Antipsychotics preserve telomere length in peripheral blood mononuclear cells after acute oxidative stress injury

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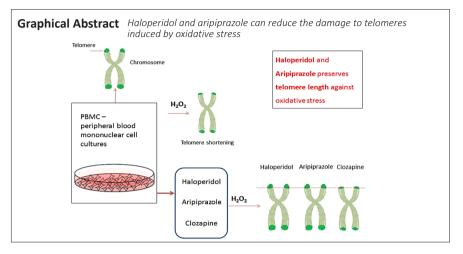
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Abstract

Antipsychotics may prolong or retain telomere length, affect mitochondrial function, and then affect the metabolism of nerve cells. To validate the hypothesis that antipsychotics can prolong telomere length after oxidative stress injury, leukocytes from healthy volunteers were extracted using FicoII-Histopaque density gradient. The mononuclear cells layer was resuspended in cell culture medium. Oxidative stress was induced with hydrogen peroxide in cultured leukocytes. Four days later, leukocytes were treated with aripiprazole, haloperidol or clozapine for 7 days. Real-time PCR revealed that treatments with aripiprazole and haloperidol increased the telomere length by 23% and 20% in peripheral blood mononuclear cells after acute oxidative stress injury. These results suggest that haloperidol and aripiprazole can reduce the damage to telomeres induced by oxidative stress. The experiment procedure was approved by the Ethics Committee of Faculty of Medicine of the University of São Paulo (FMUSP/CAAE approval No. 52622616.8.0000.0065).

Key Words: antipsychotic; aripiprazole; cell culture; clozapine; haloperidol; oxidative stress; peripheral blood mononuclear cell; telomere length

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Introduction

Telomeres are tandem repeats of the non-coding sequence TTAGGG at the end of each chromosome. Human telomeres protect the DNA from end-to-end fusion and cellular erosion, but they diminish progressively with cellular division (Manchia et al., 2020). In counterpart, telomerase is the enzyme responsible for adding new TTAGGG sequences at the end of telomeres, so that telomere erosion does not occur prematurely. Some people consider telomeres are "biological clocks", since the cell becomes senescent when they reach a critical limit (Savage, 2018). Interestingly, several conditions have been related to decreased telomere length, including cardiovascular (Yeh and Wang, 2016), neurologic (Eitan et al., 2014), and some psychiatric diseases (Lindqvist et al., 2015), probably due to chronic inflammation and oxidative stress.

Antipsychotics consist of a class of drugs used for the treatment of several neuropsychiatric conditions, including schizophrenia. The exact mechanism of action of antipsychotics is still debated and each antipsychotic has its own pharmacological features, largely represented by dopamine (DA) and serotonin (5-HT) receptor antagonism at different degrees of affinity and specificity (Farlow and Shamliyan, 2017).

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There is limited evidence that antipsychotic drugs may influence telomere dynamics. Aoyama et al. (2014) showed that atypical antipsychotics elongated telomere length in the hippocampus of mice. Conversely, Monroy-Jaramillo et al. (2017) showed decreased telomere length in leukocytes of schizophrenic patients treated with olanzapine. Yu et al. (2008) found longer telomeres in schizophrenic patients who had a good response to pharmacological treatment, but Li et al. (2015) showed that patients who responded to antipsychotic treatment had longer telomeres at baseline, in which case telomere length was a predictor of good response, not directly affected by the antipsychotic treatment itself. In an ex vivo study using cultures of peripheral blood mononuclear cells (PBMC) drawn from schizophrenic patients, haloperidol and clozapine administered for 5 days at therapeutic concentrations did not modify telomerase expression (Porton et al., 2008). In view of the conflicting reports in the literature regarding the possible influence of antipsychotics on telomere length, we designed the present study to determine whether the effect of antipsychotic drugs on telomere length may be related to the occurrence of oxidative stress. We hypothesize that the deleterious effect of oxidative stress on telomeres may be attenuated by treatment with clozapine, aripiprazole and haloperidol.

