

ASPM and the Evolution of Cerebral Cortical Size in a Community of New World Monkeys

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Abstract

The ASPM (abnormal spindle-like microcephaly associated) gene has been proposed as a major determinant of cerebral cortical size among primates, including humans. Yet the specific functions of ASPM and its connection to human intelligence remain controversial. This debate is limited in part by a taxonomic focus on Old World monkeys and apes. Here we expand the comparative context of ASPM sequence analyses with a study of New World monkeys, a radiation of primates in which enlarged brain size has evolved in parallel in spider monkeys (genus Ateles) and capuchins (genus Cebus). The primate community of Costa Rica is perhaps a model system because it allows for independent pairwise comparisons of smaller- and larger-brained species within two taxonomic families. Accordingly, we analyzed the complete sequence of exon 18 of ASPM in Ateles geoffroyi, Alouatta palliata, Cebus capucinus, and Saimiri oerstedii. As the analysis of multiple species in a genus improves phylogenetic reconstruction, we also analyzed eleven published sequences from other New World monkeys. Our exon-wide, lineage-specific analysis of eleven genera and the ratio of rates of nonsynonymous to synonymous substitutions (d_N/d_S) on ASPM revealed no detectable evidence for positive selection in the lineages leading to Ateles or Cebus, as indicated by d_N/d_S ratios of <1.0 (0.6502 and 0.4268, respectively). Our results suggest that a multitude of interacting genes have driven the evolution of larger brains among primates, with different genes involved in this process in different encephalized lineages, or at least with evidence for positive selection not readily apparent for the same genes in all lineages. The primate community of Costa Rica may serve as a model system for future studies that aim to elucidate the molecular mechanisms underlying cognitive capacity and cortical size.

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Introduction

A disproportionately large cerebral cortex is a hallmark of human evolution. It facilitates increased information processing and thus our perceived high level of intelligence and rapid rate of cultural innovation; for instance, our capacity for tool-making, social intelligence, and language [1]. As a result, considerable attention has long been focused on how and why a relatively large cerebral cortex was favored in some primate lineages. In recent years a particular emphasis has been paid to the identification of genetic variants that correlate with increased cortical size [2]. In this vein, the *ASPM* (abnormal spindle-like microcephaly associated) gene has been proposed as a major determinant of cerebral cortical size among primates [3,4].

The ASPM gene encodes a 10,434-bp-long coding sequence, in 28 exons, and spans 65 kb of genomic DNA. Exon 3 spans approximately 1500 bp while exon 18 spans approximately 4700 bp. The remaining exons of the ASPM gene span less than 200 bp each. The ASPM gene encodes a protein that is widely conserved between primates. The ASPM protein contains four distinguishable regions: a putative N-terminal microtubule-binding

domain, a calponin-homology domain, an IQ repeat domain containing multiple IQ repeats (calmodulin-binding motifs), and a C-terminal region [5]. More than half of the primate ASPM protein consists of repeated calmodulin-binding IQ domains. The major ASPM transcript contains 81 IQ domains, which are organized into IQ calmodulin-binding motifs comprised by 20–25 amino acids. The majority of these repeats are encoded in Exon 18

Calmodulin binding to IQ motifs induces a conformational change in proteins that regulate the binding of actin to the aminoterminal CH domains. It has been proposed that changes in ASPM induce changes in the orientation of the mitotic spindle of neuroblasts, which induces symmetric mitosis generating two progenitor cells; as opposed to one progenitor cell and one postmitotic neuron, typical of asymmetric mitosis. The additional rounds of symmetric duplication cause an exponential expansion of the progenitor pool. Control of this proliferative symmetry can cause dramatic alterations in cerebral cortical size, and so changes in ASPM could regulate cortical size by making subtle changes in spindle orientation [6]. Given the proposed role of ASPM in regulating divisions of

neuronal progenitors, both the number of repeats and the particular amino acid substitutions in the IQ repeats may be strongly related to brain evolution [5].

