Research Article

Prolonged Diuretic and Renoprotective Effects of a Xanthone Obtained from *Garcinia achachairu* Rusby in Normotensive and Hypertensive Rats

Luísa Nathália Bolda Mariano ,¹ Thaise Boeing ,¹ Valdir Cechinel Filho,¹ Rivaldo Niero,¹ Arquimedes Gasparotto Junior ,² Luisa Mota da Silva,¹ and Priscila de Souza ,¹

¹Programa de Pós-graduação em Ciências Farmacêuticas (PPGCF), Núcleo de Investigações Químico-Farmacêuticas (NIQFAR),

Universidade do Vale do Itajaí (UNIVALI), Rua Uruguai, 458, Centro, 88302-901, Itajaí, Brazil

²Laboratório de Farmacologia Cardiovascular (LaFaC), Faculdade de Ciências da Saúde, Universidade Federal da Grande Dourados, R. João Rosa Góes, 1761, CEP 79825-070, Dourados, Brazil

Correspondence should be addressed to Luísa Nathália Bolda Mariano; luboldamariano@gmail.com

Received 25 January 2021; Revised 28 March 2021; Accepted 30 March 2021; Published 21 April 2021

Academic Editor: Daniel Dias Rufino Arcanjo

Copyright © 2021 Luísa Nathália Bolda Mariano et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The previous study showed that 1,5,8-trihydroxy-4',5'-dimethyl-2H-pyrano(2,3:3,2)-4-(3-methylbut-2-enyl) xanthone (TDP) obtained from *Garcinia achachairu* Rusby (Clusiaceae) branches induces acute diuresis in normotensive (NTR) and spontaneously hypertensive rats (SHR) after 8 h of the experiment. In complementarity, the present study evaluated the prolonged diuretic and renoprotective effects of TDP in both NTR and SHR. The animals received, once a day, oral treatment with TDP (0.1 mg/kg), hydrochlorothiazide (10 mg/kg), or vehicle (VEH; 10 mL/kg). At the end of 7 days, the urine, blood, and kidney samples were collected for biochemical and histological analyzes. The urinary volume of both NTR and SHR after 7 days of treatment with the TDP was significantly increased, associated with augmented urinary electrolyte excretion levels. The treatments did not modify the urinary pH values nor the parameters analyzed in plasma (Na⁺, K⁺, Cl⁻, and Ca²⁺). Concerning the renal analyzes, when compared with the VEH-treated NTR group, while the activity of the enzymes catalase (CAT) and N-acetyl- β -D-glucosaminidase (NAG), as well as nitrite levels, were increased, the generation of lipid hydroperoxides and the activity of the enzyme myeloperoxidase (MPO) were unaltered. On the other hand, the activities of superoxide dismutase (SOD) and glutathione *S*-transferase (GST) and the levels of reduced glutathione (GSH) in kidney homogenates of the SHR group were decreased. However, TDP augmented the levels of GSH and GST activities and reduced the levels of nitrite and the activities of CAT and MPO, when compared with VEH-treated only SHR. Besides, the treatment with TDP alleviated the morphological changes of the renal corpuscle region of SHR. Together, these results revealed the prolonged diuretic effect of TDP and their renal protective effect by improving the antioxidative capacity.

1. Introduction

Diuretic drugs are used to treat cardiovascular and kidney disorders [1, 2]. Diuretics' main action is to increase the concentration of Na^+ and water in the renal tubules [1, 3], resulting in diuresis [4]. There are five diuretic classes, and each type differs in efficacy, mechanism of action, and location of effects in the nephron [5], so it is common to associate different categories [6]. Diuretic classes commonly

used as antihypertensives are loop diuretics, thiazide-type diuretics, and potassium-sparing agents [7].

Despite the variety of diuretics, their use is associated with the risk of developing several adverse effects, such as electrolyte disturbances, ventricular arrhythmias [8], increased risk of acquiring on-set diabetes [9], sexual dysfunction, gynecomastia [3], and ototoxicity [2]. As a result, medicinal plants and isolated compounds have been widely studied as possible alternatives to complement current therapy or reveal potential new study molecules. Some medicinal plants that are already used in folk medicine for the treatment of cardiovascular and renal disorders have scientific studies proving their diuretic effects, such as *Tropaeolum majus* L. [10], *Achillea millefolium* L. [11], *Maytenus ilicifolia* Mart ex Reissek [12], *Scutia buxifolia* Reissek [13], *Echinodorus grandiflorus* (Cham & Schltdl) [14], *Bauhinia forficata* Link [15], and *Leandra dasytricha* (A. Gray) Cong. [16].

