

Research Paper

The prognostic value of the proteasome activator subunit gene family in skin cutaneous melanoma

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Abstract

Background: The functional significance of the proteasome activator subunit (*PSME*) gene family in the pathogenesis of skin cutaneous melanoma (SKCM) remains to be elucidated.

Materials and methods: Clinical data for patients with SKCM, including expression levels of *PSME* genes, were extracted from TCGA. GO term and KEGG pathway enrichment analyses were performed. Correlations between the expression levels of *PSME* genes in SKCM were evaluated with the Pearson correlation coefficient. Functional and enrichment analyses were conducted using DAVID. Univariate and multivariate survival analyses adjusted by Cox regression were used to construct a prognostic signature. The mechanisms underlying the association between *PSME* gene expression and overall survival (OS) were explored with gene set enrichment analysis. Joint-effects survival analysis was performed to evaluate the clinical value of the prognostic signature.

Results: The median expression levels of *PSME1*, *PSME2* and *PSME3* were significantly higher in SKCM than in normal skin. *PSME1*, *PSME2*, and *PSME3* were significantly enriched in several biological processes and pathways including cell adhesion, adherens junction organization, regulation of autophagy, cellular protein localization, the cell cycle, apoptosis, and the Wnt and NF- κ B pathways. High expression levels of *PSME1* and *PSME2* combined with a low expression level of *PSME3* was associated with favorable OS.

Conclusion: Knowledge of the expression levels of the *PSME* gene family could provide a sensitive strategy for predicting prognosis in SKCM.

Key words: Proteasome activator subunit, melanoma, prognosis, nomogram, overall survival

Introduction

Skin cutaneous melanoma (SKCM) is considered one of the most aggressive and lethal cancers of the skin. In 2012, globally, there were an estimated 232,000 new cases of melanoma and 55,000 melanoma-related deaths.[1] In 2018, in the United States, there will be approximately 91,270 new cases of melanoma and 9,320 melanoma-related deaths.[2]

Tumor stage is significantly associated with prognosis in melanoma, whereby early diagnosis and treatment results in favorable overall survival (OS) rates.[3]

Proteasome activator subunit 1 (*PSME1*), proteasome activator subunit 2 (*PSME2*), proteasome activator subunit 3 (*PSME3*) and proteasome activator subunit 4 (*PSME4*) are members of the proteasome

activator subunit (PSME) gene family. Proteasome activator 28 (PA28) consists of three subunits, PA28 α , PA28 β and PA28 γ , encoded by PSME1, PSME2 and PSME3, respectively. Proteasome activators regulate proteasome function but have also been associated with several cancers and may have prognostic significance. Previous studies showed elevated expression of PSME1 in prostate cancer,[4] elevated expression of PSME2 in gastric cancer,[5] and elevated expression of PSME3 in breast cancer,[6-9] colorectal cancer,[10] and laryngeal carcinoma.[11] In some cancers, overexpression of PSME3 was associated with poor OS.[6, 12] Currently, the functional significance of PSME4 in the pathogenesis of cancer remains to be elucidated.

The objectives of the present study were to 1) identify associations between *PSME* gene expression levels in SKCM and 2) develop a risk score that includes clinical factors and the expression patterns of *PSME* genes to predict prognosis in patients with SKCM. In the present research, we were the first to analyze the prognosis value of PSME gene family in SKCM, made a nomogram model for predicting the prognosis of SKCM patients, and used whole-genome RNA-Seq dataset to explore prospective molecular mechanisms through gene set enrichment analysis (GSEA) approach.

Method and Materials

Data source

Clinical data for patients with SKCM, including gender, age, survival time, mortality, and expression levels of *PSME* genes, were extracted from The Cancer Genome Atlas (TCGA). Boxplots of expression profiles of the *PSME* genes in SKCM and healthy skin were created using Gene Expression Profiling Interactive Analysis (GEPIA, <http://gepia.cancer-pku.cn/>, accessed on June 20, 2018).[13] After exclusive the patients, which don't have gene expression data and complete prognostic information including survival status and days, 458 cases were included in our research.

PSME gene family bioinformatics analysis and correlation analysis

Gene ontology (GO) term enrichment analysis, including molecular function (MF), cellular component (CC), and biological process (BP), as well as the Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analysis were performed. *PSME* gene co-expression networks and/or pathways were predicted with GeneMANIA (<http://genemania.org/>, accessed June 22, 2018).[14] Correlations between expression levels of *PSME* genes in SKCM were evaluated with the Pearson

correlation coefficient. Functional and enrichment analyses were conducted using The Database for Annotation, Visualization, and Integrated Discovery (DAVID) v.6.8 (<https://david.ncifcrf.gov/tools.jsp>, accessed June 22, 2018).[15, 16]

Survival analysis

Prognosis of patients with SKCM was determined by OS. Correlations between expression levels of *PSME* genes in SKCM and patients' OS were evaluated using the Kaplan-Meier method and the log-rank test as well as Cox proportional hazards regression with adjustment for age and tumor stage; race was excluded as a variable due to small sample size (94% of the included patients were White). *PSME* genes were stratified by high or low expression around the median OS. The prognostic impact of high and low expression levels of each *PSME* gene was assessed.

Prognostic risk score

A prognostic risk score was developed based on the adjusted (age, tumor stage) expression levels of the *PSME1*, *PSME2* and *PSME3* genes in SKCM. Nomograms for predicting 1-, 3-, and 5-year survival were used to evaluate the association between the prognostic risk score and OS in patients with SKCM and its potential clinical application;[17] a high score was associated with poor prognosis.

Gene set enrichment analysis (GSEA)

The mechanisms underlying the association between *PSME* gene expression in SKCM and patients' OS were explored with GSEA. Pathway-based analysis in SKCM with high and low expression levels of each *PSME* gene was conducted using comparisons with the reference c5 (GO gene sets: c5.all.v6.1.symbols.gmt) and c2 (KEGG gene sets: c2.all.v6.1.symbols.gmt) gene sets from the Molecular Signatures Database (MSigDB) [18] using GSEA v.3.0 (<http://software.broadinstitute.org/gsea/msigdb/index.jsp>, accessed June 25, 2018). The number of permutations was set at 1,000. $P < 0.05$ and a false discovery rate (FDR) < 0.25 were considered statistically significant.

Joint-effects survival analysis

Associations between the expression levels of combinations of *PSME* genes in SKCM and patients' OS were assessed with joint-effects survival analysis. *PSME* genes with prognostic value on multivariate survival analysis were grouped as better OS, worse OS, or other. The prognostic value of the expression of combinations of *PSME* genes in each group was evaluated using the Kaplan-Meier method and the log-rank test.

Statistical analyses

Statistical analyses were performed with SPSS v.25.0 software (IBM, Chicago, IL, USA). Vertical scatter plots and survival curves were generated in GraphPad Prism v.7.0 (GraphPad Software, La Jolla, CA, USA) and R 3.5.1 (<http://www.R-project.org>). OS was calculated with the Kaplan-Meier method and the log-rank test. Multivariate survival analysis was evaluated with hazard ratios (HR), and 95% confidence intervals (CIs) were calculated using Cox proportional hazards regression with adjustment for influential clinical characteristics including age and tumor stage. $P < 0.05$ was considered statistically significant.

Results

Patients' clinical characteristics

Demographic and clinical data obtained from TCGA for 458 patients with SKCM are summarized. The associations between demographic and clinical characteristics and OS in patients with SKCM are summarized in **Table 1**. Race, age and tumor stage were significantly associated with median survival time (MST; $P=0.004$, $P=0.001$, and $P=0.001$, respectively).

Table 1. Clinical data for included patients.

Variables	Patients (n=458)	No. of events (%)	MST (days)	HR (95% CI)	Log-rank P
Race					0.004
White	435	208 (47.8%)	2454	Ref.	
Others	13	8 (61.5%)	636	0.348 (0.171-0.709)	
Gender					0.278
Male	284	146 (51.4%)	2454	Ref.	
Female	174	72 (41.4%)	2030	0.854 (0.642-1.136)	
Age (years)					0.001
≥60	219	102 (46.6%)	1860	Ref.	
<60	239	116 (48.3%)	3564	0.620 (0.470-2.136)	
Tumor stage					0.001
0+I+II+I/II nos	231	108 (46.8%)	3259	Ref.	
III+IV	191	91 (47.6%)	1960	0.600 (0.449-0.802)	
Missing	36				

Abbreviations: *PSME*, proteasome activator subunit; MST, median survival time; HR, hazard ratio; CI, confidence interval.

Table 2. Univariate and multivariate survival analyses.

Gene	Patients (n=458)	No. of events (%)	MST (days)	Crude HR (95% CI)	Crude P	Adjusted HR* (95% CI)	Adjusted P*
<i>PSME1</i>					0.072		0.009
Low	229	124 (54.1%)	2030	Ref.		Ref.	
High	229	94 (41.0%)	3136	0.781 (0.596-1.023)		0.685 (0.516-0.910)	
<i>PSME2</i>					0.001		0.001
Low	229	133 (58.1%)	1917	Ref.		Ref.	
High	229	85 (37.1%)	3379	0.626(0.476-0.822)		0.576 (0.431-0.769)	
<i>PSME3</i>					0.001		0.002
High	229	114 (49.8%)	1910	Ref.		Ref.	
Low	229	104 (45.4%)	3564	0.638 (0.488-0.817)		0.634 (0.477-0.842)	
<i>PSME4</i>					0.423		0.410
Low	229	100 (43.7%)	2028	Ref.		Ref.	
High	229	118 (51.5%)	2993	0.896 (0.686-1.172)		0.888(0.669-1.178)	

Notes: *, adjustment for age and tumor stage.

