



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.

Clinical Microbiology Newsletter

Vol. 23, No. 21

November 1, 2001

Antiviral Agents Against Respiratory Viruses

Cecile L. Tremblay, M.D.
*Infectious Disease Unit
Massachusetts General Hospital
GRB-05-04
Boston, MA 02114*

Introduction

Respiratory virus infections are important causes of mortality and morbidity. They also have social and economic impact, contributing to lost days from work for patients and families. Advances have been made in the development of antiviral agents to treat these infections. These advances and future prospects will be reviewed.

Respiratory Viruses

Several viruses are known to cause respiratory tract infections, including influenza A, B, and C; parainfluenza 1, 2, and 3; respiratory syncytial virus (RSV); adenovirus; rhinoviruses; and coronaviruses. Currently, antiviral agents are available for only two of these virus groups: influenza and RSV.

Influenza

Amantadine and Rimantadine

Spectrum

Amantadine (1-adamantanamine hydrochloride) and rimantadine (α -methyl-1-adamantane methylamine hydrochloride) are related, symmetric, tricyclic amines. Both inhibit replication of influenza A viruses only, at concentrations less than 1 μ g/ml. They are active against several strains of influenza A virus, including H1N1, H2N2, and H3N2 subtypes. Rimantadine is 4 to 10 times more active than amantadine in vitro. At higher concentrations, which are not achievable clinically

because of toxicity, rimantadine has in vitro activity against other enveloped viruses, such as influenza B, parainfluenza, rubella, and dengue.

Mechanism of action

At low concentrations, these drugs act by blocking the ion channel formed by the M2 protein spanning the viral membrane. This affects viral uncoating or disassembly of the virion during endocytosis (1). As hydrogen ions enter endocytotic vesicles, the pH falls. The hydrogen ions pass through the M2 channel into the interstices of the viral particle and promote the dissociation of the M1 protein from the ribonucleoprotein complexes so that the ribonucleoprotein can enter the cell nucleus and initiate replication. Amantadine and rimantadine enter the ion channel and block penetration by hydrogen ions, thereby preventing the dissociation of M1 from the ribonucleoprotein. Amantadine and rimantadine are also concentrated in the lysosomal fraction of mammalian cells, increasing lysosomal pH. This pH increase may inhibit virus-induced membrane fusion events and partly explain the broader antiviral spectrum observed at higher concentrations (2,3).

Resistance

Resistant viruses can be readily selected by in vitro virus passage in the presence of amantadine or rimantadine (4). Resistant strains can also arise following treatment with either drug. From 25 to 35% of treated patients will shed resistant virus by the fifth day of therapy (5). Single nucleotide mutations leading to amino acid changes in the M2 protein have been shown to confer

resistance to amantadine or rimantadine. The most common mutation site observed clinically is amino acid 31 (6). Amantadine and rimantadine share cross-susceptibility and cross-resistance.

Avian models have shown that resistant influenza strains are genetically stable and have the same virulence and fitness as wild-type isolates (7). In humans, transmission of resistant virus, associated with failure of drug prophylaxis, has been documented in household contacts of rimantadine-treated index cases and in nursing home residents receiving amantadine (8,9). Infections caused by a resistant strain do not lead to prolonged illness or a rebound of illness and are similar to infections caused by a susceptible strain (8). However, it is prudent to avoid use

In This Issue

Antiviral Agents Against Respiratory Viruses 163

During the last decade significant changes have occurred in the rapid diagnosis and treatment of respiratory viral infections. It is now possible to detect the most common respiratory viruses, such as influenza and respiratory syncytial virus (RSV) within hours of specimen collection. Some rapid tests can be performed point-of-care, and certain methods even allow differentiation of influenza A from influenza B when both are endemic in the community. This article reviews the older and newer antiviral agents that can be used to treat influenza and RSV and how their use relates to specific viral diagnosis.

Table 1. Antiviral drugs approved for prevention or treatment of influenza infections

| Drug | Trade name | Influenza type | Cost (US\$) ^a | Approved age (yr) | Route | Dose (for 5 d) | Half-life (h) |
|-------------|------------|----------------|--------------------------|-------------------------------------|---------|----------------|---------------|
| Amantadine | Symetrel | A | 9.83 | ≥1 | PO | 100 mg bid | 15 adults |
| | Generic | | 1.72 | | | | 30 elderly |
| Rimantadine | Flumadine | A | 18.87 | ≥1 ^b ≥18 ^c | PO | 200 mg/d | 30 |
| Zanamivir | Relenza | A and B | 44.49 | ≥7 | Aerosol | 10 mg bid | NA |
| Oseltamivir | Tamiflu | A and B | 53.00 | ≥18 | PO | 75 mg bid | 6-10 |

^a Costs for a 5 day treatment^b For prophylaxis^c For treatment

of both treatment and postexposure prophylaxis in the same household and to avoid contact between susceptible high-risk individuals and treated patients. Naturally occurring polymorphisms that confer resistance to amantadine or rimantadine have been detected in some H1N1 isolates collected between 1933 and 1945, before the introduction of amantadine, underscoring the potential for emergence of viral resistance (3).

Pharmacokinetics

Amantadine

Dosage: Amantadine is available as tablets, capsules, or syrup formulations. It is absorbed rapidly and almost completely 2 h after oral administration. The average steady-state peak concentrations range from 0.5 to 0.8 µg/ml, following the recommended dose of 100 mg twice daily (Table 1). Concentrations achieved in nasal secretions and saliva are similar to those in serum. Cerebrospinal fluid (CSF) levels are one-half of those in plasma. Plasma protein binding of amantadine is about 67%.

Metabolism: Amantadine is excreted unmetabolized in the urine through glomerular filtration and possibly through tubular secretion. The dose of amantadine should be reduced in renal insufficiency. Amantadine is not cleared by hemodialysis, therefore supplemental doses are not required (10). The plasma half-life is approximately 12 to 18 h in subjects with normal renal function but may be as long as 30 h in sub-

jects with renal insufficiency. In particular, because of age-related decrease in renal function, the half-life is increased up to twofold in the elderly, and they require only half of the dose needed for young adults to achieve trough plasma levels of 0.3 µg/ml (11).