Recently, we described the diuretic effect of the methanolic extract and fractions obtained of branches from *G. achachairu* Rusby (Clusiaceae) [17, 18], which is a plant native of Bolivia, popularly known as "achachairu" [19]. Phytochemical study permitted to isolate the 1, 5,8-trihydroxy-4',5'-dimethyl-2H-pyrano (2,3:3,2)-4-(3-methylbut-2-enyl) xanthone (TDP), which showed a significant diuretic effect in rats after a single-dose treatment [17, 18]. Xanthones are polyphenolic compounds, and their structures are recognized for their importance in discovering new active compounds [20]. Indeed, several biological activities have been described for xanthones, including anti-inflammatory, antioxidant [21], cardioprotective [22], diuretic [17, 23], and renal protective [24] effects.

Given the above and considering that TDP has already shown an acute diuretic action in our previous publication, this study aimed to investigate the diuretic and renoprotective effects of TDP after a dose-repeated treatment in both normotensive and hypertensive rats.

2. Materials and Methods

2.1. Xanthone Isolation. Details of the isolation of 1,5,8-trihydroxy-4',5'-dimethyl-2H-pyrano(2,3:3,2)-4-(3-meth-ylbut-2-enyl) xanthone (TDP) (Figure 1) obtained from branches of *G. achachairu* are described by Mariano et al. [25].

2.2. Animals. Female Wistar normotensive (NTR) and spontaneously hypertensive rats (SHR) of 3-4 months old were used in this study. The animals were provided by Universidade do Vale do Itajaí (UNIVALI) and were maintained in a controlled laboratory environment (12 h light/dark cycle and $22 \pm 2^{\circ}$ C), with free access to food and water. All methodologies were approved by the Ethical Committee for the Care and Use of animals of UNIVALI (authorization 028/17).

2.3. Prolonged Diuretic Activity Assay. The rats were randomly distributed into groups of 6–8 animals, and each group was treated daily, with vehicle (VEH; water plus 1% tween; 10 mL/kg, p.o), hydrochlorothiazide (HCTZ; 10 mg/ kg, p.o), or TDP compound (0.1 mg/kg, p.o). The animals were individually allocated in metabolic cages, the urine was collected, and the volume was recorded every day for 7 days. The cumulative urine volume was calculated to bodyweight and expressed as mL/100 g. At the end of the experiment, the organs (heart, kidney, liver, lung, and spleen) were removed and weighed. Blood samples were collected for biochemical



FIGURE 1: Molecular structure of the 1,5,8-trihydroxy-4',5'-dimethyl-2H-pyrano(2,3:3,2)-4-(3-methylbut-2-enyl) xanthone (TDP).

analysis. Besides, renal tissue samples were collected for the assessment of the tissue redox state and histological analysis.

2.4. Biochemical and Tissue Redox State Evaluation. Blood samples were examined for the content of electrolytes, creatinine, urea, aspartate aminotransferase (AST), alanine aminotransferase (ALT), and nitrite [17, 26]. The urine samples were analyzed for electrolytes excretion (Na⁺, K⁺, Ca²⁺, and Cl⁻), pH, and osmolality. Osmolality was calculated using the formula described by Bhasin and Velez, 2016 [27].

The oxidative stress analysis followed the methodologies detailed and described by De Almeida et al. [28]. Briefly, the renal tissue was homogenized in potassium phosphate buffer (200 mM with pH 6.5; 1:3 weight/volume). This homogenate was used to measure the levels of reduced glutathione (GSH) and lipid hydroperoxides (LOOH). Posteriorly, the homogenate was centrifuged (20 min at 9000 g). The supernatant was used to determine the activity of the enzymes superoxide dismutase (SOD), catalase (CAT), and glutathione *S*-transferase (GST) [28]. The pellet was used to determine the activity of the enzymes (MPO) [28] and N-acetyl- β -D-glucosaminidase (NAG) [29].

2.5. *Histological Analysis.* Kidney tissue was fixed in ALFAC solution (85% ethanol, 10% formaldehyde, and 5% acetic acid), and after 24 h, this tissue was dehydrated (alcohol and xylene), embedded in paraffin, and stained with hematoxylin/eosin (H&E). The material was examined using a stereo microscope with a magnification of 40x.

2.6. Statistical Analysis. The program GraphPad Prism version 7.00 for Mac (GraphPad Software, La Jolla, CA, USA) was used for the statistical analysis. The results were expressed as mean \pm standard error of the mean (S.E.M.) of 6–8 animals per group. The differences between means were determined by one-or two-way analysis of variance (ANOVA) followed by Dunnett's multiple comparisons test. The value was considered statistically significant when the value of p is less than 0.05.