Abbreviations: *PSME*, proteasome activator subunit; MST, median survival time; HR, hazard ratio; CI, confidence interval.

Boxplots showing the expression profiles of *PSME* genes in SKCM or healthy skin are presented in **Figure 1**. Findings showed that median expression levels of *PSME1*, *PSME2* and *PSME3* were significantly higher in SKCM than in healthy skin.

PSME gene family correlation analysis and bioinformatics analysis

GO term analysis and KEGG pathway enrichment analysis are shown in **Figure 2A**. The *PSME* gene family was involved in the MAPK cascade, NIF/NF- κ B and Wnt signaling pathways and the cell cycle, which are tumor-related processes. The pathway and co-expression prediction among *PSME1*, *PSME2* and *PSME3* is shown in **Figure 2B**. Correlations between the expression levels of individual *PSME* genes in SKCM investigated with Pearson correlation coefficient are shown in **Figure 2C**. There were correlations between the expression levels of all *PSME* genes except for *PSME1* and *PSME3* and *PSME2* and *PSME3*.

Survival analysis

Scatter plots showing the expression levels of *PSME* genes in SKCM, stratified as high expression or low expression, are shown in **Figure 3**. Survival analysis is summarized in **Table 2** and shown in **Figure 4**. On univariate survival analysis, a high expression level of *PSME2* (log-rank $P=0.001$, HR=0.626, 95%CI=0.476-0.822; **Figure 4B**) and low expression level of *PSME3* (log-rank $P=0.001$, HR=0.638, 95%CI=0.488-0.817; **Figure 4C**) in SKCM were associated with better OS. On multivariate survival analysis, a high expression level of *PSME1* (log-rank $P=0.009$ HR=0.685 95%CI=0.516-0.910), high expression level of *PSME2* (log-rank $P=0.001$ HR=0.576 95%CI=0.431-0.769), and low expression level of *PSME3* (log-rank $P=0.002$ HR=0.634 95%CI=0.477-0.842) in SKCM were associated with better OS.

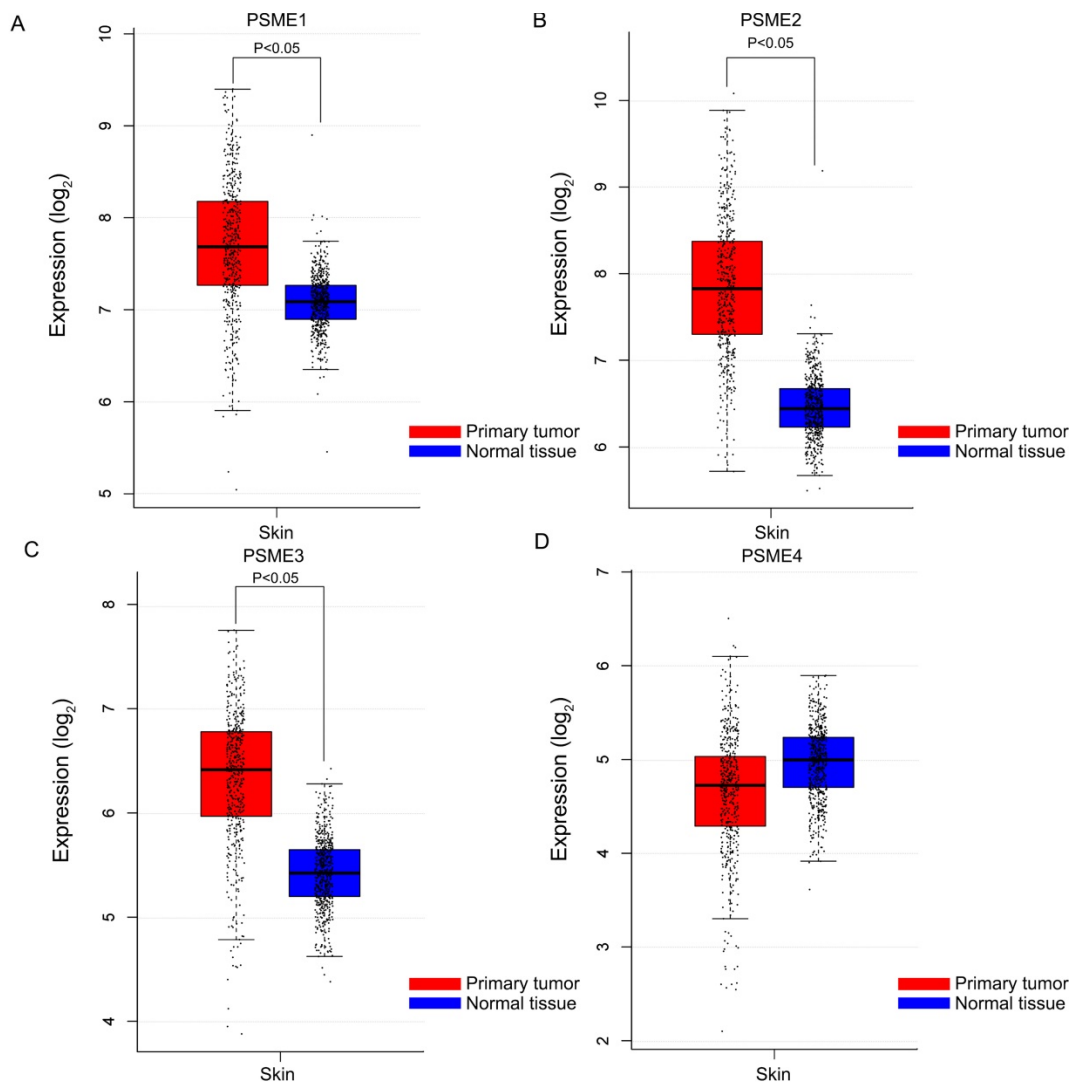


Figure 1. Boxplots showing *PSME* gene expression levels in SKCM and healthy skin. (A) *PSME1*; (B) *PSME2*; (C) *PSME3*; (D) *PSME4*. Abbreviations: *PSME*, proteasome activator subunit; GEPIA, gene expression profiling interactive analysis

Nomogram of SKCM risk score model

A nomogram substantiated that age, tumor stage, and *PSME2* and *PSME3* expression levels in SKCM created a prognostic signature that contributed the most risk (range 0–100 points) for poor OS. Each variable was assigned points based on the Cox regression coefficients. These points were summed, and the probability of survival was estimated by drawing a vertical line between the Total Points axis and the 1-year, 3-year and 5-year survival probability axes (Figure 4E).

GSEA

Pathway-based analysis in SKCM with high and low expression levels of each *PSME* gene is shown in Figure 5 (A-I), Figure 6 (A-I), Figure 7 (A-I), Figure 8 (A-I), Figure 9 (A-I) and Figure 10 (A-I). In the GO enrichment analysis, a high expression of *PSME1* was

positively correlated with the apoptotic process (Figure 5A), cell adhesion (Figure 5B), and the NF-κB (Figure 5C) and Wnt signaling pathways (Figure 5E, F). High expression of *PSME2* was negatively correlated with the apoptotic process (Figure 6B), cell adhesion (Figure 6C, F), and the NF-κB signaling pathway (Figure 6D). High expression of *PSME3* was positively correlated with the NF-κB (Figure 7C) and Wnt signaling pathways (Figure 7E, F). In the KEGG pathway, high expression of *PSME1* was positively correlated with cell adhesion (Figure 8A), apoptosis (Figure 8D, E), the cell cycle (Figure 8F), metastasis (Figure 8I) and the Wnt and NF-κB signaling pathways (Figure 8B, C and G). High expression of *PSME2* was negatively correlated with cell adhesion (Figure 9B), the cell cycle (Figure 9E), apoptosis (Figure 9F) and the Wnt signaling pathway (Figure 9G). High expression of *PSME3* was positively correlated with metastasis (Figure 10A, D), the

P53-induced cell cycle (Figure 10F, G), the cell cycle (Figure 10H), and the Wnt signaling pathway (Figure 10C, I). The remaining results were presented in Supplementary Table 1 and 2.

Joint-effects survival analysis

Based on the findings on multivariate survival analysis, a joint-effects survival analysis was performed to determine the combined effects of *PSME1*, *PSME2* and *PSME3* in SKCM on OS in

patients grouped as summarized in Table 3. Results are summarized in Table 4 and shown in Figure 11. High expression levels of *PSME1* and *PSME2* combined with low expression level of *PSME3* in SKCM in Groups I, IV, VII, and X was associated with better OS (all $P < 0.05$). In contrast, low expression levels of *PSME1* and *PSME2* combined with a high expression level of *PSME3* in SKCM in Groups III, VI, IX and XII was associated with poor OS (all $P < 0.05$).

