

Article

Zn Complex of Diaminedithiol Tetradentate Ligand as a Stable Precursor for ^{99m}Tc-Labeled Compounds

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Abstract: The diaminedithiol (N₂S₂) tetradentate ligand constitutes a useful chelating molecule for preparing ^{99m}Tc-labeled compounds of high in vivo stability in high radiochemical yields. However, since the thiol groups in the N₂S₂ ligand are easy to be oxidized to disulfide bonds, they need to be protected with an appropriate protecting group, which hinders the broad applications of the N₂S₂ ligand for radiopharmaceuticals. In this study, a Zn chelate of N₂S₂ was evaluated as a precursor for purification-free ^{99m}Tc-labeled N₂S₂ under the mild and simple procedure. Zn-N₂S₂ was prepared by reacting Zn acetate with N₂S₂, and the Zn-N₂S₂ remained stable under aerobic conditions at room temperature. ^{99m}Tc-N₂S₂ was obtained over 90% radiochemical yields at room temperature by a one-pot reaction, consisting of Zn-N₂S₂ (10⁻⁵ M), ^{99m}Tc-N₂S₂ was also obtained over 90% radiochemical yields when the reaction was conducted in the presence of an equimolar amount of IgG antibody. These findings indicate the Zn complex of N₂S₂ ligand constitutes a stable and useful precursor to prepare ^{99m}Tc-labeled N₂S₂ compounds in high yields under the mild and simple procedure.

Keywords: $^{99m}\text{Tc};$ radiopharmaceutical; Zn complex; diaminedithiol; N_2S_2

1. Introduction

The growth and broad applications of diagnostic nuclear medicine have been mainly driven by the artificial radionuclide, technetium-99m (^{99m}Tc), due to its availability from a portable ⁹⁹Mo-^{99m}Tc generator system and its almost ideal physical properties for external imaging. Although recent efforts are being made to develop radiopharmaceuticals derived from positron emitters such as ¹¹C and ¹⁸F, over 70% of diagnostic practices are still conducted with ^{99m}Tc. The cost-effectiveness of ^{99m}Tc-based radiopharmaceuticals contributes to the medical economy in both developed and developing countries. Thus, ^{99m}Tc will continue to be used as one of the essential radionuclides for diagnostic nuclear medicine. Radioactive rhenium (¹⁸⁶Re and ¹⁸⁸Re), the congener of Tc, emits beta rays appropriate to targeted radiotherapy to form radiotheranostic pairs with their ^{99m}Tc counterparts due to the chemical analogy between Tc and Re. Thus, further development of ^{99m}Tc-radiopharmaceuticals constitutes a crucial issue for cost-effective and successful patient management.

A tetradentate chelating agent containing two amines and two thiol groups, referred to as a diaminedithiol (DADT) or a bis(amino ethanethiol: BAT), forms a neutral, lipophilic, and stable complex with the pentavalent oxo-Tc and oxo-Re (TcO³⁺ and ReO³⁺) [1,2]. Such characteristics render DADT attractive as a coordination molecule for ^{99m}Tc- and ^{186/188}Re-labeled compounds [3–6]. A presentative structure of DADT ligand is shown in Scheme 1, compound **6** (N₂S₂). Indeed, ^{99m}Tc-labeled



N,N'-ethylene bis (L-cysteine ethyl ester) has been used as a radiopharmaceutical for measuring cerebral blood flow [7]. However, further applications of DADT ligands are hindered due to the inherent easily oxidized property of the thiol groups during the storage.



Scheme 1. Synthetic procedure for N₂S₂ and its Zn, Re, and ^{99m}Tc complexes.

Some acylating agents have been used to protect the thiol groups in N₂S₂- or N₃S₁-type ligands such as mercaptoacetyl glycyl-glycine. These include benzoyl [8], acetyl [9], and *m*-phthalic acid [10]. The deprotection of the benzoyl group is usually conducted at an elevated temperature (e.g., in boiling water) under alkaline conditions (pH > 10) [8,11]. The acetyl protecting group can be removed under milder conditions, whereas a long reaction time (1 h) is needed at room temperature [9]. The *m*-phthalic acid protecting group can be removed at room temperature in the presence of 1 M of NH₂OH [10], which may necessitate post-labeling removal of NH₂OH before injection to subjects. Thus, a protecting group that provides a precursor of N₂S₂ stable against oxidation and generates ^{99m}Tc-N₂S₂ in high radiochemical yields under mild conditions without post- or pre-labeling purification is highly useful for developing new ^{99m}Tc-radiopharmaceuticals.