Groups	Urine volume (mL/100g)	рН	Osmolarity (mOsm/kg H ₂ O)	Na ⁺ (mmol/L)	K ⁺ (mmol/L)	Cl⁻ (mmol/L)	Ca ²⁺ (mg/dL)
NTR							
VEH (10 mL/kg)	9.75 ± 0.77	8.39 ± 0.25	362.0 ± 13.06	129.9 ± 8.15	44.57 ± 3.04	335.3 ± 3.45	14.26 ± 0.53
HCTZ (10 mg/kg)	$23.13 \pm 0.44^{*}$	8.69 ± 0.08	$877.4 \pm 53.51^*$	$306.3 \pm 13.86^*$	$98.09 \pm 4.59^*$	$360.0 \pm 2.77^*$	$6.26 \pm 0.90^{*}$
TDP (0.1 mg/kg)	$14.91 \pm 0.69^*$	8.75 ± 0.06	$520.2 \pm 24.66^*$	$172.7 \pm 5.51^*$	$65.49 \pm 3.29^*$	324.2 ± 7.92	$10.33 \pm 0.55^{*}$
SHR							
VEH (10 mL/kg)	9.44 ± 0.73	8.58 ± 0.15	399.5 ± 25.05	122.1 ± 8.31	40.45 ± 3.43	316.1 ± 6.61	8.39 ± 0.95
HCTZ (10 mg/kg)	$17.52 \pm 1.24^*$	8.73 ± 0.02	$747.8 \pm 31.76^*$	$272.4 \pm 12.91^*$	$76.08 \pm 8.43^{*}$	$358.0 \pm 7.66^{*}$	$3.67 \pm 0.35^{*}$
TDP (0.1 mg/kg)	$15.66 \pm 0.91^{*}$	8.61 ± 0.17	$507.1 \pm 41.52^*$	$171.3 \pm 12.93^*$	46.74 ± 3.73	317.0 ± 7.13	$5.33 \pm 0.80^{*}$

TABLE 1: Urinary parameters were measured after 7 days of treatment with TDP in normotensive and hypertensive rats.

The results show the mean \pm S.E.M. of 6–8 animals per group. Statistical analysis was performed using one-way analysis of variance (ANOVA) followed by Dunnett's multiple comparisons test. *p < 0.05 when compared with the respective VEH group. NTR, normotensive rats; SHR, spontaneously hypertensive rats; VEH, vehicle (water plus 1% tween), HCTZ, hydrochlorothiazide, and TDP, 1,5,8-trihydroxy-4',5'-dimethyl-2H-pyrano(2,3:3, 2)-4-(3-methylbut-2-enyl) xanthone.

3. Results and Discussion

Bolda Mariano et al. [18] described the acute diuretic effect of the xanthone TDP (Figure 1), in which a single oral dose of 0.1 mg/kg was able to induce diuresis and increase of the urinary electrolytes excretion in both normotensive (NTR) and hypertensive (SHR) rats. Herein, to better understand the renal effects of TDP, especially in a dose-repeated treatment, the present study was performed.

The first set of results showed that the TDP (0.1 mg/kg) increased the urine volume of both NTR and SHR compared to VEH (Table 1). The treatment with HCTZ (10 mg/kg), a thiazide-type diuretic [30], as expected, was effective in the VEH group. The treatment with HCTZ increased the renal excretion of Na⁺, K⁺, and Cl⁻. The treatment with TDP increased Na⁺ excretion in NTR and SHR, confirming the compound's ability to induce diuresis [1, 3]. However, while TDP increased K⁺ excretion in NTR, the same was not detected when urine samples from SHR were analyzed. Indeed, it is necessary to be careful when using drugs that cause an increase in urinary K⁺ excretion due to the risk of developing hypokalemia, which is very common with the use of K^+ -depleting diuretics [31]. Besides, TDP was not able to increase Cl⁻ excretion neither in NTR nor SHR. HCTZ treatment, as expected, decreased the excretion of Ca^{2+} [31], while the compound TDP was also able to decrease Ca^{2+} levels in the urine. Diuretic drugs that can reduce urinary Ca²⁺ excretion are interesting to be used by patients with osteoporosis [32] or as a prophylactic treatment for kidney stones [33]. Osmolarity values were increased in the treated groups compared to the VEH animals, which was expected, since this parameter reflects the amount of ion excretion in the urine. Finally, the urinary values of pH were unaltered.

Additionally, the results obtained with plasma analysis are shown in Table 2. No statistically significant changes were found in the content of electrolytes (Na⁺, K⁺, Cl⁻ and Ca²⁺), uric acid, creatinine, urea, AST and ALT in the different experimental groups. However, nitrite levels, an indirect marker of NO production [34], were lower in SHR animals treated with VEH than VEH-treated NTR. The treatment with TDP was not able to increase the levels of nitrite in the plasma. This dataset shows that this xanthone probably does not have its diuretic and natriuretic effects related to the nitric oxide (NO) production. However, it is worth mentioning that in our previous study, indomethacin (a cyclooxygenase inhibitor) precluded TDPinduced diuresis [18], so we can suggest that the mechanisms responsible for the effects of xanthone presented here could involve direct vasodilator actions on the renal vascular bed.

The animals' weight, water, and food intake showed no differences during the 7 days of the experiment (data not shown). Besides, the weight of the kidneys and heart differed between the NTR groups and SHR treated with VEH (i.e., reduced kidney weight and increased cardiac weight in the SHR group—data not shown), which was expected, since the characteristics of SHR lineage are associated with ventricular hypertrophy, cardiac hyperplasia, and kidney damage [35]. Moreover, these results have already been described in previous studies [26, 28]. The treatment with HCTZ or TDP did not cause any changes in these tissues, suggesting that 7 days of treatment were not enough to reverse the damage already established by hypertension. Besides, the TDP did not cause any alteration in the weight of the other organs (i.e., liver, lung, and spleen—data not shown).