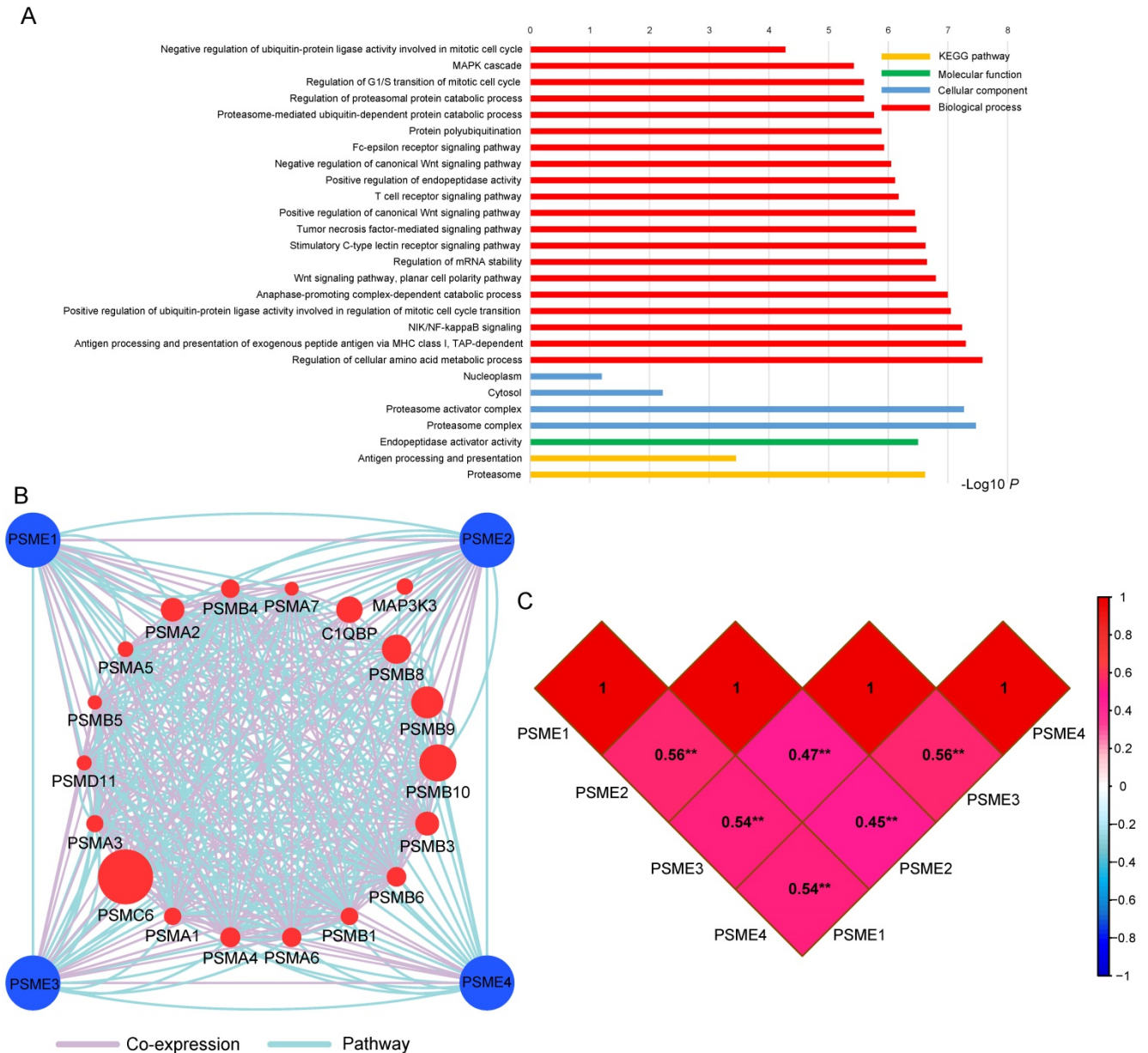


Figure 2. (A) GO enrichment and KEGG pathway analysis by DAVID; (B) Gene interaction networks among selected genes by GeneMANIA; (C) Pearson's correlation coefficients between *PSME1*, *PSME2* and *PSME3* expression levels; and $**P < 0.001$. Abbreviations: *PSME*, proteasome activator subunit; TCGA, The Cancer Genome Atlas; GeneMANIA, gene multiple association network integration algorithm; DAVID, the database for annotation, visualization, and integrated discovery; GO, gene ontology; KEGG, Kyoto encyclopedia of genes and genomes.

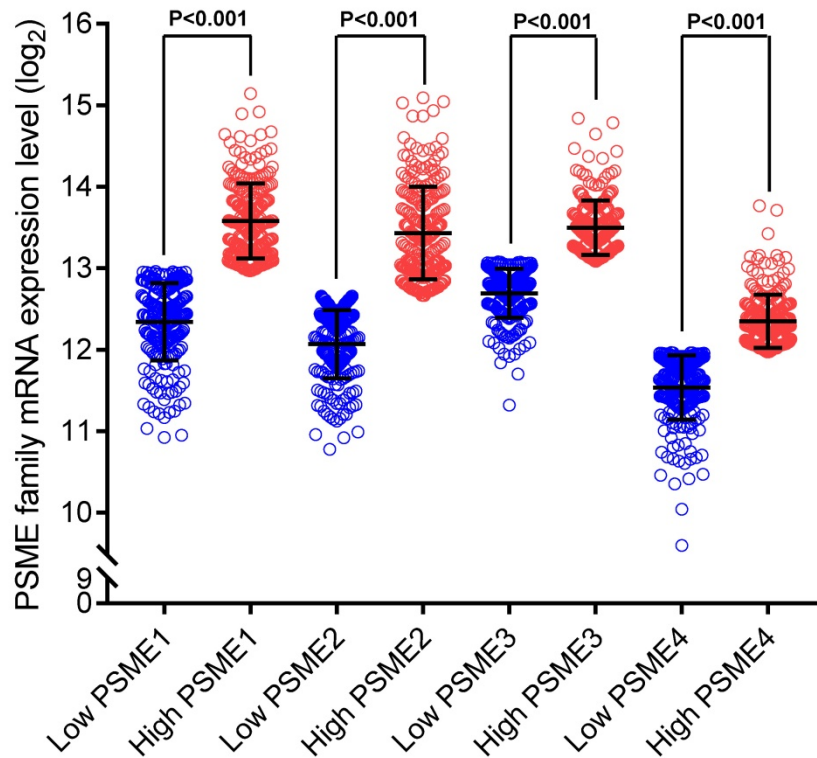


Figure 3. Scatter plots showing *PSME1*, *PSME2* and *PSME3* expression levels in SKCM Abbreviations: *PSME*, proteasome activator subunit; SKCM, skin cutaneous melanoma.

Table 3. Stratifications based on the expression levels of the *PSME1*, *PSME2* and *PSME3* genes.

Group	Composition	Group	Composition
I	high <i>PSME1</i> +high <i>PSME2</i>	X	high <i>PSME1</i> +high <i>PSME2</i> +low <i>PSME3</i>
II	low <i>PSME1</i> +high <i>PSME2</i>	XI	high <i>PSME1</i> +low <i>PSME2</i>
III	Low <i>PSME1</i> +low <i>PSME2</i>		high <i>PSME1</i> +low <i>PSME2</i> +high <i>PSME3</i>
IV	high <i>PSME1</i> + low <i>PSME3</i>		low <i>PSME1</i> +high <i>PSME2</i> +high <i>PSME3</i>
V	low <i>PSME1</i> +low <i>PSME3</i>		low <i>PSME1</i> +low <i>PSME2</i> +low <i>PSME3</i>
	high <i>PSME1</i> +high <i>PSME3</i>		high <i>PSME1</i> +high <i>PSME2</i> +high <i>PSME3</i>
VI	low <i>PSME1</i> +high <i>PSME3</i>		high <i>PSME1</i> +low <i>PSME2</i> +low <i>PSME3</i>
VII	high <i>PSME2</i> +low <i>PSME3</i>		low <i>PSME1</i> +high <i>PSME2</i> +low <i>PSME3</i>
VIII	low <i>PSME2</i> +low <i>PSME3</i>		
	high <i>PSME2</i> +high <i>PSME3</i>		
IX	low <i>PSME2</i> +high <i>PSME3</i>	XII	Low <i>PSME1</i> +low <i>PSME2</i> +high <i>PSME3</i>

Abbreviation: *PSME*, proteasome activator subunit.

Table 4. Joint-effects survival analysis.

Group	Patients (n=458)	MST (days)	Crude P	Crude HR (95% CI)	Adjusted P*	Adjusted HR* (95% CI)
I	196	3195	0.005	0.655 (0.487-0.881)	0.004	0.645 (0.479-0.867)
II	66	3869	0.075	0.702 (0.476-1.036)	0.071	0.697 (0.470-1.032)
III	196	1910	0.012	Ref.	0.009	Ref.
IV	111	4507	<0.001	0.494 (0.334-0.730)	<0.001	0.486 (0.329-0.719)
V	236	2273	0.070	0.751 (0.552-1.023)	0.036	0.718 (0.527-0.979)
VI	111	1487	0.002	Ref.	0.001	Ref.
VII	120	4570	<0.001	0.430 (0.296-0.624)	<0.001	0.428 (0.295-0.622)
VIII	218	2454	0.007	0.660 (0.488-0.893)	0.004	0.643 (0.475-0.871)
IX	120	1478	<0.001	Ref.	<0.001	Ref.
X	98	4570	<0.001	0.048	<0.001	0.440

Group	Patients (n=458)	MST (days)	Crude P	Crude HR (95% CI)	Adjusted P*	Adjusted HR* (95% CI)
XI	260	2454	0.011	0.671 (0.296-0.678)	0.006	0.645 (0.290-0.667)
XII	100	1446	0.001	Ref. (0.492-0.914)	<0.001	Ref. (0.472-0.881)

Notes: *, adjustment for age and tumor stage. Bold type highlights statistically significant values ($P \leq 0.05$).