Such claims have fueled further research on the biology and function of ASPM [7,8]. For example, Kouprina et al. [6] found that ASPM is expressed in numerous proliferating tissues outside the cerebral cortex, suggesting it has functions apart from neuroblast replication. Other authors linked ASPM to more general mechanisms such as ciliary function and spermatogenesis rather than neural development, further confusing the functional significance of ASPM during human evolution [9,10,11]. More controversially, a haplotype of ASPM was linked to recent and ongoing selective sweeps among populations of modern humans [12,13], although these conclusions were subsequently challenged [14,15] and links between ASPM genetic variants and human intelligence have been refuted [16]. Together, these studies improve our understanding of ASPM function, but do little to resolve why ASPM evinces a purported signature of positive selection in certain primate lineages.

In general, ties between ASPM and brain size evolution have been based on Old World monkeys, apes, and humans [3,4,5,17]. Such an emphasis on catarrhine primates is logical given the level of shared ancestry, but it offers limited comparative context or independent power. To address this taxonomic void, Ali and Meier [18] and Montgomery et al. [19] included New World monkeys in their tests for positive correlations between ASPM variation and brain size across primates using codon-specific maximum likelihood tests of selection. Such an approach aimed to detect mutations in specific sites that might have impacted the function of the ASPM protein in specific lineages in ways that might have positively affected the fitness of an individual that carried the mutation (such that the mutation would have swept to fixation rapidly).

Ali and Meier [18] reported signatures of positive selection associated with relatively larger cerebral cortical volumes in nine primate lineages, including humans, chimpanzees, and a family of New World monkeys, the Atelidae. In contrast, the expanded analysis of Montgomery et al. [19] found little support for such specific episodes of adaptive evolution in any given lineage. Instead, they reported a positive relation between molecular evolution in ASPM and an increase in neonatal brain size across 21 anthropoid genera based on a fraction (2.3%) of the codon sites predicted to present a significant increase in non-synonymous substitutions over synonymous substitutions. Accordingly, Montgomery et al. [19] proposed a deeper evolutionary history of ASPM, and suggested that it might be responsible for independent increases in brain size in all major clades of primates, including New World monkeys.

Montgomery et al.'s [19] analysis of 10 species of New World (platyrrhine) monkey is instructive for highlighting the complexity of ASPM evolution within primates. However, if positive selection of ASPM is related to an ancestral increase in brain size, a comparative analysis requires the inclusion of multiple related species within genera, which allows for proper reconstruction of the ancestral states of ASPM sequences. Here we expand on these earlier findings by incorporating platyrrhine genera from all previous studies and adding four species from the primate community of Costa Rica, for a total of 16 species. Our analysis not only allows for a more complete treatment of the families Atelidae and Cebidae, but it also highlights the practical value of focusing on communities as model systems for studying the evolution of cerebral cortical size.

Primates of Costa Rica

Although a sparse fossil record obscures the origins of New World monkeys, all present evidence suggests a trans-Atlantic dispersal event ca. 25-40 million years ago [20]. The subsequent adaptive radiation is remarkable for its variation in brain size, diet, and social behavior. Accordingly, New World monkeys provide fertile ground for exploring the underlying genetic mechanisms of large brain size. In this regard, the primate community of Costa Rica is a model system. It features sympatric species in the family Atelidae - the mantled howling monkey (Alouatta palliata) and the black-handed spider monkey (Ateles geoffroyi). The two species are comparable in body size, yet the brain of A. geoffroyi is nearly twofold larger (Table 1). Similarly, two species in the family Cebidae coexist in some habitats - the white-faced capuchin (Cebus capucinus) and the squirrel monkey (Saimiri oerstedii). The relative brain size of Cebus is among the largest of any nonhuman primate and 10-20% larger than that of Saimiri (Table 1) [19,21]. It also has a complicated structure. The cerebral cortex of Cebus shares with Ateles the most complex pattern of fissures among the platyrrhine primates [22]. Compellingly, such parallel increases in brain size and complexity support an expanded coterie of cognitive abilities.