Oxidative stress and the deficiency of NO may be present in hypertension or renal disease [36], and the class of xanthones is known to have antioxidant effects [21]. For this reason, we investigated the involvement of the antioxidant system in the kidneys and the possible renal protector effect of TDP. First, the levels of lipid hydroperoxides (LOOH) were measured since lipid peroxidation is related to some disorders and can trigger a variety of oxidants, which can lead to cell dysfunction and tissue damage [37], in addition to indirectly indicating the oxidative stress in the tissue [38]; however, there was no difference between the groups (Figure 2(a)). It is believed that when triggering tissue damage, the defense system itself begins to act to repair the injury [39]; however, we cannot rule out the hypothesis of other cell damage that did not cover in this study. Regarding the reduced glutathione (GSH) levels in renal tissue (Figure 2(b)), which is an antioxidant biomolecule abundant in the body [40], in SHR treated with VEH ($635 \pm 35.46 \,\mu\text{g/mg}$ of tissue), GSH levels were reduced by 40.54% compared to NTR treated with VEH $(1068 \pm 54.06 \,\mu\text{g/mg} \text{ of tissue})$, and the treatment with TDP was able to reverse this value, reaching almost baseline levels. In sequence, we analyzed the enzymes SOD, CAT, and GST, as shown in Figures 2(c)-2(e),

		•				-				
Groups	Na ⁺ (mmol/L)	K ⁺ (mmol/L)	Cl ⁻ (mmol/L)	Ca ²⁺ (mg/dL)	Uric acid (mg/dL)	Creatinine (mg/dL)	Urea (mg/dL)	AST (U/L)	ALT (U/L)	Nitrite (μ M)
NTR										
VEH (10 mL/kg)	116.6 ± 9.40	3.09 ± 0.43	224.3 ± 2.79	4.79 ± 0.04	4.61 ± 0.09	0.39 ± 0.01	36.30 ± 2.66	47.63 ± 1.27	46.91 ± 6.16	67.26 ± 3.39
HCTZ (10 mg/kg)	127.5 ± 8.50	4.12 ± 0.46	229.4 ± 4.85	4.36 ± 0.11	4.67 ± 0.17	0.45 ± 0.02	30.45 ± 2.69	45.00 ± 5.10	40.11 ± 3.06	56.48 ± 3.09
TDP (0.1 mg/kg)	117.9 ± 5.88	3.56 ± 0.23	220.8 ± 1.60	4.86 ± 0.16	4.73 ± 0.08	0.38 ± 0.01	35.49 ± 1.39	41.01 ± 1.77	36.60 ± 1.95	60.93 ± 3.62
SHR										
VEH (10 mL/kg)	123.3 ± 3.93	3.21 ± 0.87	225.2 ± 2.66	4.97 ± 0.19	4.52 ± 0.18	0.38 ± 0.01	31.91 ± 1.20	47.38 ± 1.85	46.06 ± 2.79	$54.71 \pm 3.97^{\#}$
HCTZ (10 mg/kg)	113.9 ± 7.88	2.85 ± 0.28	222.7 ± 0.57	4.47 ± 0.25	4.60 ± 0.20	0.36 ± 0.01	37.40 ± 0.78	36.27 ± 1.48	47.81 ± 3.47	62.65 ± 2.99
TDP (0.1 mg/kg)	106.3 ± 2.78	2.15 ± 0.41	227.7 ± 3.58	5.10 ± 0.18	4.60 ± 0.09	0.38 ± 0.01	35.46 ± 1.82	48.45 ± 3.99	40.55 ± 2.60	52.40 ± 3.59
The results show the 1 compared with the res	mean ± S.E.M. of 6- pective VEH group	-8 animals per grc p = 0.05 when c	ompared with the ¹	lysis was performe VEH-treated SHR	d using one-way analys group. NTR, normotens	is of variance (ANOVA) ive rats; SHR. spontaneo	followed by Dunn usly hypertensive r	ett's multiple co ats; VEH, vehicl	mparisons test. e (water plus 1%	* <i>p</i> < 0.05 when tween); HCTZ,
hydrochlorothiazide;	TDP, 1,5,8-trihydr	oxy-4',5'-dimethy	l-2H-pyrano(2,3 : î	3,2)-4-(3-methylbu	ut-2-enyl) xanthone.					