Abbreviations: *PSME*, proteasome activator subunit; MST, median survival time; HR, hazard ratio; CI, confidence interval.

Discussion

In this study, we used data from TCGA to investigate the associations between *PSME* gene expression levels in SKCM and developed a risk score that includes clinical factors and the expression patterns of *PSME* genes to predict prognosis in patients with SKCM. *PSME* genes, including *PSME1*, *PSME2* and *PSME3*, encode the PA28 α , PA28 β and PA28 γ subunits, respectively, of PA28, which regulates function of the proteasome.[19] In the present study, *PSME1*, *PSME2* and *PSME3* expression levels were significantly increased in SKCM compared to healthy skin. GO enrichment analysis showed that *PSME1* is a negative regulator of cell adhesion, *PSME2* is important for cell-cell adhesion and junction organization, and *PSME3* is associated with NF- κ B signaling. Importantly, the activation of NF- κ B can impart invasiveness and properties of cancer initiation on cells, and may act as a target for anti-cancer therapy.[20] GO term analysis also

showed that *PSME* was associated with MAPK cascade, which the pathway was found to be correlated with melanoma.[21, 22] High expression levels of *PSME1* and *PSME2* combined with a low expression level of *PSME3* in SKCM were associated with favorable prognosis. Pathway-based analysis revealed that *PSME1* is associated with KEGG and apoptosis pathways and that *PSME2* and *PSME3* are

significantly enriched in the canonical and planar cell polarity Wnt signaling pathways, which have been associated with cancer.[23, 24] Taken together, the findings from the present study suggest that expression levels of the *PSME1*, *PSME2* and *PSME3* genes in SKCM, individually and in combination, may be used as potential biomarkers to predict prognosis.

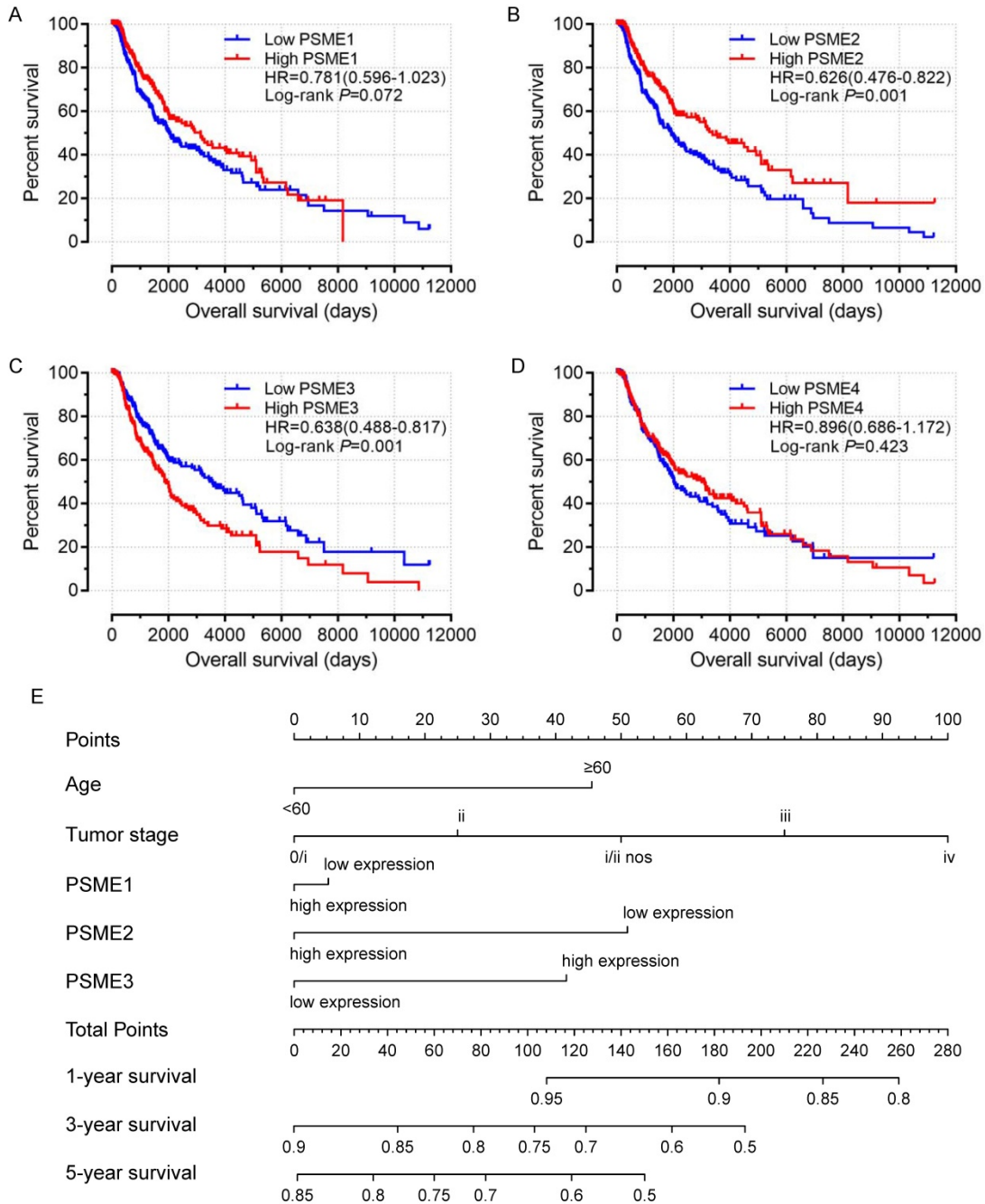


Figure 4. Univariate survival analysis and nomogram. (A) *PSME1*, (B) *PSME2*, (C) *PSME3*, (D) *PSME4*, (E) nomogram to predict survival in SKCM. Abbreviation: *PSME*, proteasome activator subunit; SKCM, skin cutaneous melanoma.

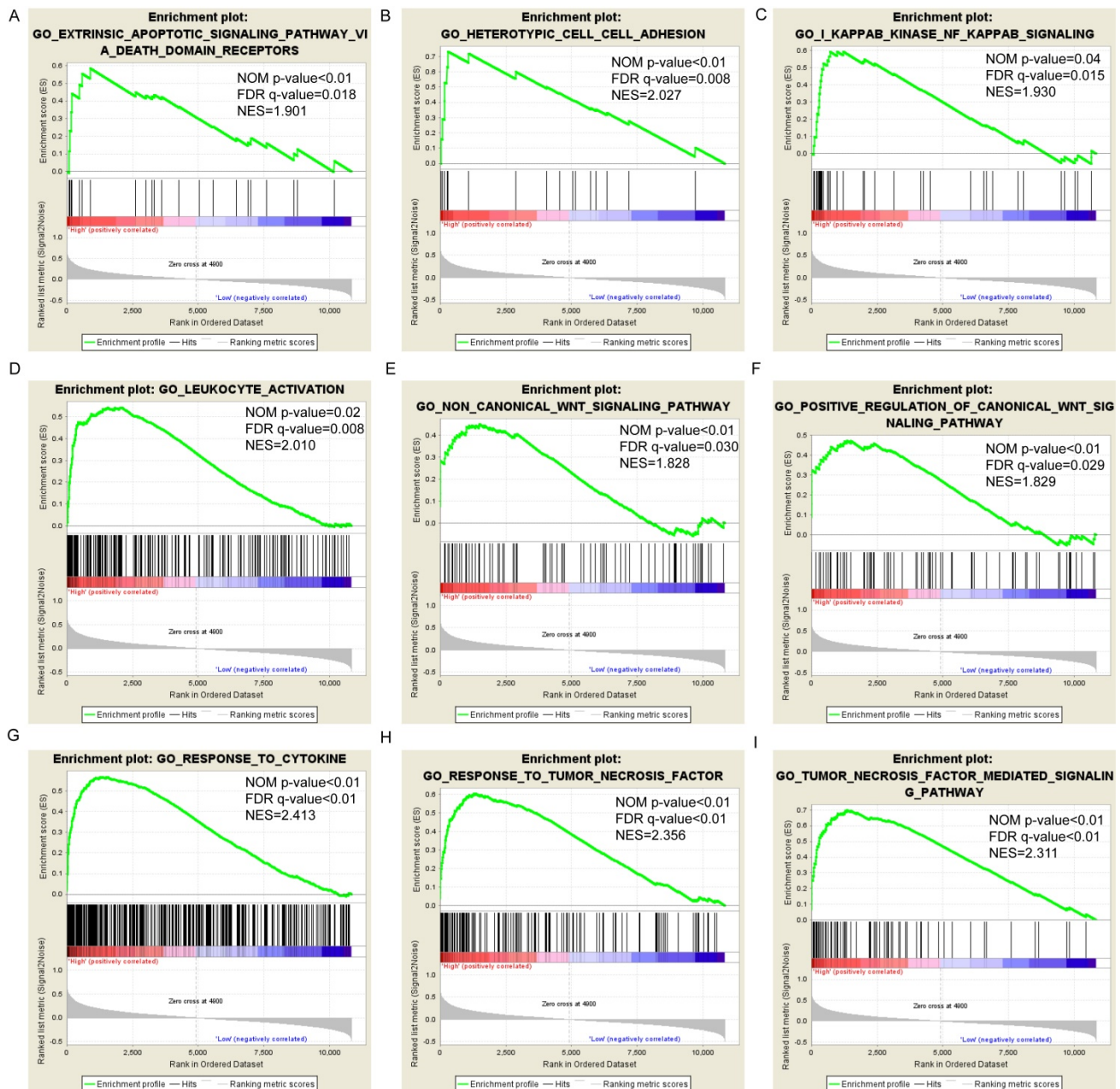


Figure 5. (A-I) GO enrichment analysis by low and high *PSME1* expression levels. Abbreviations: NES, Normalized enrichment score; FDR, false discovery rate; GSEA, gene set enrichment analysis; GO, gene ontology.