Cognitive Ecology and Brain Size

The tropical forests of Costa Rica are highly dynamic fluctuating systems that require complex foraging strategies to meet nutritional needs over all phases of an annual cycle. Despite similar body sizes, *Ateles* and *Alouatta* have evolved contrasting dietary behaviors. In northwestern Costa Rica, *Alouatta palliata* can devote up to 77% of its monthly foraging time to eating leaves [23], whereas *Ateles geoffroyi* shows a strong predilection for fruit and flowers (up to 78% and 10% of foraging time, respectively) [24]. Compared to the ubiquity of leaves, fruit and flowers are scattered spatially and temporarily; thus, for *Ateles*, the selective advantages associated with remembering the location and availability of such patchy resources has been tied to the evolution of its relatively large brain [25]. In fact, the memory skills of *Ateles* are reported to exceed those of squirrel monkeys and macaques under experimental conditions [26].

Cebus and Saimiri also share similar body sizes and morphologies, and often forage sympatrically on the same substrates [22]. However, Cebus and Saimiri differ considerably in the extent to which they acquire resources manually [27]. Whereas Saimiri tends to glean invertebrates from exposed surfaces, Cebus spends ca. 50% of its day searching for and extracting embedded foods [22]. Such extractive foraging behavior is expected to favor greater manual dexterity and a high degree of sensorimotor intelligence [28,29]. Sensorimotor intelligence, in turn, is hypothesized to drive technical innovation; indeed, Cebus manufactures the greatest variety of tools and uses them more often than all nonhuman primates excepting chimpanzees [30–32]. For Cebus, the cognitive demands and selective advantages associated with conceptualizing unseen foods and the use of tools to extract them was possibly a major contributing factor to the evolution of its large brain [33].

Given the ecological and cognitive challenges faced by *Ateles* and *Cebus*, i.e. the need to remember scarce resources or imagine and extract concealed ones, it is worth testing whether there has been convergent adaptive evolution in these lineages of the same genes believed to be responsible for human cortex size and cognitive ability. More specifically, the pair-wise coexistence of large- and small-brained atelid and cebid primates in Costa Rica provides a model system for testing molecular hypothesis focused on the evolution of brain size. In this study we compared the complete sequence of exon 18 in *ASPM* across eleven platyrrhine genera,

Table 1. Brain mass and volumes of genera in the families Atelidae and Cebidae [modified from 19].

	Adult													
Genus	Body mass (g)	Brain mass (mg)	Brain volume (mm3)	Neocortex volume (mm3)	Telencephalon volume (mm3)									
Alouatta	6400.0	52000.0	49009.0	31660.0	37388.0									
Ateles	8000.0	108000.0	101034.0	70856.0	79946.0									
Cebus	3100.0	71000.0	66939.0	46429.0	52113.0									
Saimiri	660.0	24000.0	22572.0	15541.0	17635.0									
	Neonate													
Genus	Body mass (g)	Brain mass (mg)												
Alouatta	363.9	30800.0												
Ateles	512.0	63950.0												
Cebus	232.9	33650.0												
Saimiri	153.5	15240.0												

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including multiple species of large-brained *Ateles* and *Cebus*, and small-brained *Alouatta* and *Saimiri*, and studied patterns of natural selection on this gene coding sequence using both an exon-wide lineage-specific analysis similar to Evans et al. [4] and codon-specific analyses across taxa similar to those of Ali and Meier [18] and Montgomery et al. [19]. Exon 18 alone represents 45% of the *ASPM* gene and has been shown to code for most of the critically functioning region of the ASPM protein; the Cadmodulin-binding domain, consisting of repetitive IQ domains. Early d_N/d_S analyses of anthropoid primates indicated different rates of evolution along the ASPM protein, where rapidly evolving residues were mainly concentrated in the IQ repeat domain, coded in its majority by Exon 18, making it the prime region to consider in studies of natural selection [6,10,18,19,21].