Materials and Methods

Peripheral blood mononuclear cell culture

The experiment procedure was approved by the Ethics Committee of Faculty of Medicine of University of São Paulo (FMUSP/CAAE approval No. 52622616.8.0000.0065) and conformed to the guidelines of the Declaration of Helsinki. All participants filled in the informed consent. Peripheral blood was collected from healthy volunteers (without previous psychiatric disorder and previous blood disorders and nonsmokers) for the isolation of peripheral blood mononuclear cells (PBMCs). Blood was collected (14 mL) in heparinized tubes and diluted 1:1 in sterile phosphate buffered saline (PBS). The diluted blood was carefully transferred to a conic tube containing 6 mL of Ficoll (Sigma, Sigma, St. Louis, MO, USA). The mixture was centrifuged at 700 \times q for 30 minutes at room temperature. The mononuclear cell layer was removed and centrifuged with 15 mL of sterile PBS at $700 \times g$ for 30 minutes at room temperature, for removal of contaminant platelets. The supernatant was discarded and the cell pellet was suspended with 1 mL of RPMI 1640 medium (Thermo Fisher Scientific, Waltham, MA, USA). Cell count was performed with a Neubauer chamber. For each well, we

diluted the cells to a final concentration of 5×10^5 cells/mL.

Antipsychotic preparation

The antipsychotics were diluted in the following way: haloperidol (Sigma): 2.5 mg in 1.2 mL of DMSO (Sigma); clozapine (Biogen, Cambridge, MA, USA): 25 mg in 1 mL DMSO; aripiprazole (Sigma): 2.5 mg in 500 μ L PBS. After solubilization, the clozapine and aripiprazole samples were diluted to 100 μ g/mL and the haloperidol sample to 10 μ g/mL in RPMI 1640 culture medium (Thermo fisher, USA) and stored at -20°C. The final concentration of dimethyl sulfoxide (DMSO, as vehicle) was \leq 0.02% (de Abreu Costa et al., 2017).

PBMC culture maintenance, hydrogen peroxide and antipsychotic treatment

The PBMCs were maintained in a 6-well dish, in RPMI 1640 medium supplemented with 10% bovine fetal serum (Sigma) and antibiotics (penicillin/streptomycin 1% v/v) at 37°C and an atmosphere with 5% of CO₂. To cause initial telomeric damage, we tested different hydrogen peroxide (H_2O_2) (Sigma) concentrations: 100, 200, 400, and 600 μ M. The cultures with H_2O_2 were maintained for 3 days in the same conditions described.

For each individual culture of PBMCs, we induced oxidative stress with 200 μ M of H₂O₂ for 3 days, after which the distinct antipsychotics were added to the culture medium for additional 4 days. We used working concentrations of antipsychotic drugs within the range of plasma therapeutic levels in three experimental (intervention) groups: (a) haloperidol 7 ng/mL (Van Putten et al., 1992); (b) clozapine 1000 ng/mL (Ulrich et al., 2003); (c) aripiprazole 125 ng/mL (McEvoy et al., 2014). Two comparison groups [controls and vehicle (DMSO \leq 0.02%)] were established: (d) PBMCs maintained for 7 days in culture media only (no H_2O_2 and no antipsychotics), presumably accounting for baseline in vitro effect (no/minimal undisturbed change in telomere length); and (e) PBMCs treated with H_2O_2 for 3 days but no antipsychotics added between days 4 and 7 (negative control), indicative of the effect of oxidative stress on telomeres).

Cell viability

The PBMC viability was assessed according to MTT ((3-(4, 5-dimethylthiazolyl-2)-2,5-diphenyltetrazolium bromide) (Sigma) protocol (De-Paula et al., 2016). MTT was reduced by active cells, generating purple formazan, which can be quantified by spectrophotometry.

For each sample, a 100 μ L aliquot was transferred to a 96-well dish, in triplicate and added 10 μ L of MTT reagent was added. The cells were incubated for 6 hours and 100 μ L of sodium dodecyl sulfate (SDS) 20% was added to each well. Then, the absorbance at 570 nm was recorded using a microplate reader (BioLab Inc., Lawrenceville, GA, USA).

DNA extraction and quality evaluation

The PBMCs were harvested and stored after the experiment in a refrigerator at -20°C. DNA was extracted using QIAamp DNA Mini Blood kit (QIAGEN, CA, USA) with QIAcube protocol. After extraction, DNA was diluted to 4 ng/mL and kept at -80°C. DNA quality was assessed with spectrophotometry (Nanodrop - Thermo Fisher Scientific, CA, USA). We also performed capillary electrophoresis (Fragment Analyzer – Advanced Analytical Technologies, CA, USA) to verify DNA integrity.