For *PSME1*, the findings from the present study are in contrast to those from previous reports, which demonstrated that *PSME1* expression was increased in primary and metastatic human prostate cancer, *PSME1* was a marker in mouse xenograft tumors,[4] and *PA28 α* protein was downregulated in HBV-infected well-differentiated hepatocellular carcinoma.[25] The disparate findings between the present and some previous studies suggest that *PSME1* may play different roles in different types of cancer.

Previous reports on *PSME2* are in accordance with the results from the present study. Evidence suggests that *PA28 β* protein regulates invasive-

ness and metastasis in gastric cancer, whereby the invasive abilities of gastric cancer cells were enhanced by the down-regulation of *PA28 β* and inhibited when *PA28 β* was overexpressed,[5] and that *PA28 β* is physically associated with N- α -acetyltransferase 10 protein, which regulates various pathways associated with cancer cell proliferation, metastasis, apoptosis, and autophagy.[26]

The role of *PSME3* in cancer has been well characterized. *PSME3* knockout mice treated with dextran sodium sulfate to induce acute colitis showed decreased intestinal inflammation and colitis-associated cancer compared to wild-type mice.[27] In oral squamous cell carcinoma, high

expression of *PSME3* was correlated with worse OS, while *PSME3* silencing inhibited the growth, proliferation and mobility of oral squamous cell carcinoma cells *in vitro* and reduced tumor growth and angiogenesis in mice *in vivo*. [12] Similarly, *PSME3* silencing attenuated the cell proliferation, migration and invasive abilities of endometrial cancer cells. In a model of skin tumorigenesis, *PSME3* functioned as an oncogene, whereby the TPA-induced overexpression of *PSME3* was dependent on the activation of the MAPK-p38 signaling pathway. [28] In breast cancer, 5-year disease-free survival and OS in patients with undetectable or low *PSME3* expression were significantly higher than in patients with high *PSME3* expression. [6] In colorectal cancer, *PSME3*

expression was higher in colorectal cancer tissue than in healthy tissues. [10] Other studies indicate that mutations in the *TP53* gene, which encodes the tumor suppressor protein p53, occur in various types of cancer, and that *PSME3* negatively regulates p53, whereby the elimination of endogenous *PSME3* in human cancer cells abrogates MDM2-mediated p53 degradation, increases the activity of p53, and enhances apoptosis. [29] Notably, p53 mutations show a positive correlation with *PSME3* expression in various cancer cell lines. [30] In normal endometrium, expression of *PSME3* was increased in p53-positive specimens compared to p53-negative specimens, [31] and in laryngeal carcinoma, the expression of *PSME3* was correlated with p53 and p21. [11, 32-34]

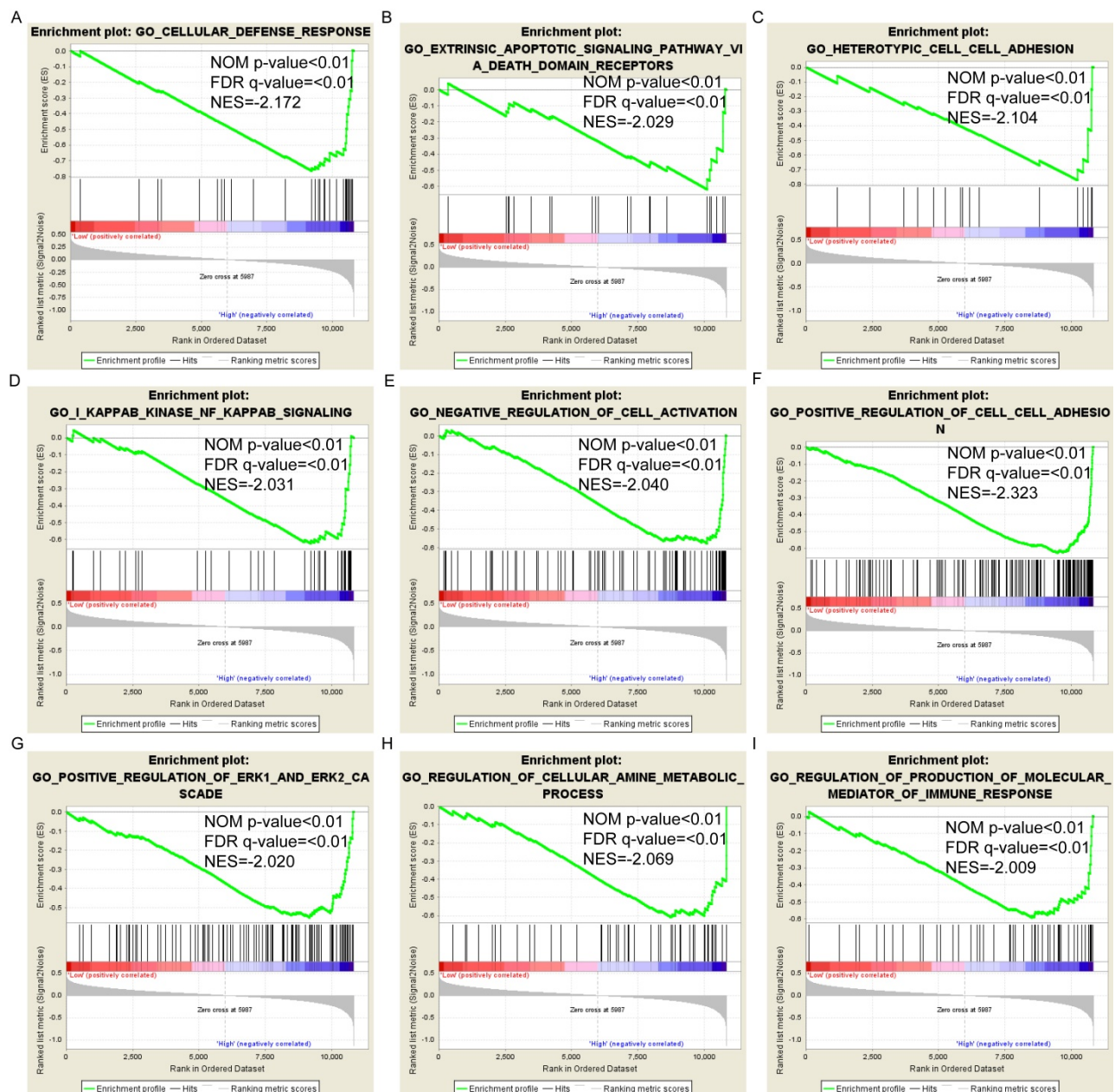


Figure 6. (A-I) GO enrichment analysis by low and high *PSME2* expression levels. Abbreviations: NES, normalized enrichment score; FDR, false discovery rate; GSEA, gene set enrichment analysis; GO, gene ontology.