Meanwhile, a transmetallation reaction between Zn chelate of dithiocarbamates (DTCs) and rhenium tricarbonyl has been reported. In this reaction, the thiol groups in the DTCs were stabilized upon complexation with Zn ion, and the direct reaction of the Zn complex with Re(CO)₃Br₃ provided Re(CO)₃-DTCs in high radiochemical yields [12]. These results suggested that Zn ion would also be applicable as a protecting agent for the thiol groups in N₂S₂ ligands removable during the complexation reaction with ^{99m}Tc and Re. Indeed, a prior study showed the formation of Zn-N₂S₂ chelate [13]. Zn is classified as the least toxic group of all metals by the ICH Harmonized Guideline [14], which is advantageous to clinical applications.

In the present study, the Zn ion was evaluated as a protecting agent for the thiol groups in N_2S_2 ligands using compound 6. After preparing the Zn complex of 6, the reaction parameters that affected the radiochemical yields of the ^{99m}Tc- N_2S_2 were investigated. The formation of ^{99m}Tc- N_2S_2 from Zn- N_2S_2 was also evaluated in the presence of an equimolar amount of IgG antibody to estimate the applicability to Zn- N_2S_2 as a chelating moiety of a bifunctional chelating agent for ^{99m}Tc-labeled polypeptides.

2. Results

2.1. Synthesis

The N_2S_2 ligand 6 was synthesized according to the procedure of Ohmomo et al. as shown in Scheme 1. 4-(Methoxyphyenyl)methanethiol 1 was reacted with ethyl 2-bromo-2-methyl propanoate 2 to prepare 3, followed by the condensation with ethylenediamine to provide 4. After reducing the amide bonds in 4 with BH₃, the thiol protecting group in the resulting compound 5 was removed with TFA/anisole/methanesulfonic acid to obtain 6.

The Zn complex of the N_2S_2 was prepared by mixing Zn acetate with compound 6 in an aqueous solution at neutral pH. The Zn- N_2S_2 was obtained by extracting the reaction solution with dichloromethane, followed by recrystallization from the mixture of chloroform and hexane in 89% yield. The Zn- N_2S_2 remained stable at room temperature under aerobic conditions over several days.

The non-radioactive ^{185/187}Re-N₂S₂ complex was synthesized by reacting 6 with ReO₄⁻ in aqueous ethanol using Sn²⁺ as a reducing agent. After silica-gel column chromatography, the oxorhenium complex of N₂S₂ was obtained in ca. 20% yield. The structure of this complex was confirmed by mass spectrometry (MS) and infrared spectroscopy (IR).

2.2. ^{99m}Tc Complexation Reaction

The reaction of Zn-N₂S₂ with ^{99m}Tc was evaluated at room temperature for future applications as a bifunctional chelating agent for labeling heat-sensitive polypeptides. The reaction parameters for preparing ^{99m}Tc-N₂S₂ from Zn-N₂S₂, which included the reaction pH, Zn-N₂S₂ concentration, presence or absence of ethylenediaminetetraacetic acid (EDTA) or glucoheptonate (GH), and the reaction time, were considered. Figures 1 and 2 show typical reversed-phase high-performance liquid chromatography (RP-HPLC) and thin-layer radiochromatography (TLC) of ^{99m}Tc-N₂S₂ and Re-N₂S₂. The RP-HPLC retention time of ^{99m}Tc-N₂S₂ (8.8 min) was similar to that of ^{185/187}Re-N₂S₂ (8.6 min) verified by MS, IR, elemental analysis, and proton nuclear magnetic resonance (¹H-NMR). These results, along with previous studies [15,16], supported that the ^{99m}Tc-N₂S₂ would possess the chemical structure shown in Scheme 1.



