



Draft Genome Sequences of Two *Clostridium* Isolates from the Poultry Gastrointestinal Tract

 Jitendra Keshri,^a Rocio Ramirez,^{a,b} Raja Chalhouni,^c Johnna K. Garrish,^d Bruce S. Seal,^e Brian B. Oakley^a

^aCollege of Veterinary Medicine, Western University of Health Sciences, Pomona, California, USA

^bGraduate College of Biomedical Sciences, Western University of Health Sciences, Pomona, California, USA

^cAnimal Science Department, College of Agriculture of Mateur, University of Carthage, Mateur, Bizerte, Tunisia

^dPoultry Microbiological Safety and Processing Research Unit, USDA, Agricultural Research Service, U.S. National Poultry Center, Athens, Georgia, USA

^eBiology Program, Oregon State University Cascades, Bend, Oregon, USA

ABSTRACT Here, we announce the draft genome sequences of two *Clostridium* strains, C8-1-8 and C2-6-12, isolated from the cecal contents of commercial broiler chickens (in Athens, GA). These strains may represent potentially novel species within the genus *Clostridium*, and these draft genomes allow further investigation into potential probiotics for poultry.

The taxonomy and systematics of the genus *Clostridium* are complex and unresolved, but a total of 231 recognized species belonging to the genus *Clostridium* were reported as of 2019 (<http://www.bacterio.net/clostridium.html>). We isolated two *Clostridium* strains, designated strains C8-1-8 and C2-6-12, from the ceca of commercial broiler chickens collected in Athens, Georgia. These strains may represent potentially novel species, as suggested by a comparison based on whole-genome sequencing. The strains were isolated from cecal contents treated with 3% chloroform to eliminate vegetative bacterial cells (1, 2) and cultured in an anaerobic environment on gut microbiota medium (3). Single colonies were subsequently propagated overnight in Luria-Bertani broth for DNA extraction from cultures using the MO BIO UltraClean microbial DNA isolation kit. Initial characterizations of the isolates were completed by Gram staining followed by 16S rRNA gene sequence analyses. The genomes of both strains were sequenced using the Illumina NovaSeq platform through the Institute for Integrative Genome Biology Genomics Core, University of California, Riverside.

Libraries with a fragment length of 150 bp were constructed using the plexWell 96 library preparation kit for Illumina sequencing platforms (seqWell, Inc., Beverly, MA, USA), and totals of 9,074 and 9,415 Mb of paired-end (150-bp) reads were generated for strains C8-1-8 and C2-6-12, respectively. Default parameters for all software were used during analyses unless otherwise specified. Quality filtering of the reads was performed using Trimmomatic v0.39 (4). High-quality reads were used for *de novo* genome assembly with SPAdes v3.12.0 using the careful assembly method (5). Scaffolds were filtered for a minimum of 200-bp read length and 30× coverage. The quality of the subsequent assemblies was assessed using QUAST (6). Genome completeness was found to be 99.1% for both strains, using the CheckM lineage (7) and Microbial Genomes Atlas (MiGA) (<http://microbial-genomes.org>) (8) workflows. The species identification tool *specl*, based on 40 universal single-copy marker genes (9), confirmed that both genomes could not be assigned to any species cluster. Similarly, neither genome could be assigned to a species when compared against reference genomes from the NCBI RefSeq database using MiGA (8). The results of assembly quality analysis by the MiGA workflow based on 106 essential genes were rated as intermediate and high for C8-1-8 and C2-6-12, respectively. The assembled genomes were annotated using the

Citation Keshri J, Ramirez R, Chalhouni R, Garrish JK, Seal BS, Oakley BB. 2020. Draft genome sequences of two *Clostridium* isolates from the poultry gastrointestinal tract. *Microbiol Resour Annu* 9:e00137-20. <https://doi.org/10.1128/MRA.00137-20>.

Editor David A. Baltrus, University of Arizona

Copyright © 2020 Keshri et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Brian B. Oakley, boakley@westernu.edu.