Telomere length measurement – real-time polymerase chain reaction

Telomere length in PBMCs was measured using the methods described previously (Cawthon, 2009), with some

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modifications. We used 36B4 for reference as a single copy gene. The primer sequences (5'–3') were: telomere forward: GGT TTT TGA GGG TGA GGG TGA GGG TGA GGG TGA GGG TGA GGG T; telomere reverse: TCC CGA CTA TCC CTA TCC CTA TCC CTA TCC CTA TCC CTA; 36B4 forward/reverse CAG CAA GTG GGA AGG TGT AAT CC; 36B4 forward/reverse CCC ATT CTA TCA TCA ACG GGT ACA A. The 36B4 reaction included 7.5 μ L of SyBr Green PCR MasterMix (Promega, UK), 0.45 μ L of primer forward, 0.75 μ L of primer reverse and 3.3 μ L of miliQ water. For telomere reaction: 7.5 μ L of SyBr Green PCR MasterMix (Promeda, 0.45 μ L of primer reverse and 3 μ L of miliQ water.

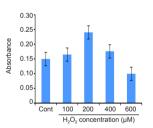
The real-time polymerase chain reaction (PCR) was run with 7500 Fast Real Time PCR System (Promega, CA, USA) at 50°C for 2 minutes, then 40 cycles of 95°C for 2 minutes, 55°C for 15 seconds and 60°C for 1 minute.

Statistical analysis

Statistical analysis was conducted using software R (version 3.2.0). For telomere length quantification, we used the ratio (average Ct telomere/average Ct 36B4) (T/S ratio). Differences in TL between control and vehicle within each treatment were investigated with one-way analysis of variance (ANOVA) followed by Tukey's *post hoc* test. The Turkey's package (*post hoc* analysis) was used in the R software. Statistical differences were considered significant if adjusted P < 0.05.

Results

After 3 days of treatment with different H_2O_2 concentrations, we harvested the cells to perform cell viability analysis and telomere mensuration. ANOVA test showed there was no difference among groups (P = 0.051). As observed, we found the tendency of diminished telomeres after H_2O_2 treatment, more pronounced after 200 μ M H_2O_2 treatment. In the light of these results, we opted for the use of concentration 200 μ M (**Figure 1**).



 $\label{eq:Figure 1} Figure 1 \mid Average cell viability (MTT absorbance at 570 nm) after treatment with different concentrations of hydrogen peroxide (H_2O_2). Cont: Control; MTT: (3-(4, 5-dimethylthiazolyl-2)-2,5-diphenyltetrazolium bromide.$

We recruited six individuals with no history of psychiatric disease or use of psychotropic medication and no smokers. Four were female and two were male subjects and the average age was 28 ± 6.2 years.

We observed significant differences in actin reactivity for only one treatment. The MTT results indicated increases at aripiprazole (13%) in comparison with vehicle (P = 0.05). Although not statistically significant, H_2O_2 treatment reduced cell viability by 11% compared to the control.

The telomere length ratio was changed to percentage to facilitate the visualization of the treatments compared with control, vehicle and H_2O_2 . The vehicle condition showed a reduction of 4% compared to the control (culture with medium only), showing that the vehicles did not cause

telomere length alteration. Cells treated with H_2O_2 showed 46% reduction in telomere length, compared to the control condition, with no statistical significance (**Figure 2**).

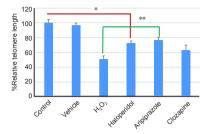


Figure 2 | Telomere length ratio in the samples studied.

The average telomere length was increased by 19% and 23% after treatment with Haloperidol and Aripiprazole, respectively compared to H_2O_2 treatment. *P < 0.05, **P < 0.01 (one-way analysis of variance followed by Tukey's *post hoc* test).

The PBMCs treated with haloperidol showed longer telomeres compared to cells with H_2O_2 toxicity. However, they showed a decrease in telomeric length when compared to the control group (P < 0.05). Telomere length in cells was significantly longer after aripiprazole treatment compared with after H_2O_2 treatment (P < 0.01).

Discussion

Currently, in psychiatry, there is a difficulty in acquiring proper cell models for a disease study, since the physiopathology of most diseases has not been completely understood. Some effort has been made to model induced pluripotent stem cells. for example, to study schizophrenia, but characteristic cell phenotypes (perhaps based on 'omics') need yet to be studied and validated (Falk et al., 2016). This is the first work in the literature to show that PBMCs subjected to oxidative damage to DNA have telomeric shortening. However, this shortening of telomeres can be avoided, since antipsychotic treatment before the event of oxidative stress protects and prevents telomere length. Here we showed that aripiprazole increased telomere length by 23% and haloperidol tended to increase telomere length in PBMC after H₂O₂ treatment. Besides enlightening novel mechanisms of action of antipsychotics, results from this study provide the basis for the use of antipsychotics in ageing-related diseases (Aviv and Shay, 2018). It has been suggested that treatment with antipsychotics in patients with schizophrenia may lengthen, or preserve the length of, the telomere and also affect mitochondrial function and consequently cell metabolism. Several cross-sectional studies have shown that psychiatric patients with longer telomeres tend to respond better to antipsychotic treatment, thus telomere length has been become a candidate for a biological marker (Polho et al., 2015; Brand et al., 2020).

