



Research Article

Impact of HIV-1 subtype and Korean Red Ginseng on AIDS progression: comparison of subtype B and subtype D

Young-Keol Cho^{1,*}, Jung-Eun Kim¹, Sun-Hee Lee², Brian T. Foley³, Byeong-Sun Choi⁴¹ Department of Microbiology, University of Ulsan College of Medicine, Seoul, Republic of Korea² Division of Infectious Diseases, Department of Internal Medicine, Pusan National University Hospital, Busan, Republic of Korea³ HIV Databases, Theoretical Biology and Biophysics Group, Los Alamos National Laboratory, Los Alamos, NM, USA⁴ Division of AIDS, Center for Immunology and Pathology, Korea National Institute of Health, Chung-buk, Republic of Korea

ARTICLE INFO

Article history:

Received 11 April 2018

Received in Revised form

11 July 2018

Accepted 16 July 2018

Available online 20 July 2018

Keywords:

AIDS

Disease progression

HIV-1 subtype D

Korean Red Ginseng

nef gene

ABSTRACT

Background: To date, no study has described disease progression in Asian patients infected with HIV-1 subtype D.

Methods: To determine whether the disease progression differs in patients infected with subtypes D and B prior to starting combination antiretroviral therapy, the annual decline (AD) in CD4⁺ T cell counts over 96 ± 59 months was retrospectively analyzed in 163 patients and compared in subtypes D and B based on the *nef* gene.

Results: CD4⁺ T cell AD was significantly higher in the six subtype D–infected patients than in the 157 subtype B–infected patients irrespective of Korean Red Ginseng (KRG) treatment ($p < 0.001$). Of these, two subtype D–infected patients and 116 subtype B–infected patients had taken KRG. AD was significantly lower in patient in the KRG-treated group than in those in the KRG-naïve group irrespective of subtype ($p < 0.05$). To control for the effect of KRG, patients not treated with KRG were analyzed, with AD found to be significantly greater in subtype D–infected patients than in subtype B–infected patients ($p < 0.01$). KRG treatment had a greater effect on AD in subtype D–infected patients than in subtype B–infected patients (4.5-fold vs. 1.6-fold). Mortality rates were significantly higher in both the 45 KRG-naïve ($p < 0.001$) and all 163 ($p < 0.01$) patients infected with subtype D than subtype B.

Conclusion: Subtype D infection is associated with a >2-fold higher risk of death and a 2.9-fold greater rate of progression than subtype B, regardless of KRG treatment.

© 2018 The Korean Society of Ginseng, Published by Elsevier Korea LLC. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

1. Introduction

The rate of AIDS progression has been reported to depend on HIV-1 subtype [1,2], with disease progressing twice as rapidly in patients infected with HIV-1 subtype D than subtype A, despite similar virus concentrations [3,4]. However, other studies report that disease progression rates do not differ significantly with HIV-1 subtype, probably because these findings were obtained in studies with short (<5 years) follow-up periods [5,6] or analysis of non-D subtypes [6]. For example, the rates of decline of CD4⁺ T cell counts were found to be 4- and 2-fold greater among individuals infected with subtype D than subtypes B [7] and A [8], respectively. Moreover, Ugandan and Zimbabwean women infected with subtype C exhibited 2.5-fold slower rates of CD4⁺ T cell count decline and higher frequencies of long-term nonprogression than those

infected with subtype A or D [9]. Studies of the mechanism underlying the faster disease progression in patients infected with subtype D show that subtype D isolates tend to have a higher frequency of syncytium formation, CXCR4 (X4) coreceptor usage, and rapidly replicating virus than isolates of other subtypes [10]. In addition, the faster progression observed in subtype D may be due to its earlier switch to X4 virus [11]. Moreover, treatment failure and drug resistance rates are more frequent in Ugandans infected with HIV-1 subtype D than subtype A [12].

Worldwide, the majority of HIV infections (70%) have been attributed to subtypes C (48%), A, and B, whereas the prevalence of subtype D has decreased from 3.2% to 2% [13]. In Asia, CRF01_AE (35%), B (33.5%), C (18%), and CRF07_BC (6.2%) are the predominant subtypes (87%), whereas only about 0.1–0.2% of patients are infected with subtype D [13–15]. In Asia and Australia, the overall

* Corresponding author. Department of Microbiology, University of Ulsan College of Medicine, 88, Olympic-ro 43-gil, Songpa-gu, Seoul 05505, Republic of Korea.
E-mail address: ykcho2@amc.seoul.kr (Y.-K. Cho).

prevalence rates of subtype D infection were approximately 0.1% based on sequences as of February 2017 and 0.4% based on patients [16], respectively, although HIV sequences deposited in the Los Alamos sequence database are not representative of the relative distributions in their countries of origin (<https://www.hiv.lanl.gov>). The paucity of information has led to a lack of reporting on disease progression in subtype D–infected patients in Asia and Australia.

Mandatory testing of overseas sailors (OSs) in Korea for antibodies to HIV-1 from 1988 to 1993 identified several who were infected with HIV-1 or HIV-2, including six who were infected with HIV-1 subtype D and the remainder with subtype B [17].

The *nef* gene is a major determinant of virulence in primate lentiviruses. Mutations in *nef* attenuate the replication capacity and pathogenicity of simian immunodeficiency viruses [18–23]. In particular, the evolutionary loss of a specific *nef* function may contribute to the high virulence of HIV-1 in humans [24]. The importance of the *nef* gene as a virulence factor led to our use of this gene to determine HIV-1 subtype in 380 infected patients and to compare disease progression in patients infected with subtypes B and D.

In a nationwide study, we found that disease progression was significantly more rapid in patients infected with subtype D than subtype B and that the disease progression rate in both groups was affected by treatment with Korean Red Ginseng (KRG).

2. Materials and methods

2.1. Study patients

Sequences of the *nef* gene were determined in 381 patients [17,25]. Of these, six were infected with subtype D and 157 with subtype B; of the latter, 36 were infected with Western subtype B and 121 with the Korean subclade of subtype B (KSB) [25] (Table 1). The six patients infected with subtype D included five OSs and one soldier (93–01) who had not gone abroad (Table 2). The phylogenetic tree indicated that these six sequences may have originated from three different sources (Supplementary Figs. 1 and 2). Additional information on these patients, including age, sex, mode of transmission, and year of diagnosis, is shown in Table 2. This study was approved by the Institutional Review Board of Asan Medical Center.