Received 10 March 2020

Accepted 15 April 2020

Published 28 May 2020

TABLE 1 Assembly and annotation metrics

| Statistic | Data for strain: | |
|---|------------------|-----------------|
| | C8-1-8 | C2-6-12 |
| No. of quality-filtered paired-end reads | 11,102,701 | 10,624,933 |
| No. of scaffolds | 116 | 121 |
| N_{50} (bp) | 177,332 | 126,571 |
| Largest scaffold length (bp) | 442,315 | 352,209 |
| Assembly length (bp) | 6,053,167 | 6,076,981 |
| Avg genome coverage (×) | 448 | 471 |
| G+C content (%) | 31.6 | 29.02 |
| No. of rRNA genes | 14 | 10 |
| No. of tRNA genes | 47 | 57 |
| No. of functional protein-coding genes | 3,956 | 4,254 |
| No. of hypothetical protein-coding genes | 1,413 | 1,373 |
| No. of genes associated with KEGG pathways | 1,209 | 1,354 |
| No. of genes associated with KEGG orthology | 3,091 | 2,350 |
| GenBank accession no. | JAABNO000000000 | JAABNP000000000 |
| SRA accession no. | SRR10903102 | SRR10903103 |

IMG/MER pipeline (10), and average coverage was calculated using the BMap tool. Assembly and annotation metrics are presented in Table 1.

Based on the compare genome analysis function in IMG, which uses the genome-wide average nucleotide identity (ANI), the maximum pairwise ANI of strain C8-1-8 with available *Clostridium* genomes ($n = 756$) was 92.6% with *Clostridium* sp. strain CT4 (GenBank accession number CP025746; aligned fraction [AF], 79.7%). The maximum ANI of strain C2-6-12 with known *Clostridium* genomes was 84.8% with *Clostridium puniceum* DSM 2619 (GenBank accession number LZM00000000.1; IMG genome identification number 2703719317; AF, 53.8%). Comparing our genomes to all available genomes of *Clostridium* type strains ($n = 80$), the maximum ANI for strain C8-1-8 was 73.3% with *Clostridium fallax* DSM 2631 (AF, 17.8%) and the maximum ANI for strain C2-6-12 was 84.8% (AF, 53.8%) with *C. puniceum* BL70/20. The ANI between our two isolates was only 72.57%, suggesting that strains C8-1-8 and C2-6-12 are distantly related to each other.

Data availability. The draft genome and raw read sequences for both strains have been deposited in DDBJ/EMBL/GenBank under the accession numbers listed in Table 1.

ACKNOWLEDGMENTS

Funding was provided by USDA NIFA grants (1015210 and 1011327) to Western University of Health Sciences College of Veterinary Medicine and B.B.O. R.C. was supported by a Non-U.S. (Visiting) Scholar Grant from the Fulbright Visiting Scholar Program (grant 68130564), Council for International Exchange of Scholars, Institute of International Education, and hosted by B.S.S. and B.B.O. at the Richard B. Russell Agricultural Research Center, Agricultural Research Service, USDA (Athens, GA) (ARS CRIS project 6612-32000-060).

REFERENCES

- Itoh K, Mitsuoka T. 1980. Production of gnotobiotic mice with normal physiological functions. I. Selection of useful bacteria from feces of conventional mice. *Z Versuchstierkd* 22:173–178.
- Atarashi K, Tanoue T, Shima T, Imaoka A, Kuwahara T, Momose Y, Cheng G, Yamasaki S, Saito T, Ohba Y, Taniguchi T, Takeda K, Hori S, Ivanov II, Umesaki Y, Itoh K, Honda K. 2011. Induction of colonic regulatory T cells by indigenous *Clostridium* species. *Science* 331:337–341. <https://doi.org/10.1126/science.1198469>.
- Goodman AL, Kallstrom G, Faith JJ, Reyes A, Moore A, Dantas G, Gordon JL. 2011. Extensive personal human gut microbiota culture collections characterized and manipulated in gnotobiotic mice. *Proc Natl Acad Sci U S A* 108:6252–6257. <https://doi.org/10.1073/pnas.1102938108>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUILT: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>.
- Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>.
- Rodríguez-R LM, Gunturu S, Harvey WT, Rosselló-Mora R, Tiedje JM, Cole JR, Konstantinidis KT. 2018. The Microbial Genomes Atlas (MiGA)

- webservice: taxonomic and gene diversity analysis of *Archaea* and *Bacteria* at the whole genome level. *Nucleic Acids Res* 46:W282–W288. <https://doi.org/10.1093/nar/gky467>.
9. Mende DR, Sunagawa S, Zeller G, Bork P. 2013. Accurate and universal delineation of prokaryotic species. *Nat Methods* 10:881–884. <https://doi.org/10.1038/nmeth.2575>.
 10. Chen IA, Chu K, Palaniappan K, Pillay M, Ratner A, Huang J, Huntemann M, Varghese N, White JR, Seshadri R, Smirnova T, Kirton E, Jungbluth SP, Woyke T, Elie-Fadrosh EA, Ivanova NN, Kyrpides NC. 2019. IMG/M v.5.0: an integrated data management and comparative analysis system for microbial genomes and microbiomes. *Nucleic Acids Res* 47:D666–D677. <https://doi.org/10.1093/nar/gky901>.