

### Atherosclerotic Calcification: Wnt Is the Hint

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C ardiovascular diseases remain the primary cause of death.<sup>1</sup> Myocardial infarction (MI), angina, and stroke take >16 million lives every year, but the underlying mechanisms by which these often-fatal cardiovascular events occur tend to escape attention.<sup>2</sup> Atherosclerosis is the underlying pathological inflammatory vascular disease not only responsible for most MIs and strokes, but it accounts for 29% of deaths worldwide.<sup>1,3</sup>

Once believed to be a passive process, it is now understood that atherosclerosis takes an active route involving several cell types, with contributions from a multitude of organ systems, molecular mechanisms, and other pathological conditions, including, but not limited to, hypertension, hyperlipidemia, and type 2 diabetes mellitus.<sup>1,3–5</sup> Atherosclerosis is characterized by the accumulation of lipids, fibrous elements, and inflammatory cells within the vascular wall of medium and large muscular and elastic arteries.<sup>1,3,6</sup> Atherosclerotic plaque formation leads to intimal thickening and luminal stenosis.<sup>1</sup> Reduction of blood flow ensues, leading to ischemia of the brain, heart, and extremities and clinically manifesting as MI, angina, or stroke.

The main features of atherosclerosis are endothelial dysfunction, intimal thickening, inflammation, and vascular calcification (VC).<sup>4,7,8</sup> These processes are considered a consequence to vascular injury.<sup>1</sup> VC is characterized by calcium deposition in the walls of the vasculature.<sup>5</sup> Expansion of the calcified lesion leads to thrombus formation and, if significant enough, vascular occlusion. Although the exact mechanisms for VC are unknown, there has been an increase in interest about the role Wingless (Wnt) signaling plays in

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disease pathogenesis. Briefly, the Wnt signaling pathway is an evolutionarily conserved pathway across kingdom Animalia and plays a crucial role in pattern formation during embryogenesis.<sup>9</sup> The Wnt signal transduction pathway plays a crucial role in organ formation in embryonic development, cell proliferation, polarity, migration, and differentiation.<sup>10</sup> Thus, deregulated Wnt signaling is associated with many human diseases.<sup>11</sup> Therefore, it is justifiable to suggest that research delineating atherogenesis, with focus on the contribution of Wnt signaling in the pathogenesis of atherosclerotic calcification, is warranted.

## Components of the Vasculature System Endothelium

The vascular endothelium has a multifaceted role, acting simultaneously as a permeability barrier, a nonthrombogenic surface, a regulator of vascular tone and transendothelial flow, and an inhibitor of vascular smooth muscle cell (VSMC) growth and migration.<sup>12</sup> Under normal circumstances, endothelial cells (ECs) release an array of vasodilators and anticoagulants, such as NO and prostacyclin, which reduce platelet aggregation, monocyte and leukocyte adhesion, and VSMC proliferation.<sup>13</sup> Under pathological conditions, ECs release factors, such as thromboxane and endothelin-1, that increase platelet aggregation, monocyte adhesion, vasoconstriction, and VSMC proliferation. Wnt signaling has been shown to be involved in EC permeability, proliferation, and survival.<sup>14–16</sup> Naturally,  $\beta$ -catenin binds to cadherins to stabilize adhesion of neighboring ECs.<sup>17</sup> During cellular migration, however,  $\beta$ -catenin localizes in the nucleus, resulting in a loss of tissue integrity.<sup>18</sup>

#### Vascular Smooth Muscle Cells

Smooth muscle cells of the vasculature are found as 1 of 2 possible phenotypes: the first is the contractile phenotype, the predominant form of VSMC, in the tunica media; and the second is the activated synthetic phenotype, in the tunica intima.<sup>12</sup> The differentiated contractile VSMCs can be

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stimulated by adrenergic receptors or angiotensin II. They have a low proliferation rate that is maintained in check by the endothelium-derived NO.<sup>12</sup> Vascular injury triggers the phenotypic change to an active synthetic phenotype with an increased proliferation rate and secretory ability.<sup>19</sup> Wnt signaling promotes VSMC survival,<sup>20</sup> whereas silencing Wnt signaling reduces cell proliferation.<sup>21</sup> Along with cellular proliferation, some data suggested that Wnt signaling may play a role in the VSMC migration during intimal thickening,<sup>2,22</sup> motility,<sup>23</sup> and osteogenesis.<sup>24</sup> Moreover, a recent study has demonstrated a role for Wnt5a in cholesterol efflux by VSMCs.<sup>25</sup>

#### **Adventitial Fibroblasts**

Fibroblasts predominate the vascular adventitia. They have low proliferation rates and can synthesize extracellular matrix proteins and ground substance. In response to injury, fibroblasts rapidly proliferate, migrate, and differentiate to myofibroblasts.<sup>12</sup> Canonical Wnt signaling has played a prominent role in fibroblast proliferation and differentiation.<sup>26,27</sup>

## Endothelial Dysfunction, Intimal Thickening, and Luminal Stenosis

Some naturally occurring stimuli within the body have the ability to injure the vessel wall and lead to endothelial dysfunction. For example, the turbulent flow that occurs in bifurcated regions of the vasculature is enough to activate endothelium, which is perhaps why atherosclerotic plaque formation often occurs in sites like carotid and coronary arteries.<sup>7</sup> Long-term exposure to risk factors, such as hyperlipidemia, hypertension, and diabetes mellitus, also contributes to endothelial injury.<sup>1,4</sup> There are 2 ways by which the endothelium can be activated. During minimal vascular injury, whereby the short-lived inflammatory response can effectively remove or solve the problem, vessel remodeling is less likely to occur.<sup>1</sup> In contrast, sustained exposure to the above mentioned risk factors increases vascular adhesiveness, leakage, and accumulation of extracellular lipid within the vessel wall, thereby inducing foam cell formation and inflammatory response propagated by sustained release of growth factors and cytokines.<sup>3,28,29</sup>

Vascular endothelium is considered to be dysfunctional when inflammation is sustained, altering the endothelial homeostatic properties.<sup>1</sup> When dysfunctional, ECs exhibit increased expression of adhesion molecules and release cytokines, resulting in monocyte, lymphocyte, and platelet adhesion, enhancing the inflammatory response.<sup>7,12</sup> Adhesion of monocytes is considered the hallmark of atherosclerosis

and considered atheromatous from this point forward.<sup>4</sup> Dysfunction also alters endothelial permeability, allowing intraplaque invasion of monocytes and macromolecules, like low-density lipoprotein.<sup>19</sup> Under normal conditions, NO inhibits VSMC migration. In endothelial dysfunction, NO levels are reduced, allowing VSMC proliferation and migration into the intima, contributing to the next key process in atheroscle-rotic progression: pathological intimal thickening.<sup>7,12</sup>

The sustained inflammatory response not only drives proliferation and migration of VSMCs, it also increases the inflammatory cell population by recruitment and intraplaque proliferation of macrophages and lymphocytes. Synthesis of extracellular matrix by VSMCs, along with inflammatory cell invasion, thickens the arterial wall.<sup>1,30</sup> When lipids begin to accumulate in the vessel wall, modification via aggregation induces lipid uptake in macrophages, creating foam cells.<sup>6,7</sup> Furthermore, macrophages can secrete cytokines to recruit more monocytes or induced proliferation of resident monocytes, creating a feed-forward cycle for an ad infinitum inflammatory state.<sup>1,31</sup>

Apoptosis of VSMCs and macrophages, along with loss of extracellular matrix, contributes to plaque instability and may lead to eventual rupture, which accounts for  $\approx$ 50% of MIs and acute coronary syndromes attributable to occlusion of the artery.<sup>1,28</sup> A plaque burden of 70% or greater has significantly increased the risk of rupture.<sup>32</sup> However, approximately half of those who died from luminal thrombosis, leading to complete artery occlusion (resulting in sudden coronary death), had <75% stenosis, and  $\approx$ 10% of sudden coronary deaths occur without atheromatous lesion formation.<sup>33,34</sup> Therefore, lesion composition may be just as relevant to risk of rupture as lesion size.<sup>35</sup>

Microvessel formation in the lesion, otherwise known as angiogenesis, contributes to plaque development and increases the risk of rupture.<sup>8</sup> In addition, there are suggestions that an obstructive calcified lesion is more stable than a nonobstructive thin-cap fibroatheroma, because the calcified plaque leads to stable angina pectoris, whereas the fibroatheroma has a tendency to rupture.<sup>36</sup> Thus, calcification may be paradoxically beneficial, despite its associations with cardiovascular morbidity and mortality.<sup>30</sup> Conversely, the increased incidence of disease and death with VC may not be a direct effect of calcification itself, but rather the fact that calcification is found in advanced atherosclerotic lesions.<sup>37</sup>

### Vascular Calcification

VC is the pathological deposition of calcium in vascular structures and is a significant area of study because it commonly affects our aging population and those experiencing diabetes mellitus, dyslipidemia, heart valve disease, and end-stage renal disease.<sup>5</sup> VC is a marker for atherosclerosis

and is associated with several cardiovascular pathological features, including hypertension, congestive heart failure, cardiac hypertrophy, and ischemia, and increased risk of MI and stroke.<sup>38,39</sup> VC is a significant risk factor for morbidity and mortality in cardiovascular disease.<sup>7,19,37,40-44</sup> Once thought to be a passive degenerative process, it is now evident that biomineralization of the arterial wall is a complex regulated process.<sup>5,45,46</sup> There are 4 histoanatomic variants of VC, which include medial artery calcification, atherosclerotic intimal calcification, cardiac valve calcification, and soft tissue calciphylaxis or calcific uremic arteriolopathy.<sup>38,47</sup> A major breakthrough in the study of VC was the realization that the ectopic deposition of calcium in vascular structures is similar to intramembranous and endochondral ossification in bone development.<sup>48</sup> As early as the 19th century, bonelike tissue was identified within atherosclerotic arteries.49

