



Draft Genome Sequence of *Stenotrophomonas bentonitica* BII-R7^T, a Selenite-Reducing Bacterium Isolated from Spanish Bentonites

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ABSTRACT The Gram-negative bacterium *Stenotrophomonas bentonitica* BII-R7^T was isolated from bentonite formations. Like other species within the genus *Stenotrophomonas*, strain BII-R7^T possesses high tolerance to numerous heavy metals, suggesting potential for bioremediation purposes. The draft genome sequence reported here comprises 4.37 Mb with a G+C content of 66.5% and 3,796 predicted protein-coding sequences.

Stenotrophomonas bentonitica BII-R7^T (= LMG 29893^T = CECT 9180^T = DSM 103927^T) is a recently described Gram-negative bacterial strain that was isolated from bentonite formations located in southern Spain (1). The genus *Stenotrophomonas* has hitherto comprised 14 established species, isolated from a large variety of environments (2–14), that are resistant to certain antibiotics and metals (15). In this sense, *S. bentonitica* has shown high uranium (1) and selenium (M. A. Ruiz-Fresneda, unpublished data) tolerance due to different interaction mechanisms, suggesting potential applicability for bioremediation purposes. Indeed, the biotechnological use of *Stenotrophomonas* spp. has already been proposed (16–21). Research on all but one of the 14 known *Stenotrophomonas* spp., *S. tumulicola* (13), counts on freely available genome sequences of the corresponding species (22). Here, we report the draft genome sequence of *S. bentonitica* strain BII-R7^T.

After cultivation on LB medium, genomic DNA of *S. bentonitica* BII-R7^T was extracted as described by Martín-Platero et al. (23). A genomic library with an insert size of 350 bp was sequenced using the Illumina HiSeq 2000 platform at Macrogen, Inc. (Seoul, Republic of Korea).

A total of 53,608,108 paired-end 101-bp reads were obtained (>1,000 × coverage). The quality of the reads was assessed using FastQC (24), and the Q20 and Q30 indices were 95.34% and 87.69%, respectively. Multiple *de novo* genome assemblies were performed using ABySS version 1.5.1 (25) with *k*-mer sizes between 19 and 95. The assemblies were merged, filtered, and further assembled into scaffolds using TransABySS (26) and GS *de novo* assembler software (Roche). We obtained 191 scaffolds with an N_{50} of 35,432 and an L_{50} of 38. The mean size of these scaffolds was 22,890 bp with the largest comprising 187,875 bp and the smallest comprising 2,262 bp. The size of the entire sequence was 4,371,992 bp with a 66.5% G+C content, which are values in accordance with described *Stenotrophomonas* spp.

Gene prediction and annotation were performed using the Rapid Annotations using Subsystems Technology server (27) and the Prokaryotic Genome Annotation Pipeline (28). The genomic features of *S. bentonitica* BII-R7^T included a total of 3,786 coding sequences (CDSs), 1 complete rRNA cluster, 44 tRNAs, 4 ncRNAs, and 158 pseudogenes. The coding sequences were classified into 431 subsystems, the most abundant of which were for the metabolism of amino acid derivatives ($n = 352$ CDSs); carbohydrates ($n =$

Received 9 June 2017 Accepted 12 June 2017 Published 3 August 2017

Citation Sánchez-Castro I, Bakkali M, Merroun ML. 2017. Draft genome sequence of *Stenotrophomonas bentonitica* BII-R7^T, a selenite-reducing bacterium isolated from Spanish bentonites. Genome Announc 5: e00719-17. <https://doi.org/10.1128/genomeA.00719-17>.

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214); protein metabolism ($n = 205$); metabolism of cofactors, vitamins, prosthetic groups, and pigments ($n = 204$); membrane transport ($n = 160$); and RNA metabolism ($n = 147$). Additionally, 115 of these coding sequences were related to stress responses, such as osmotic and oxidative stress, cold and heat shock stress, or uptake of selenate and selenite. Genes related to degradation or resistance to a variety of toxic compounds (e.g., ethidium bromide) and heavy metals (e.g., cobalt, zinc, cadmium, tellurium, copper, arsenic, or mercury) were also identified in the present draft genome. Moreover, the draft genome contains specific enzymes, such as alkaline and acid phosphatases or glutathione reductases, which could, respectively, be involved in the high levels of tolerance that *S. bentonitica* BII-R7^T (1, 17) has to uranium and selenium.

Accession number(s). This whole-genome shotgun project has been deposited at GenBank/ENA/DBJ under the accession number [MKCZ00000000](https://www.ncbi.nlm.nih.gov/nuclink/MKCZ00000000). The version described in this paper is the first version, MKCZ01000000.

