



Niacin in the Central Nervous System: An Update of Biological Aspects and Clinical Applications

Valeria Gasperi *,[†], Matteo Sibilano [†], Isabella Savini and Maria Valeria Catani *

Department of Experimental Medicine, Tor Vergata University of Rome, Via Montpellier 1, 00133 Rome, Italy; matteosibilano@libero.it (M.S.); savini@uniroma2.it (I.S.)

* Correspondence: gasperi@med.uniroma2.it (V.G.); catani@uniroma2.it (M.V.C.); Tel.: +39-06-72596465 (V.G. & M.V.C.)

+ These authors contributed equally to this work.

Received: 30 January 2019; Accepted: 20 February 2019; Published: 23 February 2019



Abstract: Niacin (also known as "vitamin B₃" or "vitamin PP") includes two vitamers (nicotinic acid and nicotinamide) giving rise to the coenzymatic forms nicotinamide adenine dinucleotide (NAD) and nicotinamide adenine dinucleotide phosphate (NADP). The two coenzymes are required for oxidative reactions crucial for energy production, but they are also substrates for enzymes involved in non-redox signaling pathways, thus regulating biological functions, including gene expression, cell cycle progression, DNA repair and cell death. In the central nervous system, vitamin B₃ has long been recognized as a key mediator of neuronal development and survival. Here, we will overview available literature data on the neuroprotective role of niacin and its derivatives, especially focusing especially on its involvement in neurodegenerative diseases (Alzheimer's, Parkinson's, and Huntington's diseases), as well as in other neuropathological conditions (ischemic and traumatic injuries, headache and psychiatric disorders).

Keywords: central nervous system; diet; NAD(P); neurodegenerative diseases; niacin; nicotinamide; nicotinic acid; vitamin B₃

1. Introduction

Niacin (also known as "vitamin B_3 " or "vitamin PP") is the generic descriptor for two vitamers, nicotinic acid (pyridine-3-carboxylic acid) and nicotinamide (nicotinic acid amide), that give rise to the biologically active coenzymes, nicotinamide adenine dinucleotide (NAD) and its phosphate analog, the nicotinamide adenine dinucleotide phosphate (NADP) [1] (Figure 1). The two coenzymes take part in redox reactions crucial for energy production: in particular, the pyridinic ring can accept and donate a hydride ion (:H⁻, the equivalent of a proton and two electrons), thus acting as an electron carrier. Nonetheless, NAD and NADP play different metabolic roles in the cytosol: the NADH/NAD⁺ ratio is small (about 8×10^{-4}), thus favoring oxidative catabolism, whereas the NADPH/NADP⁺ ratio is higher (about 75), thus providing a strongly reducing environment for biosynthetic reactions [2,3].

Maintenance of the intracellular NAD pool is not only important to fuel redox metabolism, but also to support NAD-dependent, non-redox signaling pathways. NAD is indeed a substrate of ADP-ribosyltransferases that catalyze ADP-ribose transfer reactions, thus breaking down NAD to nicotinamide and ADP-ribosyl products, which play a key role in cellular signaling cascades regulating gene expression, cell cycle progression, insulin secretion, DNA repair, apoptosis and aging [4–6]. Finally, NAD has also been recognized as an endogenous agonist of purinergic P2Y1 and P2Y11 membrane subtype receptors, through which it inhibits neurotransmission in visceral smooth muscles [7] and activates immune cells [8,9], respectively.



Figure 1. Chemical structures of niacin vitamers (**A**) and active coenzymatic forms (**B**). NAD: nicotinamide adenin dinucleotide. NADP: nicotinamide adenin dinucleotide phosphate.

2. Niacin Sources

Humans obtain niacin from both endogenous and exogenous sources. Only 2% of dietary tryptophan (Trp) is converted into niacin via a multistep pathway (see in next sections), occurring mainly in the liver [10]. Diet provides the vitamin as nicotinic acid, nicotinamide and Trp, as well as the active coenzymatic forms of niacin.

2.1. Exogenous Sources

Niacin is found in animal and vegetable foods. In meat and fish, the vitamin is present as NAD(P), whose amounts are higher in unprepared foods compared to processed foods (enzymatic hydrolysis of the coenzymes can occur during food preparation).

In mature cereal grains (particularly in corn), niacin is largely present as niacin-glycoside and, in a minor proportion, peptide-bound niacin, compounds collectively termed "niacinogens" [11]. When complexed in niacinogens, niacin is poorly available (only ~ 30%), as intestinal enzymes are not able to free niacin; nonetheless, alkali treatment of the grain increases niacin bioavailability [11].