Rimantadine

Dosage: Rimantadine is available as tablet or syrup formulations. It is nearly completely absorbed after oral administration, but the time to the peak plasma concentration is about twice that of amantadine (2 to 6 h). The average steady-state peak concentrations are 0.4 to 0.5 µg/ml in healthy young adults; however, levels are increased in elderly subjects indicating the need to lower the doses in such patients. Concentrations in nasal mucus average 50% higher than those in plasma. Rimantadine plasma protein binding is about 40% (12).

Metabolism: Rimantadine undergoes extensive metabolism by hydroxylation, conjugation, and glucuronidation, and three hydroxylated metabolites have been described. It is excreted in the urine. Its plasma half-life averages 24 to 36 h. Subjects with severe hepatic dysfunction and renal insufficiency with creatinine clearance of <10 ml/min require a dose reduction. No supplemental dose is required for hemodialysis.

Drug interactions

Antihistamines or anticholinergic drugs increase the effects of amantadine

on the central nervous system (CNS). CNS toxicity has also been associated with concurrent use of trimethoprim-sulfamethoxazole or triamterene-hydrochlorothiazide, due to decreased renal clearance (13). Patients receiving drugs likely to affect CNS function, such as antihistamines, antidepressants, and benzodiazepines, should be monitored closely. Cimetidine induces a 15 to 20% increase and aspirin or acetaminophen a 10% decrease in rimantadine concentration (14); these small changes are of unclear significance.

Toxicity

Adverse effects observed with the use of amantadine or rimantadine are usually mild in young, healthy adults. They include dose-related gastrointestinal and CNS side effects, such as nervousness, lightheadedness, difficulty concentrating, insomnia, and loss of appetite or nausea (15-17). These CNS side effects occur more frequently with the use of amantadine (5 to 33%) than rimantadine (2%) and are presumably due to amantadine's activity on the adrenergic nervous system, which affects accumulation, release, and re-uptake of catecholamines (18-20). During prophylaxis, drug discontinuation rates are usually less than 5% for rimantadine and range from 6 to 11% for amantadine (19). In elderly subjects or patients with renal failure, serious neurotoxic reactions have been reported with the use of amantadine, including tremor,

NOTE: No responsibility is assumed by the Publisher for any injury and or damage to persons or property as a matter of products liability, negligence or otherwise, or from any use or operation of any methods, products, instructions or ideas contained in the material herein. No suggested test or procedure should be carried out unless, in the reader's judgment, its risk is justified. Because of rapid advances in the medical sciences, we recommend that the independent verification of diagnoses and drug doses should be made. Discussions, views and recommendations as to medical procedures, choice of drugs and drug dosages are the responsibility of the authors.

Clinical Microbiology Newsletter (ISSN 0196-4399) is issued twice monthly in one indexed volume per year by Elsevier Science Inc., 655 Avenue of the Americas, New York, NY 10010. Subscription price per year: Personal: NLG 99 (euro 44.92) for customers in Europe; ¥6,600 for Japan; and US\$50 for all countries other than Europe and Japan. Institutional: NLG 633 (euro 287.24) for customers in Europe; ¥39,800 for Japan; and US\$321 for all countries other than Europe and Japan. Periodical postage paid at New York, NY and at additional mailing offices. Postmaster: Send address changes to *Clinical Microbiology Newsletter*, Elsevier Science Inc., 655 Avenue of the Americas, New York, NY 10010. For customer service, phone (212) 633-3950; TOLL FREE for customers in the United States and Canada: 1-888-4E5-INFO (1888 437-4636) or fax: (212) 633-3860.

hallucinations, seizures, and coma. An increased rate of seizure activity has been observed in patients with a history of epilepsy (21). At high plasma levels, cardiac arrhythmias and death can occur (22). Amantadine has anticholinergic effects that can cause dry mouth and mydriasis, and this drug is contraindicated in patients with untreated angle-closure glaucoma (15).

Clinical studies

Amantadine and rimantadine can be used for prophylaxis or treatment of influenza infection.

Treatment

Several studies have demonstrated that either amantadine or rimantadine, when administered within 48 h of the onset of illness, can reduce the duration (by about a day) and severity (approximately 50% reduction in fever and other symptoms) of uncomplicated influenza A (23-27). In most studies, there was an accompanying reduction in the amount of virus secreted. One comparative study of amantadine and rimantadine showed that the two drugs had similar efficacies (28). The benefit of these drugs in reducing the risk of complications in high-risk patients has not been established. In a study of children with influenza A H3N2 subtype infection, rimantadine treatment was associated with reduction of symptoms on days two and three of the illness but not thereafter. The mean duration of virus shedding was actually prolonged in the rimantadine arm of the study compared with the acetaminophen arm. These findings, as well as the recovery of resistant isolates on day four or later of treatment, led the authors to conclude that rimantadine therapy should be given for 3 rather than 5 days in children (29).

Prophylaxis

Influenza vaccination remains the basis of influenza prophylaxis. The Centers for Disease Control and Prevention have recommended the use of amantadine or rimantadine prophylaxis in certain circumstances (30): (i) persons at high risk for complications of influenza who receive the vaccine after influenza activity has begun in a community, since the development of antibodies can take 2 weeks; (ii) unvaccinated health care workers; (iii) persons expected to mount an inadequate response to the

vaccine, such as those with advanced HIV infection; (iv) other persons with a high risk for complications of influenza who cannot receive the vaccine; and, (v) all residents in nursing homes and long-term care facilities, for whom chemoprophylaxis is recommended during an outbreak and should be continued for at least 2 weeks or until 1 week after the end of the outbreak.

These recommendations are based on the results of several studies demonstrating the efficacy of amantadine and rimantadine in preventing symptomatic influenza virus infection. The rate of protection against influenza virus infection has ranged from 0 to 90%, averaging 50%, and both drugs are about 70 to 90% protective against clinical illness (19,20,23,25,27).