Results and Discussion

Previously, ASPM was reported to be under positive selection in the lineages leading to humans and other primates with relatively increased cortex size. In particular, a human-chimpanzee comparison revealed repeated selection on multiple sites along the ASPM gene [4]. Here we focused on two genera of New World monkey, Ateles and Cebus, which evolved increased cortical sizes independently within their taxonomic families. In contrast to their shared increased cortical sizes, amino acid replacements specific to humans, Cebus and Ateles found in exon 18 of the ASPM protein do not overlap, underlying the independent evolution of this region in these species of interest (fig. 1). Lineage-specific analyses of exon 18 sequences of the ASPM gene resulted in d_N/d_S ratios of 0.6502 for the Ateles lineage and 0.4268 for the Cebus lineage (fig. 2). Such values (<1) are consistent with functional constraint on amino acid sequences, rather than positive selection. Our exon-wide lineage-specific analysis of the ratio of the rates of nonsynonymous (amino acid-changing, potentially functional) to synonymous (presumably neutral) substitution (d_N/d_S) on ASPM thus revealed no detectable evidence for positive selection. It should be noted that the only branch to present a positive d_N/d_S ratio, was that of Alouatta palliata (3.0377, fig. 2), however, this result was driven by a small number of estimated substitutions $(\mathcal{N}=8.9, S=1.0)$, and was not significantly different from a model of neutral evolution (p = 0.235).

Although our exon-wide lineage-specific results within the Atelidae stand in contrast with those of Ali and Meier [18], we also followed their analytical approach with codon-specific models of selection. This is accomplished by allowing d_N/d_S to vary across codon positions as well as across lineages, and then recording the differences in the distribution of positively selected changes between "foreground" branches (*Cebus* and *Ateles* branches) and the rest of the phylogenetic tree. Our results from this test also failed to reject a model of neutral evolution in the *Ateles* and *Cebus* lineages (p = 0.491). The discrepancy in results may be due to the addition of an extra species within the genus *Ateles*, allowing for better phylogenetic reconstruction.

Finally, following the methodology of Montgomery et al. [19], we also implemented a branch-site model, which allows ω to vary across amino acid sites, but not across lineages. This model was used for identifying amino acid sites deviating from neutrality, which could be indicative of gradual positive selection across the entire phylogeny, as opposed to simply from the ancestral primate "root" to each terminal branch "tip". Our site test of gradual selection failed to reject a model of neutral evolution within platyrrhine monkeys (p = 0.998). It should be noted that Montgomery et al. [19] only rejected a model of neutral evolution using this test when all anthropoid primates were included, but they did not detect a signature of overall positive selection specifically within New World monkeys (as reported by a d_N/d_S value of 0.479 [19]), which is consistent with our results. Montgomery et al. [19] also predicted 2.3% of all amino acid sites in their sequences to have been under positive selection across all primates, based on the Bayes Empirical Bayes (BEB) analysis associated with the method. Our own BEB analysis, specific for New World monkeys, predicted four amino acid sites presenting significant deviations from neutrality ($\omega > 1$, p<0.05, fig. 1). Those sites however, only represent a small portion (0.3%) of all amino acid sites, and our overall model was not significantly different from neutrality. Therefore, these few positive results may not be biologically meaningful. We interpret these results to indicate no evidence of positive selection across the platyrrhines.

Conclusion

In contrast to the results of Ali and Meier [18], we found a relatively gradual tempo of neutral evolution of ASPM in the

	4	103	114	128	215	260	280	334	356	368	380	387	410	422	423	*430	434	440	462	909	*599	909	609	632	639	758	813	867	1142	204
Amino Acid (1-1333)							*									*					*									*
Ancestral state	L	I	L	Н	Q	Q	Н	L	R	M	R	K	K	Y	S	Н	K	V	K	L	R	I	T	K	R	Α	M	Н	V	Q
Homo sapiens	F	T		27		74		÷	C	V	74						ą		¥			T			W	S		Y	ų.	
Cebus clade				R			Y				K					С				H	W		1				R		1	G
Ateles clade			V		R	L	Y	S				T	E	C	R	C	Е	L	A					R						R

Figure 1. Amino acid replacements of ASPM exon 18 specific to *Homo, Cebus* **and** *Ateles.* Positions denoted by a star (*) are predicted to be under positive selection by the Bayes Empirical Bayes associated test (p<0.05). doi:10.1371/journal.pone.0044928.q001