Table 1
Demographic and HIV-1–associated characteristics of patients

Characteristic	Subtype		p-value
	B (n = 157)	D (n = 6)	
Sex, no. (%)			
Female	16 (10)	0	>0.05
Male	141 (90)	6 (100)	>0.05
Overseas sailors and spouses	10 (6)	5 (83)	<0.0001
Age, y, mean ± SD (median)	29 ± 10 (28.5)	29 ± 7 (30.5)	>0.05
Year of diagnosis of HIV-1 infection			
1986–1987	6	0	
1988–1989	17	1	
1990–1991	44	2	
1992–1993	65	2	
1994–2004	25	1	
No. (%) of long-term slow progressors	34 (22%)	0	>0.05
Follow-up months based on CD4 ⁺ T cells	97 ± 59	79 ± 50	>0.05
Amount of KRG administered to all patients, grams	3,221 ± 4,814	2,453 ± 5,093	>0.05
Amount of KRG administered to KRG-treated patients (116 vs. 2), grams	4,359 ± 5,141	7,360 ± 7,580	>0.05
Amount of KRG per year, grams	539 ± 453	643 ± 600	>0.05
CD4 ⁺ T cell count at diagnosis/μL (mean ± SD) (range)	496 ± 254 (131–1,484)	557 ± 362 (7–926)	>0.05
HIV-1 load, copies/mL, mean ± SD (n = 109 vs. 5)	23,911 ± 55,871	47,156 ± 59,206	> 0.05
β2-microglobulin (mg/L), mean ± SD	2.44 ± 0.94	2.50 ± 0.59	>0.05
Study endpoint reached, no. (%)			
Death	56 (36)	5 (83)	<0.05
Survival from diagnosis to death or last CD4 ⁺ T cell count before cART, months, mean ± SD	105 ± 58	82 ± 51	>0.05
AD in CD4 ⁺ T cells >200/μL	44 ± 58	251 ± 132	<0.0001
AD in CD4 ⁺ T cells <200/μL	77 ± 72	105 ± 133	>0.05

AD, annual decline; KRG, Korean Red Ginseng; SD, standard deviation; cART, combinational antiretroviral therapy.

Table 2
Characteristics of the seven patients infected with HIV-1 subtype D.

Patient code	Sex/age at diagnosis	Transmission mode	Visited country	Time of foreign visit	Presumed primary infection	Diagnosis of HIV-1 infection	Plasma RNA copy (/mL)	SD (months)	AD
88–05	M/25	Unknown	Kenya, JP	Feb 86–Jul 1987	Feb 1987	Feb 1988	7,831–3,262	128	77
90–15	M/29	Heterosexual	Kenya	After Nov 1988	Aug 1989	Apr 1990	122,712	59	198
90–16	M/35	Bisexual	Africa	1986–Jul 1988	Jan 1988	Apr 1990	4,459 ¹⁾	28	234
92–65	M/31	Heterosexual	Kenya	1992	July 1992	Oct 1992	>100,000	110	95
93–01	M/21	Homosexual	None	1988 ²⁾	Dec 1988	Dec 1992	781–2,561	140	–12
94–29	M/37	Heterosexual	JP, other	After 1989	Jan 1990	May 1994	ND	24	229

JP, Japan; NA, not applicable; AD, annual decline in CD4⁺ T cell count; SD, survival duration from diagnosis to death (0/μL) or last CD4⁺ T cell count before starting combination antiretroviral therapy; ND, not determined.

¹⁾ On the 5th day of zidovudine treatment.

²⁾ First sexual contact when he was high school student in 1988.

2.2. Treatment with KRG and data collection

Data on CD4⁺ T cell count were from the National Registry, which was established by the division of AIDS, Center for Immunology and Pathology, Korea National Institute of Health (KNIH). Study of KRG treatment in HIV-1 infected patients was initiated at the KNIH in 1991 [26–27] and lasted until February 1996. In parallel, research on KRG from HIV-1 infections began in University of Ulsan College of Medicine in May 1993 and lasted until 2018. Male and female were instructed to take orally six and three capsules of KRG three times daily, respectively [26–30]. One capsule contained 300 mg powder without any additives. There were several interruptions prior to July 2000 including an interruption in KRG intake for 4–5 months after the first 6-month pilot study initiated in November 1991. Therefore, amount and duration of KRG treatment are varied. KRG-treated patients were defined as those who had taken at least 150 g KRG per year. In subtype B–infected and subtype D–infected patients, yearly intake of KRG in the 116 and two KRG-treated patients was 539 ± 453 g and 643 ± 600 g, respectively (Table 1).

2.3. CD4⁺ and CD8⁺ T cell measurements

Peripheral blood mononuclear cells were obtained from patients every 6 months. The peripheral blood mononuclear cell preparations were incubated with phycoerythrin- and fluorescein isothiocyanate-conjugated antibodies against CD4 and CD8 antigens, respectively (Simultest reagents; Becton Dickinson, San Jose, CA, USA), and CD4⁺ and CD8⁺ T cells were counted by flow cytometry (FACScan; Becton Dickinson) [26–30].

2.4. RNA copy measurement

HIV-1 RNA copy numbers (/mL) in serum (before 1997) and plasma (beginning in 1997) were measured using AMPLICOR HIV-1 monitor kits (Roche Diagnostics Systems, Branchburg, NJ, USA). Because RNA copy numbers are 0.28 logs lower in serum than in plasma [31], copy numbers in sera were converted to plasma-equivalent numbers by the addition of 0.28 logs.

2.5. Statistical analysis

All data were expressed as mean ± standard deviation. Statistical significance was estimated by Student's two-tailed *t* test using MedCalc Software (Ostend, Belgium). Kaplan–Meier survival analyses were used to explore the relationship between subtypes (MedCalc Software). Statistical significance was defined as *p* < 0.05.

2.6. Sequence data

The GenBank accession numbers for the sequences in this study are AF063920, AF462775-7, AF584794-9, AY121473, AY221703, AY260808, AY899400-01, DQ054367, KU588843-5, KU588847-50, KY557555-8, KY557830-5, KY558144-7, KY558156-70, KY683964-72, KY683993-5, MF176214-19, and MH396195-36.