Neuraminidase Inhibitors

Zanamivir (4-guanidino-2,4-dideoxy-2,3-dehydro-N-acetylneuraminic) and oseltamivir

Spectrum

Zanamivir (4-guanidino-2,4-dideoxy-2,3-dehydro-N-acetylneuraminic) and oseltamivir (GS4104, the ethyl ester prodrug of GS4071) are related antiviral agents, that are potent and specific inhibitors of the neuraminidases of influenza A and B viruses. These sialic acid analogs competitively and reversibly interact with the active enzyme site to inhibit neuraminidase activity. Zanamivir is approved by the United States Food and Drug Administration (FDA) for treatment of influenza in persons at least 7 years of age who have been symptomatic for less than 2 days. Oseltamivir is approved for treatment in persons at least 18 years of age, and approval for use in those at least 1 year of age is pending (31). It is also approved for prophylaxis in persons at least 13 years of age. A recent study has also shown zanamivir to be efficacious in treating mice infected with various strains of avian influenza viruses responsible for infections in humans in Hong Kong (H5N1, H6N1, and H9N2) (32).

Mechanism of action

Influenza viruses possess two surface glycoproteins with either hemagglutinin (HA) or neuraminidase (NA) activity. These glycoproteins mediate the interaction of influenza A and B with N-acetylneuraminic acid-contain-

ing cellular receptors. HA initiates infection by binding to these cellular receptors on respiratory epithelial cell surfaces, whereas NA acts at a later stage to release viruses from infected cells. It does so by cleaving a terminal sialic acid (N-acetylneuraminic acid) residue from an oligosaccharide chain, thereby destroying the HA receptors. NA inhibitors mimic the structure of N-acetylneuraminic acid, a receptor determinant recognized by both influenza A and B viruses. They competitively inhibit neuraminidase. This inhibition prevents the progeny virions from self-aggregating and binding to the surface of infected cells, which prevents spread from cell to cell (2,33-35). NA also prevents the entrapment of virus by sialic acid-containing mucoproteins in respiratory secretions; virus binds to the mucus, but elution resulting from the activity of NA allows the virus to circumvent this barrier and penetrate the cells. NA inhibitors prevent this from occurring (3,36).

Resistance

Because they target a different virus protein, zanamivir and oseltamivir are active against influenza strains resistant to amantadine and rimantadine. In vitro, resistance to NA inhibitors can occur in two steps. The first step consists of a reduction of the virus' dependence on NA activity because of changes in the HA. Mutations in HA at or near the site that binds to sialic acid reduce the affinity of the HA for its receptor. As a result, these variants are less dependent on NA activity for release from cells and subsequent spread to other cells. Variants with these mutations are cross-resistant to other NA inhibitors. The other pathway of resistance development involves the acquisition of mutations in the active site of the NA, which decrease binding of the drugs (Glu119Gly/Ala/Asp); or within the catalytic framework of the NA (Arg292Lys) (37,38). Oseltamivir retains activity against variants with the Glu119 mutation but has less activity than zanamivir against Arg292 variants (37). These mutations may be associated with significantly reduced enzyme activity, and the mutated viruses have decreased infectivity in animal models.

One case of a resistant virus isolated from an immunosuppressed child receiving zanamivir under a compassionate use protocol has been reported

to date (39). So far, resistant viruses have not been isolated from humans who have received zanamivir in clinical trials. Mutations leading to resistance to oseltamivir have occurred in about 1.5% of treated persons (3). In a study of volunteers experimentally infected with influenza virus (H1N1) and treated with oseltamivir, 2 of 54 (4%) developed resistant virus with substitution at codon His274Tyr of the NA (40).

Pharmacokinetics

Zanamivir has poor oral bioavailability (<5%) and must be administered by inhalation through the mouth or intranasally (41). The inhaler device used for delivery of the drug is breath actuated and requires a cooperative patient. When inhaled at a high flow rate through the mouth, 78% of the dose is deposited in the oropharynx, whereas only 15% reaches the tracheobronchial airways and the lung (42,43). At recommended doses the concentration in secretions should exceed the 90% inhibitory concentration of drug for most strains of influenza virus. Bioavailability ranges from 10 to 20% (44). When administered intravenously, the plasma half-life averages 1.6 h. 90% is excreted unchanged in the urine (45).

Oseltamivir is administered orally. It is converted into its active metabolite, GS4071, by esterases in the gastrointestinal tract or blood. GS4071 has an estimated bioavailability of 80%, and its half-life averages 7 to 9 h. The time to peak plasma concentration is about 3 to 4 h. Administration with food may slightly delay absorption but does not affect peak plasma concentrations. In ferret models, widespread tissue distribution has been demonstrated, whereas in humans, distribution is not well characterized (46). The prodrug and GS4071 are eliminated unchanged in the urine. Guidelines for use in subjects with renal insufficiency are not available, but reductions in the dose are recommended for those with a creatinine clearance of less than 30 ml per min.

Drug interactions

No drug interactions have been observed so far with either zanamivir or oseltamivir.

Toxicity

Zanamivir and oseltamivir are both well tolerated. In initial evaluation of patients with mild-to-moderate asthma,

zanamivir did not reduce pulmonary function or increase airway responsiveness to methacholine (47). However, a recent report of respiratory distress in a patient with chronic obstructive pulmonary disease (COPD) following zanamivir inhalation (48), as well as preliminary data from a placebo-controlled trial suggesting reduced airflow in patients with COPD or asthma (49), has led to a warning from the manufacturer to use caution when administering zanamivir in these populations.

Oseltamivir is associated with nausea and vomiting in 10 to 15% of recipients. These side effects are transient and may be ameliorated by ingestion of food (50-53).

Clinical studies

Zanamivir and oseltamivir are effective for treatment and prophylaxis of influenza.

Treatment

Both drugs have been approved for treatment of influenza in persons who have been symptomatic for less than 2 days. Several studies of zanamivir and oseltamivir have demonstrated a clinical benefit, illustrated by a 1- to 1.5-day decrease in the duration of symptoms associated with a reduction in virus shedding in respiratory secretions (51,53-57). The ability of these drugs to decrease the frequency of pneumonia associated with influenza remains to be determined, although some studies indicated a decreased frequency of complications such as sinusitis, purulent bronchitis, and otitis media (3).

Prophylaxis

Only oseltamivir is FDA approved for prophylaxis of influenza, although three recent clinical trials have shown efficacy of zanamivir in preventing new cases of influenza in families or close contacts of infected individuals. In contrast to what had been reported with rimantadine prophylaxis, there was no emergence of resistant viruses in family contacts who received prophylaxis (58-60). In different clinical trials, oseltamivir was evaluated as prophylaxis when given before exposure, after exposure to infected family members, and after exposure in nursing home settings. It was effective in preventing influenza in contacts and preventing outbreaks within households. The benefit was sustained even in a vaccinated nursing

home population (61-63).