Atelidae, and no evidence of strong positive selection in any other lineage of New World monkeys. We also found no evidence of gradual positive selection over the platyrrhine clade, despite evidence for independent cerebral enlargement in the lineages leading to Ateles and Cebus. This finding is somewhat contradictory to Montgomery et al.'s [19] conclusion of shared genetic bases for encephalization across anthropoid primates (which includes platyrrhine monkeys). However, it should be noted that both Ali and Meier [18] and Montgomery et al. [19] included exon 3 of ASPM into their analysis, so it is possible that our results are consistent with positive selection occurring on exon 3. Because changes in exon 18 are most likely to affect neural cell duplication, we expected positive selection associated with brain size increase to occur in this region. It is possible that positive selection of ASPM is associated with processes other than brain development, at least in New World monkeys, thus explaining the inconsistencies between our results and previously published results.

We note that it is also possible that our tests had insufficient power to detect evidence of positive selection on any specific changes in ASPM exon 18, even if such changes did in fact partly account for the large differences in brain anatomy among New World monkeys. It is also plausible that changes in other ASPM exons that we did not study, or in ASPM regulatory regions, may harbor evidence of positive selection in these taxa. Of course, brain development and higher cognitive skills are almost certainly affected by a multitude of interacting genes, such that convergent

phenotypic features may be achieved through different modifications on separate lineages.

The primate community of Costa Rica, featuring coexisting small- and large-brained species with remarkable behavioral diversity, has the potential to be an instructive model system for testing hypotheses relevant to human evolution. By contrasting candidate genes such as ASPM with species under similar selective pressures, one might be able to provide insight into human neurogenetics. Species such as Cebus and Ateles offer independent examples of encephalization and relatively increased intelligence among primates.

Materials and Methods

Ethics Statement

We affirm that our research methods were approved by all relevant government agencies and the Chancellor's Animal Research Committee of the University of California, Santa Cruz (approval no. 0906). UC-Santa Cruz was the institutional affiliation of the corresponding author at the time of the research. Samples were collected with the approval of Costa Rica's Ministerio de Ambiente y Energía (permit no. 069-2001 OFAU). Buccal swabs containing DNA were exported from Costa Rica (Ministerio de Agricultura y Ganaderia permiso fitosanitario no. 331) and imported into the USA (CITES permit no. 06US130146/9).

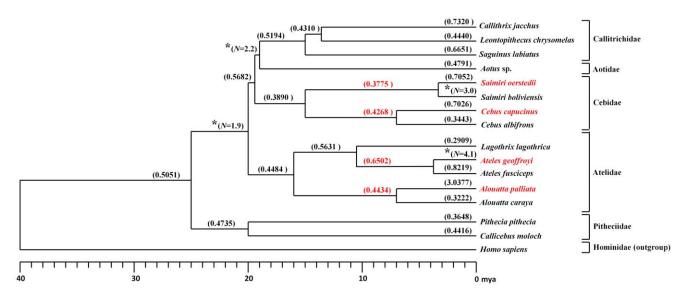


Figure 2. Exon-wide lineage-specific test of neutrality: d_N/d_S ratios for eleven new world monkey species. Genera with novel added species are marked in red, d_N/d_S values for the ancestral branch of those genera are also marked in red. Ratios significantly greater than 1 suggest positive selection while ratios significantly less than 1 are consistent with purifying selection on nonsynonymous mutations. Ratios non-significantly different from 1 are consistent with neutrality. Asterisks represent a missing value; between these lineages there are no synonymous substitutions $(d_S = 0)$ and thus PAML is unable to properly estimate d_N/d_S . In these cases, the number of nonsynonymous substitutions (N) is reported. doi:10.1371/journal.pone.0044928.g002

Samples and DNA Analysis

One of the authors (G.A.G.-E.) tranquilized wild individuals [34] of four species (Alouatta palliata, Ateles geoffroyi, Cebus capucinus, Saimiri oerstedii) in the Pacific region of Costa Rica (Ministerio de Ambiente y Energía permit no. 069-2001 OFAU). Buccal swabs containing DNA were exported from Costa Rica (Ministerio de Agricultura y Ganaderia permiso fitosanitario no. 331) and imported into the USA (CITES permit no. 06US130146/9). The DNA was extracted using a standard Qiagen DNeasy blood and tissue extraction kit. This protocol was reviewed and approved by the Chancellor's Animal Research Committee of the University of California, Santa Cruz (approval no. 0906).