In general, arterial calcification occurs in the intimal layer of the wall and in the aortic valve, and it is commonly found in the aorta and coronary, carotid, and renal arteries.<sup>50,51</sup> However, the prevalence of arterial calcification varies widely between populations.<sup>51</sup> Both vascular and cardiac valve calcification are considered complications of atherosclerosis because risk factors that predispose and/or potentiate plaque formation may also predict progression of calcification.<sup>52–54</sup> Intimal calcification itself is a key contributive process of advanced atherosclerotic lesion formation, and vascular calcium detection by computed tomography can be used as a subclinical marker of atherosclerosis. The fact that coronary artery and aortic valve calcification.<sup>4,7,33,42,55,56</sup>

The progression of atherosclerosis results in a structural and cellular remodeling of the vasculature.4,7,41,57 Changes that alter mineral metabolism, modify calcification regulators, or modulate vascular cell phenotype can have catastrophic consequences in vascular structures.40,55,58-60 Advanced calcific lesions can develop woven bone and even commence hematopoiesis. This remodeling of the vasculature can extend to the point of acquiring some functional characteristics of bone, offering the perspective that VC occurs in a manner similar to bone development.<sup>48,60–62</sup> Briefly, osteogenesis involves formation of mesenchymal cell condensations, which differentiate into bone-forming cells for intramembranous or endochondral ossification.63,64 Direct differentiation of mesenchymal cells to osteoblasts in intramembranous (IM) bone development allows deposition of mineralization-ready bone matrix, whereas EC ossification first requires the deposition of a cartilage template by chondrocytes, on which osteoblasts will elaborate to form bone.<sup>65</sup> Mineralization of bone is initiated by shedding of matrix vesicles by osteoblasts and chondrocytes.<sup>66,67</sup> These matrix vesicles contain hydroxyapatite crystals that act as nucleation sites for the mineralization cascade.<sup>66,67</sup> Several genetic studies support that both IM and EC mechanisms are regulated by multiple signaling factors and pathways, including bone morphogenic proteins (BMPs), muscle segment homeobox protein homolog 2 (Msx2), Runt-related transcription factor 2 (Runx2), sex determining region Y-box 9 (Sox-9), and Wnt.<sup>5,68,69</sup> BMPs regulate bone formation and osteogenic differentiation.<sup>70</sup> Msx2 expression is crucial for cranial bone development and IM ossification.<sup>55</sup> Runx2 is a master transcription factor also required for osteoblast differentiation and chondrocyte maturation.<sup>65</sup> Sox-9 is a master regulator of chondrogenic mesenchymal condensations.55,65 Wnt signaling mechanisms regulate stem cell proliferation and organogenesis; they also regulate bone mass.<sup>64</sup> Altered expression of these factors impairs proper bone development by disturbing osteogenic cell differentiation or tissue mineralization.

There are 2 pathways for bone formation: intramembranous ossification or endochondral ossification.<sup>64</sup> Intramembranous ossification requires the formation of mesenchymal condensations that eventually directly differentiate into osteoblasts and mineralize into bone.<sup>64</sup> Most bones of the body, however, undergo endochondral ossification, whereby a cartilage template is formed first, followed by mineralization. Low levels of  $\beta$ -catenin signaling allow increased expression of SOX-9, a regulatory factor of cartilage, resulting in endochondral formation, whereas increased  $\beta$ -catenin levels drive Runx2 expression and intramembranous ossification. This suggests a regulatory factor controlling VC.

The mechanism of VC seems to involve several pathways as well as exogenous influence. It has been established that Msx2 drives osteogenic differentiation by activating Wnt signals and upregulating alkaline phosphatase.<sup>5,61</sup> Moreover, Msx2 upregulates Wnt/β-catenin-dependent T-cell factor/ lymphoid enhancer-binding factor transcription, as well as upregulating Wnt3a and Wnt7a while downregulating the Wnt inhibitor Dickkhopf homologue-1.<sup>5</sup> The study of bone metabolism has progressed much further than that of calcified vascular tissue,<sup>48</sup> and the mechanisms underlying the active process and pathogenesis of VC remain unclear. The crucial role of Wnt signaling in bone metabolism makes the Wnt signal transduction pathways a promising field of study in determining the molecular mechanisms underlying VC. Developing a better understanding of the pathogenesis of VC will help contribute to the advancement of treatments and therapies for this vascular pathological feature.

#### Wnt Signaling Pathways

The name Wnt originates from the fusion of the words *wingless*, the Drosophila segment polarity gene, and *integrated* or *int-1*, the mammalian homolog of *wingless*.<sup>71</sup> The Wnt family of proteins consists of 19 secreted lipid-modified

glycoproteins that bind to the Frizzled receptor family.<sup>11</sup> There are 10 Frizzled isoforms in humans, and these Frizzled proteins have 7 transmembrane domains and are classified as a distinct family of G-protein–coupled receptors.<sup>72</sup> Once Wnt binds to Frizzled and the associated coreceptor complex, the signal is transduced to the cytoplasmic protein Disheveled. At this level, the cascade branches off into 3 major signal transduction pathways, the canonical or Wnt/ $\beta$ -catenin pathway and the noncanonical planar cell polarity and Ca<sup>2+</sup>dependent pathways (Wnt/Ca<sup>2+</sup> pathway).<sup>2</sup>

The canonical Wnt/ $\beta$ -catenin signaling pathway is the most studied Wnt pathway and has played an essential role in many biological processes, such as development, cell proliferation, and bone metabolism.<sup>73</sup> At rest, under unstimulated conditions, a protein scaffold of Axin, adenomatous polyposis coli,<sup>74</sup> and glycogen synthase kinase-3 $\beta$  (GSK3) mediate the phosphorylation of  $\beta$ -catenin. This results in the polyubiquitination and proteasomal degradation of  $\beta$ -catenin.<sup>75,76</sup> In active Wnt

signaling, Wnt binds Frizzled receptors and low-density lipoprotein receptor-related protein (LRP) 5/6 coreceptors, resulting in the activation of Disheveled, which inhibits GSK3 activity and disrupts the adenomatous polyposis coli/Axin/ GSK3 complex that targets  $\beta$ -catenin for destruction.<sup>76–78</sup> As a result,  $\beta$ -catenin is stabilized, accumulates in the cytoplasm, and subsequently translocates to the nucleus.<sup>73</sup> In the nucleus, β-catenin associates with members of the lymphoid enhancer-binding factor/T-cell factor family of transcription factors and several other factors, enabling this complex to act as a transcriptional activator or repressor of Wnt target genes.<sup>76,79,80</sup> Canonical Wnt signaling is illustrated in Figure 1. As mentioned previously, Wnt molecules interact with several different surface receptors and can produce a variety of cellular outcomes, depending on the presence of coreceptors and which receptor complexes can be formed.<sup>81</sup>

The 2 main noncanonical/ $\beta$ -catenin-independent Wnt signaling cascades are the Wnt planar cell polarity and the



**Figure 1.** Schematic representation of inactive and activated canonical Wht signaling. Without activation of the Frizzled receptor and the lowdensity lipoprotein receptor–related protein (LRP) 5/6 coreceptor, the Axin, adenomatous polyposis coli (APC), and glycogen synthase kinase-3 (GSK3) complex forms and phosphorylates  $\beta$ -catenin, leading to its ubiquitination and subsequent degradation. Activation of canonical Wht signaling leads to  $\beta$ -catenin nuclear localization and change in expression of Wht target genes. LEF/TCF indicates lymphoid enhancer-binding factor/T-cell factor.

Wnt/Ca<sup>2+</sup> signaling pathways. However, there are other noncanonical Wnt signaling pathways, such as the Wnt-Rasrelated protein 1 (RAP1) pathway, the Wnt/receptor tyrosine kinaselike orphan receptor 2 receptor pathway, the Wntprotein kinase A pathway, the Wnt-GSK3-microtubule pathway, the Wnt-c-Jun N-terminal kinase signaling pathway, the Wnt-related to tyrosine kinases pathway, and the Wntmammalian target of rapamycin pathway.<sup>47</sup> There is a great deal of overlap between these noncanonical pathways and between noncanonical Wnt signaling and the canonical Wntβ-catenin pathway.<sup>82</sup> The traditional view of Wnt signaling suggests that Wnt1, Wnt3a, Wnt8, and Wnt8b are involved in canonical Wnt signaling, whereas Wnt4 and Wnt5a act in the noncanonical Wnt signaling pathway. However, classification of these molecules is difficult because of the significant cross talk between the pathways.<sup>83</sup> In the noncanonical planar cell polarity pathway, Wnt signaling leads to the activation of the small GTPases Rho and Rac, which mediate cytoskeletal rearrangements, resulting in lateral asymmetry.<sup>77</sup> In the Wnt/