Figure 1. Reversed-phase high-performance liquid chromatography (HPLC) radio-ultraviolet (UV) chromatograms: (**a**) ^{99m}Tc-N₂S₂; (**b**) Re-N₂S₂.

Figure 3 shows the results from a preliminary experiment as a function of reaction parameters. ^{99m}Tc-N₂S₂ was obtained in high radiochemical yields only when the reaction was conducted in the presence of EDTA. The reaction did not proceed without EDTA, while the presence of GH in place of EDTA resulted in low radiochemical yields (Figure 3a). The increase in EDTA concentration in the reaction mixture increased the radiochemical yields of ^{99m}Tc-N₂S₂ (Figure 3b). The radiochemical yields of ^{99m}Tc-N₂S₂ decreased as the reaction pH increased from pH = 5.5 to 7.5. However, the radiochemical yields reached similar to one another at 30 min (Figure 3c).

Figure 4 shows the radiochemical yields of 99m Tc-N₂S₂ under the optimal conditions in the presence or absence of an IgG equimolar amount to that of Zn-N₂S₂. No significant differences were observed in the radiochemical yields of 99m Tc-N₂S₂ between the two experimental conditions.



Figure 2. Thin-layer radiochromatography (TLC) of ^{99m}Tc-N₂S₂.



Figure 3. The radiochemical yields of ^{99m}Tc-N₂S₂ as a function of (**a**) ethylenediaminetetraacetic acid (EDTA) or glucoheptonate (GH) (pH = 7.5, Zn-N₂S₂: 1.0×10^{-5} M, EDTA: 1.5×10^{-4} M, GH: 1.5×10^{-4} M); (**b**) EDTA concentration (pH = 7.5, Zn-N₂S₂: 1.0×10^{-5} M); (**c**) reaction pH (Zn-N₂S₂: 1.0×10^{-5} M, EDTA: 1.5×10^{-4} M).



Figure 4. The radiochemical yields of 99m Tc-N₂S₂ in the presence or absence of an equimolar amount of IgG (EDTA: 1.5×10^{-4} M, pH = 5.5, Zn-N₂S₂: 1.0×10^{-5} M, IgG: 0, or 1.0×10^{-5} M).

3. Discussion

 ^{99m}Tc radiopharmaceuticals are usually prepared under sterile conditions by mixing a solution of $^{99m}\text{TcO}_4^-$ with a kit formulation consisting of a ligand and a reducing agent. The in situ deprotection of N₂S₂ ligand and subsequent ^{99m}Tc complexation reaction is preferable for clinical applications. In this study, the one-pot synthesis of $^{99m}\text{Tc}\text{-N}_2\text{S}_2$ from Zn-N₂S₂ was investigated at room temperature for the applications to heat-sensitive molecules. The Zn-N₂S₂ concentration of 1.0×10^{-5} M was selected for future applications to imaging the saturable systems of the body [16].

Since the direct reaction of 99m TcO₄⁻ and Zn-N₂S₂ in the presence of a reducing agent, Sn²⁺, failed to produce 99m Tc-N₂S₂ (Figure 3a), GH was added to the reaction mixture to stabilize the

pentavalent ^{99m}TcO³⁺ against hydrolysis [16]. Under the conditions, small amounts of ^{99m}Tc-N₂S₂ (ca. 10%) were obtained with unchanged radiochemical yields with the reaction time (Figure 3a). These results suggested that GH might rather act as a weak demetallation agent to produce N₂S₂ from Zn-N₂S₂. The concentration of GH was 15-times higher than that of Zn-N₂S₂. The stability constant for Zn-GH was assumed to be close to the stability constant for Zn-gluconic acid (1.70) [17], considering the similar chemical structures and acid deprotonation constants between GH and gluconic acid [18]. Thus, a deprotection agent of higher stability constant with Zn ion was then investigated to assess the working hypothesis.