Standard PCR conditions were used to amplify ASPM. Primers were designed specifically for exon 18 (Table S1). Invitrogen Standard TOPO cloning was used with some amplicons (Vector primers available in Table S1). In these cases, three clones were sequenced per amplicon and a consensus of all three was used to rule out replication errors. All sequencing was done using standard dye terminator chemistry on PCR products at UC Berkeley sequencing facilities with an Applied Biosystems 96 capillary-3730xi genetic analyzer.

Sequence Analysis

Comparative exon 18 sequence for *Homo sapiens* was taken directly from the UCSC Genome Browser (http://www.genome.ucsc.edu/, NCBI36/hg18, March 2006). All new sequences were deposited into GenBank with the following accession numbers: *Alouatta palliata* [GQ221757], *Ateles geoffroyi* [GQ221758], *Cebus capucinus* [GQ221759], *Saimiri oerstedii* [GQ221760]. Sequences for the remaining species were acquired from GenBank: *Saimiri boliviensis* [AY485419]; *Aotus* sp. [AY485422; [4]]; *Saguinus labiatus* [AY497015; [5]]; *Ateles fusciceps* [FJ013122]; *Lagothrix lagotricha* [FJ013130; [18]]; *Callithrix jacchus* [HQ540102]; *Cebus albifrons* [HQ540103]; *Alouatta caraya* [HQ540104]; *Leontopithecus chrysomelas* [HQ540105]; *Callicebus moloch* [HQ540106]; *Pithecia pithecia* [HQ540107; [19]].

Sequence alignment was performed in Bioedit [35]. A species tree was assembled based on the primate phylogeny published in Perelman et al. [36]. Next, MEGA 4.0 beta 4020 [37] and DAMBE [38] were used to prepare input data for PAML 4 [39]. The d_N/d_S ratios were estimated in PAML using model 1. In this model, ω is the average d_N/d_S within lineages with independent ω 's between lineages. In the absence of synonymous substitutions, the d_N/d_S ratio cannot be calculated; as a result, some of our branches lack a d_N/d_S ratio (fig. 1). Because synonymous substitutions were often absent between closely related species within genera, $(d_S = 0)$, PAML is unable to properly estimate d_N/d_S . In these cases d_N was reported individually. To test if the high value of d_N/d_S in Alouatta palliata is significantly different from neutrality, a specific model in which ω evolves at a different rate for this branch was tested against a neutral model using a likelihood ratio test.

As suggested by Ali and Meier [18], a branch-site test of neutrality (to potentially identify positive selection) was performed

References

- Gibson KR (2002) Evolution of human intelligence: The roles of brain size and mental construction. Brain Behav Evol 59: 10–20.
- Pollard KS, Salama SR, Lambert N, Lambot M-A, Coppens S, et al. (2006) An RNA gene expressed during cortical development evolved rapidly in humans. Nature 443: 167–172.
- Bond J, Roberts E, Mochida GH, Hampshire DJ, Scott S, et al. (2002) ASPM is a major determinant of cerebral cortical size. Nat Genet 32: 316–320.

as recommended by Yang [39] using model 2 in PAML. This branch site model was specifically designed to test for episodes of positive selection (as deviations from neutrality) acting on a small number of branches [40], by using a maximum likelihood approach to detect codon-specific positive selection and allowing ω to vary across codon positions as well as across priori-assigned foreground and background lineages. In this case, *Ateles* and *Cebus* branches were selected as foreground branches based on behavioral and morphological observations. The cut-off for statistical significance was assigned at the 0.05 level of probability by testing twice the difference of the log-likelihood (2 Δ I) output from PAML compared against a chi-square distribution (d.f. = 1).

