



Draft Genome Sequence of Cercospora arachidicola, Causal Agent of Early Leaf Spot in Peanuts

Valerie A. Orner,^a Emily G. Cantonwine,^b Xinye Monica Wang,^a Amr Abouelleil,^c James Bochicchio,^c Chad Nusbaum,^c Albert K. Culbreath,^d Zaid Abdo,^{e*} Renee S. Arias^a

National Peanut Research Laboratory, Dawson, Georgia, USAa; Department of Biology, Valdosta State University, Valdosta, Georgia, USAb; Broad Institute of MIT and Harvard, Cambridge, Massachusetts, USAc; Department of Plant Pathology, University of Georgia, Tifton, Georgia, USAd; USDA, ARS, SEA, Athens, Georgia, USAc

Cercospora arachidicola, causal agent of early leaf spot, is an economically important peanut pathogen. Lack of genetic information about this fungus prevents understanding the role that potentially diverse genotypes may have in peanut breeding programs. Here, we report for the first time a draft genome sequence of *C. arachidicola*.

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Address correspondence to Renee S. Arias, renee.arias@ars.usda.gov, or Valerie A. Orner, valerie.orner@ars.usda.gov.

arly leaf spot caused by Cercospora arachidicola S. Hori (teleomorph Mycosphaerella arachidis Deighton) is one of two important leaf spot diseases in peanut (Arachis hypogaea L.) responsible for significant economic loss to the industry (1, 2). Infections by C. arachidicola appear as small necrotic lesions on the leaves, petioles, or stems, which may be followed by premature defoliation, and, if left unmanaged on susceptible cultivars, can severely decrease yield (1). An effective, yet expensive, disease management strategy consists of multiple fungicide applications throughout the growing season (3). Other strategies such as strip-tillage instead of conventional tillage (4) or weather forecast models that predict disease outbreaks (5) can help minimize the number of fungicide treatments. However, the development of leaf-spotresistant cultivars that require no fungicide application would be the most desirable means of control (6). The recent completion of the peanut genome (http://www.peanutbase.org) will aid breeding programs, but the negligible amount of *C. arachidicola* genetic information hinders progress. Currently, C. arachidicola entries in the NCBI-GenBank database total 8,077 bp in 21 sequences, with half of these entries corresponding to rRNA and the rest only 61 bp each. The genome sequence of C. arachidicola will provide relevant information for the advancement of leaf-spot resistant cultivars, be a useful resource to aid in the selection of target genes for disease control, and contribute to the study of genetic diversity of C. arachidicola.

A single-spore isolate of *C. arachidicola* from an infected peanut plant near Tifton, Georgia, USA, was grown on potato dextrose agar (Difco, Franklin Lakes, NJ, USA) for 6 months. The fungus was removed from the agar and ground using a Kleco tissue pulverizer (Garcia Machine, Visalia, CA, USA). Genomic DNA was extracted using phenol/chloroform/isoamyl alcohol followed by isopropanol precipitation (7) and cleaned using the GeneJET gel Extraction kit (Thermo, Fisher Scientific, Waltham, MA, USA). Sequencing and assembly were performed by the

Broad Institute of MIT and Harvard using an Illumina HiSeq 2500 whole-genome shotgun approach. A total of 465,511,514 reads with an estimated genome coverage of $>100\times$ were assembled *de novo* using ALLPATHS (8) and generated 796 contigs (>400 bp each) with an average size of 40,930 bp that assembled into 491 scaffolds (>1,000 bp each). The average size of the scaffolds was 67,710 bp with a total of 33,245,410 bp and a maximum length of 1,387,526 bp. Most hits from BLAST analysis corresponded to the genera *Pseudocercospora* and *Passalora*.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited in DDBJ/ENA/GenBank under the accession number LIHB000000000. The version described in this paper is the first version, LIHB01000000.

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REFERENCES

- Nutter FW, Jr, Shokes FM. 1995. Management of foliar diseases caused by fungi, p 65–73. *In* Melouk HA, Shokes FM (ed), Peanut health management. APS Publishing, St. Paul, MN.
- 2. Leidner J. 2012. The peanut genomics initiative. Southeastern Peanut Farmer 50:15.
- Smith DH, Littrell RH. 1980. Management of peanut foliar diseases with fungicides. Plant Dis 64:356–361. http://dx.doi.org/10.1094/PD-64-356.
- Cantonwine EG, Culbreath AK, Stevenson KL. 2007. Characterization of early leaf spot suppression by strip tillage in peanut. Phytopathology 97: 187–194. http://dx.doi.org/10.1094/PHYTO-97-2-0187.
- Olatinwo RO, Prabha TV, Paz JO, Hoogenboom G. 2012. Predicting favorable conditions for early leaf spot of peanut using output from the

^{*} Present address: Zaid Abdo, Department of Microbiology, Immunology and Pathology, Colorado State University, Fort Collins, Colorado, USA

- weather research and forecasting (WRF) model. Int J Biometeorol $\bf 56: 259-268. \ http://dx.doi.org/10.1007/s00484-011-0425-6.$
- Branch WD, Culbreath AK. 2013. Yield performance and pest resistance among peanut genotypes when grown without fungicides or insecticides. Crop Protect 52:22–25. http://dx.doi.org/10.1016/j.cropro.2013.05.005.
- 7. Sambrook J, Fritsch EF, Maniatis T. 1989. Molecular cloning: a laboratory
- manual, 2nd ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.
- 8. Butler J, MacCallum I, Kleber M, Shlyakhter IA, Belmonte MK, Lander ES, Nusbaum C, Jaffe DB. 2008. ALLPATHS: de novo assembly of wholegenome shotgun microreads. Genome Res 18:810–820. http://dx.doi.org/10.1101/gr.7337908.