	23
	5
	5
	1
	5
•	Ξ.
	2
	Η.
	Ξ.
	5
	ĕ
	рч
	2
-	9
	5
-	÷.
	22
	≓
	2
	a.
	Ξ.
	2
	È
	Ξ.
	ŏ.
	5
	q
	ц
	a
	•
	ž
	1
	Ś
	ц.
	e
1	Ξ.
	З.
	Ξ.
	Ξ.
	0
	Ξ.
•	-
	S
	≻
	g
1	σ
I	~
I	r /
	or /
۱ ر	tor /
۱ ر	for /
	JP tor 7
	DP for 7
	TUP for 7
	TUP for 7
	h TUP for /
	ith TDP for 7
	with TDP for 7
	with TDP for 7
	it with TDP for 7
	int with TDP for 7
	ent with TDP for 7
	nent with TDP for 7
	tment with TDP for 7
	atment with TDP for 7
	eatment with TDP for 7
	creatment with TDP for 7
	treatment with TDP for 7
	r treatment with TDP for 7
	cer treatment with TDP for 7
	tter treatment with TDP for 7
	after treatment with TDP for 7
	s after treatment with TDP for 7
	rs after treatment with TDP for 7
	ers atter treatment with TDP for 7
	eters after treatment with TDP for 7
	neters after treatment with TDP for 7
	meters after treatment with TDP for 7
	ameters after treatment with TUP for 7
	trameters after treatment with TDP for 7
	barameters after treatment with TDP for 7
	parameters after treatment with TDP for 7
	a parameters after treatment with TDP for /
	na parameters atter treatment with TDP for 7
	ima parameters atter treatment with TDP for 7
	asma parameters after treatment with TDP for 7
	lasma parameters after treatment with LDP for /
	Plasma parameters after treatment with TDP for 7
	: Plasma parameters after treatment with TDP for /
	2: Plasma parameters after treatment with TDP for 7
	3.2: Plasma parameters after treatment with TDP for 7
	JE 2: Plasma parameters after treatment with TDP for /
	3LE 2: Plasma parameters atter treatment with TDP for 7
	(BLE 2: Plasma parameters after treatment with TDP for /
	ABLE 2: Plasma parameters after treatment with IDP for /

4





FIGURE 2: Effect of TDP on renal markers of oxidative stress, endogenous antioxidants factors, and cell biomarkers after 7 days of treatment in rats. (a) Lipid hydroperoxides (LOOH) content, (b) reduced glutathione (GSH) levels, (c) superoxide dismutase (SOD) activity, (d) catalase (CAT) activity, (e) glutathione S-transferase (GST) activity, (f) myeloperoxidase (MPO) activity, (g) N-acetyl- β -Dglucosaminidase (NAG) activity, and (h) nitrite levels in kidney samples collected from normotensive rats (NTR) and spontaneously hypertensive rats (SHR). The results show the mean ± S.E.M. of 6–8 animals per group. Statistical analysis was performed using one-way analysis of variance (ANOVA) followed by Dunnett's multiple comparisons test. *p < 0.05 when compared with the respective VEH group. #p < 0.05 when compared with the VEH-treated SHR group. TDP, 1,5,8-trihydroxy-4',5'-dimethyl-2H-pyrano(2, 3:3, 2)-4-(3-methylbut-2-enyl) xanthone.



FIGURE 3: Representative images of renal tissue morphology stained by hematoxylin and eosin (H&E). (a) Normotensive rats and (b) spontaneously hypertensive rats. The blue and red arrows indicate Bowman's capsule region and the renal glomerulus, respectively. VEH, vehicle (water plus 1% tween). HCTZ, hydrochlorothiazide, and TDP, 1,5,8-trihydroxy-4',5'-dimethyl-2H-pyrano(2, 3:3, 2)-4-(3-methylbut-2-enyl) xanthone.

respectively. The animals of the SHR group treated with VEH showed decreased SOD and GST activities and increased CAT activity when compared to the NTR group treated with VEH. Briefly, these results demonstrated that the antioxidant system defense is lower in SHR animals treated with VEH only. On the other hand, TDP partially restored the antioxidant system (i.e., decreasing CAT activity and increasing GST activity), suggesting a possible antioxidant effect of this compound on renal tissue.

Additionally, we analyzed the effect of TDP on MPO and NAG activities in renal tissue, respectively (Figures 2(f) and 2(g)). The MPO is a biomarker of cell infiltration, mainly neutrophils, and a marker of acute inflammation in the tissue [41]. Likewise, NAG is an indicator of cell infiltration highly specific for macrophages [42]. Unlike neutrophils, macrophages remain in the tissue longer and are among the cells involved in chronic inflammation [43]. In this group of experiments, the results obtained show that the MPO activity did not change between the SHR and NTR groups treated with VEH only (Figure 2(f)). On the other hand, the NAG activity was higher in the SHR group treated with VEH than the NTR group treated with VEH only (Figure 2(g)), suggesting that the disease already well installed in the body; cells of the chronic inflammatory process are generally present. However, TDP treatment did not change NAG activity in renal tissue than the VEH group, although it reduced MPO activity in the SHR group. These data suggest that TDP may have an anti-inflammatory effect related to reducing neutrophil infiltration; however, further experiments are needed to confirm this effect. According to literature data, the class of xanthones can present an antiinflammatory potential [21, 44].