Following the recommendations of Montgomery et al. [19] we adopted a branch site model, which allows ω to vary across sites, but not across lineages, and tests for deviations from neutrality across a phylogeny. Site models M1a (null model of neutral evolution) and M2a (alternative model of positive selection) were compared using a likelihood ratio test statistic with a cut-off for statistical significance assigned at the 0.05 level of probability by testing twice the difference of the log-likelihood (2 Δ l) output from PAML compared against a chi-square distribution (d.f. = 2) [19,39]. In addition, this model utilizes Yang's Bayes Empirical Bayes (BEB) model to predict amino acid sites where $\omega > 1$ at the 0.05 and 0.01 levels of probability. Because some of the results in Montgomery et al. [19] were only significant when the callitrichids were removed from their analysis, we repeated all analysis after removing Saguinus labiatus, Callithrix jacchus and Leontopithecus chrysomelas from our sample. The removal of this family led to no differences in results for the remaining species.

Supporting Information

Table S1 ASPM Exon 18 primers and Invitrogen TOPO cloning vector primers.

(XLS)

File S1 Abstract (Spanish).

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Author Contributions

Conceived and designed the experiments: FAV NJD GHP. Performed the experiments: FAV. Analyzed the data: FAV. Contributed reagents/materials/analysis tools: GG. Wrote the paper: FAV NJD GHP.

- Evans PD, Anderson JR, Vallender EJ, Gilbert SL, Malcom CM, et al. (2004)
 Adaptive evolution of ASPM, a major determinant of cerebral cortical size in
 humans. Hum Mol Genet 13: 489–494.
- Kouprina N, Pavlicek A, Mochida GH, Solomon G, Gersch W, et al. (2004) Accelerated evolution of the ASPM gene controlling brain size begins prior to human brain expansion. PLoS Biol 2: e126.
- Kouprina N, Pavlicek A, Collins NK, Nakano M, Noskov VN, et al. (2005) The microcephaly ASPM gene is expressed in proliferating tissues and encodes for a mitotic spindle protein. Hum Mol Genet 14: 2155–2165.

- Fish JL, Kosodo Y, Enard W, Paabo S, Huttner WB (2006) ASPM specifically maintains symmetric proliferative divisions of neuroepithelial cells. Proc Natl Acad Sci USA 103: 10438–10443.
- van der Voet M, Berends CWH, Perreault A, Nguyen-Ngoc T, Gonczy P, et al. (2009) NuMA-related LIN-5, ASPM-1, calmodulin and dynein promote meiotic spindle rotation independently of cortical LIN-5/GPR/G. Nat Cell Biol 11: 260-277
- 9. Ponting CP (2006) A novel domain suggests a ciliary function for ASPM, a brain size determining gene. Bioinformatics 22: 1031-1035.
- Ponting CP, Jackson AP (2005) Evolution of primary microcephaly genes and the enlargement of primate brains. Curr Opin Genet Dev 15: 241–248.
- Pulvers JN, Bryk J, Fish JL, Wilsch-Bräuninger M, Arai Y, et al. (2010) Mutations in mouse ASPM (abnormal spindle-like microcephaly associated) cause not only microcephaly but also major defects in the germline. Proc Natl Acad Sci USA 107: 16595–16600.
- Mekel-Bobrov N, Gilbert SL, Evans PD, Vallender EJ, Anderson JR, et al. (2005) Ongoing adaptive evolution of ASPM, a brain size determinant in Homo sapiens. Science 309: 1720–1722.
- Mekel-Bobrov N, Lahn BT (2007) Response to comments by Timpson, et al. and Yu, et al. Science 317: 1036b.
- Timpson N, Heron J, Smith GD, Enard W (2007) Comment on papers by Evans, et al. and Mekel-Bobrov, et al. on evidence for positive selection of MCPH1 and ASPM. Science 317: 1036a.
- Yu F, Hill RS, Schaffner SF, Sabeti PC, Wang ET, et al. (2007) Comment on "Ongoing adaptive evolution of ASPM, a brain size determinant in Homo sapiens". Science 316: 370b.
- Mekel-Bobrov N, Posthuma D, Gilbert SL, Lind P, Gosso MF, et al. (2007) The ongoing adaptive evolution of ASPM and Microephalin is not explained by increased intelligence. Hum Mol Genet 16: 600–608.
- Zhang J (2003) Evolution of the human ASPM gene, a major determinant of brain size. Genetics 165: 2063–2070.
- Ali F, Meier R (2008) Positive selection in ASPM is correlated with cerebral cortex evolution across primates but not with whole-brain size. Mol Biol Evol 25: 2247–2250.
- Montgomery SH, Capellini I, Venditti C, Barton RA, Mundy NI (2011) Adaptive evolution of four microcephaly genes and the evolution of brain size in anthropoid primates. Mol Biol Evol 28: 625–638.
- Goodman M, Porter CA, Czelusniak J, Page SL, Schneider H, et al. (1998)
 Toward a phylogenetic classification of primates based on DNA evidence complemented by fossil evidence. Mol Phylogenet Evol 9: 585–598.
- Woods CG, Bond J, Enard W (2005) Autosomal recessive primary microcephaly (MCPH): A review of clinical, molecular, and evolutionary findings. Am J Hum Genet 76: 717–728.
- Janson CH, Boinski S (1992) Morphological and behavioral adaptations for foraging in generalist primates: The case of the cebines. Am J Phys Anthropol 88: 483-408