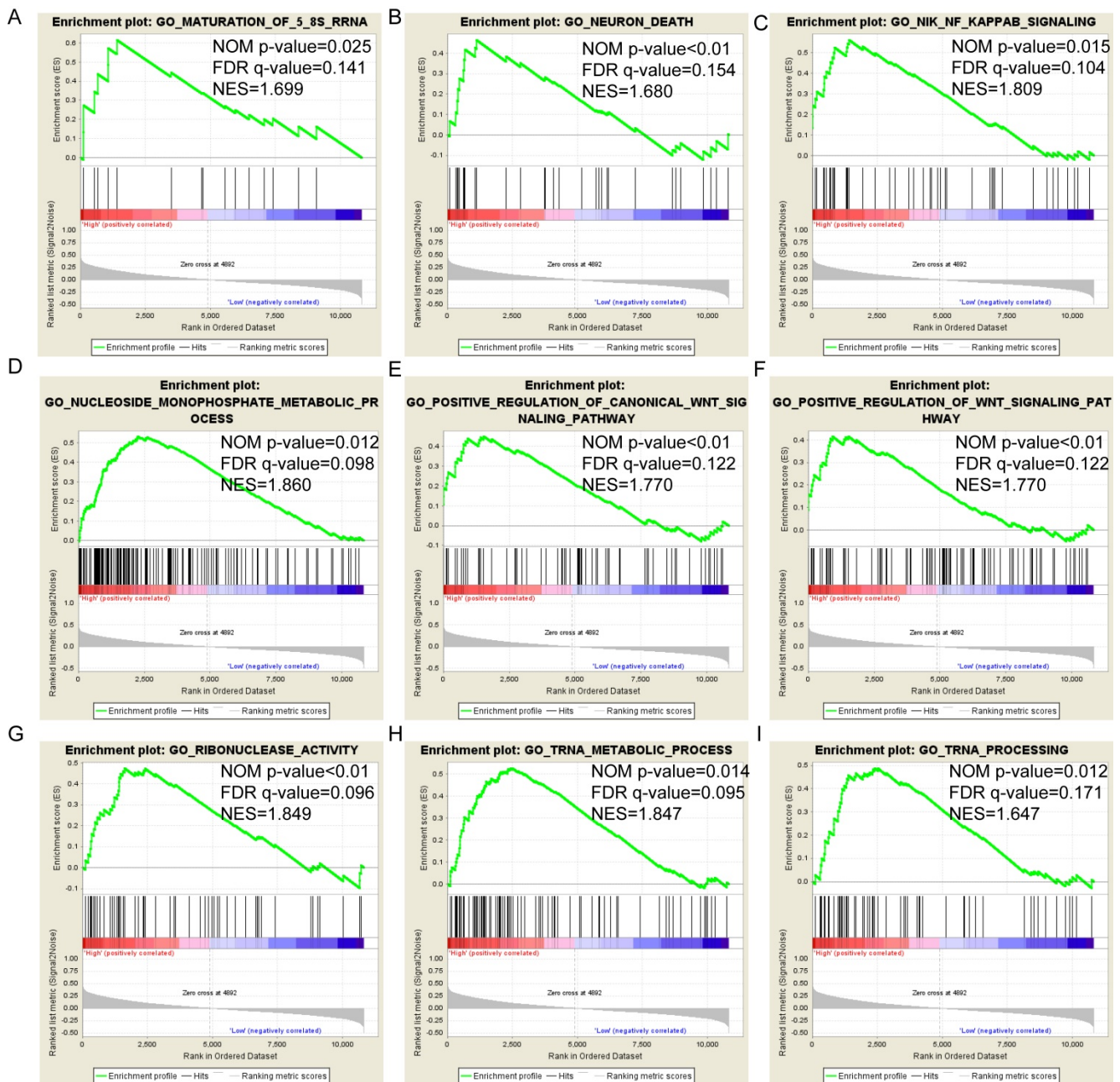


Figure 7. (A-I) GO enrichment analysis by low and high *PSME3* expression levels. Abbreviations: NES, normalized enrichment score; FDR, false discovery rate; GSEA, gene set enrichment analysis; GO, gene ontology.

Despite the wealth of literature on the role of *PSME* genes in cancer, to the authors' knowledge, the present study is the first to develop a risk score that includes clinical factors and the expression patterns of *PSME* genes to predict prognosis in patients with SKCM. The risk score can be used to stratify patients with SKCM into groups at high or low risk for poor prognosis. Univariate survival analysis showed that a high expression level of *PSME2* and low expression level of *PSME3* in SKCM were correlated with favorable OS. Multivariate survival analysis showed that high expression level of *PSME1*, adjusted by age and tumor stage, in SKCM was also correlated with

favorable prognosis. Joint-effects survival analysis showed that high expression levels of *PSME1* and *PSME2* combined with a low expression level of *PSME3* in SKCM was associated with favorable OS. In contrast, low expression levels of *PSME1* and *PSME2* combined with a high expression level of *PSME3* was associated with poor OS.

This study had several limitations. First, the sample size was small. In particular, a more ethnically diverse study population is required. In the present study, the majority of subjects were White. Second, clinical information, including information on sun exposure and genetic factors, was lacking. Third, the

patients in the current study were from a single cohort, which may introduce bias. Findings from the present study should be verified in a larger and more diverse set of patients. Forth, our current study is a bioinformatics research and most of the findings were generated from public database and bioinformatics analysis, which lack of verification through *in vitro* and *in vivo* experiments. Finally, SKCM is the melanoma of skin is a fairly rare disease and the related resources are also rare, so this study lake of validation methods to confirm the results including

independent cohort. Therefore, results of current study still need further verified.

Despite these limitations, to the authors' knowledge, this is the first study to demonstrate that high expression levels of *PSME1* and *PSME2* combined with a low expression level of *PSME3* is associated with favorable prognosis in SKCM. These findings may have prognostic significance in SKCM. The prognostic model constructed in this study may have value in clinical applications.

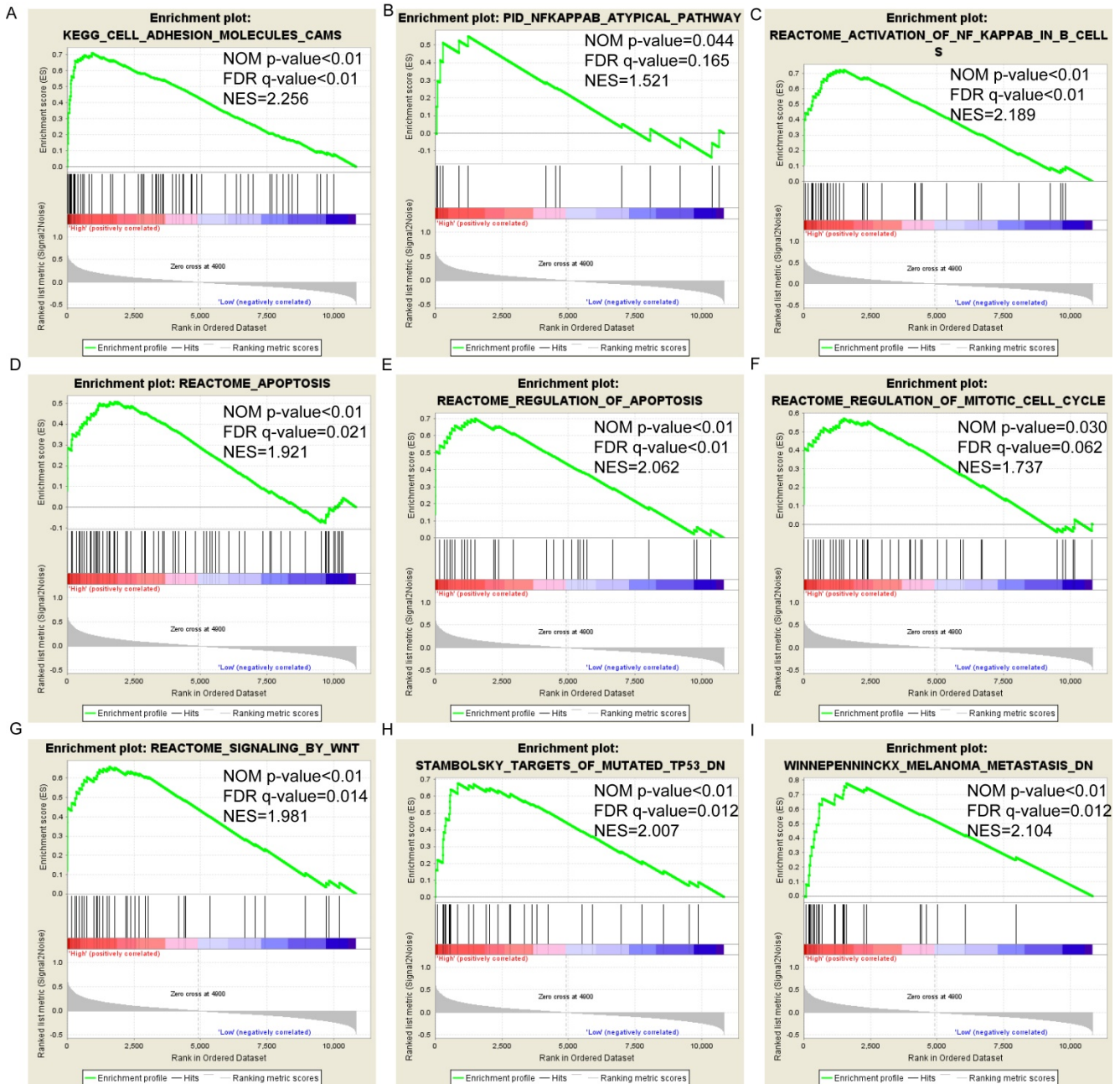


Figure 8. (A-I) KEGG pathway analysis by low and high *PSME1* expression levels. Abbreviations: NES, normalized enrichment score; FDR, false discovery rate; GSEA, gene set enrichment analysis; KEGG, Kyoto encyclopedia of genes and genomes.