Once ingested, free niacin can be adsorbed in the stomach, although the small intestine absorbs it faster. The mechanism of transport across the enterocyte brush border membrane is not fully clarified yet. Several transporters, indeed, appear to be involved in intestinal niacin uptake; among them, the most common are the human organic anion transporter-10 (hOAT-10, a proton-driven carrier that also mediates the transport of urate and *p*-aminohippurate) [12], responsible for niacin uptake at physiological concentrations [13], and the sodium-coupled monocarboxylate transporter (SMCT1 or SLC5A8, a transporter for lactate, pyruvate and short-chain fatty acids), specifically active at high pharmacological doses of nicotinic acid [14,15].

NAD and NADP are quickly hydrolyzed, by intestinal mucosa and liver glycohydrolases, to nicotinamide that is subsequently transported to tissues, where it is converted into coenzymatic forms as necessary. It seems noteworthy that nicotinamide moves freely into or out of the brain [16] and, as discussed in the next sections, such a property has important neurobiological implications.

2.2. Endogenous Synthesis

Starting from dietary Trp, niacin is synthesized via the kynurenine pathway (KP) (Figure 2), occurring mainly in the liver and, to a lesser extent, in extrahepatic tissues (especially upon immune cell activation) [17–19].



Figure 2. De novo synthesis of NAD(P) from tryptophan, nicotinamide and nicotinic acid. (1) Two iron porphyrin metalloproteins, tryptophan 2,3 dioxygenase (TDO, in the liver) and indolamine-pyrrole 2-3 dioxygenase (IDO, in extrahepatic tissues), oxidize the pyrrole moiety of Tryptophan (Trp), thus forming N-L-formylkynurenine. (2) Arylformamidase (AFMID) hydrolytically removes the formyl group producing kynurenine and is then (3) hydroxylated to 3-hydroxykynurenine by kynurenine-3 monooxygenase (KMO), a mitochondrial flavo-enzyme that uses O₂ as a substrate and NADPH as a cofactor. The action of (4) kynureninase B (KYNU, a vitamin B_6 -dependent enzyme) and (5) 3-hydroxyanthranilic dioxygenase (HAAO, a nonheme iron-dependent dioxygenase) leads to production of 2-amino-3-carboxymuconic-6-semialdehyde acid, an unstable product that (6) spontaneously condensates and rearranges to form quinolinic acid; then, (7) quinolinic acid is decarboxylated and converted to nicotinic acid mononucleotide by quinolinic acid phosphoribosyltransferase (QPRT). Nicotinic acid mononucleotide is also produced through the "salvage pathway", via the action of (8) nicotinic acid phosphoribosyltransferase (NPRT). The subsequent action of (9) nicotinamide/nicotinic acid-mononucleotide-adenylyltransferases (NMNAT1-3) and (10) NAD synthetase (NADSYN1) leads to the generation of NAD, which is then (11) phosphorylated to produce NADP. NAD can also derive directly from nicotinamide through the action of (12) nicotinamide phosphoribosyltransferase (NAMPT) and (13) nicotinamide/nicotinic acid-mononucleotide-adenylyltransferase (NMNAT1-3). Red frames: dietary precursors of NAD(P). Ala: alanine; Gln: glutamine; Glu: glutamate; PLP pyridoxal phosphate; PRPP: 5-phosphoribosyl-1- pyrophosphate.

Tryptophan 2,3 dioxygenase (TDO), catalyzing the first reaction, is the rate-limiting enzyme. Several nutritional, hormonal and physio-pathological factors affect the efficiency of this anabolic pathway. Deficiencies of vitamin B_6 , riboflavin, iron and heme (all essential cofactors for specific enzymes), as well as of vitamin B_1 and Trp itself, slow the reaction rate [18,20]. Overall: (i) a protein-enriched diet (particularly, consumption of foods with high concentrations of leucine, such as maize or sorghum) decreases niacin biosynthesis; (ii) unsaturated fatty acid-enriched diet increases it, while saturated fatty acids do not exert any effect; (iii) the transformation ratio is higher in diets containing starch with respect to sucrose-rich diets; (iv) caloric restriction drastically suppresses biosynthesis [18,21–26].

