hepatic transferrin plays a role in systemic iron homeostasis and liver ferroptosis," *Blood*, vol. 136, no. 6, pp. 726–739, 2020.

- [36] J. Yin, N. Hu, L. Yi et al., "Identification of ferroptosis biomarker in AHH-1 lymphocytes associated with low dose radiation," *Health Physics*, vol. 120, no. 5, pp. 541–551, 2021.
- [37] H. Zhu, A. Santo, Z. Jia, and Y. Robert Li, "GPx4 in bacterial infection and polymicrobial sepsis: involvement of ferroptosis and pyroptosis," *Reactive Oxygen Species*, vol. 7, no. 21, pp. 154–160, 2019.
- [38] H. Mao, Y. Zhao, H. Li, and L. Lei, "Ferroptosis as an emerging target in inflammatory diseases," *Progress in Biophysics* and Molecular Biology, vol. 155, pp. 20–28, 2020.
- [39] M. Herscovitch, W. Comb, T. Ennis et al., "Intermolecular disulfide bond formation in the NEMO dimer requires Cys54 and Cys347," *Biochemical and Biophysical Research Communications*, vol. 367, no. 1, pp. 103–108, 2008.
- [40] "Crosstalk of reactive oxygen species and NF-κB signaling -PubMed," August 2021, https://pubmed.ncbi.nlm.nih.gov/ 21187859/.
- [41] S. S. Singh, S. N. Rai, H. Birla, W. Zahra, A. S. Rathore, and S. P. Singh, "NF-κB-mediated neuroinflammation in Parkinson's disease and potential therapeutic effect of polyphenols," *Neurotoxicity Research*, vol. 37, no. 3, pp. 491– 507, 2020.
- [42] C. Wang, W. Yuan, A. Hu et al., "Dexmedetomidine alleviated sepsis-induced myocardial ferroptosis and septic heart injury," *Molecular Medicine Reports*, vol. 22, no. 1, pp. 175– 184, 2020.
- [43] W. S. Yang, K. J. Kim, M. M. Gaschler, M. Patel, M. S. Shchepinov, and B. R. Stockwell, "Peroxidation of polyunsaturated fatty acids by lipoxygenases drives ferroptosis," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 113, no. 34, pp. E4966–E4975, 2016.
- [44] F. Ito, Y. Sono, and T. Ito, "Measurement and clinical significance of lipid peroxidation as a biomarker of oxidative stress: oxidative stress in diabetes, atherosclerosis, and chronic inflammation," *Antioxidants*, vol. 8, no. 3, p. 72, 2019.
- [45] K. S. Fritz and D. R. Petersen, "An overview of the chemistry and biology of reactive aldehydes," *Free Radical Biology & Medicine*, vol. 59, pp. 85–91, 2013.
- [46] "Arachidonic acid metabolism in brain physiology and pathology: lessons from genetically altered mouse models -PubMed," August 2021, https://pubmed.ncbi.nlm.nih.gov/ 17403135/.
- [47] S. Nogawa, F. Zhang, M. E. Ross, and C. Iadecola, "Cyclooxygenase-2 gene expression in neurons contributes to ischemic brain damage," *The Journal of Neuroscience*, vol. 17, no. 8, pp. 2746–2755, 1997.
- [48] Y.-G. Park, S.-K. Kang, S.-H. Noh et al., "PGE₂ induces IL-1β gene expression in mouse osteoblasts through a cAMP-PKA signaling pathway," *International Immunopharmacology*, vol. 4, no. 6, pp. 779–789, 2004.
- [49] "Cyclooxygenase-2 regulates NLRP3 inflammasome-derived IL-1β production - PubMed," August 2021, https://pubmed .ncbi.nlm.nih.gov/25294243/.
- [50] "Involvement of 15-lipoxygenase in the inflammatory arthritis - PubMed," August 2021, https://pubmed.ncbi.nlm.nih .gov/22573549/.