3. Results

3.1. Patient characteristics

HIV-1 disease progression was analyzed in all patients infected with subtypes D and B, as determined by *nef* sequences, and with at least two measurements prior to starting combinational antiretroviral therapy (cART). CD4⁺ T cell counts were followed up for 96 ± 59 months. The mean time from diagnosis to last CD4⁺ T cell

count before cART or death was 104 ± 58 months. The demographic characteristics of the 163 patients infected with HIV-1 subtypes B and D are shown in Table 1. The proportion of OSs and their spouses was significantly higher in patients infected with subtype D than subtype B (83% vs. 6%, *p* < 0.0001). Sex and age at diagnosis did not differ in these two groups (Table 1).

3.2. Viral load level

Plasma RNA copy numbers were measured in 95 subtype B–infected patients and five subtype D–infected patients. Although initial viral loads were 1.8-fold higher in subtype D–infected patients (47,156 ± 59,206 copies/mL) than in the subtype B–infected patients (23,911 ± 55,871 copies/mL) (Table 1), the RNA copy number at the last time point was higher in patients infected with subtype B (123,241 ± 187,987 copies/mL) than in those infected with subtype D (53,246 ± 55,214 copies/mL) at the same level of CD4⁺ T cells (303 ± 208/μL vs. 305 ± 383/μL). This finding showed that there was no significant difference in plasma viral load between patients infected with subtypes B and D. The two subtype D–infected patients with the highest viral loads, patients 90-15 and 92-65, rapidly progressed to AIDS (Table 2, Supplementary Fig. 3). By contrast, three subtype D–infected patients (88-05, 90-16, and 93-01) had lower viral loads (mean: 2,007 ± 2,123 copies/mL) before the introduction of KRG or zidovudine (Table 2). Following diagnosis, two subtype D–infected patients with low virus concentrations (88-05 and 93-01) survived for more than 10 years in the absence of cART (Supplementary Fig. 3). Both patients were CCR5 virus carriers. In particular, patient 93-01 had a virus concentration as low as 781/mL beginning in April 1993, and this low viral load was maintained for over 10 years after taking KRG with a normal range of high-sensitivity C-reactive protein concentrations (0.3, 0.2, and 0.3 mg/L in December 1992, October 1993, and August 2004, respectively). Despite his first sexual contact in 1988 and a low viral load, his earlier decline in CD4⁺ T cell counts to 200/μL was very rapid (Supplementary Fig. 3).

3.3. Definition of AD, AD >200, and AD <200

Since 1988, Korean OSs had undergone compulsory testing for anti-HIV-1 antibody [17]. If each OS returns to Korea once every 2 or 3 years, the time of primary infection can be estimated relatively precisely in conjunction with previous negative results for anti-HIV-1 antibody. Setting a baseline CD4⁺ T cell count at 1,000/μL, the annual decline (AD) from this level to about 200/μL was calculated (AD >200). Subsequently, the AD from a CD4⁺ T cell of about 200/μL to the last CD4⁺ T cell count or death (0/μL) was also calculated (AD <200). The overall AD was defined as the rate from the first to the last measurement of CD4⁺ T cell count or to death (0/μL).

3.4. Comparison of AD in patients infected with subtypes B and D

Patients infected with subtype D showed a rapid decline in CD4⁺ T cell counts and progressed to AIDS within 10 years, although their exact times of initial infection were not known (Supplementary Fig. 3). Specifically, patients 90-15, 90-16, and 94-29 progressed to AIDS within 4–6 years. Their mean AD (*n* = 6) was 140 ± 97 cells/μL over 79 ± 50 months (Table 2 and Fig. 1). By comparison, the 157 patients infected with subtype B had a mean AD of 49 ± 58 cells/μL over 97 ± 59 months (Fig. 1). Taken together, the AD of subtype D–infected patients was 2.9-fold faster than that of subtype B–infected patients (*p* < 0.001; Fig. 1).

Of the six patients with subtype D, two were treated with KRG (643 ± 600 g/y) and four were not, with AD being mild significantly (4.5-fold) lower in the former (*p* = 0.07) (Fig. 2). Of the 157 patients

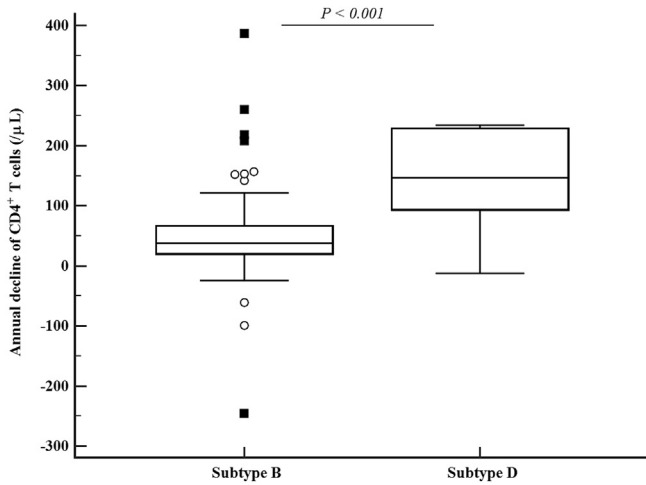


Fig. 1. Annual decline (AD) in CD4⁺ T cell counts in patients infected with HIV-1 subtypes B and D. The intervals between the first and last CD4⁺ T cell measurements before combination antiretroviral therapy (cART) or death in patients with subtypes D and B were 79 ± 50 months and 97 ± 59 months, respectively. In Table 2, the AD of subtype D–infected patients was 140 ± 97/µL, which was significantly higher than that of subtype B–infected patients (49 ± 58). Thus, the AD of subtype D–infected patients was 2.9-fold faster than that of subtype B–infected patients.