To facilitate the removal of Zn ion from Zn-N₂S₂, EDTA was selected due to its high stability constant of 16.44 with Zn ion [19]. As shown in Figure 3a, over 90% radiochemical yields were achieved under the Zn-N₂S₂ concentration of 10⁻⁵ M at room temperature for 30 min. The radiochemical yields of ^{99m}Tc-N₂S₂ increased with an increase in the EDTA concentration (Figure 3b), indicating that the demetallation of Zn from Zn-N2S2 constituted the rate-determining step for the synthesis of ^{99m}Tc-N₂S₂ from Zn-N₂S₂. It should be noted that EDTA also forms a complex with the trivalent Tc [20,21]. However, since much higher EDTA concentrations (ca. $>10^{-3}$ M) are needed to prepare $^{99m}\text{Tc-EDTA}$ [20,21], EDTA acted as a demetallation agent to generate N_2S_2 from Zn- N_2S_2 under the present reaction conditions. These results also indicated that N2S2 preferentially provided its ^{99m}Tc complex in high radiochemical yields under the presence of higher concentrations of labile chelating molecules. Indeed, ^{99m}Tc-N₂S₂ was obtained in high radiochemical yields in the presence of an equimolar amount of IgG, as shown in Figure 4. The formation of ^{99m}Tc-N₂S₂ under a wide range of reaction pH rendered Zn-N₂S₂ applicable to a variety of biomolecules of interest (Figure 3c). The gathered findings indicate that the Zn-N₂S₂ constitutes a useful precursor to prepare a variety of 99m Tc-N₂S₂-based radiopharmaceuticals at low Zn-N₂S₂ concentrations under mild reaction conditions by a simple procedure.

4. Materials and Methods

4.1. Materials

All chemicals and an antibody were reagent grade and used without further purification. The pertechnetate-99m solution was obtained from a commercial ⁹⁹Mo-^{99m}Tc Generator (Ultra-Techne Kow, FUJIFILM Toyama Chemical Co., Ltd., Tokyo, Japan).

4.2. Equipment

Radiochemical purities were determined with a Radio-Thin Layer Chromatography (TLC) Analyzer (GITA-STAR, Elysia-RAYTEST, Straubenhardt, Germany). High-performance liquid chromatography (HPLC) analyses were performed using a SHIMADZU model LC-20AD (Kyoto, Japan).

4.3. Syntheses

The N_2S_2 ligand, 1,1'-(ethane-1,2-diylbis(azanediyl))bis(2-methylpropane-2-thiol) dihydro-chloride, was synthesized according to the procedure of Ohmomo et al. [22].

Zn complex of N_2S_2 ligand: Under an argon atmosphere, 1,1'-(ethane-1,2-diylbis-(azanediyl))bis(2-methylpropane-2-thiol) dihydrochloride 50.0 mg (0.162 mmol) and anhydrous sodium acetate 19.1 mg (0.233 mmol) were dissolved in H_2O (1.3 mL) at room temperature. A mixture of Zn(II) aceteate dehydrate 59.1 mg (0.269 mmol) and anhydrous sodium acetate 19.0 mg (0.232 mmol) in 1.3 mL of H_2O was added dropwise to the solution. The reaction mixture was stirred at room temperature for 1.5 h. The solution was extracted with 10 mL of dichloromethane three times. The organic solution was dried over Na₂SO₄. After filtration, the filtrate was evaporated, and the residue was recrystallized from chloroform and hexane to afford Zn-N₂S₂ as a white powder. Yield 43.4 mg (89.4%). ESI-MS, $C_{10}H_{22}N_2S_2Zn [M + H]^+ m/z$ 299.06, found: 299.11. Anal. C₁₀H₂₂N₂S₂Zn·0.6Na₂SO₄: C, 31.19; H, 5.76; N, 7.28%, found: C, 31.59; H, 5.75; N, 7.09%.