- [51] C. Abrial, S. Grassin-Delyle, H. Salvator, M. Brollo, E. Naline, and P. Devillier, "15-Lipoxygenases regulate the production of chemokines in human lung macrophages," *British Journal* of *Pharmacology*, vol. 172, no. 17, pp. 4319–4330, 2015.

- [52] Q. Yang and J. Zhou, "Neuroinflammation in the central nervous system: symphony of glial cells," *Glia*, vol. 67, no. 6, pp. 1017–1035, 2019.
- [53] E. H. Kim, S.-W. Wong, and J. Martinez, "Programmed necrosis and disease: we interrupt your regular programming to bring you necroinflammation," *Cell Death & Differentiation*, vol. 26, no. 1, pp. 25–40, 2019.
- [54] B. Proneth and M. Conrad, "Ferroptosis and necroinflammation, a yet poorly explored link," *Cell Death & Differentiation*, vol. 26, no. 1, pp. 14–24, 2019.
- [55] G. W. Konat, T. Kielian, and I. Marriott, "The role of Toll-like receptors in CNS response to microbial challenge," *Journal of Neurochemistry*, vol. 99, no. 1, pp. 1–12, 2006.
- [56] S. Lehnardt, "Innate immunity and neuroinflammation in the CNS: the role of microglia in Toll-like receptor-mediated neuronal injury," *Glia*, vol. 58, no. 3, pp. 253–263, 2009.
- [57] J. Minkiewicz, J. P. de Rivero Vaccari, and R. W. Keane, "Human astrocytes express a novel NLRP2 inflammasome: the NLRP2 inflammasome activated by ATP," *Glia*, vol. 61, no. 7, pp. 1113–1121, 2013.
- [58] C. S. Jack, N. Arbour, J. Manusow et al., "TLR signaling tailors innate immune responses in human microglia and astrocytes," *Journal of Immunology*, vol. 175, no. 7, pp. 4320– 4330, 2005.
- [59] M. Sarhan, W. G. Land, W. Tonnus, C. P. Hugo, and A. Linkermann, "Origin and consequences of necroinflammation," *Physiological Reviews*, vol. 98, no. 2, pp. 727–780, 2018.
- [60] J. Li, F. Cao, H. Yin et al., "Ferroptosis: past, present and future," Cell Death & Disease, vol. 11, no. 2, p. 88, 2020.
- [61] L. Jiang, J. H. Hickman, S.-J. Wang, and W. Gu, "Dynamic roles of p53-mediated metabolic activities in ROS-induced stress responses," *Cell Cycle*, vol. 14, no. 18, pp. 2881–2885, 2015.
- [62] R. Kang, G. Kroemer, and D. Tang, "The tumor suppressor protein p53 and the ferroptosis network," *Free Radical Biology & Medicine*, vol. 133, pp. 162–168, 2019.
- [63] T. Thomas and T. J. Thomas, "Polyamine metabolism and cancer," *Journal of Cellular and Molecular Medicine*, vol. 7, no. 2, pp. 113–126, 2003.
- [64] S. Mandal, A. Mandal, and M. H. Park, "Depletion of the polyamines spermidine and spermine by overexpression of spermidine/spermine N¹-acetyltransferase 1 (SAT1) leads to mitochondria-mediated apoptosis in mammalian cells," *The Biochemical Journal*, vol. 468, no. 3, pp. 435–447, 2015.
- [65] Y. Ou, S.-J. Wang, D. Li, B. Chu, and W. Gu, "Activation of SAT1 engages polyamine metabolism with p53-mediated ferroptotic responses," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 113, no. 44, pp. E6806–E6812, 2016.
- [66] R. U. Macías-Rodríguez, M. E. Inzaugarat, A. Ruiz-Margáin, L. J. Nelson, C. Trautwein, and F. J. Cubero, "Reclassifying hepatic cell death during liver damage: ferroptosis—a novel form of non-apoptotic cell death?," *International Journal of Molecular Sciences*, vol. 21, no. 5, article 1651, 2020.