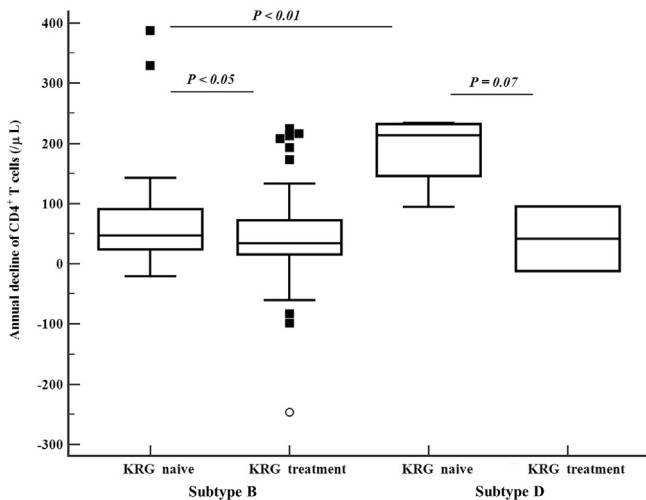


Fig. 2. Korean Red Ginseng (KRG) significantly affects the annual decline (AD) in CD4⁺ T cell counts in patients with HIV-1 subtypes B and D. AD was significantly higher in KRG-naïve patients with subtype D than in those with subtype B and was significantly lower in KRG-treated patients than in KRG-naïve patients with subtype B. These findings suggest that the effect of KRG treatment might be greater in patients infected with subtype D than subtype B (4.5-fold vs. 1.6-fold).

infected with subtype B, 116 were treated with KRG (539 ± 453 g/y) and 41 were not (Table 1). Mean AD was significantly lower in those treated (44 ± 61 cells/µL for 107 ± 59 months) than in those not (70 ± 82 cells/µL over 80 ± 44 months) treated with KRG ($p < 0.05$; Fig. 2). A comparison in patients not treated with KRG showed that AD was 2.8-fold higher in patients infected with subtype D than subtype B ($p < 0.01$; Fig. 2). Moreover, these findings suggest that the effect of KRG treatment might be greater in patients infected with subtype D than subtype B (4.5-fold vs. 1.6-fold).

3.5. Comparison of AD by CD4⁺ T cell counts in patients infected with subtypes B and D

A CD4⁺ T cell count below 200/µL is defined as AIDS. Therefore, the rate of progression may differ in patients experiencing declines

in CD4⁺ T cell counts from 1,000 to 200/µL and from 200 to 0/µL. Independent of KRG treatment, AD during the decline from 1,000 to 200/µL was significantly higher (4.9-fold) in patients infected with subtype D (251 ± 132 cells/µL) than in those infected with subtype B (52 ± 68 cells/µL) ($p < 0.0001$), whereas the difference was not significant (1.4-fold) during the decline from <200 to 0/µL (Fig. 3). This may due to the difference in the proportion of KRG-treated patients among subtype D–infected patients between both periods.

3.6. Effect of KRG treatment on AD

Of the six patients infected with subtype D, two had taken KRG, patient 92-65 when CD4⁺ T cell count was >200/µL and patient 93-01 when CD4⁺ T cell count was <200/µL. Patient 93-01 maintained CD4⁺ T cell counts for 13 years without cART (Supplementary Fig. 3). Of the 157 patients infected with subtype B, 116 had taken a mean 4,359 ± 5,141 g of KRG (median, 2,400 g; range, 30–25,602 g), beginning in November 1991 (Table 1).

In the KRG-naïve patients, AD was significantly greater in patients infected with subtype D than subtype B during the decline in CD4⁺ T cell counts from 1,000 to 200/µL (3.9-fold, $p < 0.0001$; Fig. 4A). In the same way, AD was greater in patients infected with subtype D than in those infected with subtype B during the decline in CD4⁺ T cell counts from <200 to 0/µL (1.8-fold, $p > 0.05$; Fig. 4B).

Regarding the KRG effect, during the decline in CD4⁺ T cell counts from 1,000 to 200/µL, AD was significantly lower in subtype B–infected patients treated with KRG than in those not treated with KRG (KRG-naïve patients) ($p < 0.05$; Fig. 4A). During the decline in CD4⁺ T cell counts from 200 to 0/µL, AD was lower in the one KRG-treated than in the four KRG-naïve patients infected with subtype D (Fig. 4B).

3.7. Survival analysis

The survival durations (SDs) from diagnosis to death or last CD4⁺ T cell count in patients infected with subtypes D and B were 82 ± 51 months (range, 24–140 months) and 105 ± 58 months (range, 4–339 months), respectively (Table 1). Kaplan–Meier survival analysis showed that mortality rates were significantly higher

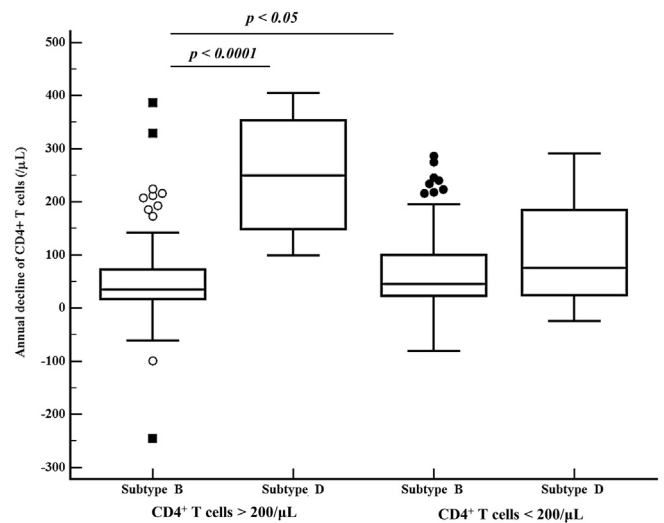


Fig. 3. Effect of CD4⁺ T cell counts and HIV-1 subtypes on the annual decline (AD) in CD4⁺ T cell counts in patients with HIV-1 subtypes B and D. AD was significantly higher in subtype D–infected patients than in subtype B–infected patients when CD4⁺ T cell counts were >200/µL.