Oxorhenium(V) complex of N₂S₂ ligand: The N₂S₂ ligand 62.3 mg (2.01 mmol) in 7 mL of 50% (*v*/*v*) ethanol/water was added to a solution of potassium perrhenate 58.2 mg (0.201 mmol) in 30 mL of 50% (*v*/*v*) ethanol/water. Tin(II) chloride dehydrate 45.5 mg (0.202 mmol) in 7 mL of 50% (*v*/*v*) ethanol/water was then added to the mixture, and the reaction mixture was stirred at room temperature for 2 h. After removing ethanol by evaporation, an aqueous solution of 1 M potassium carbonate was added to bring the reaction pH neutral. The solution was extracted with 100 mL of ethyl acetate three times. The organic solution was dried over Na₂SO₄. After filtration, the filtrate was evaporated to dryness. The residue was purified by silica gel chromatography eluted with a mixture of dichloromethane and methanol (95:5) to provide a purple crystal of ReO-N₂S₂. Yield 16.8 mg (19.2%). ESI-MS, Mass C₁₀H₂₁N₂OReS₂ [M + H]⁺ *m*/*z* 437.07, found 437.19. Anal. C₁₀H₂₁N₂OReS₂: C, 27.57; H, 4.86; N, 6.43%, found: C, 27.34; H, 4.73; N, 6.22%. IR (KBr) 920 cm⁻¹ (Re=O). ¹H-NMR (400 MHz, CD₃OD) δ 1.46 (s, 6H), 1.60 (s, 3H), 1.80 (s, 3H), 2.99 (br, 2H), 3.37 (br, 2H), 3.76 (br, 4H).

4.4. Technetuim-99m Complexatoin Reaction

All the solutions were bubbled with a stream of N₂ before use. A 1-mL solution of pertechnetate-99m (^{99m}TcO₄⁻, 581 ± 185 MBq) was added to 0.5 mL of 0.2 M of sodium phosphate buffer (pH = 5.5, 6.5 and 7.5) and 0.1 mL of Zn-N₂S₂ (6.39, 38.3 and 230 µg) ethanol solution. Then, 0.5 mL of SnCl₂·2H₂O (2.1×10^{-4} M) solution or SnCl₂·2H₂O (2.1×10^{-4} M) containing a transfer ligand (EDTA·2Na·2H₂O 6.4 × 10⁻⁴ M or GH sodium salt 6.4 × 10⁻⁴ M) aqueous solution was added to the mixture. The mixture was stood at room temperature for 5, 15, and 30 min. The effect of EDTA concentrations (1.0, 6.1, and 37 × 10⁻⁵ M) on the radiochemical yields was also evaluated.

The radiochemical yields of 99m Tc-N₂S₂ were also determined in the presence of an equimolar amount of IgG at pH = 5.5 in the presence of 1.5×10^{-4} M of EDTA. The concentration of Zn-N₂S₂ and the IgG were 1.0×10^{-5} M.

4.5. Measurement of Radiochemical Yields of 99m Tc-N₂S₂

Radiochemical yields of ^{99m}Tc-N₂S₂ were determined by TLC method with the C18 reversed-phase TLC plate (NAGEL RP-18W/UV254) eluted with a mixture of acetone and 0.5 M of ammonium acetate (65:35). Under the conditions, ^{99m}Tc-N₂S₂ had a Rf value of 0.6–0.7, while the Rf values of ^{99m}TcO₄⁻ and ^{99m}TcO₂.nH₂O were 0.9 to 1.0 and 0, respectively.

4.6. Characterization of 99m Tc-N₂S₂ and Re-N₂S₂

The HPLC retention time of 99m Tc-N₂S₂ and Re-N₂S₂ was compared with SHISEIDO CAPCELL PAK UG120 (5 μ m, 4.6 × 150 mm) eluted with 45% (v/v) aqueous methanol at 40 °C under a flow rate of 1 mL/min.

4.7. Statistical Analysis

Results were statistically analyzed using EXSUS Version 8.1. Differences were considered statistically significant when p was < 0.05. A Shapiro–Wilk test was used to determine normality. When data were normally distributed for two groups, a student's *t*-test was used. If that was not the case, the nonparametric Wilcoxon's rank sum test was used.

5. Conclusions

The Zn-N₂S₂ was easily synthesized, remained stable under aerobic conditions, and provided 99m Tc-N₂S₂ in high radiochemical yields under a mild one-pot reaction at the Zn-N₂S₂ concentration of 10^{-5} M. The formation of 99m Tc-N₂S₂ was not hindered by the presence of labile chelators, such as EDTA and an IgG antibody. The gathered findings would facilitate the development of cost-effective kit formulations for 99m Tc-radiopharmaceuticals using the N₂S₂ ligand as the chelating moiety.