NO is involved in several physiological processes in the kidneys, including diuresis and natriuresis [36]. Our kidney sample results reveal no differences in the nitrite content between the SHR and NTR treated with VEH only, a result similar to that found by Almeida et al. [28]. Interestingly, the SHR groups treated with HCTZ and TDP showed a decreased value of nitrite in the renal tissue (Figure 2(h)), a result that can be directly linked to the local inflammatory response [45].

Finally, the histological results of the kidneys obtained from SHR animals treated with VEH (Figure 3(b)) showed a disruption of the mesangial space, an increase in the glomerular size, and an increase in the thickening of the Bowman capsule when compared with the histology of VEH-treated NTR (Figure 3(a)). The changes found in the SHR group treated with HCTZ and TDP are less evident when compared to the group treated with VEH. Besides, histological analysis of renal tissue did not show any changes in NTR animals treated with HCTZ or TDP in NTR (Figure 3(a)). This result is significant because it shows that after 7 days of treatment, there was no glomerular or tubular damage induced by TDP, supporting the absence of adverse effects.

4. Conclusion

The prolonged treatment with TDP-induced significant diuretic and natriuretic effects, restored the imbalance of the

antioxidant system in the renal tissue, and mitigated hypertensive rats' renal damage, revealing an important renoprotective effect.

Abbreviations

- ALT: Alanine aminotransferase
- AST: Aspartate aminotransferase
- CAT: Catalase
- GSH: Reduced glutathione
- HCTZ: Hydrochlorothiazide
- H & E: Hematoxylin and eosin
- iNOS: Inducible nitric oxide synthase
- LOOH: Lipid hydroperoxides
- MPO: Myeloperoxidase
- NAG: N-acetyl- β -D-glucosaminidase
- NO: Nitric oxide
- NTR: Normotensive rats
- SHR: Spontaneously hypertensive rats
- SOD: Superoxide dismutase
- GST: Glutathione S-transferase
- TDP: 1,5,8-Trihydroxy-4',5'-dimethyl-2H-pyrano(2,3:3,2)-4-(3-methylbut-2-enyl) xanthone
- VEH: Vehicle.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors' Contributions

Priscila de Souza designed the study. Luísa Nathália Bolda Mariano and Thaise Boeing performed the experiments. Luísa Nathália Bolda Mariano, Rivaldo Niero, and Valdir Cechinel Filho performed the phytochemical analysis. Luísa Mota da Silva contributed to the renal analysis. Arquimedes Gasparotto Junior contributed to biochemical analysis. Luísa Nathália Bolda Mariano and Priscila de Souza prepared the article. All authors read and approved the final version of the article.

Acknowledgments

The authors acknowledge the support received from the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES), Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), and Universidade do Vale do Itajaí (UNIVALI).

References

 P. A. Sarafidis, P. I. Georgianos, and A. N. Lasaridis, "Diuretics in clinical practice. Part I: mechanisms of action, pharmacological effects and clinical indications of diuretic compounds," *Expert Opinion on Drug Safety*, vol. 9, no. 2, pp. 243–257, 2010.