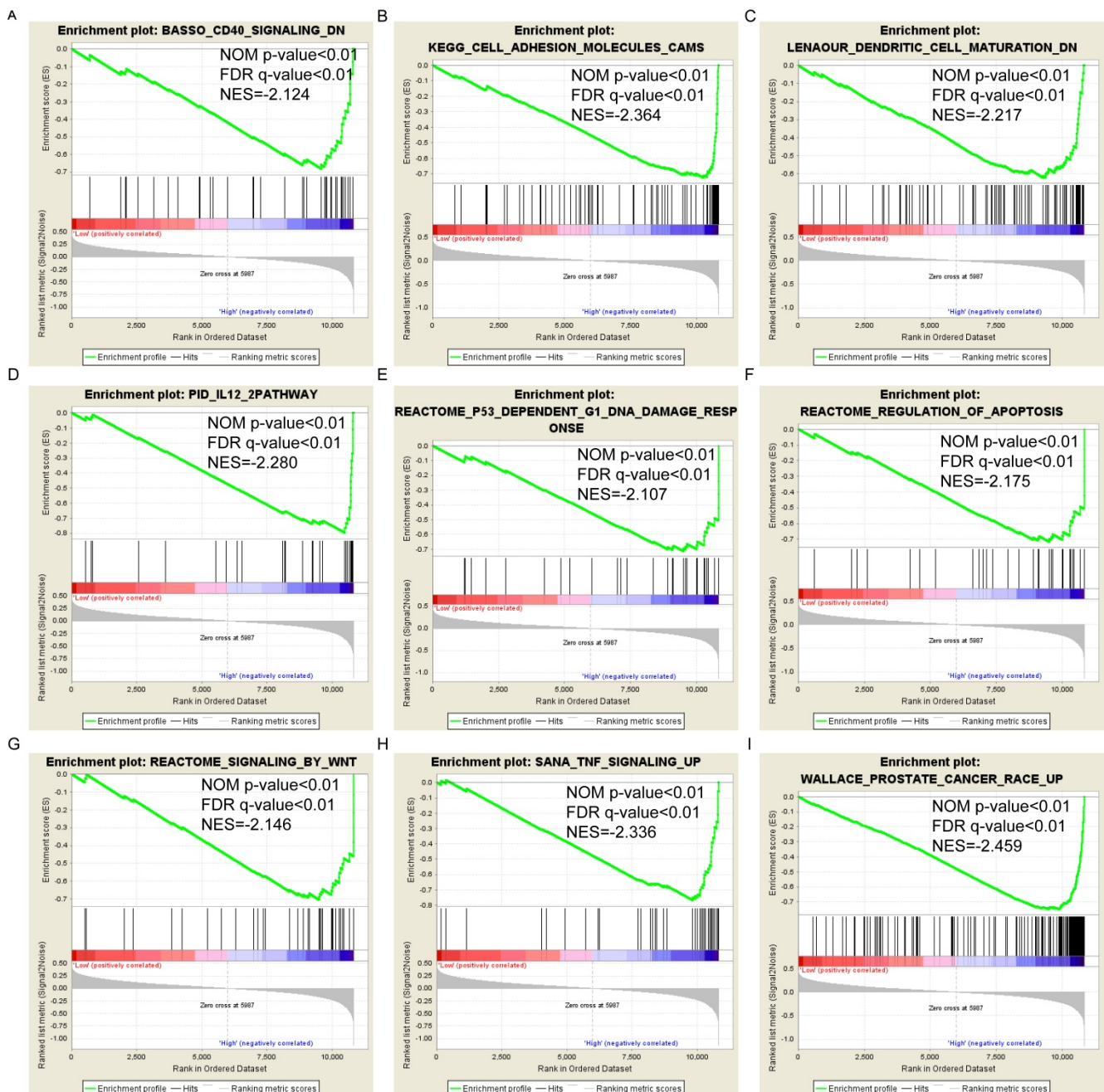


Figure 9. (A-I) KEGG pathway analysis by low and high *PSME2* expression levels. Abbreviations: NES, normalized enrichment score; FDR, false discovery rate; GSEA, gene set enrichment analysis; KEGG, Kyoto encyclopedia of genes and genomes.

Conclusion

Findings from the present study indicate that a high expression of *PSME1* and *PSME2* and low expression of *PSME3* are associated with favorable prognosis and may act as potential prognostic biomarkers in SKCM. The combined expression levels of these genes could provide a sensitive strategy for predicting prognosis in SKCM.

Supplementary Material

Supplementary table 1.

<http://www.jcancer.org/v10p2205s1.xlsx>

Supplementary table 2.

<http://www.jcancer.org/v10p2205s2.xlsx>

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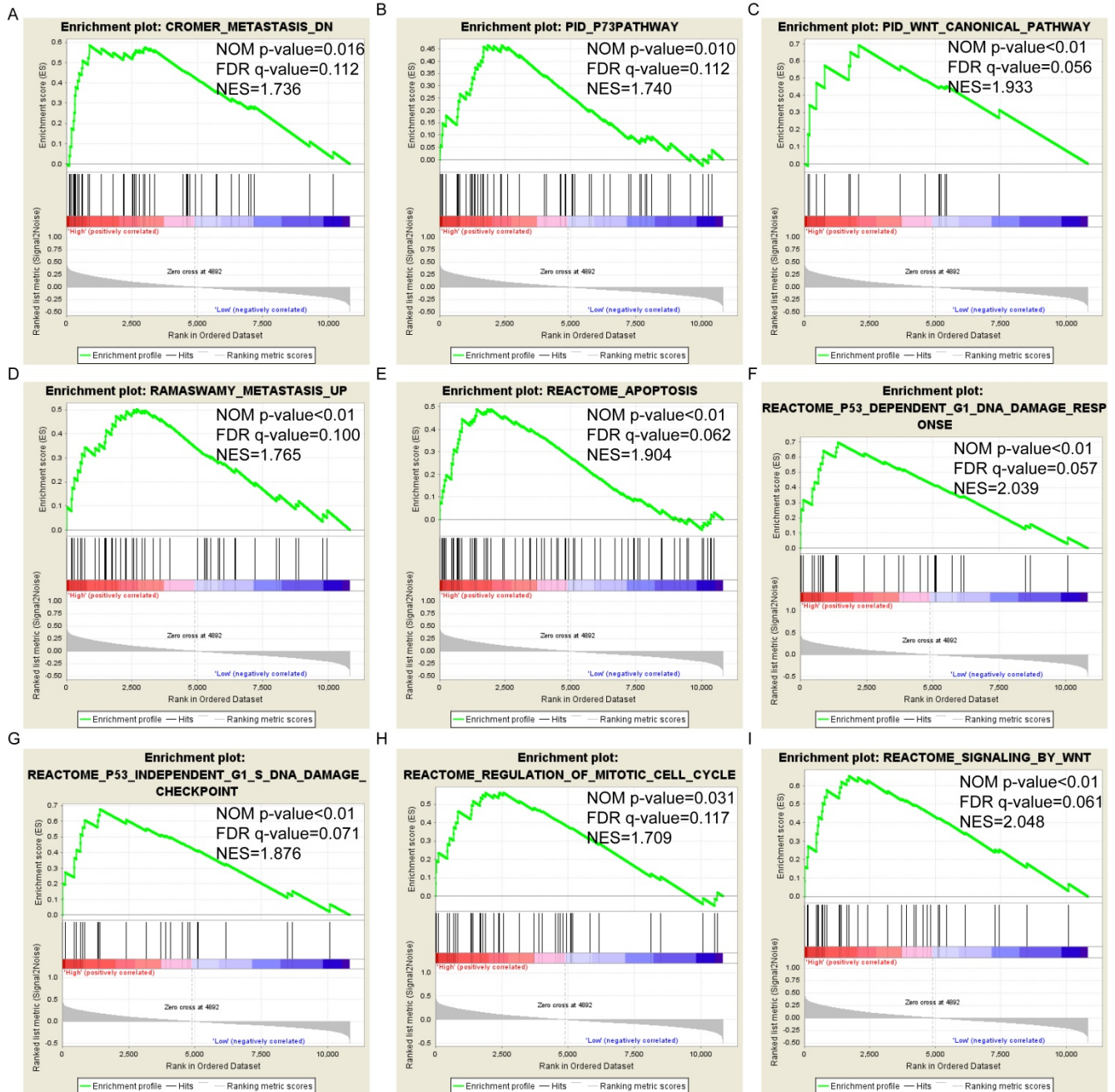
Ethics approval and consent to participate

This article does not contain any studies with

human participants or animals performed by any of the authors. Since all datasets included in the present study were downloaded from TCGA, additional approval by an Ethics Committee was not needed. The procedures were in accordance with the Helsinki declaration of 1964 and its later amendments.

Competing Interests

The authors have declared that no competing interest exists.