Author Contributions: S.O. conceived, designed, performed experiments on syntheses of Zn, Re-N₂S₂ and ^{99m}Tc labeling, wrote paper; T.U. performed experiments on synthesis of N₂S₂ and Zn-N₂S₂, revised the paper; H.S. performed experiments on synthesis of N₂S₂; M.K.-S. performed statistical analysis on ^{99m}Tc-N₂S₂ labeling with non IgG and IgG; A.H. designed, revised the paper; Y.A. participated in the design and execution of the studies, and critically revised the paper. All authors have read and agreed to the published version of the manuscript.

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References

- Kung, H.F.; Guo, Y.-Z.; Yu, C.-C.; Billings, J.; Subramanyam, V.; Calabrese, J.C. New brain perfusion imaging agents based on ^{99m}Tc-bis(aminoethanethiol) complexes: Stereoisomers and biodistribution. *J. Med. Chem.* **1989**, *32*, 433–437. [CrossRef] [PubMed]
- Mahmood, A.; Baidoo, K.E.; Lever, S.Z. Stereoisomera of neutral oxotechnetium(V) and oxorhenium(V) complexes. In *Technetium and Rhenium in Chemistry and Nuclear Medicine 3*; Nicolini, M., Bandoli, G., Mazzi, U., Eds.; CortinaInternational, Verona, Raven Press: New York, NY, USA, 1990; pp. 119–124.
- 3. Kung, H.F.; Molnar, M.; Billings, J.; Wicks, R.; Blau, M. Synthesis and biodistribution of neutral lipid-soluble Tc-99m complexes that cross the blood-brain barrier. *J. Mucl. Med.* **1984**, *25*, 326–332.
- 4. Yamamura, N.; Magata, Y.; Arano, Y.; Kawaguchi, T.; Ogawa, K.; Konishi, J.; Saji, H. Technetium-99m-labeled medium-chain fatty acid analogues metabolized by beta-oxidation: Radiopharmaceutical for assessing liver function. *Bioconjug. Chem.* **1999**, *10*, 489–495. [CrossRef] [PubMed]
- Santos-Cuevas, C.L.; Ferro-Flores, G.; Rojas-Calderon, E.L.; García-Becerra, R.; Ordaz-Rosado, D.; Arteaga de Murphy, C.; Pedraza-López, M. ^{99m}Tc-N₂S₂-Tat (49-57)-bombesin internalized in nuclei of prostate and breast cancer cells: Kinetics, dosimetry and effect on cellular proliferation. *Nucl. Med. Commun.* **2011**, *32*, 303–313. [CrossRef] [PubMed]
- Demoin, D.W.; Dame, A.N.; Minard, W.D.; Gallazzi, F.; Seickman, G.L.; Rold, T.L.; Bernskoetter, N.; Fassbender, M.E.; Hoffman, T.J.; Deakyne, C.A.; et al. Monooxorhenium(V) complexes with 222-N₂S₂ MAMA ligands for bifunctional chelator agents: Syntheses and preliminary in vivo evaluation. *Nucl. Med. Biol.* 2016, 43, 802–811. [CrossRef] [PubMed]
- Orlandi, C.; Crane, P.D.; Platts, S.H.; Walovitch, R.C. Regional cerebral blood flow and distribution of [^{99m}Tc]ethyl cysteinate dimer in nonhuman primates. *Stroke* 1990, 21, 1059–1063. [CrossRef] [PubMed]
- Thakur, M.L.; Aruva, M.R.; Gariepy, J.; Acton, P.; Rattan, S.; Prasad, S.; Wickstrom, E.; Alavi, A. PET imaging of oncogene overexpression using ⁶⁴Cu-vasoactive intestinal peptide (VIP) analog: Comparison with ^{99m}Tc-VIP analog. *J. Mucl. Med.* 2004, 45, 1381–1389.
- 9. Hnatowich, D.J.; Qu, T.; Chang, F.; Ley, A.C.; Ladner, R.C.; Rusckowski, M. Labeling peptides with technetium-99m using a bifunctional chelator of a N-hydroxysuccinimide ester of mercaptoacetyltriglycine. *J. Nucl. Med.* **1998**, *39*, 56–64. [PubMed]
- Weber, R.W.; Boutin, R.H.; Nedelman, M.A.; Lister-James, J.; Dean, R.T. Enhanced kidney clearance with an ester-linked ^{99m}Tc-radiolabeled antibody Fab'-chelator conjugate. *Bioconjug. Chem.* **1990**, *1*, 431–437. [CrossRef] [PubMed]
- Rajagopalan, R.; Grummon, G.D.; Bugaj, J.; Hallemann, L.S.; Webb, E.G.; Marmion, M.E.; Vanderheyden, J.L.; Srinivasan, A. Preparation, characterization, and biological evaluation of technetium(V) and rhenium(V) complexes of novel heterocyclic tetradentate N3S ligands. *Bioconjug. Chem.* 1997, *8*, 407–415. [CrossRef] [PubMed]
- Lecina, J.; Carrer, A.; Alvarez-Larena, A.; Mazzi, U.; Melendez-Alafort, L.; Suades, J. New Bioconjugated Rhenium Carbonyls by Transmetalation Reaction with Zn Derivatives. *Organometallics* 2012, *31*, 5884–5893. [CrossRef]
- 13. Potenza, M.N.; Stibrany, R.T.; Potenza, J.A.; Schugar, H.J. Structures of Zinc(II) with Tetradentate N₂S₂ Ligation. *Acta Crystallogr. C* **1992**, *48*, 454–457. [CrossRef] [PubMed]
- 14. PMDA. Available online: http://www.pmda.go.jp/files/000197758.pdf (accessed on 3 April 2017).
- Meegalla, S.K.; Plössl, K.; Kung, M.P.; Stevenson, D.A.; Mu, M.; Kushner, S.; Liable-Sands, L.M.; Rheingold, A.L.; Kung, H.F. Specificity of diastereomers of [^{99m}]TRODAT-1 as dopamine transporter imaging agents. *J. Med. Chem.* **1998**, *41*, 428–436. [CrossRef] [PubMed]