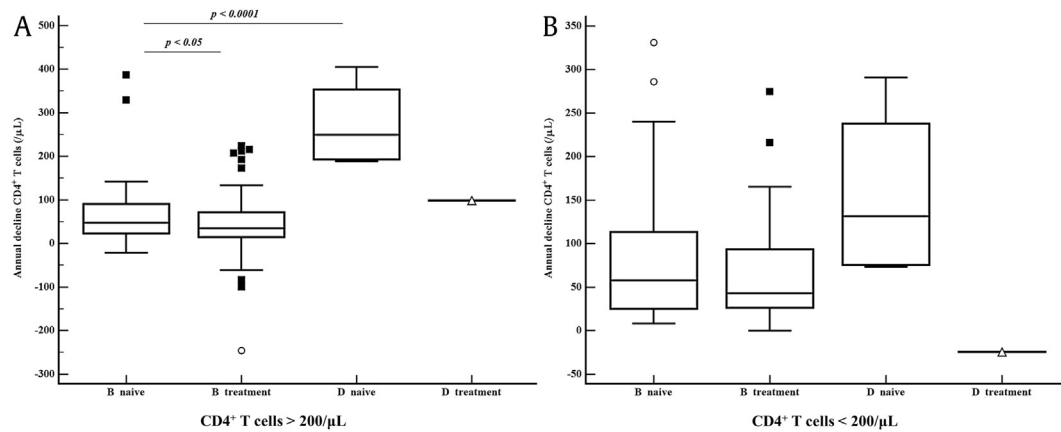


Fig. 4. Effect of Korean Red Ginseng (KRG) treatment and CD4⁺ T cell counts on the annual decline (AD) in CD4⁺ T cell counts. (A) CD4⁺ T cell counts were >200/μL. (B) CD4⁺ T cell counts were <200/μL. KRG treatment reduced AD significantly in subtype B–infected patients when CD4⁺ T cell counts were >200/μL ($p < 0.05$). Excluding patients treated with KRG, AD was 3.9-fold ($p < 0.0001$) and 1.7-fold more rapid in patients with subtype D than subtype B when CD4⁺ T cell counts were >200/μL and <200/μL, respectively.

in subtype D–infected patients than subtype B–infected patients, both for the 45 who were KRG-naïve ($p < 0.001$; Fig. 5) and for all patients ($p < 0.01$).

Among patients infected with subtype B, SD was significantly longer in those treated with KRG (112 ± 59 months) than in the KRG-naïve group (88 ± 48 months) ($p < 0.05$) (Fig. 6). Among patients infected with subtype D, SD was 2.1-fold longer in those who were treated with KRG than in the KRG-naïve group (125 ± 21 months vs. 60 ± 48 months; Fig. 6). Taken together, the prolongation of SD due to KRG intake was 2.7-fold longer in subtype D–infected patients (65 months) than in subtype B–infected patients (24 months), although infection with subtype D was associated with a >2-fold higher risk of death than infection with subtype B ($p < 0.05$; Table 1).

3.8. CXCR4 versus CCR5 tropism

We determined *env* tropism using the program Geno2pheno [coreceptor] 2.5 (<http://coreceptor.bioinf.mpi-inf.mpg.de/>). Patient 93-01 had dual tropism (CCR5 > CXCR4), patients 90-15 and 92-65 had CXCR4 tropism, and patients 88-05 and 90-16 had CCR5 tropism with a false positive rate of 10%. However, the 11/25

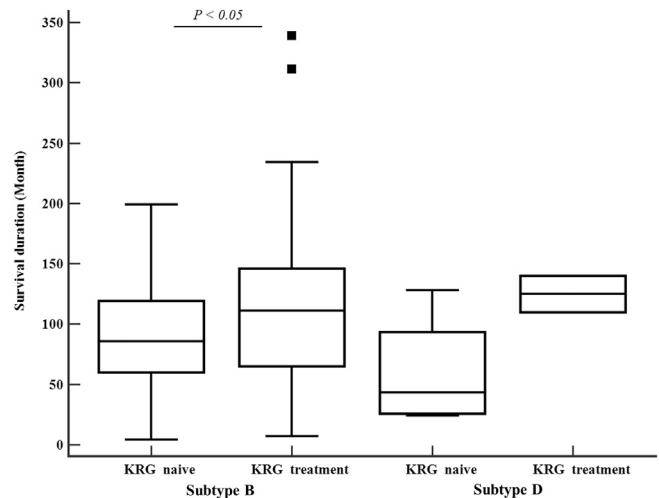


Fig. 6. Effects of HIV-1 subtype and treatment with Korean Red Ginseng (KRG) on mean survival duration (SD). SD was significantly longer in KRG-treated (B treatment) than in KRG-naïve patients with subtype B (B naïve) and tended to be longer in KRG-treated patients (D treatment) than in KRG-naïve patients with subtype D (D naïve), with KRG treatment enhancing SD more in patients with subtype D than with subtype B.

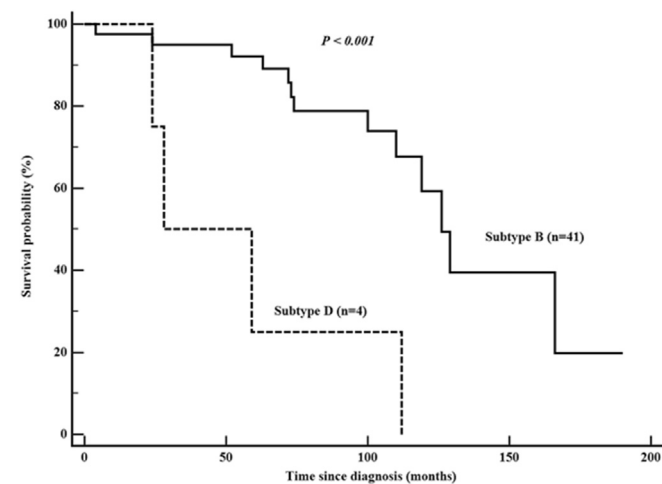


Fig. 5. Kaplan–Meier survival curves for 45 Korean Red Ginseng (KRG)-naïve patients with HIV-1 subtypes B and D from the time of diagnosis of HIV-1 infection to death or start of combination antiretroviral treatment.

[32] would predict the sequence from four patients (90-15, 90-16, 92-65, and 93-01) to be from X4 virus. Interestingly, the two patients who survived for more than 10 years (patients 88-05 and 93-01) were CCR5 carriers (Supplementary Fig. 3).

4. Discussion

This study of HIV-1–infected Korean men followed since 1988 showed that infection with subtype D was associated with a more rapid decline in CD4⁺ T cell counts and a 2.8-fold higher mortality rate, regardless of CD4⁺ T cell counts and KRG treatment than infection with subtype B. This study is unique, in that all of the patients with subtype D were men, and HIV-1 subtyping was based on the *nef* gene although the *env* gene was also determined in five patients. In addition, subtype D–infected patients in this study were followed up for a long period of time. Moreover, SD in subtype B–infected patients who were not treated with KRG (88 ± 48 months) was significantly longer than in our previous study of KRG-naïve patients in whom the HIV-1 subtype was not determined (59 ± 40 months) ($p < 0.001$) [28]. To our knowledge, this report is

the first to compare disease progression in Asian patients infected with HIV-1 subtypes B and D. Our results are similar to those of Esterbrook et al [7], who reported that subtype D–infected patients had a 4-fold higher rate of CD4⁺ T cell decline than subtype B–infected patients in England. The faster rate of CD4 decline in subtype D–infected patients remained significant after adjustment for ethnicity, gender, and baseline CD4⁺ T cell count [7]. In the present study, the AD of subtype D–infected patients was 2.9-fold faster than that of subtype B–infected patients, which was slightly slower than that among English patients described earlier. This is probably due to the fact that the efficacy of KRG in subtype D–infected patients was higher (4.5-fold) than that in subtype B–infected patients (1.6-fold), as shown in Fig. 2.