Evidence-Based Complementary and Alternative Medicine

- [2] D. Ding, H. Liu, W. Qi et al., "Ototoxic effects and mechanisms of loop diuretics," *Journal of Otology*, vol. 11, no. 4, pp. 145–156, 2016.
- [3] G. C. Roush, R. Kaur, and M. E. Ernst, "Diuretics," *Journal of Cardiovascular Pharmacology and Therapeutics*, vol. 19, no. 1, pp. 5–13, 2014.
- [4] D. Wile, "Diuretics: a review," Annals of Clinical Biochemistry, vol. 49, no. 5, pp. 419–431, 2012.
- [5] D. C. Brater, "Update in diuretic therapy: clinical pharmacology," *Seminars in Nephrology*, vol. 31, no. 6, pp. 483–494, 2011.
- [6] A. H. Qavi, R. Kamal, and R. W. Schrier, "Clinical use of diuretics in heart failure, cirrhosis, and nephrotic syndrome," *International Journal of Nephrology*, vol. 2015, Article ID 975934, 9 pages, 2015.
- [7] D. L. Blowey, "Diuretics in the treatment of hypertension," *Pediatric Nephrology*, vol. 31, no. 12, pp. 2223–2233, 2016.
- [8] G. C. Roush and D. A. Sica, "Diuretics for hypertension: a review and update," *American Journal of Hypertension*, vol. 29, no. 10, pp. 1130–1137, 2016.
- [9] P. Reddy and L. Dupree, "Approach to antihypertensive therapy," *American Journal of Therapeutics*, vol. 23, no. 2, pp. e451–473, 2016.
- [10] A. Gasparotto, M. A. Boffo, E. L. B. Lourenço, M. E. A. Stefanello, C. A. L. Kassuya, and M. C. A. Marques, "Natriuretic and diuretic effects of Tropaeolum majus (Tropaeolaceae) in rats," *Journal of Ethnopharmacology*, vol. 122, no. 3, pp. 517–522, 2009.
- [11] P. De Souza, S. Crestani, R. D. C. V. Da Silva et al., "Involvement of bradykinin and prostaglandins in the diuretic effects of Achillea millefolium L. (Asteraceae)," *Journal of Ethnopharmacology*, vol. 149, no. 1, pp. 157–161, 2013.
- [12] T. Dos Santos Vilhena Leme, T. Bruno Lima Prando, F. Mourão Gasparotto et al., "Role of prostaglandin/cAMP pathway in the diuretic and hypotensive effects of purified fraction of Maytenus ilicifolia mart ex reissek (Celastraceae)," *Journal of Ethnopharmacology*, vol. 150, no. 1, pp. 154–161, 2013.
- [13] R. D. C. V. D. A. F. Da Silva, P. De Souza, S. Crestani et al., "Hypotensive and diuretic effect of the butanolic soluble fraction of the hydroethanolic extract of bark of Scutia buxifolia Reissek in rats," *Journal of Ethnopharmacology*, vol. 172, pp. 395–401, 2015.
- [14] T. B. L. Prando, L. N. Barboza, V. d. O. Araújo et al., "Involvement of bradykinin B2 and muscarinic receptors in the prolonged diuretic and antihypertensive properties of Echinodorus grandiflorus (Cham. & Schltdl.) Micheli," *Phytomedicine*, vol. 23, no. 11, pp. 1249–1258, 2016.
- [15] P. de Souza, L. M. da Silva, T. Boeing et al., "Influence of prostanoids in the diuretic and natriuretic effects of extracts and kaempferitrin fromBauhinia forficataLink leaves in rats," *Phytotherapy Research*, vol. 31, no. 10, pp. 1521–1528, 2017.
- [16] C. L. B. de Almeida, T. Boeing, L. B. Somensi et al., "Diuretic, natriuretic and potassium-sparing effect of nothofagin isolated from Leandra dasytricha (A. Gray) Cogn. leaves in normotensive and hypertensive rats," *Chemico-Biological Interactions*, vol. 268, pp. 103–110, 2017.
- [17] L. N. Bolda Mariano, T. Boeing, R. de Cássia Melo Vilhena de Andrade Fonseca da Silva et al., "1, 3, 5, 6-tetrahydroxyxanthone, a natural xanthone, induces diuresis and saluresis in normotensive and hypertensive rats," *Chemico-Biological Interactions*, vol. 311, Article ID 108778, 2019.

- [18] L. N. Bolda Mariano, T. Boeing, V. Cechinel-Filho, R. Niero, L. Mota da Silva, and P. de Souza, "The acute diuretic effects with low-doses of natural prenylated xanthones in rats," *European Journal of Pharmacology*, vol. 884, Article ID 173432, 2020.
- [19] M. M. D. Molin, S. Silva, D. R. Alves et al., "Phytochemical analysis and antinociceptive properties of the seeds of garcinia achachairu," *Archives of Pharmacal Research*, vol. 35, no. 4, pp. 623–631, 2012.
- [20] B. Lesch and S. Bräse, "A short, atom-economical entry to tetrahydroxanthenones," Angewandte Chemie International Edition, vol. 43, no. 1, pp. 115–118, 2004.
- [21] Shagufta and I. Ahmad, "Recent insight into the biological activities of synthetic xanthone derivatives," *European Journal* of Medicinal Chemistry, vol. 116, pp. 267–280, 2016.
- [22] O. Goshain and B. Ahmed, "Antihypertensive activity, toxicity and molecular docking study of newly synthesized xanthon derivatives (xanthonoxypropanolamine)," *PLoS One*, vol. 14, no. 8, pp. 1–13, 2019.
- [23] P. K. Jyotshna, P. Khare, and K. Shanker, "Mangiferin: a review of sources and interventions for biological activities," *BioFactors*, vol. 42, no. 5, pp. 504–514, 2016.
- [24] M. N. Rana, J. Tangpong, and M. A. Rahman, "Xanthones protects lead-induced chronic kidney disease (CKD) via activating Nrf-2 and modulating NF-kB, MAPK pathway," *Biochemistry and Biophysics Reports*, vol. 21, Article ID 100718, 2020.
- [25] L. N. B. Mariano, D. B. Vendramini-Costa, A. L. T. G. Ruiz et al., "In vitro antiproliferative activity of uncommon xanthones from branches of Garcinia achachairu," *Pharmaceutical Biology*, vol. 54, no. 9, pp. 1697–1704, 2016.
- [26] C. C. Cechinel-Zanchett, L. N. Bolda Mariano, T. Boeing et al., "Diuretic and renal protective effect of kaempferol 3- O-alpha- 1 -rhamnoside (afzelin) in normotensive and hypertensive rats," *Journal of Natural Products*, vol. 83, no. 6, pp. 1980–1989, 2020.
- [27] B. Bhasin and J. C. Q. Velez, "Evaluation of polyuria: the roles of solute loading and water diuresis," *American Journal of Kidney Diseases*, vol. 67, no. 3, pp. 507–511, 2016.
- [28] C. L. B. de Almeida, V. Cechinel-Filho, T. Boeing et al., "Prolonged diuretic and saluretic effect of nothofagin isolated from Leandra dasytricha (A. Gray) Cogn. leaves in normotensive and hypertensive rats: role of antioxidant system and renal protection," *Chemico-biological Interactions*, vol. 279, pp. 227–233, 2017.
- [29] P. J. Bailey, "Sponge implants as models," *Methods in En*zymology, vol. 162, pp. 327–334, 1988.
- [30] K. Y. Na, Y. K. Oh, J. S. Han et al., "Upregulation of Na+ transporter abundances in response to chronic thiazide or loop diuretic treatment in rats," *American Journal of Physi*ology-Renal Physiology, vol. 284, pp. 133–143, 2003.
- [31] A. Greenberg, "Diuretic complications," The American Journal of the Medical Sciences, vol. 319, no. 1, pp. 10–24, 2000.
- [32] I. Legroux-Gerot, L. Catanzariti, X. Marchandise, B. Duquesnoy, and B. Cortet, "Bone mineral density changes in hypercalciuretic osteoporotic men treated with thiazide diuretics," *Joint Bone Spine*, vol. 71, no. 1, pp. 51–55, 2004.
- [33] P. Singh, J. J. Knoedler, A. E. Krambeck, J. C. Lieske, and E. J. Bergstralh, "Thiazide diuretic prophylaxis for kidney stones and the risk of diabetes mellitus," *Bone*, vol. 23, no. 1, pp. 1–7, 2008.
- [34] E. M. Kurowska, "Nitric oxide therapies in vascular diseases," *Current Pharmaceutical Design*, vol. 8, no. 6, pp. 155–166, 2002.