Other neuraminidase inhibitors in development

RWJ-270201 is the most potent compound of a novel series of cyclopentane derivatives discovered through structure-based drug design. It is still in pre-clinical development. In vitro studies have shown comparable or slightly higher anti-influenza activity than that of zanamivir and oseltamivir. In a murine influenza model, protection was observed when the drug was administered before or within 48 h of viral challenge. It is active against both influenza A and B (64,65).

Respiratory Syncytial Virus

Respiratory syncytial virus is the single most important cause of lower respiratory tract infection during infancy and early childhood and causes significant morbidity in immunocompromised adults, especially bone marrow transplant recipients. So far, the only agent approved by the FDA for the treatment of RSV lower respiratory tract infection is ribavirin, which can be used alone or in association with specific RSV immune globulin.

Ribavirin

Spectrum

Ribavirin (1- β -D-ribofuranosyl-1,2,4-thiazole-3-carboxamide) is a guanosine analog. It is active in vitro and in vivo against a wide range of RNA and DNA viruses, including myxo-, paramyxo-, arena-, bunya-, adeno-, pox-, retro-, herpes-, and viruses (66-68). In RSV infection, its aerosol administration is more effective than parenteral dosing, and enhanced activity is observed when ribavirin is combined with immunoglobulin (69,70). Parenteral ribavirin has antiviral and therapeutic activity against hepatitis C virus, Lassa virus, other arenavirus, and bunyavirus infections (68).

Mechanism of action

Ribavirin is phosphorylated intracellularly into mono-, di-, and triphosphate derivatives by host-cell enzymes. Ribavirin monophosphate competitively inhibits inosine-5'-phosphate dehydrogenase and interferes with the synthesis of guanosine triphosphate and, therefore, with nucleic acid synthesis. Ribavirin triphosphate may inhibit influenza virus RNA polymerase activity and competitively inhibit the guanosine

triphosphate-dependent 5'-capping of viral messenger RNA (2). Ribavirin diphosphates and triphosphates have also been shown to inhibit HIV reverse transcriptase activity (71). In vitro, combinations of ribavirin and zidovudine show antagonistic interactions. However, ribavirin enhances the activity of purine dideoxynucleosides (72,73).

Resistance

There is no evidence that RSV develops resistance to ribavirin either in vitro or in clinical use. The only reported virus to have developed resistance to ribavirin is Sindbis virus.

Pharmacokinetics

The pharmacokinetics of ribavirin are complex. Bioavailability ranges from 33 to 45% after oral administration. The peak plasma concentrations after oral dosing occur 1 to 2 h after administration. When the drug is administered intravenously, peak plasma levels are 10 times higher than following oral intake. At steady state, CSF levels are 70% of those in plasma. Its elimination occurs in two phases: α -phase with a half-life of 2 h and a terminal phase with a half-life of 18 to 36 h. Its triphosphate concentrates in erythrocytes, and these are eliminated with a half-life of approximately 40 days. Hepatic metabolism is an important route of elimination. After oral administration, 4% is recovered unchanged in the urine, and 39% is excreted as the metabolite 1,2,4-triazole-3-carboxamide (74-76).

With aerosol delivery, plasma levels increase with the duration of exposure. The half-life of ribavirin in respiratory secretions ranges from 1.4 to 2.5 h (77). Proper delivery necessitates the use of a specialized aerosol generator to reduce the size of the particles in order to reach the lower respiratory tract. Age, as well as several other factors, influence dosage (78).

Toxicity

Ribavirin administered systemically causes anemia by two mechanisms: extravascular hemolysis and bone marrow suppression of the erythroid lineage. Severe anemia may require dose reduction or cessation (79). During short-term oral administration, reversible increases in serum bilirubin, iron, and uric acid concentrations occur frequently. Other side effects include pru-

ritus, rash, nausea, depression, cough, and respiratory symptoms and, when given by bolus intravenous infusion, rigors (80,81). When administered by aerosol, mild conjunctival irritation, rash, bronchospasm, and rarely water intoxication have been observed (82). Aerosolized ribavirin is not associated with hematologic toxicity.

There have been concerns about potential toxicity for health care workers exposed during administration of aerosolized ribavirin. Recommendations have been made to decrease the level of exposure, including the use of aerosol containment systems except during mechanical ventilation, turning off the aerosol generator before providing routine care, and use of protective equipment (83-85). Ribavirin is teratogenic, embryotoxic, mutagenic, tumor promoting, and gonadotoxic. Therefore, pregnant women should not directly care for patients receiving ribavirin aerosols (79).

Clinical Studies

Treatment of RSV bronchiolitis in infancy

A meta-analysis of 11 randomized trials of ribavirin for the treatment of lower respiratory tract infection in infancy has been published (86). In six controlled, randomized studies, ribavirin reduced the severity of RSV illness as demonstrated by a decrease in viral shedding, improvement in oxygen saturation, and improvement in clinical scores. However, ribavirin did not lead to improvement in clinically important outcomes, such as shortened duration of hospitalization (87-92). Studies looking at long-term outcome following ribavirin therapy for RSV infection have shown conflicting results. In several controlled studies, children treated with ribavirin for RSV lower respiratory tract infection had improved clinical and pulmonary evaluations on follow-up compared to controls (93-97). However, in a recent retrospective study of children with or without treatment with ribavirin, no differences in wheezing or other pulmonary function measures were seen at 6 to 8 years of follow-up (98). Diminished levels of RSV-specific IgE and IgA antibodies, which have been associated with more severe clinical illness, have been demonstrated in the secretions of infants treated with ribavirin (99). The growing concern about the

efficacy of ribavirin and the high cost associated with its use have led the American Academy of Pediatrics to change the wording of their recommendation from "should be used" to "may be considered" for selected infants and young children at high risk for serious RSV disease, such as those with underlying cardiac, pulmonary, and immunosuppressive conditions (100).