Ca<sup>2+</sup> signaling cascade (Figure 2), Wnt binds to the extracellular cysteine-rich domain on the N terminus of its cognate Frizzled receptor.<sup>72</sup> This promotes the interaction of Frizzled with Disheveled-Axin-GSK3, and GSK3 mediates the phosphorylation of receptor tyrosine kinaselike orphan receptor 1/ 2, a Wnt-Frizzled coreceptor.83 The phosphorylation of the receptor tyrosine kinaselike orphan receptor coreceptor leads to the activation of phospholipase C, which mediates a transient increase in inositol 1,4,5-triphosphate and 1,2diacylglycerol from membrane-bound phospholipid phosphatidylinositol 4,5-bisphosphate.83 Inositol 1,4,5-triphosphate subsequently interacts with the calcium channels on the endoplasmic reticulum membrane. This results in a release of Ca<sup>2+</sup>, which interact with calmodulin to activate calmodulin-dependent protein kinase II.83,84 The mobilized calcium ions from the endoplasmic reticulum also interact with 1,2-diacylglycerol to activate protein kinase C.85 Calmodulin-dependent protein kinase II and protein kinase C then activate various nuclear transcription factors (eg, nuclear



**Figure 2.** Schematic representation of the cell polarity and Ca<sup>2+</sup> noncanonical Wnt signaling pathways. In the cell polarity pathway, activation of particular Frizzled receptors leads to cytoskeletal rearrangement via activation of Rho and Rac GTPases. In the Ca<sup>2+</sup> pathway, Wnt-Frizzled receptor binding leads to increased intracellular Ca<sup>2+</sup> concentrations from the endoplasmic reticulum. Under these conditions, nuclear factor- $\kappa$ B (NF- $\kappa$ B), cAMP responsive element binding, and nuclear factor associated with T cells transcription factors can translocate to the nucleus and promote an osteogenic gene profile. CamKII indicates Ca<sup>2+</sup>/ calmodulin-dependent protein kinase II; GSK3, glycogen synthase kinase-3; IP3, inositol 1,4,5-triphosphate; PIP2, phosphatidylinositol 4,5-bisphosphate 2; PKC, protein kinase C; PLC, phospholipase C; and Ror2, receptor tyrosine kinaselike orphan receptor 2.

factor- $\kappa$ B and cAMP responsive element binding), and the released calcium ions can activate phosphatase calcineurin, a widely expressed protein that can then dephosphorylate and thus activate cytoplasmic nuclear factor associated with T cells.<sup>83,86</sup> Nuclear factor- $\kappa$ B, cAMP responsive element binding, and nuclear factor associated with T cells can translocate to the nucleus and regulate the transcription of Wnt target genes.<sup>83</sup> The noncanonical cell polarity and Ca<sup>2+</sup> Wnt signaling pathways are illustrated in Figure 2.

### Implication of Wnt Signaling in Atherosclerosis Pathological Features

Wnt signaling plays an important role in several key biological processes, including cardiac development and angiogenesis. More recently, evidence is emerging for the role of Wnt signaling in atherosclerosis and, in particular, VC (summarized in Figure 3).<sup>4</sup> Behind the skeleton, the arterial vasculature is the second most extensively calcified structure in the human body.<sup>39</sup> Recent evidence demonstrates that Wnt ligands and their receptors play a role in calcification itself as well as the atherosclerotic events that predispose vessels to this pathological calcification. As mentioned previously, monocyte adhesion to ECs is a hallmark of atherosclerosis. Canonical Wnt signaling has been demonstrated to directly increase this monocyte adhesion.<sup>87</sup> Interestingly, Wnt signaling in ECs has been shown to activate proinflammatory gene expression via

inhibition of canonical Wnt signaling and activation of the noncanonical Ca<sup>2+</sup>-dependent pathway via ligand Wnt5a.<sup>88,89</sup> Several studies have demonstrated a direct link between canonical Wnt signaling via ligands Wnt1, Wnt2, and Wnt3a and arterial smooth muscle cell proliferation, which underlies the intimal thickening stage of atherosclerosis. 20,21,90,91 Canonical Wnt signaling and LRP5 have also been directly implicated in the promotion of foam cell formation.<sup>6</sup> Sclerostin, an inhibitor of canonical Wnt signaling, has also been of interest to prevent atherosclerosis. A recent study used human aortic aneurysm samples and transgenic mice overexpressing sclerostin in apolipoprotein E-deficient mice.<sup>92</sup> Sclerostin was downregulated in aortic aneurysms, and overexpressing sclerostin in atherogenic mice resulted in a decreased inflammatory response that inhibits development of atherosclerosis and aortic aneurysms.

## Implication of Wnt Signaling in Atherosclerotic Calcification

VC in diabetic  $LDLR^{-/-}$  mice is associated with upregulation of Wnt signaling via a mechanism involving BMP2.<sup>5</sup> The latter has been demonstrated in atherosclerotic lesions, particularly in vascular myofibroblasts and endothelium.<sup>93</sup> BMP2 expression is activated by pathogenic stimuli, such as tumor necrosis factor- $\alpha$ ,<sup>94</sup> oxidized lipids,<sup>94</sup> and hyperglycemia<sup>95,96</sup> More important, BMP2 has been demonstrated to regulate



Figure 3. Involvement of Wnt signaling in the pathogenesis of atherosclerosis. Wnt signaling is associated with monocyte attachment, inflammation, cell proliferation, foam cell formation, and atherosclerotic calcification, ultimately leading to arterial stenosis. VSMC indicates vascular smooth muscle cell.

osteogenic programs contributing to VC.<sup>55,62</sup> One of the main ways in which BMP2 regulates osteogenic gene expression programs is through the induction of transcription factor Msx2, which controls craniofacial mineralization.<sup>97</sup> Msx2 has also promoted osteogenic differentiation, and it suppresses adipogenesis of vascular myofibroblasts.<sup>96</sup> Msx2 enhances Wnt signaling through the upregulation of several Wnt ligands<sup>98</sup> and the downregulation of dickkopf homologue 1,<sup>99</sup> a canonical Wnt pathway antagonist. Indeed, Msx2 and Wnt seem to regulate each other's expression in a bidirectional way.<sup>100,101</sup> For instance, Msx2 stimulates Wnt-dependent T-cell factor/lymphoid enhancer-binding factor transcription, increases  $\beta$ -catenin nuclear localization, and increases the expression of canonical Wnt ligands, Wnt3a and Wnt7, and noncanonical ligand, Wnt5.

#### Wnt Signaling in Pericytes

Wnt/β-catenin signaling has also contributed to VC and atherosclerosis by controlling the aberrant differentiation of pericytes in the vasculature.<sup>102</sup> Although pericytes were initially thought to be a feature of the microvasculature, they have been found to be present across the entire vasculature.<sup>103</sup> Their recruitment to the intima, media, and sites of arterial calcification in several pathological conditions, such as atherosclerosis, suggests their involvement in these diseases.<sup>102,104,105</sup> It has been demonstrated that the activation of Wnt/ $\beta$ -catenin signaling by transforming growth factor- $\beta$ 3 induces the chondrogenic and inhibits the adipogenic differentiation of pericytes,<sup>102</sup> which are thought to be uncommitted mesenchymal progenitor cells.<sup>106</sup> Studies have shown calcifying vascular cells to express pericyte markers, such as  $\alpha$ -smooth muscle actin and 3G5.<sup>93</sup> These studies, in combination with the knowledge that pericytes can undergo chondrogenic, osteogenic, and adipogenic differentiation,<sup>107,108</sup> contrasts the function of vascular cells, which can only undergo chondrogenic and osteogenic differentiation.<sup>109</sup> This prompted the hypothesis that calcifying vascular cells originate from pericytes that have been exposed to Wnt signaling.<sup>102</sup>

### Implication of Canonical Wnt Signaling in VC

In support of its role in VC, Wnt/ $\beta$ -catenin signaling plays a crucial role in bone development and homeostasis. During development, canonical ligand Wnt3a maintains apical ecto-dermal cells of limb buds in a highly proliferative state and is, thus, essential to embryonic skeletal development.<sup>110,111</sup> Loss-of-function mutations of Wnt coreceptor LRP5 cause reduced bone mass, skeletal fragility, and osteoporosis.<sup>112</sup> LRP5<sup>-/-</sup> mice have increased tibial fractures because of an

overall decrease in bone mass and reduced osteoblast proliferation and function with no effect on bone resorption, whereas single-allele knockouts of LRP5 lead to an intermediate phenotype of bone mass. This suggests a dosedependent response for LRP5 alleles on bone mass density and that LRP5 regulates osteogenesis through its effects on osteoblast proliferation, differentiation, and function.<sup>113,114</sup> Wnt coreceptor LRP6 is thought to control postnatal bone mass,<sup>113</sup> and LRP6<sup>+/-</sup> mice lacking LRP5 had an even further decrease in bone mass density.<sup>115</sup> LRP5/6 knockout models in hypercholesterolemic (apolipoprotein E-deficient) mice developed less calcification in the aortic valve compared with control models.<sup>116</sup> On the other hand, gain-of-function mutations of LRP5 are associated with a high bone mass phenotype.<sup>117,118</sup> Interestingly, no change in osteoblast activity was observed in mice with increased expression of LRP5.<sup>119</sup> These mice did, however, have decreased osteoblast apoptosis, suggesting that the high bone mass phenotype is because of increased osteoblast cell count rather than function. Genomic sequencing of unrelated families with high bone density identified 6 different gain-of-function mutations in the LRP5 gene.<sup>120</sup> Loss-of-function mutations in LRP5 were identified to be associated with osteoporosis.<sup>112</sup> In the context of atherosclerosis, a study of a family with early coronary artery disease, metabolic syndrome, and osteoporosis showed an LRP6 mutation that impairs Wnt signaling,