Among hormones, estrogens, glucorticoids and thyroxine are the best characterized modulators of the KP. Estrogens enhance TDO activity; enzyme activity is triplicated in women who are pregnant or are taking oral contraceptives [27,28]. Glucocorticoids stimulate de novo synthesis, by inducing TDO via a mechanism potentiated by glucagon and inhibited by insulin and adrenaline [18,29,30]. The effects of thyroxine on TDO activity are still controversial, as some studies suggested a positive action, while others did not observe any effect [31–34].

Due to individual differences, it has been estimated that, in human healthy individuals, Trp is converted to niacin with an average conversion efficiency of 60:1 [35]. Therefore, niacin intakes are expressed as niacin equivalents (NE; 1 mg NE = 1 mg niacin or 60 mg Trp): Recommended Dietary Allowance for adults is 16 mg NE/day for men and 14 mg NE/day for women, with a Tolerable Upper Intake Level of 35 mg/day, based on flushing as the critical adverse effect [36].

3. Vitamin Catabolism

The tight intracellular regulation of NAD is guaranteed not only at biosynthetic but also at catabolic level; in the latter case, NAD can be either recycled or metabolized and eliminated via urine (Figure 3) [37–39].



Figure 3. Schematic representation of distinct catabolic pathways. (1) NAD is hydrolyzed onto nicotinamide mononucleotide via the action of specific pyrophosphatases belonging to Nudix (nucleoside diphosphate linked to moiety X) family. (2) Nicotinamide mononucleotide is then dephosphorylated by Isn1 and Sdt1 cytosolic nucleotidases, which release the corresponding riboside cleaved to nicotinamide by a purine nucleoside phosphorylase (PNP) (3). Alternatively, NAD becomes a substrate of sirtuins (4), ADP-ribosyltransferases (ARTC) (5) and diphtheria toxin-like ADP-ribosyltransferases (ARTD) (6). Nicotinamide can be either re-converted to NAD by specific enzymes (7) (see also Figure 2) or methylated by nicotinamide-*N*-methyl transferase (NNMT) to N^1 -methylnicotinamide (8) that, in turn, (9) is oxidized to N^1 -methyl-4-pyridone-3-carboxamide (4-Py) and N^1 -methyl-2-pyridone-5-carboxamide (2-Py) by aldehyde oxidases. 2-OAADPr: O-acetyl-ADP ribose; NAMPT: nicotinamide phosphoribosyltransferase; SAH: *S*-adenosylhomocysteine; SAM: *S*-adenosyl-methionine.

In the recycling pathways, NAD is metabolized to nicotinamide through the action of different ADP-ribosyltransferases. Sirtuins (SIRT) are NAD-dependent deacetylases and mono-ADP-ribosyl transferases belonging to the highly conserved family of silent information regulator-2 like proteins [40–42]. During deacetylation, NAD is hydrolyzed and the ε -acetyl lysine residues of the target protein is transferred onto the ADP-ribose moiety, thus forming *O*-acetyl-ADP ribose (Figure 3), which is a ligand of calcium channels in the plasma membrane [43]. SIRTs deacetylate a broad spectrum of proteins, thus modulating their activity, stability or localization. Depending on the targeted protein, these enzymes affect several biological processes, including transcription, cell cycle progression, genome stability, cell death and mitochondrial biogenesis [42,44,45]. ADP-ribosyltransferases (ARTC) and diphtheria toxin-like ADP-ribosyltransferases (ARTD) catalyze mono- and poly-ADP-ribosylation, respectively, of specific amino acids (arginine, cysteine, asparagine, histidine) of membrane proteins (Figure 3), thus regulating innate immunity and cell-to-cell cross-talk, as well as cell cycle, cell death and energy metabolism [46–48].

Finally, NAD(P) can be hydrolyzed to nicotinamide by two ADP-ribose cyclases, namely CD38 and CD157, which also release cyclic ADP-ribose, an endogenous activator of ryanodine receptor-mediated calcium release [49–52] and suggested to be involved in pathological diseases such as cancer, neurodegeneration and autoimmune diseases [53–56].

If not recycled, nicotinamide is methylated, by the cytosolic nicotinamide *N*-methyltransferase (NNMT) that uses *S*-adenosyl-methionine (SAM) as a methyl donor, and eliminated as oxidized metabolites (Figure 3). Altered enzyme activity has been linked to several pathological conditions, including neurodegenerative diseases, obesity, type 2 diabetes and cancer [57–64]. It should be recalled that, beside nicotinamide by-products, also those deriving from conjugation of nicotinic acid to glycine (nicotinuric acid) or from its methylation (1-methylnicotinic acid) can be found in urine [65–67].