Treatment of RSV infection in immunocompromised patients

Several uncontrolled studies of ribavirin administered by intravenous, oral, or aerosolized routes in various immunocompromised groups with RSV pneumonia have been conducted. Intravenous or aerosolized ribavirin alone appeared to be ineffective in bone marrow transplant subjects (101,102,104). However, combinations of high-titered RSV immunoglobulin plus aerosolized ribavirin appeared to be beneficial in bone marrow transplant recipients with RSV pneumonia, reducing the mortality rate to 50% (105). Early initiation of therapy (at least 1 day prior to the onset of respiratory failure) is important, since mortality rates are 100% in patients treated after the onset of respiratory failure (106). There are insufficient data to evaluate the efficacy of ribavirin for the treatment of RSV infections in the elderly (107).

Treatment of other viral respiratory infections with ribavirin

Although use of intravenous and aerosolized ribavirin has been attempted in severe influenza virus as well as parainfluenza and adenovirus infections, results have been inconsistent, and ribavirin is not approved for these indications (108-113).

In Development

New therapies in development to combat RSV infections include sulfated polysaccharide compounds, miscellaneous organic compounds, a variety of proteins (some with known enzymatic activities), and nucleosides other than ribavirin, including antisense oligodeoxyribonucleotides (114).

References

1. Hay, A.J. 1996. Amantadine and rimantadine: mechanisms, p. 43. *In* D.D. Richman (ed.). Antiviral drug resistance. John Wiley & Sons, New York.
2. Hayden, F.G. 2000. Antiviral drugs

- (other than antiretrovirals), p. 460-488. *In* Mandell, G.L. et al. (ed.). Principles and practice of infectious diseases, 5th ed. Churchill Livingstone, Philadelphia, PA.
3. Couch, R.B. 2000. Prevention and treatment of influenza. *N. Engl. J. Med.* 343:1778-1787.
 4. Oxford, J.S. and A. Galraith. 1980. Antiviral activity of amantadine: a review of laboratory and clinical data. *Pharmacol. Ther.* 11:181-262.
 5. Hayden, F.G. 1996. Amantadine and rimantadine - clinical aspects, p. 59-77. *In* D.D. Richman (ed.), Antiviral drug resistance. John Wiley & Sons, New York.
 6. Belshe, R.B. et al. 1988. Genetic basis of resistance to rimantadine emerging during treatment of influenza virus infection. *J. Virol.* 62:1508.
 7. Bean, W.J., S.C. Threlkild, and R.G. Webster. 1989. Biologic potential of amantadine-resistant influenza A in an avian model. *J. Infect. Dis.* 159:1050-1056.
 8. Hayden, F.G. et al. 1989. Emergence and apparent transmission of rimantadine-resistant influenza A virus in families. *N. Engl. J. Med.* 321:166-702.
 9. Degelau, J. et al. 1992. Amantadine-resistant influenza A in a nursing facility. *Arch. Intern. Med.* 15:362.
 10. Aoki, F.Y. and D.S. Sitar. 1988. Clinical pharmacokinetics of amantadine hydrochloride. *Clin. Pharmacokinet.* 14:35-51.
 11. Aoki, F.Y. and D.S. Sitar. 1985. Amantadine kinetics in healthy elderly men: implications for influenza prevention. *Clin. Pharmacol. Ther.* 28:216-221.
 12. Wills, R.J. et al. 1987. Rimantadine pharmacokinetics after single and multiple doses. *Antimicrob. Agents Chemother.* 31:826-828.
 13. Speeg, K.V., J.A. Leighton, and A.L. Maldonado. 1989. Toxic delirium in a patient taking amantadine and trimethoprim-sulfamethoxazole. *Am. J. Med. Sci.* 298:410-412.
 14. Wills, R.J. 1989. Update on rimantadine's clinical pharmacokinetics. *J. Respir. Dis.* 10(Suppl.):S20-S25.
 15. Symmetrel, p. 1040. *In* Physicians' Desk Reference, 2000, 54th ed. Medical Economics Co., Inc., Montvale, NJ.
 16. Flumadine, p. 1078. *In* Physicians' Desk Reference, 2000, 54th ed. Medical Economics Co., Inc., Montvale, NJ.
 17. Soo, W. 1989. Adverse effects of rimantadine: summary from clinical trials. *J. Respir. Dis.* 10(Suppl.):S26-S31.
 18. Vernier, V.G. et al. 1969. The toxicologic and pharmacologic properties of amantadine hydrochloride. *Toxicol. Appl. Pharmacol.* 15:642-665.
 19. Dolin, R. et al. 1982. A controlled trial of amantadine and rimantadine in the prophylaxis of influenza A infection. *N. Engl. J. Med.* 307:580.
 20. Pettersson, R.F. et al. 1980. Evaluation of amantadine in the prophylaxis of influenza A (H1N1) virus infection: a controlled field trial among young adults and high-risk patients. *J. Infect. Dis.* 42:377.
 21. Atkinson, W.L. et al. 1986. Amantadine prophylaxis during an institutional outbreak of type A (H1N1) influenza. *Arch. Intern. Med.* 16:1751-1756.
 22. M. Sartori, C.M. Pratt, and J.B. Young. 1984. Torsade de pointe: malignant cardiac arrhythmia induced by amantadine poisoning. *Am. J. Med.* 77:388-391.
 23. Zlydnkov, D.M. et al. 1981. Study of rimantadine in the USSR: a review of the literature. *Rev. Infect. Dis.* 3:408.
 24. Hirsch, M.S. and M.N. Swartz. 1980. Drug therapy: antiviral agents (Part 1). *N. Engl. J. Med.* 302:903.
 25. Wingfield, W.L., D. Pollac, and R.R. Grunert. 1969. Therapeutic efficacy of amantadine HCl and rimantadine HCl in naturally occurring influenza A2 respiratory illness in man. *N. Engl. J. Med.* 281:579.
 26. Hayden, F.G. 1997. Antivirals for pandemic influenza. *J. Infect. Dis.* 17(Suppl. 1):S56.
 27. Couch, R.B. 1997. Respiratory virus infections, p. 369-413. *In* G.J. Galasso, R.J. Whitley, and T.C. Merigan (ed.), Antiviral agents and human viral diseases, 4th ed. Lippincott-Raven, Philadelphia, PA.
 28. Van Voris, L.P. et al. 1981. Successful treatment of naturally occurring influenza A/USSR/77 H1N1. *JAMA* 245:1128-1131.
 29. Hall, C.B. et al. 1987. Children with influenza A infection: treatment with rimantadine. *Pediatrics* 80:275.
 30. Centers for Disease Control and Prevention. 2000. Prevention and control of influenza: recommendations of the advisory committee on immunization practices. *Morb. Mortal. Wkly. Rep.* 49(RR03):1.
 31. Couch, R.B. 2000. Influenza: prospects for control. *Ann. Intern. Med.* 133:992-998.
 32. Leneva, I.A. et al. 2001. Efficacy of zanamivir against avian influenza A viruses that possess genes encoding H5N1 internal proteins and are pathogenic in mammals. *Antimicrob. Agents Chemother.* 45:1216-1224.
 33. Von Itzstein, M. et al. 1993. Rational design of potent sialidase-based inhibitors of influenza virus replication. *Nature* 363:418-423.
 34. Kim, C.U. et al. 1997. Influenza neuraminidase inhibitors possessing a novel hydrophobic interaction in the enzyme active site: design, synthesis, and structural analysis of carbocyclic sialic acid analogues with potent anti-influenza activity. *J. Am. Chem. Soc.* 119:681-690.
 35. Woods, J.M. et al. 1993. 4-guanidino-2,4-dideoxy-2,3-dehydro-n-acetylneuraminic acid is a highly effective inhibitor both of the sialidase (neuraminidase) and of growth of a wide range of influenza A and B viruses in vitro. *Antimicrob. Agents Chemother.* 37:1473-1479.
 36. Lamb RA. and R.M. Krug. 1996. Orthomyxoviridae: the viruses and their replication, p. 1353-1395. *In* B.M. Fields, D.M. Knipe, and P.M. Howley (ed.), Fields virology, 3rd ed., vol. 1. Lippincott-Raven, Philadelphia, PA.
 37. Gubareva, L.V. et al. 1997. Catalytic and framework mutations in the neuraminidase active site of influenza viruses that are resistant to 4-guanidino-Neu5Ac2en. *J. Virol.* 71:3385-3390.
 38. Gubareva, L.V. et al. 1996. Characterization of mutants of influenza A virus selected with the neuraminidase inhibitor 4-guanidino-Neu5Ac2en. *J. Virol.* 70:1818-1827.
 39. Gubareva, L.V. et al. 1998. Evidence for zanamivir resistance in an immunocompromised child infected with influenza B virus. *J. Infect. Dis.* 178:157-162.
 40. Gubareva, L.V. et al. 2001. Selection of influenza virus mutants in experimentally infected volunteers treated with oseltamivir. *J. Infect. Dis.* 183:523-531.
 41. Dunn, C.J. and K.L. Goa. 1999. Zanamivir: a review of its use in influenza. *Drugs* 58:761.
 42. Peng, A.W., S. Milleri, and D.S. Stein. 2000. Direct measurement of the anti-influenza agent zanamivir in the respiratory tract following inhalation. *Antimicrob. Agents Chemother.* 44:1974.
 43. Cass, L.M.R. et al. 1999. Pharmacoscintigraphic evaluation of lung deposition of inhaled zanamivir in healthy volunteers. *Clin. Pharmacokinet.* 36:21-31.
 44. Cass, L.M.R., C. Efthymiopoulos, and A. Bye. 1999. Pharmacokinetics of zanamivir after intravenous, oral, inhaled or intranasal administration to healthy volunteers. *Clin. Pharmacokinet.* 36(Suppl. 1):1-11.
 45. Waghorn, S.L. and K.L. Goa. 1998. Zanamivir. *Drugs* 55:722-725.
 46. Bardsley-Elliott, A. and S. Noble. 1999. Oseltamivir. *Drugs* 58:61.
 47. Cass, L.M. et al. 2000. Pulmonary func-