# Implication of Noncanonical Wnt Signaling in VC

suggesting a genetic link between LRP6 and Wnt signaling

with coronary artery disease.<sup>121</sup>

Despite the fact that there is evidence implicating Wnt signaling in osteogenic differentiation and development, until recently, little was known about the role of noncanonical Wnt ligands in vascular and valvular calcification and atherosclerosis. The prototypical noncanonical ligand Wnt5a has been implicated in mouse bone homeostasis.<sup>122</sup> Additional studies have shown that  $Wnt5a^{+/-}$  mice have reduced osteogenesis and increased adipogenesis in bone marrow mesenchymal progenitors and low bone mass.<sup>123</sup> Through the calciumdependent Wnt signaling pathway, Wnt5a is thought to activate histone methyltransferase SET domain bifurcated 1, leading to the formation of a corepressor complex that suppresses peroxisome proliferator-activated receptor  $\gamma$ (PPAR $\gamma$ ) function and thus mediates the osteoblastic differentiation of mesenchymal stem cells (MSCs).<sup>123</sup> A recent study has demonstrated a link between Wnt5a, toll-like receptor 4, and pathological inflammation in human macrophages.<sup>124</sup> Furthermore, the expression of Wnt5a has been demonstrated in murine and human atherosclerotic tissue.<sup>19</sup> In addition to finding strong Wnt5a expression in macrophagerich regions of atherosclerotic lesions of apolipoprotein E– deficient mice on atherogenic diets, Wnt5a was also expressed in other regions of developed human plaques, such as in smooth muscle cells.<sup>19</sup>

The potential involvement of Wnt5a in VC supports the investigation of other noncanonical Wnt ligands, particularly those working through the Wnt/Ca<sup>2+</sup> pathway, and their roles in the molecular mechanisms underlying atherosclerosis. In particular, Wnt5b is an interesting candidate to study, because it is a paralog of Wnt5a, with 80% amino acid identity to the human Wnt5a protein.<sup>125</sup> By a recent bone mineral density genome-wide association study, Wnt5b (along with other Wnt signaling molecules, such as AXIN1, LRP5, Catenin beta-1 (CTNNB1), WNT3, and several others) has been shown to be involved in key biological pathways involving the skeleton.<sup>126</sup> Both Wnt5a and Wnt5b have been involved in coordinating chondrocyte proliferation and differentiation during endochondral ossification of long bones in mice.<sup>127</sup> More specifically, both Wnt5a and Wnt5b are expressed in the chondrocytes of developing long bones, and Wnt5a signaling regulated proliferation and differentiation of zone I epiphysis chondrocytes and the highly proliferative zone II chondrocytes. Although Wnt5a inhibits the transition of cells from zone I to zone II, Wnt5b promotes the transition from zone I to zone II and thus has an antagonistic effect in regulating chondrocyte proliferation.<sup>127</sup> A functional role for Wnt5b in adipogenesis was elucidated in 3T3-L1 preadpiocytes, where Wnt5b overexpression promoted adipogenesis by antagonizing the Wnt3a-mediated Wnt/β-catenin pathway.<sup>128</sup> In subsequent studies in the same preadipocyte cell line, both Wnt5a and Wnt5b inhibited β-catenin-dependent Wnt signaling; however, Wnt5b overexpression, in particular, was determined to be a potent stimulator of adipogenesis via inhibition of canonical Wnt signaling and increased PPAR $\gamma$ expression.<sup>129</sup> This increased PPAR $\gamma$  expression is of particular interest because PPARy involvement in lipid metabolism is particularly relevant in the study of atherosclerosis.

PPARγ is expressed primarily in adipose tissue and is also found in the heart.<sup>130</sup> PPARγ is expressed in ECs, VSMCs, and macrophages.<sup>131</sup> In addition to the well-established role of PPARγ in lipid metabolism, there is increasing support for its role in modulating inflammation, macrophage differentiation, and atherosclerosis.<sup>132</sup> PPARγ has had inhibitory effects on VSMC proliferation and migration<sup>133,134</sup> as well as EC growth and migration.<sup>135</sup> PPARγ activation reduces monocyte recruitment to atherosclerotic plaques.<sup>136</sup> PPARγ also has a modulatory effect on inflammation and is known to decrease monocyte/macrophage production of several inflammatory cytokines.<sup>137</sup> There is a large amount of evidence supporting the antiatherosclerotic properties of PPARγ, whether indirectly by improving control of lipid and blood glucose levels or directly by increasing plaque stability and direct actions on ECs, VSMCs, and macrophages.<sup>135</sup> This evidence suggests the potential of PPAR $\gamma$  in the treatment of atherosclerotic disease<sup>133</sup> and makes its relationship with Wnt5b and Wnt signaling a crucial area of study.

Another interesting Wnt ligand in the study of atherosclerosis is Wnt11. The embryonic expression pattern of Wnt11, particularly its restricted expression to the perichondrium of the developing skeleton, has long suggested its role in skeletal development.<sup>138</sup> Interestingly, the chromosomal location of Wnt11 to 11q13.5 is near previously linked markers of high bone mass.<sup>138,139</sup> In addition, Wnt11 expression is upregulated during mesenchymal osteogenesis.<sup>140</sup> Wnt11 activation has been associated with bone canonical Wnt/ $\beta$ -catenin signaling and noncanonical signaling.<sup>141</sup> Activation of Wnt11 through β-catenin-independent Wnt signaling is essential for embryonic cardiac development.<sup>142</sup> In terms of its role in osteogenic development, Wnt11 activation in MC3T3E1 preosteoblasts increases  $\beta$ -catenin accumulation, promoting BMP-induced osteoblast maturation and mineralization through canonical Wnt signaling.143 Wnt11 induces the expression of R-spondin 2, a secreted enhancer of Wnt signaling, which stimulates osteoblastogenesis through increasing the expression of osteogenic factors, such as Dmp1 (dentin matrix protein 1), Phex (phosphate-regulating endopeptidase homolog), and *Bsp* (bone sialoprotein).<sup>143</sup>

When studying the effects of Wnt signaling on the osteogenic differentiation of adult human MSCs, an interesting model of cross regulation between canonical and noncanonical Wnt signaling in the regulation of osteogenesis was proposed.<sup>140</sup> In this model, the MSCs exist in a proliferating progenitor cell stage or in a lineage-committed differentiated stage. Canonical Wnt ligands Wnt3a and Wnt9a promote MSC proliferation and maintenance of the progenitor cell pool. Meanwhile, noncanonical signaling, mediated by Wnt5a and Wnt11, is upregulated during MSC osteogenic and chondrogenic differentiation<sup>144</sup> and is involved in regulating and promoting MSC differentiation.<sup>140</sup> In this model, MSCs are maintained in the progenitor state by canonical signaling and will undergo differentiation on an exogenous signal involving noncanonical Wnt signaling that will release the cells from suppression of differentiation. These cells are then able to undergo osteogenic, chondrogenic, or adipogenic lineage commitment.<sup>140</sup> This model suggests that Wnt/ $\beta$ -catenin signaling can promote osteogenesis by expansion of the progenitor cell compartment.<sup>140</sup>

# Involvement of Wnt Signaling in Aortic Valve Calcification

There is strong evidence suggesting the involvement of Wnt signaling cascades in cardiac valve calcification associated with human degenerative valve disease.<sup>145</sup> For example, LRP5 Wnt

coreceptor regulates bone matrix protein expression in the aortic valves and vasculature of mice with hypercholesterolemia.<sup>46</sup> Through its interaction with Wnt proteins, LRP5 activates  $\beta$ -catenin and is, thus, an important factor in mediating osteogenesis. Calcified aortic valves had a bonelike phenotype, whereas mitral valves had a cartilage phenotype; LRP5/Wnt3 signaling markers were present in both types of disease valves, but expression was more prominent in aortic valves.<sup>145</sup> These findings suggest that the LRP5/Wnt3 signaling pathway could be mediating an endochondral ossification process in the progression of valvular heart disease.<sup>145</sup>

Recently, we studied the role of noncanonical ligands Wnt5a, Wnt5b, and Wnt11 in aortic valve calcification.<sup>146</sup> Calcified valves had significantly elevated mRNA expression of all 3 Wnt ligands compared with noncalcified valves. Immunohistochemical analyses in atherosclerotic aortic valves demonstrate elevated Wnt5a expression within and surrounding areas of calcification, whereas Wnt5b and Wnt11 are found in fibrosis, inflammation, lipid, and calcification. Both Wnt5b and Wnt11 immunostaining had significant positive correlations with aortic jet velocity, mean and maximum pressures across the aortic valve. In addition, in vitro studies of the effects of Wnt5a, Wnt5b, and Wnt11 on human aortic valve interstitial cells demonstrated that all 3 Wnts increased apoptosis, osteogenic gene expression, and calcification (via alkaline phosphatase activity) in these cells.