Among Asian patients infected with non-B subtypes, CRF01_AE is the dominant subtype, with subtype D being very rare [13–15]. To date, no report has compared Asian patients infected with subtypes D and B, although one study compared disease progression rates in Asian patients infected with CRF01_AE and subtype B [33]. CD4⁺ T cell decline is generally significantly more rapid (4-fold) in patients infected with subtype D than with other subtypes, a decline reported to result from subtype differences in the levels of replicative fitness and virulence in the order D > A > C ($p < 0.001$) [34]. That is, patients infected with subtype D would be expected to have a high plasma viral load accompanying the rapid decline in CD4⁺ T cell counts. The replication capacity is significantly higher for subtype B than subtype C [35]. However, only one report to date has compared the progression rates of subtypes D and B, although several have compared their replication capacity [36].

In the present study, all subtype D patients showed rapid declines in CD4⁺ T cell counts from diagnosis or primary infection to around 200/μL and slow decline from a CD4⁺ T cell count of about 200/μL to the last CD4⁺ T cell count or death. Three subtype D–infected patients (patients 88-05, 90-16, and 93-01) had relatively low viral loads raising questions regarding their very rapid reductions in CD4⁺ T cell counts to <200 cells/μL. Furthermore, patients 88-05 and 93-01, with low viral loads at baseline, survived for >10 years after diagnosis in the absence of cART. Their low viral load and long-term survival may have resulted from CCR5 tropism. Patient 93-01 had a greater degree of dual tropism than other patients, which explains, at least in part, the finding that subtype D was associated with a faster rate of disease progression than other HIV-1 subtypes [37]. Additionally, patient 93-01 was treated with KRG. Finally, he had the same 15-bp deletion in the *nef* gene (Δ15-bp; amino acids 9–13), which was present from the earliest samples in December 1992 (AY584798), prior to KRG treatment, to the final samples (Supplementary Fig. 2). Similarly, patient 87-05, who had survived 30 years in the absence of antiretroviral treatment, harbored a 9-bp deletion in *nef* (amino acids 9–11), irrespective of KRG treatment [29,30].

We also determined the sequences of *vif* and *env* genes in six subtype D–infected patients, with the exception of patient 94-29 (Supplementary Figs. 4 and 5). In 1988–1992, various non-B subtypes were transmitted to Korea through OSs [17]. As some of these were homosexual, recombinant strains of KSB and non-B may have arisen, as in patient 92-65, who was infected with subtype D based on the *nef* gene. Some of the OSs are likely to be bisexual, suggesting that they may have spread HIV-1 subtype D to homosexuals who were not OSs, such as patient 93-01. In detail, the *pol* and *vif* genes and 5' two-thirds of the *env* gene (MH425163-64) of patient 92-65 were found to be KSB, with the 3' terminal one-third of the *env* gene being subtype D, suggesting that the strain in this patient may be a recombinant of KSB and subtype D. In contrast to the rapid reduction in CD4⁺ T cell counts observed in other patients infected with subtype D, patient 92-65 experienced a gradual reduction over 9 years, suggesting that the biologically important *env* gene

was replaced by KSB. The *vif* and *env* genes in the remaining four patients were of subtype D. Further research is needed to determine whether the six patients classified as having subtype D were infected with full-length HIV-1 subtype D.

The present study provides further evidence that subtype D may be more pathogenic than subtype B. Interestingly, recent studies suggest that subtype D may be decreasing in prevalence [38], consistent with the notion that more pathogenic variants may be less fit for transmission [39]. To date, the biological mechanisms that cause such differences between subtypes are unknown. This study, as well as a previous study [3], showed that increased pathogenesis of subtype D is not simply the result of increased replication because the differences in disease progression were independent of plasma HIV-1 load. Subtype D has been reported more likely to utilize the coreceptor CXCR4 than other subtypes [10] and to develop CXCR4 tropism earlier after infection [2,34]. The presence of a more facile coreceptor switch provides an attractive model for the increased virulence of subtype D because the emergence of CXCR4 variants has been linked to disease progression in patients infected with subtype B [39]. Such differences in coreceptor use could also explain differences in transmissibility [3]. More detailed studies of subtype D variants may help elucidate the biological properties that contribute to HIV-1 virulence.

The mechanism of the anti-HIV action of KRG was mentioned in previous studies [25–29]. In brief, significant decreases in serum p24 antigen as well as soluble CD8 antigen were observed [40–42]. The decrease in soluble CD8 antigen results in the potentiation of cytotoxic CD8⁺ T cell activity [28]. Furthermore, KRG treatment induces significant genetic defects in the proviral DNA of HIV-1 subtype B [28–30]. In addition, it is well known that many components of ginseng attenuate inflammatory cytokine production [43]. We also observed significant decreases in or a normal range of high-sensitivity C-reactive protein in HIV-1 patients treated with KRG, as evidenced in patient 93-01.

This study had several limitations, including the small number of patients infected with HIV-1 subtype D and the unknown primary infection date. In addition, full-length HIV-1 sequences were not determined. Despite these limitations, treatment with KRG affected the progression rate of both subtypes, with greater efficacy in subtype D patients than in subtype B.

In conclusion, the progression rate to AIDS was about 2.9-fold more rapid in patients infected with HIV-1 subtype D than subtype B. Further studies are needed to determine whether more rapid progression is due to other factors such as socioeconomic factors.

Conflicts of interest

All contributing authors declare no conflicts of interest.

Acknowledgments

This work was supported by a grant from the Korean Society of Ginseng (2016–2017).

Appendix A. Supplementary data

Supplementary data related to this article can be found at <https://doi.org/10.1016/j.jgr.2018.07.006>.