- tion and airway responsiveness in mild to moderate asthmatics given repeated doses of zanamivir. *Respir. Med.* 94:166-173.
48. Williamson, J.C. and P.S. Pegram. 2000. Respiratory distress associated with zanamivir. *N. Engl. J. Med.* 342:661. (Letter.)
 49. Neuraminidase inhibitors for treatment of influenza A and B infections. 1999. *MMWR Morb. Mortal. Wkly. Rep.* 48(RR-14):1.
 50. Yamey, G. 2000. Drug company issues warning about flu drug (news). *Br. Med. J.* 320:334.
 51. Treanor, J. et al. 2000. Efficacy and safety of the oral neuraminidase inhibitor oseltamivir in treating acute influenza: a randomized controlled trial. *JAMA* 283:1016.
 52. Hayden, F.G. et al. 1999. Use of the oral neuraminidase inhibitor oseltamivir in experimental human influenza: randomized controlled trials for prevention and treatment. *JAMA* 282:1240.
 53. Nicholson K.G. et al. 2000. Efficacy and safety of oseltamivir in treatment of acute influenza: a randomised controlled trial. *Lancet* 355:518-545.
 54. Hayden, F.G. et al. 1997. Efficacy and safety of the neuraminidase inhibitor zanamivir in the treatment of influenza virus infections. *N. Engl. J. Med.* 337:874.
 55. Monto, A.S. et al. 1999. Efficacy and safety of the neuraminidase inhibitor zanamivir in the treatment of influenza A and B virus infections. *J. Infect. Dis.* 180:254.
 56. The MIST (Management of Influenza in the Southern Hemisphere Trialists) Study Group. 1998. Randomised trial of efficacy and safety of inhaled zanamivir in treatment of influenza infections. *Lancet* 352:1877.
 57. Hedrick, J.A. et al. 2000. Zanamivir for treatment of symptomatic influenza A and B infection in children five to twelve years of age: a randomized controlled trial. *Pediatr. Infect. Dis. J.* 19:410.
 58. Monto, A.S. et al. 1999. Zanamivir in the prevention of influenza among healthy adults: a randomized controlled trial. *JAMA* 282:231.
 59. Kaiser, L. et al. 2000. Short-term treatment with zanamivir to prevent influenza: results of a placebo-controlled study. *Clin. Infect. Dis.* 30:587.
 60. Hayden, F.G. et al. 2000. Inhaled zanamivir for the prevention of influenza in families. *N. Engl. J. Med.* 343:1282.
 61. Hayden, F.G. et al. 1999. Use of the selective oral neuraminidase inhibitor oseltamivir to prevent influenza. *N. Engl. J. Med.* 341:1336.
 62. Munoz, F.M. et al. 2000. Current research on influenza and other respiratory viruses: II. International Symposium. *Antivir. Res.* 46:91-124.
 63. Welliver, R. et al. 2001. Effectiveness of oseltamivir in preventing influenza in household contacts: a randomized controlled trial. *JAMA* 285:748-754.
 64. Smee, D.F. et al. 2001. Cyclopentane neuraminidase inhibitors with potent in vitro anti-influenza virus activities. *Antimicrob. Agents Chemother.* 45:743-748.
 65. Bantia, S. et al. 2001. Comparison of the anti-influenza virus activity of RWJ-270201 with those of oseltamivir and zanamivir. *Antimicrob. Agents Chemother.* 45:1162-1167.
 66. Hruska, J.F. et al. 1982. In vivo inhibition of respiratory syncytial virus by ribavirin. *Antimicrob. Agents Chemother.* 21:125-130.
 67. Wyde, P.R. et al. 1987. Efficacy of high dose-short duration ribavirin aerosol in the treatment of respiratory syncytial virus infected cotton rats and influenza B virus infected mice. *Antivir. Res.* 7:211-220.
 68. Jahrling, P.B. et al. 1980. Lassa virus infection of rhesus monkeys: pathogenesis and treatment with ribavirin. *J. Infect. Dis.* 141:580-589.
 69. Gruber, W.C. et al. 1987. Immunoglobulin administration and ribavirin therapy: efficacy in respiratory syncytial virus infection of the cotton rat. *Pediatr. Res.* 21:270-274.
 70. Hayden, F.G. 1996. Combination antiviral therapy for respiratory virus infections. *Antivir. Res.* 29:45-48.
 71. Fernandez-Larsson, R. and J.L. Patterson. 1990. Ribavirin is an inhibitor of human immunodeficiency virus reverse transcriptase. *Mol. Pharmacol.* 38:766-770.
 72. Vogt, M.W. et al. 1987. Ribavirin antagonizes the effect of azidothymidine on HIV replication. *Science* 235:1376-1379.
 73. Baba, M. et al. 1987. Ribavirin antagonizes inhibitory effects of pyrimidine 2',3'-dideoxynucleosides but enhances inhibitory effects of uridine 2',3'-dideoxynucleosides on replication of human immunodeficiency virus in vitro. *Antimicrob. Agents Chemother.* 31:1613-1617.
 74. Laskin, O.L. et al. 1987. Ribavirin disposition in high-risk patients for acquired immunodeficiency syndrome. *Clin. Pharmacol. Ther.* 41:546-555.
 75. Connor, E. et al. 1993. Safety, tolerance, and pharmacokinetics of systemic ribavirin in children with human immunodeficiency virus infection. *Antimicrob. Agents Chemother.* 37:532-539.
 76. Paroni, R. et al. 1989. Pharmacokinetics of ribavirin and urinary excretion of the major metabolite 1,2,4-triazole-3-carboxamide in normal volunteers. *Int. J. Clin. Pharmacol. Ther. Toxicol.* 27:302-307.
 77. Connor, J.D., M. Hintz, and R. Van Dyke. 1984. Ribavirin pharmacokinetics in children and adults during therapeutic trials. p. 107-123. *In* R.A. Smith, V. Knight, and J. Smith (ed.), *Clinical applications of ribavirin*. Academic Press, Orlando, FL.
 78. Knight, V. et al. 1988. Estimating the dosage of ribavirin aerosol according to age and other variables. *J. Infect. Dis.* 158:443-448.
 79. Hillyard, I.W. 1980. The preclinical toxicology and safety of ribavirin, p. 59. *In* R.A. Smith, and W. Kirkpatrick (ed.), *Ribavirin: a broad spectrum antiviral agent*. Academic Press, New York.
 80. Di Bisceglie, A.M. et al. 1995. Ribavirin as therapy for chronic hepatitis C: a randomized, double-blind, placebo-controlled trial. *Ann. Intern. Med.* 123:897-903.
 81. Fisher-Hoch, S.P., S. Gborie, and L. Parker. 1992. Unexpected adverse reactions during a clinical trial in rural West Africa. *Antivir. Res.* 19:139-147.
 82. Titus, B.J., A.F. Perez, and B.I. Arcala. 1995. Water intoxication after nebulised ribavirin. *Lancet* 345:1116.
 83. Rodriguez, W.J. et al. 1987. Environmental exposure of primary care personnel to ribavirin aerosol when supervising treatment of infants with respiratory syncytial virus infections. *Antimicrob. Agents Chemother.* 31:1143-1146.
 84. Bradley, J.S. et al. 1990. Exposure of health care workers to ribavirin during therapy for respiratory syncytial virus infections. *Antimicrob. Agents Chemother.* 34:668-670.
 85. Shults, R.A. et al. 1996. Health care worker exposure to aerosolized ribavirin: biological and air monitoring. *J. Occup. Environ. Med.* 38:257-263.
 86. Randolph, A.G. and E.E.L. Wang. 1996. Ribavirin for respiratory syncytial virus lower respiratory tract infection: a systemic overview. *Arch. Pediatr. Adolesc. Med.* 150:942-947.
 87. Barry, W. et al. 1985. Ribavirin aerosol for acute bronchiolitis. *Arch. Dis. Child.* 61:593-597.