More recently, we have shown that lipoprotein (a), known to be genetically associated with aortic valve calcification, caused increased deposition of minerals and microvessel formation in human aortic valve interstitial cells through induction of Wnt signaling pathways.<sup>147</sup>

### Conclusion

Investigating the role of Wnt signaling in vascular and valvular calcification is fascinating given the crucial role that both canonical and noncanonical Wnt ligands play in skeletal development and the accumulating evidence suggesting that atherosclerotic calcification recapitulates bone development. The current body of evidence is suggestive of both canonical and noncanonical Wnt ligands contributing to pathological calcification of the aortic valves and vasculature. Further research into the exact signaling pathways underlying these processes is warranted to better understand this phenomenon. However, it is more likely that multiple pathways contribute to calcification, making it difficult to identify a single molecular target to treat VC. Current pharmacological therapies targeting Wnt signaling are still in their infancy.<sup>148</sup> Indeed, several pharmaceutical molecules that are known to modulate the Wnt signaling pathway have been developed and tested in cancer and neurologic disorder therapy.<sup>149,150</sup> A couple of others are used in the treatment of osteoporosis.<sup>151</sup> Indeed, on the basis of the evidence we provided above, there is still tremendous potential to use Wnts and their effectors in treating VC. Thus, we suggest that future experiments attempt to identify novel targets that either regulate or contribute to Wnt signaling. In doing so, these new targets, as well as those that are in current use clinically, may ultimately contribute to reducing cardiovascu-

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lar morbidity and mortality.

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#### **Disclosures**

None.

#### References

- 1. Ross R. Atherosclerosis: an inflammatory disease. N Engl J Med. 1999;340:115–126.
- Mill C, George SJ. Wnt signalling in smooth muscle cells and its role in cardiovascular disorders. *Cardiovasc Res.* 2012;95:233–240.
- Ramsey SA, Gold ES, Aderem A. A systems biology approach to understanding atherosclerosis. *EMBO Mol Med*. 2010;2:79–89.
- Marinou K, Christodoulides C, Antoniades C, Koutsilieris M. Wnt signaling in cardiovascular physiology. *Trends Endocrinol Metab.* 2012;23:628–636.
- Shao JS, Cheng SL, Pingsterhaus JM, Charlton-Kachigian N, Loewy AP, Towler DA. Msx2 promotes cardiovascular calcification by activating paracrine Wnt signals. J Clin Invest. 2005;115:1210–1220.
- Borrell-Pages M, Romero JC, Juan-Babot O, Badimon L. Wnt pathway activation, cell migration, and lipid uptake is regulated by low-density lipoprotein receptor-related protein 5 in human macrophages. *Eur Heart J.* 2011;32:2841–2850.
- Tsaousi A, Mill C, George SJ. The Wnt pathways in vascular disease: lessons from vascular development. *Curr Opin Lipidol*. 2011;22:350–357.
- Tsai JS, Chen CY, Chen YL, Chuang LM. Rosiglitazone inhibits monocyte/ macrophage adhesion through de novo adiponectin production in human monocytes. J Cell Biochem. 2010;110:1410–1419.
- 9. Komiya Y, Habas R. Wnt signal transduction pathways. *Organogenesis*. 2008;4:68-75.
- Logan CY, Nusse R. The Wnt signaling pathway in development and disease. *Annu Rev Cell Dev Biol*. 2004;20:781–810.
- 11. George SJ. Wnt pathway: a new role in regulation of inflammation. *Arterioscler Thromb Vasc Biol.* 2008;28:400–402.
- Leopold JA, Loscalzo J. Clinical importance of understanding vascular biology. Cardiol Rev. 2000;8:115–123.
- Forstermann U, Munzel T. Endothelial nitric oxide synthase in vascular disease: from marvel to menace. *Circulation*. 2006;113:1708–1714.
- Skaria T, Schoedon G. Inflammatory Wnt5A signalling pathways affecting barrier function of human vascular endothelial cells. J Inflamm (London). 2017;14:15.
- Masckauchan TN, Shawber CJ, Funahashi Y, Li CM, Kitajewski J. Wnt/betacatenin signaling induces proliferation, survival and interleukin-8 in human endothelial cells. *Angiogenesis*. 2005;8:43–51.
- Dejana E. The role of Wnt signaling in physiological and pathological angiogenesis. *Circ Res.* 2010;107:943–952.
- Stepniak E, Radice GL, Vasioukhin V. Adhesive and signaling functions of cadherins and catenins in vertebrate development. *Cold Spring Harb Perspect Biol.* 2009;1:a002949.
- Heuberger J, Birchmeier W. Interplay of cadherin-mediated cell adhesion and canonical Wnt signaling. *Cold Spring Harb Perspect Biol.* 2010;2:a002915.

- Christman MA II, Goetz DJ, Dickerson E, McCall KD, Lewis CJ, Benencia F, Silver MJ, Kohn LD, Malgor R. Wnt5a is expressed in murine and human atherosclerotic lesions. *Am J Physiol Heart Circ Physiol*. 2008;294:H2864– H2870.
- Wang X, Xiao Y, Mou Y, Zhao Y, Blankesteijn WM, Hall JL. A role for the betacatenin/T-cell factor signaling cascade in vascular remodeling. *Circ Res.* 2002;90:340–347.
- Wang X, Adhikari N, Li Q, Hall JL. LDL receptor-related protein LRP6 regulates proliferation and survival through the Wnt cascade in vascular smooth muscle cells. Am J Physiol Heart Circ Physiol. 2004;287:H2376–H2383.
- Slater SC, Koutsouki E, Jackson CL, Bush RC, Angelini GD, Newby AC, George SJ. R-cadherin:beta-catenin complex and its association with vascular smooth muscle cell proliferation. *Arterioscler Thromb Vasc Biol.* 2004;24:1204–1210.
- Pena E, Arderiu G, Badimon L. Tissue factor induces human coronary artery smooth muscle cell motility through Wnt-signalling. J Thromb Haemost. 2013;11:1880–1891.
- Menini S, Iacobini C, Ricci C, Blasetti Fantauzzi C, Salvi L, Pesce CM, Relucenti M, Familiari G, Taurino M, Pugliese G. The galectin-3/RAGE dyad modulates vascular osteogenesis in atherosclerosis. *Cardiovasc Res.* 2013;100:472–480.
- Qin L, Hu R, Zhu N, Yao HL, Lei XY, Li SX, Liao DF, Zheng XL. The novel role and underlying mechanism of Wnt5a in regulating cellular cholesterol accumulation. *Clin Exp Pharmacol Physiol.* 2014;41:671–678.
- Carthy JM, Garmaroudi FS, Luo Z, McManus BM. Wnt3a induces myofibroblast differentiation by upregulating TGF-beta signaling through SMAD2 in a beta-catenin-dependent manner. *PLoS One*. 2011;6:e19809.
- Liu J, Wang Y, Pan Q, Su Y, Zhang Z, Han J, Zhu X, Tang C, Hu D. Wnt/betacatenin pathway forms a negative feedback loop during TGF-beta1 induced human normal skin fibroblast-to-myofibroblast transition. *J Dermatol Sci.* 2012;65:38–49.
- Virmani R, Kolodgie FD, Burke AP, Farb A, Schwartz SM. Lessons from sudden coronary death: a comprehensive morphological classification scheme for atherosclerotic lesions. *Arterioscler Thromb Vasc Biol.* 2000;20:1262–1275.
- Goldschmidt-Clermont PJ, Creager MA, Losordo DW, Lam GK, Wassef M, Dzau VJ. Atherosclerosis 2005: recent discoveries and novel hypotheses. *Circulation*. 2005;112:3348–3353.
- Tsaousi A, Williams H, Lyon CA, Taylor V, Swain A, Johnson JL, George SJ. Wnt4/beta-catenin signaling induces VSMC proliferation and is associated with intimal thickening. *Circ Res.* 2011;108:427–436.
- Oh DY, Olefsky JM. Medicine: Wnt fans the flames in obesity. Science. 2010;329:397–398.
- Stone GW, Maehara A, Lansky AJ, de Bruyne B, Cristea E, Mintz GS, Mehran R, McPherson J, Farhat N, Marso SP, Parise H, Templin B, White R, Zhang Z, Serruys PW. A prospective natural-history study of coronary atherosclerosis. *N Engl J Med.* 2011;364:226–235.
- Burke AP, Taylor A, Farb A, Malcom GT, Virmani R. Coronary calcification: insights from sudden coronary death victims. *Z Kardiol.* 2000;89(suppl 2):49– 53.
- Tavora F, Cresswell N, Li L, Ripple M, Fowler D, Burke A. Sudden coronary death caused by pathologic intimal thickening without atheromatous plaque formation. *Cardiovasc Pathol.* 2011;20:51–57.
- Hwang DS, Shin ES, Kim SJ, Lee JH, Kim JM, Lee SG. Early differential changes in coronary plaque composition according to plaque stability following statin initiation in acute coronary syndrome: classification and analysis by intravascular ultrasound-virtual histology. *Yonsei Med J.* 2013;54:336–344.
- Broedl UC, Lebherz C, Lehrke M, Stark R, Greif M, Becker A, von Ziegler F, Tittus J, Reiser M, Becker C, Goke B, Parhofer KG, Leber AW. Low adiponectin levels are an independent predictor of mixed and non-calcified coronary atherosclerotic plaques. *PLoS One*. 2009;4:e4733.
- Geng Y, Hsu JJ, Lu J, Ting TC, Miyazaki M, Demer LL, Tintut Y. Role of cellular cholesterol metabolism in vascular cell calcification. *J Biol Chem.* 2011;286:33701–33706.
- Demer LL, Tintut Y. Vascular calcification: pathobiology of a multifaceted disease. *Circulation*. 2008;117:2938–2948.
- Johnson RC, Leopold JA, Loscalzo J. Vascular calcification: pathobiological mechanisms and clinical implications. *Circ Res.* 2006;99:1044–1059.
- Chen NX, Moe SM. Vascular calcification: pathophysiology and risk factors. Curr Hypertens Rep. 2012;14:228–237.
- Leopold JA. Cellular mechanisms of aortic valve calcification. *Circ Cardiovasc Interv*. 2012;5:605–614.
- Wasilewski J, Mirota K, Wilczek K, Glowacki J, Polonski L. Calcific aortic valve damage as a risk factor for cardiovascular events. *Pol J Radiol.* 2012;77:30– 34.