- [67] A. Franceschini, M. Capece, P. Chiozzi et al., "The P2X7 receptor directly interacts with the NLRP3 inflammasome scaffold protein," *The FASEB Journal*, vol. 29, no. 6, pp. 2450–2461, 2015.
- [68] A. Gustin, M. Kirchmeyer, E. Koncina et al., "NLRP3 inflammasome is expressed and functional in mouse brain

microglia but not in astrocytes," *PLoS One*, vol. 10, no. 6, article e0130624, 2015.

- [69] C. Agulhon, M.-Y. Sun, T. Murphy, T. Myers, K. Lauderdale, and T. A. Fiacco, "Calcium signaling and gliotransmission in normal vs. reactive astrocytes," *Frontiers in Pharmacology*, vol. 3, p. 139, 2012.
- [70] P. R. Angelova, M. L. Choi, A. V. Berezhnov et al., "Alpha synuclein aggregation drives ferroptosis: an interplay of iron, calcium and lipid peroxidation," *Cell Death & Differentiation*, vol. 27, no. 10, pp. 2781–2796, 2020.
- [71] M. A. Smith, G. Perry, and W. A. Pryor, "Causes and consequences of oxidative stress in Alzheimer's disease1, 2," *Free Radical Biology & Medicine*, vol. 32, no. 11, p. 1049, 2002.
- [72] W. R. Markesbery and J. M. Carney, "Oxidative alterations in Alzheimer's disease," *Brain Pathology*, vol. 9, no. 1, pp. 133– 146, 1999.
- [73] B. Hallgren and P. Sourander, "The non-haemin iron in the cerebral cortex in Alzheimer's disease," *Journal of Neurochemistry*, vol. 5, pp. 307–310, 1960.
- [74] D. J. Dedman, A. Treffry, J. M. Candy et al., "Iron and aluminium in relation to brain ferritin in normal individuals and Alzheimer's-disease and chronic renal-dialysis patients," *The Biochemical Journal*, vol. 287, no. 2, pp. 509–514, 1992.
- [75] H. Svobodová, D. Kosnáč, Z. Balázsiová et al., "Elevated agerelated cortical iron, ferritin and amyloid plaques in APP(swe)/PS1(deltaE9) transgenic mouse model of Alzheimer's disease," *Physiological Research*, vol. 68, Suppl 4, pp. S445– S451, 2019.
- [76] Z.-Y. Zhu, Y.-D. Liu, Y. Gong et al., "Mitochondrial aldehyde dehydrogenase (ALDH2) rescues cardiac contractile dysfunction in an APP/PS1 murine model of Alzheimer's disease via inhibition of ACSL4-dependent ferroptosis," *Acta Pharmacol Sin*, 2021.
- [77] D. G. Smith, R. Cappai, and K. J. Barnham, "The redox chemistry of the Alzheimer's disease amyloid beta peptide," *Biochimica et Biophysica Acta*, vol. 1768, no. 8, pp. 1976–1990, 2007.
- [78] A. Tsatsanis, B. X. Wong, A. P. Gunn et al., "Amyloidogenic processing of Alzheimer's disease β-amyloid precursor protein induces cellular iron retention," *Molecular Psychiatry*, vol. 25, no. 9, pp. 1958–1966, 2020.
- [79] W. S. Hambright, R. S. Fonseca, L. Chen, R. Na, and Q. Ran, "Ablation of ferroptosis regulator glutathione peroxidase 4 in forebrain neurons promotes cognitive impairment and neurodegeneration," *Redox Biology*, vol. 12, pp. 8–17, 2017.
- [80] W.-D. Bao, P. Pang, X.-T. Zhou et al., "Loss of ferroportin induces memory impairment by promoting ferroptosis in Alzheimer's disease," *Cell Death & Differentiation*, vol. 28, no. 5, pp. 1548–1562, 2021.