ACKNOWLEDGMENTS

This study was supported by the ERDF-cofinanced grants CGL2012-36505 and CGL2014-59616-R (Ministerio de Ciencia e Innovación, Spain; 80% funded by FEDER). We also acknowledge funding received from the Euratom research and training program 2014–2018 under grant agreement no. 661880.

REFERENCES

- López-Fernández M, Fernández-Sanfrancisco O, Moreno-García A, Martín-Sánchez I, Sánchez-Castro I, Merroun ML. 2014. Microbial communities in bentonite formations and their interactions with uranium. *Appl Geochem* 49:77–86. <https://doi.org/10.1016/j.apgeochem.2014.06.022>.
- Palleroni NJ, Bradbury JF. 1993. *Stenotrophomonas*, a new bacterial genus for *Xanthomonas maltophilia* (Hugh 1980) Swings et al. 1983. *Int J Syst Bacteriol* 43:606–609. <https://doi.org/10.1099/00207713-43-3-606>.
- Finkmann W, Altendorf K, Stackebrandt E, Lipski A. 2000. Characterization of N₂O-producing *Xanthomonas*-like isolates from biofilters as *Stenotrophomonas nitritireducens* sp. nov., *Luteimonas mephitis* gen. nov., sp. nov. and *Pseudoxanthomonas broegbermensis* gen. nov., sp. nov. *Int J Syst Evol Microbiol* 50:273–282. <https://doi.org/10.1099/00207713-50-1-273>.
- Assih EA, Ouattara AS, Thierry S, Cayol JL, Labat M, Macarie H. 2002. *Stenotrophomonas acidaminiphila* sp. nov., a strictly aerobic bacterium isolated from an upflow anaerobic sludge blanket (UASB) reactor. *Int J Syst Evol Microbiol* 52:559–568. <https://doi.org/10.1099/00207713-52-2-559>.
- Wolf A, Fritze A, Hagemann M, Berg G. 2002. *Stenotrophomonas rhizophila* sp. nov., a novel plant-associated bacterium with antifungal properties. *Int J Syst Evol Microbiol* 52:1937–1944. <https://doi.org/10.1099/00207713-52-6-1937>.
- Yang HC, Im WT, Kang MS, Shin DY, Lee ST. 2006. *Stenotrophomonas koreensis* sp. nov., isolated from compost in South Korea. *Int J Syst Evol Microbiol* 56:81–84. <https://doi.org/10.1099/ijs.0.63826-0>.
- Heylen K, Vanparys B, Peirsegaale F, Lebbe L, De Vos P. 2007. *Stenotrophomonas terrae* sp. nov. and *Stenotrophomonas humi* sp. nov., two nitrate-reducing bacteria isolated from soil. *Int J Syst Evol Microbiol* 57:2056–2061. <https://doi.org/10.1099/ijs.0.65044-0>.
- Kaparullina E, Doronina N, Chistyakova T, Trotsenko Y. 2009. *Stenotrophomonas chelatiphaga* sp. nov., a new aerobic EDTA-degrading bacterium. *Syst Appl Microbiol* 32:157–162. <https://doi.org/10.1016/j.syapm.2008.12.003>.
- Kim HB, Srinivasan S, Sathiyaraj G, Quan LH, Kim SH, Bui TPN, Liang ZQ, Kim YJ, Yang DC. 2010. *Stenotrophomonas ginsengisoli* sp. nov., isolated from a ginseng field. *Int J Syst Evol Microbiol* 60:1522–1526. <https://doi.org/10.1099/ijs.0.014662-0>.
- Yi H, Srinivasan S, Kim MK. 2010. *Stenotrophomonas panacihumi* sp. nov., isolated from soil of a ginseng field. *J Microbiol* 48:30–35. <https://doi.org/10.1007/s12275-010-0006-0>.
- Lee M, Woo SG, Chae M, Shin MC, Jung HM, Ten LN. 2011. *Stenotrophomonas daejeonensis* sp. nov., isolated from sewage. *Int J Syst Evol Microbiol* 61:598–604. <https://doi.org/10.1099/ijs.0.017780-0>.
- Ramos PL, Van Trappen S, Thompson FL, Rocha RC, Barbosa HR, De Vos P, Moreira-Filho CA. 2011. Screening for endophytic nitrogen-fixing bacteria in Brazilian sugar cane varieties used in organic farming and description of *Stenotrophomonas pavanii* sp. nov. *Int J Syst Evol Microbiol* 61:926–931. <https://doi.org/10.1099/ijs.0.019372-0>.