References

- [1] Kanki PJ, Hamel DJ, Sankalé JL, Hsieh Cc, Thior I, Barin F, Woodcock SA, Guèye-Ndiaye A, Zhang E, Montano M, et al. Human immunodeficiency virus type 1 subtypes differ in disease progression. *J Infect Dis* 1999;179:68–73.

- [2] Kaleebu P, French N, Mahe C, Yirrell D, Watera C, Lyagoba F, Nakiyingi J, Rutebemberwa A, Morgan D, Weber J, et al. Effect of human immunodeficiency virus (HIV) type 1 envelope subtypes A and D on disease progression in a large cohort of HIV-1-positive persons in Uganda. *J Infect Dis* 2002;185:1244–50.
- [3] Baeten JM, Chohan B, Lavreys L, Chohan V, McClelland RS, Certain L, Mandaliya K, Jaoko W, Overbaugh J. HIV-1 subtype D infection is associated with faster disease progression than subtype A in spite of similar plasma HIV-1 loads. *J Infect Dis* 2007;195:1177–80.
- [4] Morgan D, Kaleebu P, Whitworth J, Yirrell D, Rutebemberwa A, Shier R, Sebabi B, Gilmour J, Eotu H, Omoding N, et al. The stability between two HIV-1 RNA measurements one year apart and the relationship with HIV subtype in rural Uganda. *Int J STD AIDS* 2001;12:116–21.
- [5] Alaeus A, Lidman K, Björkman A, Giesecke J, Albert J. Similar rate of disease progression among individuals infected with HIV-1 genetic subtypes A-D. *AIDS* 1999;13:901–7.
- [6] Laurent C, Bourgeois A, Faye MA, Mougnotou R, Seydi M, Gueye M, Liégeois F, Kane CT, Butel C, Mbuagbaw J, et al. No difference in clinical progression between patients infected with the predominant human immunodeficiency virus type 1 circulating recombinant form (CRF) 02_AG strain and patients not infected with CRF02_AG, in Western and West-Central Africa: a four-year prospective multicenter study. *J Infect Dis* 2002;186:486–92.
- [7] Easterbrook PJ, Smith M, Mullen J, O'Shea S, Chrystie I, de Ruiter A, Tatt ID, Geretti AM, Zuckerman M. Impact of HIV-1 viral subtype on disease progression and response to antiretroviral therapy. *J Int AIDS Soc* 2010;13:4. <https://doi.org/10.1186/1758-2652-13-4>.
- [8] Amornkul PN, Karita E, Kamali A, Rida WN, Sanders EJ, Lakhi S, Price MA, Kilembe W, Cormier E, Anzala O, et al. IAVI Africa HIV Prevention Partnership. Disease progression by infecting HIV-1 subtype in a seroconverter cohort in sub-Saharan Africa. *AIDS* 2013 13;27:2775–86.
- [9] Bousheri S, Burke C, Ssewanyana I, Harrigan R, Martin J, Hunt P, Bangsberg DR, Cao H. Infection with different HIV subtypes is associated with CD4 activation-associated dysfunction and apoptosis. *J Acquir Immune Defic Syndr* 2009;52:548–52.
- [10] Tscherning C, Alaeus A, Fredriksson R, Björndal A, Deng H, Littman DR, Fenyö EM, Albert J. Differences in chemokine coreceptor usage between genetic subtypes of HIV-1. *Virology* 1998;241:181–8.
- [11] Kaleebu P, Nankya IL, Yirrell DL, Shafer LA, Kyosiimire-Lugemwa J, Lule DB, Morgan D, Beddows S, Weber J, Whitworth JA. Relation between chemokine receptor use, disease stage, and HIV-1 subtypes A and D: results from a rural Ugandan cohort. *J Acquir Immune Defic Syndr* 2007;45:28–33.
- [12] Keyune F, Nankya I, Metha S, Akao J, Ndashimye E, Tebit DM, Rodriguez B, Kityo C, Salata RA, Mugenyi P, et al. JCRRC Drug Resistance Working Group. Treatment failure and drug resistance is more frequent in HIV-1 subtype D versus subtype A-infected Ugandans over a 10-year study period. *AIDS* 2013;27:1899–909.
- [13] Hemelaar J, Gouws E, Ghys PD, Osmanov S. Global trends in molecular epidemiology of HIV-1 during 2000–2007. *AIDS* 2011;25:679–89.
- [14] Buonaguro L, Tornesello M, Buonaguro FM. Human immunodeficiency virus type 1 subtype distribution in the worldwide epidemic: pathogenetic and therapeutic implications. *J Virol* 2007;81:10209–19.
- [15] Fujisaki S, Ibe S, Hattori J, Shigemitsu U, Fujisaki S, Shimizu K, Nakamura K, Yokomaku Y, Mamiya N, Utsumi M, et al. An 11-year surveillance of HIV type 1 subtypes in Nagoya, Japan. *AIDS Res Hum Retroviruses* 2009;25:15–21.
- [16] Castley A, Sawleshwarkar S, Varma R, Herring B, Thapa K, Dwyer D, Chibo D, Nguyen N, Hawke K, Ratcliff R, et al. A national study of the molecular epidemiology of HIV-1 in Australia 2005–2012. *PLoS One* 2017;12:e0170601.
- [17] Cho YK, Kim JE, Foley BT. High prevalence of Non-B HIV-1 subtypes in overseas sailors and prostitutes in Korea. *AIDS Res Hum Retroviruses* 2018;34:347–53.
- [18] Chakrabarti L, Baptiste V, Khatissian E, Cumont MC, Aubertin AM, Montagnier L, Hurtrel B. Limited viral spread and rapid immune response in lymph nodes of macaques inoculated with attenuated simian immunodeficiency virus. *Virology* 1995;213:535–48.
- [19] Desrosiers RC, Lifson JD, Gibbs JS, Czajak SC, Howe AY, Arthur LO, Johnson RP. Identification of highly attenuated mutants of simian immunodeficiency virus. *J Virol* 1998;72:1431–7.
- [20] Hofmann-Lehmann R, Vlasak J, Williams AL, Chenine AL, McClure HM, Anderson DC, O'Neil S, Rupprecht RM. Live attenuated, *nef*-deleted SIV is pathogenic in most adult macaques after prolonged observation. *AIDS* 2003;17:157–66.
- [21] Iafraite AJ, Carl S, Bronson S, Stahl-Hennig C, Swigut T, Skowronski J, Kirchhoff F. Disrupting surfaces of *nef* required for downregulation of CD4 and for enhancement of virion infectivity attenuates simian immunodeficiency virus replication in vivo. *J Virol* 2000;74:9836–44.