- van Gils MJ, Bodde MC, Cremers LG, Dippel DW, van der Lugt A. Determinants of calcification growth in atherosclerotic carotid arteries: a serial multidetector CT angiography study. *Atherosclerosis*. 2013;227:95–99.
- 44. Tsuda H, Moritsuchi Y, Almeida FR, Lowe AA, Tsuda T. The relationship between cephalometric carotid artery calcification and Framingham Risk Score profile in patients with obstructive sleep apnea. *Sleep Breath*. 2013;17:1003–1008.
- Shames S, Gillam LD. Sex differences in aortic valve calcification. Circ Cardiovasc Imaging. 2013;6:8–10.
- Rajamannan NM, Subramaniam M, Caira F, Stock SR, Spelsberg TC. Atorvastatin inhibits hypercholesterolemia-induced calcification in the aortic valves via the Lrp5 receptor pathway. *Circulation*. 2005;112:1229–1234.
- Semenov MV, Habas R, Macdonald BT, He X. SnapShot: noncanonical Wnt signaling pathways. *Cell*. 2007;131:1378.
- Shao JS, Cai J, Towler DA. Molecular mechanisms of vascular calcification: lessons learned from the aorta. *Arterioscler Thromb Vasc Biol.* 2006;26:1423–1430.
- Virchow R. Cellular pathology: as based upon physiological and pathological histology: lecture XVI—atheromatous affection of arteries: 1858. *Nutr Rev.* 1989;47:23–25.
- Qasim AN, Rafeek H, Rasania SP, Churchill TW, Yang W, Ferrari VA, Jha S, Master SM, Mulvey CK, Terembula K, Dailing C, Budoff MJ, Kawut SM, Reilly MP. Cardiovascular risk factors and mitral annular calcification in type 2 diabetes. *Atherosclerosis*. 2013;226:419–424.
- Nasir K, Katz R, Takasu J, Shavelle DM, Detrano R, Lima JA, Blumenthal RS, O'Brien K, Budoff MJ. Ethnic differences between extra-coronary measures on cardiac computed tomography: Multi-Ethnic Study of Atherosclerosis (MESA). *Atherosclerosis*. 2008;198:104–114.
- Askevold ET, Gullestad L, Aakhus S, Ranheim T, Tonnessen T, Solberg OG, Aukrust P, Ueland T. Secreted Wnt modulators in symptomatic aortic stenosis. J Am Heart Assoc. 2012;1:e002261. DOI: 10.1161/JAHA.112.002261.
- Poggio P, Grau JB, Field BC, Sainger R, Seefried WF, Rizzolio F, Ferrari G. Osteopontin controls endothelial cell migration in vitro and in excised human valvular tissue from patients with calcific aortic stenosis and controls. *J Cell Physiol.* 2011;226:2139–2149.
- Alexopoulos A, Kaoukis A, Papadaki H, Pyrgakis V. Pathophysiologic mechanisms of calcific aortic stenosis. *Ther Adv Cardiovasc Dis.* 2012;6:71–80.
- 55. Tyson KL, Reynolds JL, McNair R, Zhang Q, Weissberg PL, Shanahan CM. Osteo/chondrocytic transcription factors and their target genes exhibit distinct patterns of expression in human arterial calcification. *Arterioscler Thromb Vasc Biol.* 2003;23:489–494.
- Bos D, Ikram MA, Isaacs A, Verhaaren BF, Hofman A, van Duijn CM, Witteman JC, van der Lugt A, Vernooij MW. Genetic loci for coronary calcification and serum lipids relate to aortic and carotid calcification. *Circ Cardiovasc Genet*. 2013;6:47–53.
- Cagirci G, Cay S, Canga A, Karakurt O, Yazihan N, Kilic H, Topaloglu S, Aras D, Demir AD, Akdemir R. Association between plasma asymmetrical dimethylarginine activity and severity of aortic valve stenosis. *J Cardiovasc Med* (*Hagerstown*). 2011;12:96–101.
- Awan Z, Denis M, Bailey D, Giaid A, Prat A, Goltzman D, Seidah NG, Genest J. The LDLR deficient mouse as a model for aortic calcification and quantification by micro-computed tomography. *Atherosclerosis*. 2011;219:455–462.
- Liu Y, Shanahan CM. Signalling pathways and vascular calcification. Front Biosci (Landmark Ed). 2011;16:1302–1314.
- Sallam T, Cheng H, Demer LL, Tintut Y. Regulatory circuits controlling vascular cell calcification. *Cell Mol Life Sci.* 2013;70:3187–3197.
- Cheng SL, Shao JS, Halstead LR, Distelhorst K, Sierra O, Towler DA. Activation of vascular smooth muscle parathyroid hormone receptor inhibits Wnt/beta-catenin signaling and aortic fibrosis in diabetic arteriosclerosis. *Circ Res.* 2010;107:271–282.
- Towler DA, Bidder M, Latifi T, Coleman T, Semenkovich CF. Diet-induced diabetes activates an osteogenic gene regulatory program in the aortas of low density lipoprotein receptor-deficient mice. J Biol Chem. 1998;273:30427–30434.
- Liu TM, Lee EH. Transcriptional regulatory cascades in Runx2-dependent bone development. *Tissue Eng Part B Rev.* 2013;19:254–263.
- 64. Tompkins KA. Wnt proteins in mineralized tissue development and homeostasis. *Connect Tissue Res.* 2011;52:448–458.
- de Crombrugghe B, Lefebvre V, Nakashima K. Regulatory mechanisms in the pathways of cartilage and bone formation. *Curr Opin Cell Biol.* 2001;13:721– 727.
- Anderson HC. Molecular biology of matrix vesicles. *Clin Orthop Relat Res.* 1995;5:266–280.

- Golub EE. Biomineralization and matrix vesicles in biology and pathology. Semin Immunopathol. 2011;33:409–417.
- Shao JS, Aly ZA, Lai CF, Cheng SL, Cai J, Huang E, Behrmann A, Towler DA. Vascular Bmp Msx2 Wnt signaling and oxidative stress in arterial calcification. Ann N Y Acad Sci. 2007;1117:40–50.
- Li YL, Xiao ZS. Advances in Runx2 regulation and its isoforms. *Med Hypotheses*. 2007;68:169–175.
- Liberman M, Johnson RC, Handy DE, Loscalzo J, Leopold JA. Bone morphogenetic protein-2 activates NADPH oxidase to increase endoplasmic reticulum stress and human coronary artery smooth muscle cell calcification. *Biochem Biophys Res Commun.* 2011;413:436–441.
- Wodarz A, Nusse R. Mechanisms of Wnt signaling in development. Annu Rev Cell Dev Biol. 1998;14:59–88.
- Schulte G, Bryja V. The Frizzled family of unconventional G-protein-coupled receptors. Trends Pharmacol Sci. 2007;28:518–525.
- He X, Semenov M, Tamai K, Zeng X. LDL receptor-related proteins 5 and 6 in Wnt/beta-catenin signaling: arrows point the way. *Development*. 2004;131:1663–1677.
- Veeman MT, Axelrod JD, Moon RT. A second canon: functions and mechanisms of beta-catenin-independent Wnt signaling. *Dev Cell*. 2003;5:367–377.
- Zeng L, Fagotto F, Zhang T, Hsu W, Vasicek TJ, Perry WL III, Lee JJ, Tilghman SM, Gumbiner BM, Costantini F. The mouse Fused locus encodes Axin, an inhibitor of the Wnt signaling pathway that regulates embryonic axis formation. *Cell*. 1997;90:181–192.
- 76. Gordon MD, Nusse R. Wnt signaling: multiple pathways, multiple receptors, and multiple transcription factors. J Biol Chem. 2006;281:22429–22433.
- 77. Nusse R. Wnt signaling. Cold Spring Harb Perspect Biol. 2012;4:a011163.
- Hatsell S, Rowlands T, Hiremath M, Cowin P. Beta-catenin and Tcfs in mammary development and cancer. J Mammary Gland Biol Neoplasia. 2003;8: 145–158.
- Brunner E, Peter O, Schweizer L, Basler K. Pangolin encodes a Lef-1 homologue that acts downstream of Armadillo to transduce the Wingless signal in Drosophila. *Nature*. 1997;385:829–833.
- van de Wetering M, Cavallo R, Dooijes D, van Beest M, van Es J, Loureiro J, Ypma A, Hursh D, Jones T, Bejsovec A, Peifer M, Mortin M, Clevers H. Armadillo coactivates transcription driven by the product of the Drosophila segment polarity gene dTCF. *Cell.* 1997;88:789–799.
- Grumolato L, Liu G, Mong P, Mudbhary R, Biswas R, Arroyave R, Vijayakumar S, Economides AN, Aaronson SA. Canonical and noncanonical Wrts use a common mechanism to activate completely unrelated coreceptors. *Genes Dev.* 2010;24:2517–2530.
- Weidinger G, Moon RT. When Wnts antagonize Wnts. J Cell Biol. 2003;162:753–755.
- De A. Wnt/Ca2+ signaling pathway: a brief overview. Acta Biochim Biophys Sin. 2011;43:745–756.
- Kuhl M, Sheldahl LC, Malbon CC, Moon RT. Ca(2+)/calmodulin-dependent protein kinase II is stimulated by Wnt and Frizzled homologs and promotes ventral cell fates in Xenopus. J Biol Chem. 2000;275:12701–12711.
- Sheldahl LC, Park M, Malbon CC, Moon RT. Protein kinase C is differentially stimulated by Wnt and Frizzled homologs in a G-protein-dependent manner. *Curr Biol.* 1999;9:695–698.
- Hogan PG, Chen L, Nardone J, Rao A. Transcriptional regulation by calcium, calcineurin, and NFAT. *Genes Dev.* 2003;17:2205–2232.
- Lee DK, Nathan Grantham R, Trachte AL, Mannion JD, Wilson CL. Activation of the canonical Wnt/beta-catenin pathway enhances monocyte adhesion to endothelial cells. *Biochem Biophys Res Commun.* 2006;347:109–116.
- Kim J, Kim J, Kim DW, Ha Y, Ihm MH, Kim H, Song K, Lee I. Wnt5a induces endothelial inflammation via beta-catenin-independent signaling. *J Immunol.* 2010;185:1274–1282.
- Ueland T, Otterdal K, Lekva T, Halvorsen B, Gabrielsen A, Sandberg WJ, Paulsson-Berne G, Pedersen TM, Folkersen L, Gullestad L, Oie E, Hansson GK, Aukrust P. Dickkopf-1 enhances inflammatory interaction between platelets and endothelial cells and shows increased expression in atherosclerosis. *Arterioscler Thromb Vasc Biol.* 2009;29:1228–1234.
- Takahashi J, Orcholski M, Yuan K, de Jesus Perez V. PDGF-dependent betacatenin activation is associated with abnormal pulmonary artery smooth muscle cell proliferation in pulmonary arterial hypertension. *FEBS Lett.* 2016;590:101–109.
- Williams H, Mill CA, Monk BA, Hulin-Curtis S, Johnson JL, George SJ. Wnt2 and WISP-1/CCN4 induce intimal thickening via promotion of smooth muscle cell migration. *Arterioscler Thromb Vasc Biol.* 2016;36:1417–1424.