- Handa Y, Tazato N, Nagatsuka Y, Koide T, Kigawa R, Sano C, Sugiyama J. 2016. *Stenotrophomonas tumulicola* sp. nov., a major contaminant of the stone chamber interior in the Takamatsuzuka Tumulus. *Int J Syst Evol Microbiol* 66:1119–1124. <https://doi.org/10.1099/ijsem.0.000843>.
- Sánchez-Castro I, Ruiz-Fresneda MA, Bakkali M, Kämpfer P, Glaeser SP, Busse H-J, López-Fernández M, Martínez-Rodríguez P, Merroun ML. *Stenotrophomonas bentonitica* sp. nov., isolated from bentonite formations. *Int J Syst Evol Microbiol*, in press.
- Ryan RP, Monchy S, Cardinale M, Taghavi S, Crossman L, Avison MB, Berg G, van der Lelie D, Dow JM. 2009. The versatility and adaptation of bacteria from the genus *Stenotrophomonas*. *Nat Rev Microbiol* 7:514–525. <https://doi.org/10.1038/nrmicro2163>.
- Binks PR, Nicklin S, Bruce NC. 1995. Degradation of hexahydro-1,3,5-trinitro-1,3,5-triazine (RDX) by *Stenotrophomonas maltophilia* PB1. *Appl Environ Microbiol* 61:1318–1322.
- Merroun ML, Selenska-Pobell S. 2008. Bacterial interactions with uranium: an environmental perspective. *J Contam Hydrol* 102:285–295. <https://doi.org/10.1016/j.jconhyd.2008.09.019>.
- Song HP, Li XG, Sun JS, Xu SM, Han X. 2008. Application of a magnetotactic bacterium, *Stenotrophomonas* sp. to the removal of Au(III) from contaminated wastewater with a magnetic separator. *Chemosphere* 72:616–621. <https://doi.org/10.1016/j.chemosphere.2008.02.064>.
- Morel MA, Ubalde MC, Olivera-Bravo S, Callejas C, Gill PR, Castro-Sowinski S. 2009. Cellular and biochemical response to Cr(VI) in *Stenotrophomonas* sp. *FEMS Microbiol Lett* 291:162–168. <https://doi.org/10.1111/j.1574-6968.2008.01444.x>.
- Ghosh A, Das Saha PD. 2013. Optimization of copper bioremediation by *Stenotrophomonas maltophilia* PD2. *J Environ Chem Eng* 1:159–163. <https://doi.org/10.1016/j.jece.2013.04.012>.
- Ge S, Ge SC. 2016. Simultaneous Cr(VI) reduction and Zn(II) biosorption by *Stenotrophomonas* sp. and constitutive expression of related genes. *Biotechnol Lett* 38:877–884. <https://doi.org/10.1007/s10529-016-2057-8>.
- Patil PP, Midha S, Kumar S, Patil PB. 2016. Genome sequence of type strains of genus *Stenotrophomonas*. *Front Microbiol* 7:309. <https://doi.org/10.3389/fmicb.2016.00309>.
- Martín-Platero AM, Valdivia E, Maqueda M, Martínez-Bueno M. 2007. Fast, convenient and economical method for isolating genomic DNA from lactic-acid bacteria using a modification of the “protein salting-out”

- procedure. *Anal Biochem* 366:102–104. <https://doi.org/10.1016/j.ab.2007.03.010>.
24. Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. <http://www.bioinformatics.babraham.ac.uk/projects/fastqc>.
25. Simpson JT, Wong K, Jackman SD, Schein JE, Jones SJ, Birol I. 2009. ABySS: a parallel assembler for short read sequence data. *Genome Res* 19:1117–1123. <https://doi.org/10.1101/gr.089532.108>.
26. Robertson G, Schein J, Chiu R, Corbett R, Field M, Jackman SD, Mungall K, Lee S, Okada HM, Qian JQ, Griffith M, Raymond A, Thiessen N, Cezard T, Butterfield YS, Newsome R, Chan SK, She R, Varhol R, Kamoh B, Prabhu AL, Tam A, Zhao Y, Moore RA, Hirst M, Marra MA, Jones SJ, Hoodless PA, Birol I. 2010. *De novo* assembly and analysis of RNA-seq data. *Nat Methods* 7:909–912. <https://doi.org/10.1038/nmeth.1517>.
27. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.
28. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. *Nucleic Acids Res* 44: 6614–6624. <https://doi.org/10.1093/nar/gkw569>.