- [22] Kestler 3rd HW, Ringler DJ, Mori K, Panicali DL, Sehgal PK, Daniel MD, Desrosiers RC. Importance of the *nef* gene for maintenance of high virus loads and for development of AIDS. *Cell* 1991;65:651–62.
- [23] Homann S, Tibroni N, Baumann I, Sertel S, Keppler OT, Fackler OT. Determinants in HIV-1 Nef for enhancement of virus replication and depletion of CD4+ T lymphocytes in human lymphoid tissue *ex vivo*. *Retrovirology* 2009;6:6. <https://doi.org/10.1186/1742-4690-6-6>.
- [24] Schindler M, Münch J, Kutsch O, Li H, Santiago ML, Bibollet-Ruche F, Müller-Trutwin MC, Novembre FJ, Peeters M, Courgnaud V, et al. Nef-mediated suppression of T cell activation was lost in a lentiviral lineage that gave rise to HIV-1. *Cell* 2006;125:1055–67.
- [25] Cho YK, Kim JE, Foley BT. Phylogenetic analysis of the earliest *nef* gene from hemophiliacs and local controls in Korea. *Biores Open Access* 2012;1:41–9.
- [26] Cho YK, Kim YK, Lee I, Choi MH, Shin YO. The effect of Korean red ginseng (KRG), zidovudine (ZDV), and the combination of KRG and ZDV on HIV-infected patients. *J Korean Soc Microbiol* 1996;31:353–60.
- [27] Cho YK, Lee HJ, Kim YB, Oh WI, Kim YK. Sequence analysis of C2/V3 region of human immunodeficiency virus type 1 gp120 and its correlation with clinical significance: the effect of long-term intake of KRG on *env* gene variation. *J Korean Soc Microbiol* 1997;32:611–23.
- [28] Cho YK, Kim JE. Effect of Korean red ginseng intake on the survival duration of human immunodeficiency virus type 1 patients. *J Ginseng Res* 2017;41:222–6.
- [29] Cho YK, Lim JY, Jung YS, Oh SK, Lee HJ, Sung H. High frequency of grossly deleted *nef* genes in HIV-1 infected long-term slow progressors treated with Korean red ginseng. *Curr HIV Res* 2006;4:447–57.
- [30] Cho YK, Kim JE, Woo JH. Genetic defects in the *nef* gene are associated with Korean red ginseng intake: monitoring of *nef* sequence polymorphisms over 20 years. *J Ginseng Res* 2017;41:144–50.
- [31] Griffith BP, Rigsby MO, Garner RB, Gordon MM, Chacko TM. Comparison of the Amplicor HIV-1 monitor test and the nucleic acid sequence-based amplification assay for quantitation of human immunodeficiency virus RNA in plasma, serum, and plasma subjected to freeze-thaw cycles. *J Clin Microbiol* 1997;35:3288–91.
- [32] de Jong JJ, Goudsmit J, Keulen W, Klaver B, Krone W, Tersmette M, de Ronde A. Human immunodeficiency virus type 1 clones chimeric for the envelope V3 domain differ in syncytium formation and replication capacity. *J Virol* 1992;66:757–65.
- [33] Lee LK, Lin L, Chua A, Leo YS, Ng OT. Poorer immunologic outcome on treatment among patients infected with HIV-1 non-B subtypes compared with subtype B in Singapore. *Clin Infect Dis* 2012;54:1818–20.
- [34] Vasan A, Renjifo B, Hertzmark E, Chaplin B, Msamanga G, Essex M, Fawzi W, Hunter D. Different rates of disease progression of HIV type 1 infection in Tanzania based on infecting subtype. *Clin Infect Dis* 2006;42:843–52.
- [35] Kiguoya MW, Mann JK, Chopera D, Gounder K, Lee GQ, Hunt PW, Martin JN, Ball TB, Kimani J, Brumme ZL, et al. Subtype-specific differences in gag-protease-driven replication capacity are consistent with intersubtype differences in HIV-1 disease progression. *J Virol* 2017;91. e00253-17.
- [36] Abrahama A, Nankya IL, Gibson R, Demers K, Tebit DM, Johnston E, Katzenstein D, Siddiqui A, Herrera C, Fischetti L, et al. CCR5- and CXCR4-tropic subtype C human immunodeficiency virus type 1 isolates have a lower level of pathogenic fitness than other dominant group M subtypes: implications for the epidemic. *J Virol* 2009;83:5592–605.
- [37] Huang W, Eshleman SH, Toma J, Franssen S, Stawiski E, Paxinos EE, Whitcomb JM, Young AM, Donnell D, Mmoro F, et al. Coreceptor tropism in human immunodeficiency virus type 1 subtype D: high prevalence of CXCR4 tropism and heterogeneous composition of viral populations. *J Virol* 2007;81:7885–93.
- [38] Rainwater S, DeVange S, Sagar M, Ndinya-Achola J, Mandaliya K, Kreiss JK, Overbaugh J. No evidence for rapid subtype C spread within an epidemic in which multiple subtypes and intersubtype recombinants circulate. *AIDS Res Hum Retroviruses* 2005;21:1060–5.
- [39] Overbaugh J, Bangham CR. Selection forces and constraints on retroviral sequence variation. *Science* 2001;292:1106–9.
- [40] Cho YK, Kim YB, Choi BS, Kim YK, Choi MH, Jang YS, Shin YO. Effect of Korean red ginseng on the levels of serum p24 antigen, β_2 -microglobulin, and CD4+ T cells in HIV infected patients treated with AZT (I). *J Korean Soc Microbiol* 1993;28:409–317.
- [41] Sung HS, Kang SM, Lee MS, Kim TG, Cho YK. Korean Red Ginseng slows depletion of CD4 T cells in HIV type 1-infected patients. *Clin Diagn Lab Immunol* 2005;12:497–501.
- [42] Cho Y, Sung H. Effect of Korean Red Ginseng on serum soluble CD8 in HIV-1-infected patients. *J Ginseng Res* 2007;31:175–80.
- [43] Kim JH, Yi YS, Kim MY, Cho JY. Role of ginsenosides, the main active components of *Panax ginseng*, in inflammatory responses and diseases. *J Ginseng Res* 2017;41:435–43.