The aripiprazole has demonstrated neuroprotective effects through increasing transcripts and protein level of brainderived neurotrophic factor, phosphorylation and GSK-3 β in SH-SY5Y cells (Gibson et al., 1999; Xu et al., 2015), and decreasing presynaptic-like glutamate release in synaptosomes (Yu et al, 2008). Although the mechanism through which aripiprazole exhibits neuroprotective effects is unknown, aripiprazole is distinct from all other antipsychotics in that it has partial agonist activity at the dopamine D2 receptor and serotonin 5-HT1A receptor (Cardenas et al., 2019).

Atypical antipsychotics are associated with a lower risk of many reasons for mortality and extrapyramidal symptoms compared with conventional antipsychotics. However, atypical antipsychotics have been associated with a higher risk of stroke compared with conventional antipsychotics (Farlow and Shamliyan, 2017). peroxidation in schizophrenia patients (Gubert et al., 2013). Considering that PBMC results should reflect the status of the brain, and then increased neuronal oxidative stress could produce impairments in signal transduction and neuronal plasticity (Lindqvist et al., 2015). Biomarkers of oxidative stress allow for the identification of cellular impairment via measurable parameters. Arvindakshan et al. (2003) found that the level of lipid peroxidation was certainly correlated with symptom severity in drug-free schizophrenic patients. Casademont et al. (2007) described a stronger effect of haloperidol, on decreasing complex I activity and increasing lipid peroxidation when associated with atypical agents, such as clozapine and risperidone, in PBMCs from schizophrenia. Some studies showed the relationship between telomere length and treatment response to psychiatric medications (Honig et al., 2006; Khan et al., 2017; Polho et al., 2015). It has been shown that shortened telomere length in schizophrenia terms. is an indicator of oxidative stress, and the subsequent cellular dysfunction may be a factor contributing to the progressive deterioration in treatment resistant schizophrenia (Yu et al, 2008). These results indicate that telomere length is shortened largely in a subgroup of chronic schizophrenia patients who respond poorly to treatment. Interestingly, it has been demonstrated that haloperidol decreases the viability of neuronal cells in vitro (Brinholi et al., 2016), which increases the activity of apoptosis proteins. On the other hand, studies in vitro have shown that while haloperidol is apparently neurotoxic, secondgeneration antipsychotics offer protection against serum withdrawal-induced apoptosis in SK-N-SH cells (Gassó et

A significant decrease was observed in mitochondrial complex

I activity in PBMCs obtained from patients with chronic

and stabilized schizophrenia when compared with healthy controls. An increase in plasma level of TBARS suggests lipid

al., 2012), or against cytotoxin-induced apoptosis in the pheochromocytoma cell line (Lindqvist et al., 2015). In vitro experiments using haloperidol separately and above the therapeutic dosage, showed decreased telomerase expression in leukocytes of healthy individuals (Porton et al., 2008). Perhaps the tendency of haloperidol increasing telomere length observed in our experiments may only represent a selection of the leukocyte population with long telomeres.

Our study has some limitations. The number of subjects was small, so the lack of statistical significance may be explained by underpowered sample. These results demonstrate the effects of antipsychotics on oxidative stress and only three antipsychotics were tested and on a single dose. Since different antipsychotics have different pharmacodynamics, new studies must be made to demonstrate other effects on telomere length. Oxidative stress is a well described phenomenon in schizophrenia and some authors suggest it may contribute to the pathology. Although schizophrenia may be better represented by chronic oxidative stress, our purpose was to verify only DNA damage (more specifically, telomere attrition) (Bitanihirwe and Woo, 2011). As expected, H₂O₂ treatment reduced telomere length in PBMCs of healthy subjects.

In conclusion, our results showed that aripiprazole can reduce the damage caused by oxidative stress and preserve telomere length and cell viability.

Author contributions: Experimental design, isolation and culture of mononuclear cells, analysis of telomere length, and preparation of the manuscript: GBP, VJDP. Sample collection and preparation, standardization of the qPCR protocol, interpretation of results, and writing the discussion section: GMC, DSK, TC. Analysis of results and participating in writing the discussion section: WFG, OVF, HPB. All authors approved the final version

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