- Glander KE (1978) Howling monkey feeding behavior and plant secondary compounds: A study of strategies. In: Montgomery GG, editor. The ecology of arboreal folivores. Washington D.C.: Smithsonian Institution Press. 561–574.
- Chapman CA (1987) Flexibility in diets of three species of Costa Rican primates.
 Folia Primatol 49: 90–105.
- Dew JL (2005) Foraging, food choice, and food processing by sympatric ripefruit specialists: Lagothrix lagotricha poeppigii and Ateles belzebuth belzebuth. Int J Primatol 26: 1107–1135.
- Laska M, Salazar LTH, Luna ER (2003) Successful acquisition of an olfactory discrimination paradigm by spider monkeys, *Ateles geoffroyi*. Physiol Behav 78: 321–329.
- MacNeilage PF (1990) Grasping in modern primates: The evolutionary context.
 In: Goodale MA, editor. Vision and action: The control of grasping. Norwood: Ablex. 1–13.
- Parker ST, Gibson KR (1977) Object manipulation, tool use and sensorimotor intelligence as feeding adaptations in cebus monkeys and great apes. J Hum Evol 6: 623–641.
- Gibson KR (1986) Cognition, brain size and the extraction of embedded food resources. In: Else JG, Lee PC, editors. Primate ontogeny, cognition and social behaviour. Cambridge: Cambridge University Press. 93–103.
- Panger MA (1998) Object-use in free-ranging white-faced capuchins (Cebus capucinus) in Costa Rica. Am J Phys Anthropol 106: 311–321.
- Panger MA, Perry S, Rose L, Gros-Louis J, Vogel E, et al. (2002) Cross-site differences in foraging behavior of white-faced capuchins (*Cebus capucinus*). Am J Phys Anthropol 119: 52–66.
- Moura AC, Lee PC (2004) Capuchin stone tool use in Caatinga dry forest. Science 306: 1909.
- Reader SM, Laland KN (2002) Social intelligence, innovation, and enhanced brain size in primates. Proc Natl Acad Sci USA 99: 4436–4441.
- Glander KE, Fedigan LM, Fedigan L, Chapman CA (1991) Field methods for capture and measurement of three monkey species in Costa Rica. Folia Primatol 57: 70–82.
- Hall TA (1999) BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nucleic Acids Symp Ser 41: 95–98.
- Perelman P, Johnson WE, Roos C, Seuánez HN, Horvath JE, et al. (2011) A molecular phylogeny of living primates. PLoS Genet 7: e1001342.
- Kumar S, Nei M, Dudley J, Tamura K (2008) MEGA: A biologist-centric software for evolutionary analysis of DNA and protein sequences. Brief Bioinform 9: 299–306.
- 38. Xia X, Xie Z (2001) DAMBE: Software package for data analysis in molecular biology and evolution. J Hered 92: 371–373.
- Yang Z (2007) PAML 4: A program package for phylogenetic analysis by maximum likelihood. Mol Biol Evol 24: 1586–1591.
- 40. Anisimova M, Yang Z (2007) Multiple hypothesis testing to detect lineages under positive selection that affects only a few sites. Mol Biol Evol 24: 1219–1228.