- Krishna SM, Seto SW, Jose RJ, Li J, Morton SK, Biros E, Wang Y, Nsengiyumva V, Lindeman JH, Loots GG, Rush CM, Craig JM, Golledge J. Wnt signaling pathway inhibitor sclerostin inhibits angiotensin II-induced aortic aneurysm and atherosclerosis. *Arterioscler Thromb Vasc Biol.* 2017;37:553–566.
- Bostrom K, Watson KE, Horn S, Wortham C, Herman IM, Demer LL. Bone morphogenetic protein expression in human atherosclerotic lesions. *J Clin Invest.* 1993;91:1800–1809.
- Cola C, Almeida M, Li D, Romeo F, Mehta JL. Regulatory role of endothelium in the expression of genes affecting arterial calcification. *Biochem Biophys Res Commun.* 2004;320:424–427.
- Zhang L, Zalewski A, Liu Y, Mazurek T, Cowan S, Martin JL, Hofmann SM, Vlassara H, Shi Y. Diabetes-induced oxidative stress and low-grade inflammation in porcine coronary arteries. *Circulation*. 2003;108:472–478.
- Cheng SL, Shao JS, Charlton-Kachigian N, Loewy AP, Towler DA. MSX2 promotes osteogenesis and suppresses adipogenic differentiation of multipotent mesenchymal progenitors. *J Biol Chem.* 2003;278:45969–45977.
- Satokata I, Ma L, Ohshima H, Bei M, Woo I, Nishizawa K, Maeda T, Takano Y, Uchiyama M, Heaney S, Peters H, Tang Z, Maxson R, Maas R. Msx2 deficiency in mice causes pleiotropic defects in bone growth and ectodermal organ formation. *Nat Genet*. 2000;24:391–395.
- Goodwin AM, D'Amore PA. Wnt signaling in the vasculature. *Angiogenesis*. 2002;5:1–9.
- Mao B, Wu W, Davidson G, Marhold J, Li M, Mechler BM, Delius H, Hoppe D, Stannek P, Walter C, Glinka A, Niehrs C. Kremen proteins are Dickkopf receptors that regulate Wnt/beta-catenin signalling. *Nature*. 2002;417:664–667.
- Hussein SM, Duff EK, Sirard C. Smad4 and beta-catenin co-activators functionally interact with lymphoid-enhancing factor to regulate graded expression of Msx2. J Biol Chem. 2003;278:48805–48814.
- Willert J, Epping M, Pollack JR, Brown PO, Nusse R. A transcriptional response to Wnt protein in human embryonic carcinoma cells. *BMC Dev Biol.* 2002;2:8.
- 102. Kirton JP, Crofts NJ, George SJ, Brennan K, Canfield AE. Wnt/beta-catenin signaling stimulates chondrogenic and inhibits adipogenic differentiation of pericytes: potential relevance to vascular disease? *Circ Res.* 2007;101:581– 589.
- Andreeva ER, Pugach IM, Gordon D, Orekhov AN. Continuous subendothelial network formed by pericyte-like cells in human vascular bed. *Tissue Cell*. 1998;30:127–135.
- Liu Y, Wilkinson FL, Kirton JP, Jeziorska M, Iizasa H, Sai Y, Nakashima E, Heagerty AM, Canfield AE, Alexander MY. Hepatocyte growth factor and c-Met expression in pericytes: implications for atherosclerotic plaque development. J Pathol. 2007;212:12–19.
- Collett GD, Canfield AE. Angiogenesis and pericytes in the initiation of ectopic calcification. *Circ Res.* 2005;96:930–938.
- 106. Kirton JP, Wilkinson FL, Canfield AE, Alexander MY. Dexamethasone downregulates calcification-inhibitor molecules and accelerates osteogenic differentiation of vascular pericytes: implications for vascular calcification. *Circ Res.* 2006;98:1264–1272.
- Doherty MJ, Ashton BA, Walsh S, Beresford JN, Grant ME, Canfield AE. Vascular pericytes express osteogenic potential in vitro and in vivo. J Bone Miner Res. 1998;13:828–838.
- Farrington-Rock C, Crofts NJ, Doherty MJ, Ashton BA, Griffin-Jones C, Canfield AE. Chondrogenic and adipogenic potential of microvascular pericytes. *Circulation*. 2004;110:2226–2232.
- Tintut Y, Alfonso Z, Saini T, Radcliff K, Watson K, Bostrom K, Demer LL. Multilineage potential of cells from the artery wall. *Circulation*. 2003;108:2505–2510.
- 110. Kengaku M, Capdevila J, Rodriguez-Esteban C, De La Pena J, Johnson RL, Izpisua Belmonte JC, Tabin CJ. Distinct WNT pathways regulating AER formation and dorsoventral polarity in the chick limb bud. *Science*. 1998;280:1274–1277.
- 111. Kawakami Y, Capdevila J, Buscher D, Itoh T, Rodriguez Esteban C, Izpisua Belmonte JC. WNT signals control FGF-dependent limb initiation and AER induction in the chick embryo. *Cell*. 2001;104:891–900.
- 112. Gong Y, Slee RB, Fukai N, Rawadi G, Roman-Roman S, Reginato AM, Wang H, Cundy T, Glorieux FH, Lev D, Zacharin M, Oexle K, Marcelino J, Suwairi W, Heeger S, Sabatakos G, Apte S, Adkins WN, Allgrove J, Arslan-Kirchner M, Batch JA, Beighton P, Black GC, Boles RG, Boon LM, Borrone C, Brunner HG, Carle GF, Dallapiccola B, De Paepe A, Floege B, Halfhide ML, Hall B, Hennekam RC, Hirose T, Jans A, Juppner H, Kim CA, Keppler-Noreuil K, Kohlschuetter A, LaCombe D, Lambert M, Lemyre E, Letteboer T, Peltonen L, Ramesar RS, Romanengo M, Somer H, Steichen-Gersdorf E, Steinmann B, Sullivan B, Superti-Furga A, Swoboda W, van den Boogaard MJ, Van Hul W, Vikkula M, Votruba M, Zabel B, Garcia T, Baron R, Olsen BR, Warman ML. LDL receptor-related protein 5 (LRP5) affects bone accrual and eye development. *Cell.* 2001;107:513–523.