- [81] V. Calsolaro and P. Edison, "Neuroinflammation in Alzheimer's disease: current evidence and future directions," *Alzheimers Dement*, vol. 12, no. 6, pp. 719–732, 2016.
- [82] M. T. Heneka, M. J. Carson, J. El Khoury et al., "Neuroinflammation in Alzheimer's disease," *The Lancet Neurology*, vol. 14, no. 4, pp. 388–405, 2015.
- [83] M. Obulesu and M. Jhansilakshmi, "Neuroinflammation in Alzheimer's disease: an understanding of physiology and pathology," *The International Journal of Neuroscience*, vol. 124, no. 4, pp. 227–235, 2014.
- [84] T. Jonsson, H. Stefansson, S. Steinberg et al., "Variant of TREM2 associated with the risk of Alzheimer's disease,"

The New England Journal of Medicine, vol. 368, no. 2, pp. 107–116, 2013.

- [85] T. T. Rohn, "The Triggering Receptor Expressed on Myeloid Cells 2: "TREM-ming" the Inflammatory Component Associated with Alzheimer's Disease," Oxidative Medicine and Cellular Longevity, vol. 2013, Article ID 860959, 8 pages, 2013.
- [86] D. J. DiSabato, N. Quan, and J. P. Godbout, "Neuroinflammation: the devil is in the details," *Journal of Neurochemistry*, vol. 139, pp. 136–153, 2016.
- [87] M. T. Heneka, M. P. Kummer, and E. Latz, "Innate immune activation in neurodegenerative disease," *Nature Reviews Immunology*, vol. 14, no. 7, pp. 463–477, 2014.
- [88] F. Leng and P. Edison, "Neuroinflammation and microglial activation in Alzheimer disease: where do we go from here?," *Nature Reviews Neurology*, vol. 17, no. 3, pp. 157–172, 2021.
- [89] G. D. Femminella, S. Ninan, R. Atkinson, Z. Fan, D. J. Brooks, and P. Edison, "Does microglial activation influence hippocampal volume and neuronal function in Alzheimer's disease and Parkinson's disease dementia?," *Journal of Alzheimer's Disease*, vol. 51, no. 4, pp. 1275–1289, 2016.
- [90] E. E. Spangenberg and K. N. Green, "Inflammation in Alzheimer's disease: lessons learned from microglia-depletion models," *Brain, Behavior, and Immunity*, vol. 61, pp. 1–11, 2017.
- [91] J. M. Graham, M. N. Paley, R. A. Grünewald, N. Hoggard, and P. D. Griffiths, "Brain iron deposition in Parkinson's disease imaged using the PRIME magnetic resonance sequence," *Brain*, vol. 123, no. 12, pp. 2423–2431, 2000.
- [92] E. C. Hirsch and S. Hunot, "Neuroinflammation in Parkinson's disease: a target for neuroprotection?," *The Lancet Neurology*, vol. 8, no. 4, pp. 382–397, 2009.
- [93] P. L. McGeer, S. Itagaki, B. E. Boyes, and E. G. McGeer, "Reactive microglia are positive for HLA-DR in the substantia nigra of Parkinson's and Alzheimer's disease brains," *Neurology*, vol. 38, no. 8, pp. 1285–1291, 1988.
- [94] E. Bertrand, W. Lechowicz, G. M. Szpak, E. Lewandowska, A. Członkowska, and J. Dymecki, "Quantitative study of pathological forms of astroglia in Wilson's disease," *Folia Neuropathologica*, vol. 35, no. 4, pp. 227–232, 1997.
- [95] P. Damier, E. C. Hirsch, P. Zhang, Y. Agid, and F. Javoy-Agid, "Glutathione peroxidase, glial cells and Parkinson's disease," *Neuroscience*, vol. 52, no. 1, pp. 1–6, 1993.