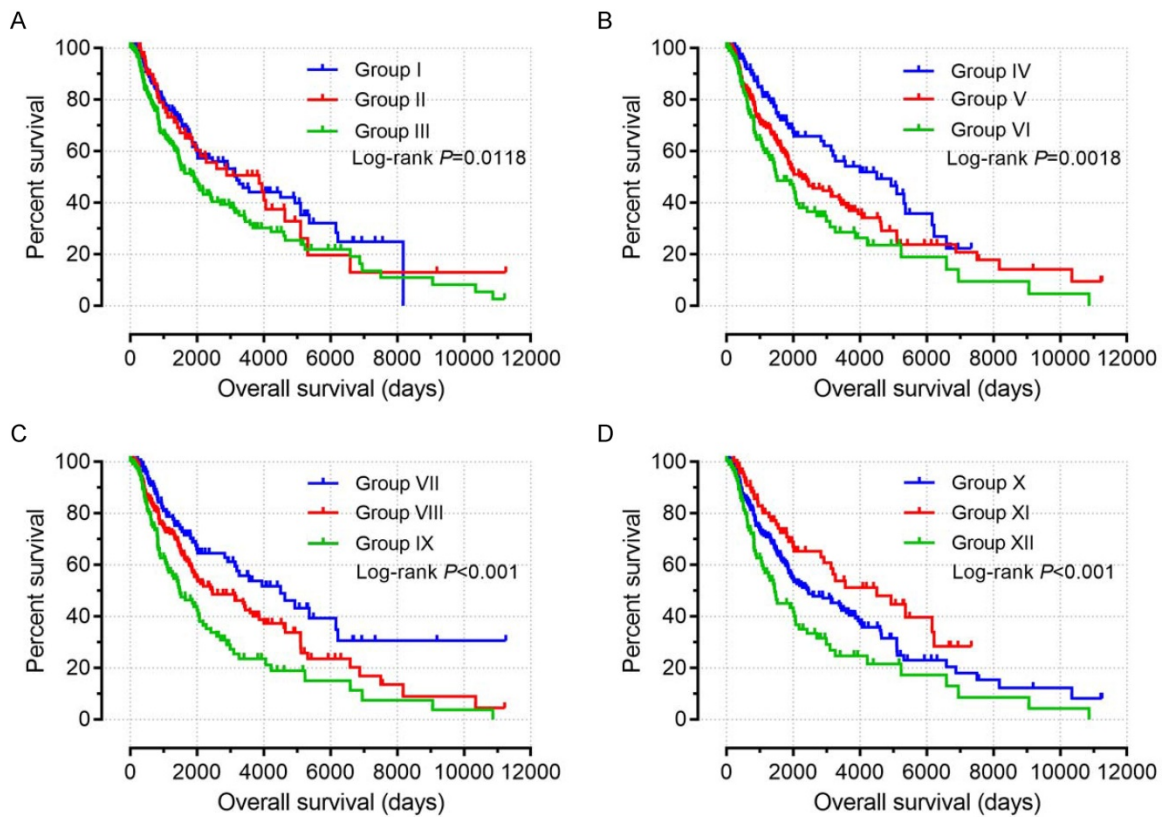


Figure 11. Joint-effects survival analysis of the influence of combined PSME gene expression on OS stratified for PSME1, PSME2 and PSME3 expression levels. (A) PSME1 and PSME2; (B) PSME1 and PSME3; (C) PSME2 and PSME3; (D) PSME1, PSME2 and PSME3. I, high PSME1+high PSME2; III, Low PSME1+low PSME2; IV, high PSME1+ low PSME3; VI, low PSME1+high PSME3; VII, high PSME2+low PSME3; IX, low PSME2+high PSME3; X, high PSME1+high PSME2+low PSME3; XII, Low PSME1+low PSME2+high PSME3. The combinations of genes and unlisted combinations are shown in Table 1. Abbreviation: PSME, proteasome activator subunit.

References

- Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer*. 2015; 136: E359-86.
- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2018. *CA Cancer J Clin*. 2018; 68: 7-30.
- Gershenwald JE, Scolyer RA, Hess KR, Sondak VK, Long GV, Ross MI, et al. Melanoma staging: Evidence-based changes in the American Joint Committee on Cancer eighth edition cancer staging manual. *CA Cancer J Clin*. 2017; 67: 472-92.
- Sanchez-Martin D, Martinez-Torrecuadrada J, Teesalu T, Sugahara KN, Alvarez-Cienfuegos A, Ximenez-Embun P, et al. Proteasome activator complex PA28 identified as an accessible target in prostate cancer by in vivo selection of human antibodies. *Proc Natl Acad Sci U S A*. 2013; 110: 13791-6.
- Zheng DL, Huang QL, Zhou F, Huang QJ, Lin JY, Lin X. PA28beta regulates cell invasion of gastric cancer via modulating the expression of chloride intracellular channel 1. *J Cell Biochem*. 2012; 113: 1537-46.
- Chai F, Liang Y, Bi J, Chen L, Zhang F, Cui Y, et al. High expression of REGgamma is associated with metastasis and poor prognosis of patients with breast cancer. *Int J Clin Exp Pathol*. 2014; 7: 7834-43.
- Shi Y, Luo X, Li P, Tan J, Wang X, Xiang T, et al. miR-7-5p suppresses cell proliferation and induces apoptosis of breast cancer cells mainly by targeting REGgamma. *Cancer Lett*. 2015; 358: 27-36.
- Chai F, Liang Y, Bi J, Chen L, Zhang F, Cui Y, et al. REGgamma regulates ERalpha degradation via ubiquitin-proteasome pathway in breast cancer. *Biochem Biophys Res Commun*. 2015; 456: 534-40.
- Wang X, Tu S, Tan J, Tian T, Ran L, Rodier JF, et al. REG gamma: a potential marker in breast cancer and effect on cell cycle and proliferation of breast cancer cell. *Med Oncol*. 2011; 28: 31-41.
- Chen D, Yang X, Huang L, Chi P. The expression and clinical significance of PA28 gamma in colorectal cancer. *J Investig Med*. 2013; 61: 1192-6.
- Li LP, Cheng WB, Li H, Li W, Yang H, Wen DH, et al. Expression of proteasome activator REGgamma in human laryngeal carcinoma and associations with tumor suppressor proteins. *Asian Pac J Cancer Prev*. 2012; 13: 2699-703.
- Li J, Feng X, Sun C, Zeng X, Xie L, Xu H, et al. Associations between proteasomal activator PA28gamma and outcome of oral squamous cell carcinoma: Evidence from cohort studies and functional analyses. *EBioMedicine*. 2015; 2: 851-8.
- Tang Z, Li C, Kang B, Gao G, Li C, Zhang Z. GEPIA: a web server for cancer and normal gene expression profiling and interactive analyses. *Nucleic Acids Res*. 2017; 45: W98-W102.
- Warde-Farley D, Donaldson SL, Comes O, Zuberi K, Badrawi R, Chao P, et al. The GeneMANIA prediction server: biological network integration for gene prioritization and predicting gene function. *Nucleic Acids Res*. 2010; 38: W214-20.
- Huang DW, Sherman BT, Lempicki RA. Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nature Protocols*. 2009; 4: 44.
- Huang da W, Sherman BT, Lempicki RA. Bioinformatics enrichment tools: paths toward the comprehensive functional analysis of large gene lists. *Nucleic Acids Res*. 2009; 37: 1-13.
- Balachandran VP, Gonen M, Smith JJ, Dematteo RP. Nomograms in Oncology - More than Meets the Eye. *Lancet Oncology*. 2015; 16: e173.
- Subramanian A, Tamayo P, Mootha VK, Mukherjee S, Ebert BL, Gillette MA, et al. Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci U S A*. 2005; 102: 15545-50.
- Nomura N, Nagase T, Miyajima N, Sazuka T, Tanaka A, Sato S, et al. Prediction of the coding sequences of unidentified human genes. II. The coding sequences of 40 new genes (KIAA0041-KIAA0080) deduced by analysis of cDNA clones from human cell line KG-1 (supplement). *DNA Res*. 1994; 1: 251-62.
- Vlahopoulos SA, Cen O, Hengen N, Agan J, Moschovi M, Critselis E, et al. Dynamic aberrant NF-kappaB spurs tumorigenesis: a new model encompassing the microenvironment. *Cytokine Growth Factor Rev*. 2015; 26: 389-403.
- Wellbrock C. MAPK pathway inhibition in melanoma: resistance three ways. *Biochem Soc Trans*. 2014; 42: 727-32.
- Konieczkowski DJ, Johannessen CM, Abudayyeh O, Kim JW, Cooper ZA, Piris A, et al. A melanoma cell state distinction influences sensitivity to MAPK pathway inhibitors. *Cancer Discov*. 2014; 4: 816-27.
- Goessling W, North TE, Loewer S, Lord AM, Lee S, Stoick-Cooper CL, et al. Genetic interaction of PGE2 and Wnt signaling regulates developmental specification of stem cells and regeneration. *Cell*. 2009; 136: 1136-47.
- Anastas JN, Moon RT. WNT signalling pathways as therapeutic targets in cancer. *Nat Rev Cancer*. 2013; 13: 11-26.

25. Zhang D, Lim SG, Koay ES. Proteomic identification of down-regulation of oncoprotein DJ-1 and proteasome activator subunit 1 in hepatitis B virus-infected well-differentiated hepatocellular carcinoma. *Int J Oncol.* 2007; 31: 577-84.
26. Min L, Xu H, Wang J, Qu L, Jiang B, Zeng Y, et al. N-alpha-acetyltransferase 10 protein is a negative regulator of 28S proteasome through interaction with PA28beta. *FEBS Lett.* 2013; 587: 1630-7.
27. Xu J, Zhou L, Ji L, Chen F, Fortmann K, Zhang K, et al. The REGgamma-proteasome forms a regulatory circuit with IkappaBvarepsilon and NFkappaB in experimental colitis. *Nat Commun.* 2016; 7: 10761.
28. Li L, Dang Y, Zhang J, Yan W, Zhai W, Chen H, et al. REGgamma is critical for skin carcinogenesis by modulating the Wnt/beta-catenin pathway. *Nat Commun.* 2015; 6: 6875.
29. Zhang Z, Zhang R. Proteasome activator PA28 gamma regulates p53 by enhancing its MDM2-mediated degradation. *EMBO J.* 2008; 27: 852-64.
30. Ali A, Wang Z, Fu J, Ji L, Liu J, Li L, et al. Differential regulation of the REGgamma-proteasome pathway by p53/TGF-beta signalling and mutant p53 in cancer cells. *Nat Commun.* 2013; 4: 2667.
31. Wang H, Bao W, Jiang F, Che Q, Chen Z, Wang F, et al. Mutant p53 (p53-R248Q) functions as an oncogene in promoting endometrial cancer by up-regulating REGgamma. *Cancer Lett.* 2015; 360: 269-79.
32. Liu J, Yu G, Zhao Y, Zhao D, Wang Y, Wang L, et al. REGgamma modulates p53 activity by regulating its cellular localization. *J Cell Sci.* 2010; 123: 4076-84.
33. Gao G, Wong J, Zhang J, Mao I, Shrivah J, Wu Y, et al. Proteasome activator REGgamma enhances coxsackieviral infection by facilitating p53 degradation. *J Virol.* 2010; 84: 11056-66.
34. Tian M, Xiaoyi W, Xiaotao L, Guosheng R. Proteasomes reactivator REG gamma enhances oncogenicity of MDA-MB-231 cell line via promoting cell proliferation and inhibiting apoptosis. *Cell Mol Biol (Noisy-le-grand).* 2009; 55 Suppl: OL1121-31.