- [35] M. A. Alam, C. Sernia, and L. Brown, "Ferulic acid improves cardiovascular and kidney structure and function in hypertensive rats," *Journal of Cardiovascular Pharmacology*, vol. 61, no. 3, pp. 240–249, 2013.
- [36] M. Majzunova, M. Kvandova, A. Berenyiova, P. Balis, I. Dovinova, and S. Cacanyiova, "Chronic NOS inhibition affects oxidative state and antioxidant response differently in the kidneys of young normotensive and hypertensive rats," *Oxidative Medicine and Cellular Longevity*, vol. 2019, Article ID 5349398, 10 pages, 2019.
- [37] K. V. Ramana, S. Srivastava, and S. S. Singhal, "Lipid peroxidation products in human health and disease 2016," Oxidative Medicine and Cellular Longevity, vol. 2017, Article ID 2163285, 2 pages, 2017.
- [38] Ş. Coşkun, E. G. G. Peker, B. Balabanlı, S. Ahıska, and F. Acartürk, "Effect of transforming growth factor beta 1 (TGF-beta 1) on nitric oxide production and lipid peroxidation in oral mucosal wound healing," *Medicinal Chemistry Research*, vol. 20, no. 1, pp. 23–28, 2011.
- [39] J. P. Kehrer and L.-O. Klotz, "Free radicals and related reactive species as mediators of tissue injury and disease: implications for Health," *Critical Reviews in Toxicology*, vol. 45, no. 9, pp. 765–798, 2015.
- [40] G. F. Rushworth and I. L. Megson, "Existing and potential therapeutic uses for N-acetylcysteine: the need for conversion to intracellular glutathione for antioxidant benefits," *Pharmacology & Therapeutics*, vol. 141, no. 2, pp. 150–159, 2014.
- [41] A. L. T. Ribeiro, A. L. B. Shimada, C. B. Hebeda et al., "In vivo hydroquinone exposure alters circulating neutrophil activities and impairs LPS-induced lung inflammation in mice," *Toxicology*, vol. 288, no. 1-3, pp. 1–7, 2011.
- [42] P. J. Bailey, A. Sturm, and B. Lopez-Ramos, "A biochemical study of the cotton pellet granuloma in the rat," *Biochemical Pharmacology*, vol. 31, no. 7, pp. 1213–1218, 1982.
- [43] N. Fujiwara and K. Kobayashi, "Macrophages in inflammation," Current Drug Target -Inflammation & Allergy, vol. 4, no. 3, pp. 281–286, 2005.
- [44] S. Saha, P. Sadhukhan, and P. C. Sil, "Mangiferin: a xanthonoid with multipotent anti-inflammatory potential," *BioFactors*, vol. 42, no. 5, pp. 459–474, 2016.
- [45] M. L. Lo Faro, B. Fox, J. L. Whatmore, P. G. Winyard, and M. Whiteman, "Hydrogen sulfide and nitric oxide interactions in inflammation," *Nitric Oxide*, vol. 41, pp. 38–47, 2014.