- 113. Kubota T, Michigami T, Ozono K. Wnt signaling in bone metabolism. J Bone Miner Metab. 2009;27:265–271.
- 114. Kato M, Patel MS, Levasseur R, Lobov I, Chang BH, Glass DA II, Hartmann C, Li L, Hwang TH, Brayton CF, Lang RA, Karsenty G, Chan L. Cbfa1-independent decrease in osteoblast proliferation, osteopenia, and persistent embryonic eye vascularization in mice deficient in Lrp5, a Wnt coreceptor. *J Cell Biol.* 2002;157:303–314.
- 115. Holmen SL, Giambernardi TA, Zylstra CR, Buckner-Berghuis BD, Resau JH, Hess JF, Glatt V, Bouxsein ML, Ai M, Warman ML, Williams BO. Decreased BMD and limb deformities in mice carrying mutations in both Lrp5 and Lrp6. J Bone Miner Res. 2004;19:2033–2040.
- Rajamannan NM. The role of Lrp5/6 in cardiac valve disease: experimental hypercholesterolemia in the ApoE-/-/Lrp5-/- mice. J Cell Biochem. 2011;112:2987–2991.
- 117. Boyden LM, Mao J, Belsky J, Mitzner L, Farhi A, Mitnick MA, Wu D, Insogna K, Lifton RP. High bone density due to a mutation in LDL-receptor-related protein 5. N Engl J Med. 2002;346:1513–1521.
- 118. Little RD, Carulli JP, Del Mastro RG, Dupuis J, Osborne M, Folz C, Manning SP, Swain PM, Zhao SC, Eustace B, Lappe MM, Spitzer L, Zweier S, Braunschweiger K, Benchekroun Y, Hu X, Adair R, Chee L, FitzGerald MG, Tulig C, Caruso A, Tzellas N, Bawa A, Franklin B, McGuire S, Nogues X, Gong G, Allen KM, Anisowicz A, Morales AJ, Lomedico PT, Recker SM, Van Eerdewegh P, Recker RR, Johnson ML. A mutation in the LDL receptor-related protein 5 gene results in the autosomal dominant high-bone-mass trait. *Am J Hum Genet*. 2002;70:11–19.
- Babij P, Zhao W, Small C, Kharode Y, Yaworsky PJ, Bouxsein ML, Reddy PS, Bodine PV, Robinson JA, Bhat B, Marzolf J, Moran RA, Bex F. High bone mass in mice expressing a mutant LRP5 gene. J Bone Miner Res. 2003;18:960–974.
- 120. Van Wesenbeeck L, Cleiren E, Gram J, Beals RK, Benichou O, Scopelliti D, Key L, Renton T, Bartels C, Gong Y, Warman ML, De Vernejoul MC, Bollerslev J, Van Hul W. Six novel missense mutations in the LDL receptor-related protein 5 (LRP5) gene in different conditions with an increased bone density. *Am J Hum Genet.* 2003;72:763–771.
- 121. Mani A, Radhakrishnan J, Wang H, Mani A, Mani MA, Nelson-Williams C, Carew KS, Mane S, Najmabadi H, Wu D, Lifton RP. LRP6 mutation in a family with early coronary disease and metabolic risk factors. *Science*. 2007;315:1278–1282.
- Hu H, Hilton MJ, Tu X, Yu K, Ornitz DM, Long F. Sequential roles of Hedgehog and Wnt signaling in osteoblast development. *Development*. 2005;132:49– 60.
- 123. Takada I, Mihara M, Suzawa M, Ohtake F, Kobayashi S, Igarashi M, Youn MY, Takeyama K, Nakamura T, Mezaki Y, Takezawa S, Yogiashi Y, Kitagawa H, Yamada G, Takada S, Minami Y, Shibuya H, Matsumoto K, Kato S. A histone lysine methyltransferase activated by non-canonical Wnt signaling suppresses PPAR-gamma transactivation. *Nat Cell Biol.* 2007;9:1273–1285.
- 124. Blumenthal A, Ehlers S, Lauber J, Buer J, Lange C, Goldmann T, Heine H, Brandt E, Reiling N. The Wingless homolog WNT5A and its receptor Frizzled-5 regulate inflammatory responses of human mononuclear cells induced by microbial stimulation. *Blood*. 2006;108:965–973.
- Katoh M, Katoh M. Comparative genomics on Wnt5a and Wnt5b genes. Int J Mol Med. 2005;15:749–753.
- 126. Hsu YH, Kiel DP. Clinical review: genome-wide association studies of skeletal phenotypes: what we have learned and where we are headed. J Clin Endocrinol Metab. 2012;97:E1958–E1977.
- 127. Yang Y, Topol L, Lee H, Wu J. Wnt5a and Wnt5b exhibit distinct activities in coordinating chondrocyte proliferation and differentiation. *Development*. 2003;130:1003–1015.
- 128. Kanazawa A, Tsukada S, Kamiyama M, Yanagimoto T, Nakajima M, Maeda S. Wht5b partially inhibits canonical wht/beta-catenin signaling pathway and promotes adipogenesis in 3T3-L1 preadipocytes. *Biochem Biophys Res Commun.* 2005;330:505–510.
- 129. van Tienen FH, Laeremans H, van der Kallen CJ, Smeets HJ. Wnt5b stimulates adipogenesis by activating PPARgamma, and inhibiting the beta-catenin dependent Wnt signaling pathway together with Wnt5a. *Biochem Biophys Res Commun.* 2009;387:207–211.
- Zandbergen F, Plutzky J. PPARalpha in atherosclerosis and inflammation. Biochem Biophys Acta. 2007;1771:972–982.
- Hamblin M, Chang L, Fan Y, Zhang J, Chen YE. PPARs and the cardiovascular system. *Antioxid Redox Signal*. 2009;11:1415–1452.

- 132. Berger J, Moller DE. The mechanisms of action of PPARs. Annu Rev Med. 2002;53:409-435.
- Verma S, Szmitko PE. The vascular biology of peroxisome proliferatoractivated receptors: modulation of atherosclerosis. *Can J Cardiol.* 2006;22 (suppl B):12b–17b.
- Ricote M, Huang JT, Welch JS, Glass CK. The peroxisome proliferatoractivated receptor(PPARgamma) as a regulator of monocyte/macrophage function. J Leukoc Biol. 1999;66:733–739.
- Soskic SS, Dobutovic BD, Sudar EM, Obradovic MM, Nikolic DM, Zaric BL, Stojanovic SD, Stokic EJ, Mikhailidis DP, Isenovic ER. Peroxisome proliferator-activated receptors and atherosclerosis. *Angiology*. 2011;62:523–534.
- 136. Marx N, Sukhova G, Murphy C, Libby P, Plutzky J. Macrophages in human atheroma contain PPARgamma: differentiation-dependent peroxisomal proliferator-activated receptor gamma(PPARgamma) expression and reduction of MMP-9 activity through PPARgamma activation in mononuclear phagocytes in vitro. *Am J Pathol.* 1998;153:17–23.
- Jiang C, Ting AT, Seed B. PPAR-gamma agonists inhibit production of monocyte inflammatory cytokines. *Nature*. 1998;391:82–86.
- Lako M, Strachan T, Bullen P, Wilson DI, Robson SC, Lindsay S. Isolation, characterisation and embryonic expression of WNT11, a gene which maps to 11q13.5 and has possible roles in the development of skeleton, kidney and lung. *Gene*. 1998;219:101–110.
- 139. Johnson ML, Gong G, Kimberling W, Recker SM, Kimmel DB, Recker RB. Linkage of a gene causing high bone mass to human chromosome 11 (11q12-13). Am J Hum Genet. 1997;60:1326–1332.
- Boland GM, Perkins G, Hall DJ, Tuan RS. Wnt 3a promotes proliferation and suppresses osteogenic differentiation of adult human mesenchymal stem cells. J Cell Biochem. 2004;93:1210–1230.
- 141. Tao Q, Yokota C, Puck H, Kofron M, Birsoy B, Yan D, Asashima M, Wylie CC, Lin X, Heasman J. Maternal Wnt11 activates the canonical Wnt signaling pathway required for axis formation in Xenopus embryos. *Cell*. 2005;120:857–871.
- 142. Pandur P, Lasche M, Eisenberg LM, Kuhl M. Wnt-11 activation of a noncanonical Wnt signalling pathway is required for cardiogenesis. *Nature*. 2002;418:636–641.
- 143. Friedman MS, Oyserman SM, Hankenson KD. Wnt11 promotes osteoblast maturation and mineralization through R-spondin 2. J Biol Chem. 2009;284:14117–14125.
- 144. Sekiya I, Vuoristo JT, Larson BL, Prockop DJ. In vitro cartilage formation by human adult stem cells from bone marrow stroma defines the sequence of cellular and molecular events during chondrogenesis. *Proc Natl Acad Sci U S A*. 2002;99:4397–4402.
- 145. Caira FC, Stock SR, Gleason TG, McGee EC, Huang J, Bonow RO, Spelsberg TC, McCarthy PM, Rahimtoola SH, Rajamannan NM. Human degenerative valve disease is associated with up-regulation of low-density lipoprotein receptor-related protein 5 receptor-mediated bone formation. J Am Coll Cardiol. 2006;47:1707–1712.
- 146. Albanese I, Yu B, Al-Kindi H, Barratt B, Ott L, Al-Refai M, de Varennes B, Shum-Tim D, Cerruti M, Gourgas O, Rheaume E, Tardif JC, Schwertani A. Role of noncanonical Wnt signaling pathway in human aortic valve calcification. *Arterioscler Thromb Vasc Biol.* 2017;37:543–552.
- 147. Yu B, Anouar H, George T, Leah O, Nial F, Marta C, Gourgas O, Shum-Tim D, Al-Kindi H, de Varennes B, Alsheikh-Ali A, Genest J, Schwertani A. Lipoprotein (a) induces human aortic valve interstitial cell calcification. *JACC Basic Transl Sci.* 2017;2:358–371.
- 148. Kahn M. Can we safely target the WNT pathway? *Nat Rev Drug Discov.* 2014;13:513–532.
- 149. Takebe N, Miele L, Harris PJ, Jeong W, Bando H, Kahn M, Yang SX, Ivy SP. Targeting Notch, Hedgehog, and Wnt pathways in cancer stem cells: clinical update. *Nat Rev Clin Oncol.* 2015;12:445–464.
- Eldar-Finkelman H, Martinez A. GSK-3 inhibitors: preclinical and clinical focus on CNS. Front Mol Neurosci. 2011;4:32.
- 151. Larsson S. Anti-sclerostin: is there an indication? Injury. 2016;47(suppl 1): S31–S35.

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