- [96] M. Mogi, M. Harada, P. Riederer, H. Narabayashi, K. Fujita, and T. Nagatsu, "Tumor necrosis factor- α (TNF- α) increases both in the brain and in the cerebrospinal fluid from parkinsonian patients," *Neuroscience Letters*, vol. 165, no. 1–2, pp. 208–210, 1994.
- [97] S. Hands, M. U. Sajjad, M. J. Newton, and A. Wyttenbach, "In Vitro and in Vivo Aggregation of a Fragment of Huntingtin Protein Directly Causes Free Radical Production," The Journal of Biological Chemistry, vol. 286, no. 52, pp. 44512– 44520, 2011.
- [98] S.-Y. Chou, J.-Y. Weng, H.-L. Lai et al., "Expanded-polyglutamine huntingtin protein suppresses the secretion and production of a chemokine (CCL5/RANTES) by astrocytes," *The Journal of Neuroscience*, vol. 28, no. 13, pp. 3277–3290, 2008.
- [99] T. L. Spires, H. E. Grote, S. Garry et al., "Dendritic spine pathology and deficits in experience-dependent dendritic plasticity in R6/1 Huntington's disease transgenic mice,"

The European Journal of Neuroscience, vol. 19, no. 10, pp. 2799–2807, 2004.

- [100] M. A. Ariano, C. Cepeda, C. R. Calvert et al., "Striatal potassium channel dysfunction in Huntington's disease transgenic mice," *Journal of Neurophysiology*, vol. 93, no. 5, pp. 2565– 2574, 2005.
- [101] G. J. Hankey, "Stroke," *The Lancet*, vol. 389, no. 10069, pp. 641–654, 2017.
- [102] C. Iadecola and J. Anrather, "The immunology of stroke: from mechanisms to translation," *Nature Medicine*, vol. 17, no. 7, pp. 796–808, 2011.
- [103] S. Ahmad, N. M. Elsherbiny, R. Haque et al., "Sesamin attenuates neurotoxicity in mouse model of ischemic brain stroke," *Neurotoxicology*, vol. 45, pp. 100–110, 2014.
- [104] Q. Tuo, P. Lei, K. A. Jackman et al., "Tau-mediated iron export prevents ferroptotic damage after ischemic stroke," *Molecular Psychiatry*, vol. 22, no. 11, pp. 1520–1530, 2017.
- [105] I. Alim, J. T. Caulfield, Y. Chen et al., "Selenium drives a transcriptional adaptive program to block ferroptosis and treat stroke," *Cell*, vol. 177, no. 5, pp. 1262–1279.e25, 2019.
- [106] E. D. Ponomarev, T. Veremeyko, and H. L. Weiner, "Micro-RNAs are universal regulators of differentiation, activation, and polarization of microglia and macrophages in normal and diseased CNS," *Glia*, vol. 61, no. 1, pp. 91–103, 2013.
- [107] E. Parada, J. Egea, I. Buendia et al., "The microglial α7-acetylcholine nicotinic receptor is a key element in promoting neuroprotection by inducing heme oxygenase-1 via nuclear factor erythroid-2-related factor 2," *Antioxidants & Redox Signaling*, vol. 19, no. 11, pp. 1135–1148, 2013.
- [108] S. S. Karuppagounder, L. Alin, Y. Chen et al., "N-Acetylcysteine targets 5 lipoxygenase-derived, toxic lipids and can synergize with prostaglandin E2to inhibit ferroptosis and improve outcomes following hemorrhagic stroke in mice," *Annals of Neurology*, vol. 84, no. 6, pp. 854–872, 2018.
- [109] W.-N. Jin, S. X.-Y. Shi, Z. Li, K. Wood, R. J. Gonzales, and Q. Liu, "Depletion of microglia exacerbates postischemic inflammation and brain injury," *Journal of Cerebral Blood Flow and Metabolism*, vol. 37, no. 6, pp. 2224–2236, 2017.