- Taira, Y.; Uehara, T.; Tsuchiya, M.; Takemori, H.; Mizuno, Y.; Takahashi, S.; Suzuki, H.; Hanaoka, H.; Akizawa, H.; Arano, Y. Coordination-Mediated Synthesis of Purification-Free Bivalent ^{99m}Tc-Labeled Probes for in Vivo Imaging of Saturable System. *Bioconjug. Chem.* **2018**, *29*, 459–466. [CrossRef] [PubMed]
- 17. Martell, A.E.; Smith, R.M. *Critical Stability Constants Volume 3: Other Organic Ligands*; Plenum Press: New York, NY, USA, 1977.
- 18. Escandar, G.M.; Sala, L.F. Complexes of Cu(II) with D-aldonic and D-alduronic acids in aqueous solution. *Can. J. Chem.* **1992**, *70*, 2053–2057. [CrossRef]
- 19. Martell, A.E.; Smith, R.M. *Critical Stability Constants Volume 5: First Supplement*; Plenum Press: New York, NY, USA, 1982.
- Seifert, S.; Künstler, J.U.; Schiller, E.; Pietzsch, H.J.; Pawelke, B.; Bergmann, R.; Spies, H. Novel procedures for preparing ^{99m}Tc(III) complexes with tetradentate/monodentate coordination of varying lipophilicity and adaptation to 188Re analogues. *Bioconjug. Chem.* 2004, *15*, 856–863. [CrossRef] [PubMed]
- 21. Kunstler, J.U.; Seidel, G.; Pietzsch, H.J. Efficient preparation of (^{99m})Tc(III) '4+1' mixed-ligand complexes for peptide labeling with high specific activity. *Appl. Radiat. Isot.* **2010**, *68*, 1728–1733. [CrossRef] [PubMed]
- 22. Ohmomo, Y.; Francesconi, L.; Kung, M.P.; Kung, H.F. New conformationally restricted 99mTc N2S2 complexes as myocardial perfusion imaging agents. *J. Med. Chem.* **1992**, *35*, 157–162. [CrossRef] [PubMed]

Sample Availability: Samples of the compounds are not available from the authors.



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