88. Hall, C.B., J.T. McBride, and E.E. Walsh. 1998. Aerosolized ribavirin treatment of infants with respiratory syncytial virus infection. *N. Engl. J. Med.* 308:1443-1447.
89. Hall, C.B. et al. 1985. Ribavirin treatment of respiratory syncytial viral infection in infants with underlying cardiopulmonary disease. *JAMA* 254:3047-3051.
90. Rodriguez, W.J., H.W. Kim and C.D. Brandt. 1987. Aerosolized ribavirin in the treatment of patients with respiratory syncytial virus disease. *Pediatr. Infect. Dis.* 6:159-163.
91. Smith, D.W. et al. 1991. A controlled trial of aerosolized ribavirin in infants receiving mechanical ventilation for severe respiratory syncytial virus infection. *N. Engl. J. Med.* 325:24-29.
92. Taber, L.H. et al. 1983. Ribavirin aerosol treatment of bronchiolitis due to respiratory syncytial virus infection in infants. *Pediatrics* 72:613-618.
93. Long, C. et al. 1997. Long term follow-up of children hospitalized with respiratory syncytial virus lower respiratory tract infection and randomly treated with ribavirin or placebo. *Pediatr. Infect. Dis. J.* 16:1023-1028.
94. McConnochie, K.M. et al. 1985. Normal pulmonary function measurement and airway reactivity in childhood after mild bronchiolitis. *Pediatrics* 107:54-58.
95. Voter, K.Z., C. Long, and C.B. Hall. 1996. Respiratory illnesses and lung function following ribavirin therapy in infancy. *Pediatr. Res.* 39:392A. (Abstract.)
96. Rodriguez, W.J. et al. 1996. Prospective (7 yrs) follow up and pulmonary functions from a placebo controlled randomized trial of ribavirin in RSV bronchiolitis. *Pediatr. Res.* 39:183A. (Abstract.)
97. Edell, D. et al. 1998. Reduced long-term respiratory morbidity after treatment of respiratory syncytial virus bronchiolitis with ribavirin in previously healthy infants: a preliminary report. *Pediatr. Pulmonol.* 25:154-158.
98. Krilov, L. et al. 1997. The Bronchiolitis Study Group. Follow-up of children with respiratory syncytial virus bronchiolitis in 1986-1987: potential effect of ribavirin on long term pulmonary function. *Pediatr. Infect. Dis. J.* 16:273-276.
99. Rosner, I. et al. 1987. Effect of ribavirin therapy on RSV-specific IgE and IgA responses after infection. *J. Infect. Dis.* 155:1043-1047.
100. Committee on Infectious Disease, American Academy of Pediatrics. 1996. Reassessment of the indications for ribavirin therapy in respiratory syncytial virus infections. *Pediatrics* 97:137-140.
101. Harrington, R.D. et al. 1992. An outbreak of respiratory syncytial virus in bone marrow transplant center. *J. Infect. Dis.* 165:987-993.
102. Hertz, M.I. et al. 1989. Respiratory syncytial virus-induced acute lung injury in adult patients with bone marrow transplants: a clinical approach and review of the literature. *Medicine* 68:269-281.
103. Lewinson, D.M. et al. 1996. Phase I study of intravenous ribavirin treatment of respiratory syncytial virus pneumonia after marrow transplantation. *Antimicrob. Agents Chemother.* 40:2555-2557.
104. Whimbey, E. et al. 1995. Combination therapy with aerosolized ribavirin and intravenous immunoglobulin for respiratory syncytial virus disease in adult bone marrow transplant recipients. *Bone Marrow Transplant.* 16:393-399.
105. Whimbey, E. et al. 1996. Community respiratory virus infections among hospitalized adult bone marrow transplant recipients. *Clin. Infect. Dis.* 22:778-782.
106. Falsey, A.R. and E.E. Walsh. 2000. Respiratory syncytial virus infection in adults. *Clin. Microbiol. Rev.* 13:371-384.
107. Knight, V. and B.E. Gilbert. 1987. Ribavirin aerosol treatment of influenza (review). *Infect. Dis. Clin. N. Am.* 1:441-457.
108. Ray, C.G. et al. 1989. The use of intravenous ribavirin to treat influenza virus-associated acute myocarditis. *J. Infect. Dis.* 159:829-836. (Erratum; 160:564; 1989).
109. Stein, D.S. et al. 1987. Oral ribavirin treatment of influenza A and B. *Antimicrob. Agents Chemother.* 31:1285-1287.
110. Wendt, C.H. et al. 1992. Parainfluenza virus respiratory infection after bone marrow transplantation. *N. Engl. J. Med.* 326:921-926.
111. Kaplan, L.J. et al. 1992. Severe measles in immunocompromised patients. *JAMA* 267:1237-1241.
112. Gururangan, S., R.F. Stevens, and D.J. Morris. 1990. Ribavirin response in measles pneumonia. *J. Infect.* 20:219-221.
113. Maslo, C. et al. 1997. Ribavirin therapy for adenovirus pneumonia in an AIDS patient. *Am. J. Respir. Crit. Care Med.* 156:1263-1264.
114. Domachowske, J.B. and H.F. Rosenberg. 1999. Respiratory syncytial virus infection: immune response, immunopathogenesis, and treatment. *Clin. Microbiol. Rev.* 12:298-309.

Editors:

Mary Jane Ferraro
Paul A. Granato
Josephine A. Morello
R.J. Zabransky

© 2001 Elsevier Science Inc.

ISSN 0196-4399
CMNEEJ 23(21)163-170, 2001

Elsevier



0196-4399(20011101)23:21;1-B

General Information

Subscription information can be found inside the front cover.

This newsletter has been registered with the Copyright Clearance Center, Inc. Consent is given for copying articles for personal or internal use, or for the personal or internal use of specific clients. This consent is given on the condition that the copier pay through the Center the per-page fee stated in the code on the first page for copying beyond that permitted by the US Copyright Law. If no code appears on an article, the author has not given broad consent to copy and permission to copy must be obtained directly from the author. This consent does not extend to other kinds of copying, such as for general distribution, resale, advertising and promotional purposes, or for creating new collective works.

Address orders, changes of address, and claims for missing issues to Journal Fulfillment Department, Elsevier Science Inc., 655 Avenue of the Americas, New York, NY 10010. Claims for missing issues can be honored only up to three months for domestic addresses and six months for foreign addresses. Duplicate copies will not be sent to replace ones undelivered due to failure to notify Elsevier of change of address.

Postmaster: Send address changes to *Clinical Microbiology Newsletter*, Elsevier Science Inc., 655 Avenue of the Americas, New York, NY 10010.

Editorials and letters printed in this newsletter are published for the interest of the readers and do not necessarily reflect the opinions of the editors.

Clinical Microbiology Newsletter is abstracted in *Tropical Diseases Bulletin*, *Abstracts on Hygiene and Communicable Diseases*, *EMBASE/Excerpta Medica*, and *Current AIDS Literature*.

Full text of this and previous issues can viewed online using the ScienceDirect or ScienceDirect Web-Editions services. Visit <http://www.sciencedirect.com> or <http://www.web-editions.com> for more details and access.