Due to the multiplicity of NAD-dependent biological events, which lead to NAD degradation, cells need to replenish their intracellular NAD(P) pools; inhibition of NAD biosynthesis, for example, decreases intracellular NAD content within a few hours [68].

4. Severe Vitamin Deficiency

Severe niacin and/or Trp deficiency leads to a variety of clinical symptoms, including diarrhea, dermatitis and dementia, collectively known as "pellagra" or "the three D disease" [69]; although this disease has become rare in developed countries, it remains endemic in underdeveloped countries [70]. Pellagra is common in people who mostly eat maize [71], as well as in malnourished and alcoholic men [26]; other risk factors leading to vitamin B₃ deficiency are nervous anorexia [72], AIDS [73], cancer [74] and chemotherapy [75], as well as malabsorptive disorders, such as Crohn's disease [76].

Light sensitivity is high: dermatitis derives from deficits in poly(ADP-ribose) polymerase activity that leads to impaired DNA repair. Patients can show psychiatric symptoms (i.e., depression, paranoid behaviors, suicide and aggressive tendencies) that disappear when they take niacin [77,78]; some of these symptoms are also related to deficit of serotonin that derives from Trp [78].

5. Pharmacological Effects of Niacin

When supplemented at physiological amounts, nicotinic acid (15–20 mg/day) and nicotinammide (300 mg/day) are effective in treating traditional pellagra [77,78]; nonetheless, at higher concentrations, they display separate additional pharmacological activities, ranging from anti-dyslipidemic to anti-inflammatory action. The first evidence of lipid-altering effects of niacin dates back to 1955, when Altschul and co-workers reported the ability of 3000 mg/day nicotinic acid (but not nicotinamide) to reduce serum cholesterol in humans [79]. An every growing body of experimental data points to beneficial effects of nicotinic acid as an anti-hyperlipidemic agent. It is now well established that nicotinic acid efficaciously: (i) inhibits free fatty acid mobilization and lipolysis; (ii) reduces hepatic triglyceride synthesis and very low density lipoprotein (VLDL) secretion; (iii) inhibits VLDL conversion into low density lipoprotein (LDL); (iv) increases serum high-density-lipoprotein (HDL) levels;

(v) triggers LDL conversion from small, dense particles to large, low density particles, (vi) reduces serum lipoprotein concentrations; and (vii) increases apolipoprotein A1 [80,81].

To date, the underlying mechanisms are still speculative; in particular, nicotinic acid (at levels higher than those achieved with diet) has been reported to bind to and activate GPR109A and GPR109B, two G_0/G_i -coupled membrane receptors highly expressed in adipose tissue; nonetheless, these receptors are absent, or present only at low levels, in the liver [82]. Therefore, it is conceivable that nicotinic acid might exert its lowering-lipid effects through receptor-independent and -dependent mechanisms.

Due to the above mentioned positive effects, in 2008, nicotinic acid was commercially available as Trevaclyn[®], Tredaptive[®] or Pelzont[®], at the dose of 1.0 g (in combination with laropipram, an anti-flushing agent); this prescription product has been used to treat mixed dyslipidemic and/or primary hypercholesterolemic adults receiving statins [83]. However, results from the Atherothrombosis Intervention in Metabolic Syndrome with Low HDL/High Triglycerides: Impact on Global Health Outcomes (AIM-HIGH) trial [84], together with the Heart Protection Study 2-Treatment of HDL to Reduce the Incidence of Vascular Events (HPS2-THRIVE) trial [85,86], reported no clinical benefits (i.e., reduced risk of heart attack and stroke) from the long-lasting usage of niacin. A lack of efficacy, together with the onset of recurrent serious side effects (gastrointestinal, musculoskeletal, and skin-related), has led to drug withdrawal from the EU market.

In vitro and in vivo studies have also demonstrated that nicotinic acid (or activation of its molecular targets) exerts significant anti-inflammatory, anti-oxidant and anti-apoptotic activities in a variety of cells and tissues [87], thus being potentially beneficial for the management of several pathological conditions, including type-2 diabetes [88,89], obesity [90,91], atherosclerosis [92], kidney and lung injury [93–95], and hyperalgesia [96].

Also nicotinamide at high doses can exert specific pharmacological activities, particularly those related to cancer management. Indeed, several experimental and clinical studies have shown the ability of nicotinamide to sensitize tumors to radiation or chemotherapy [97–100]. Such an activity depends on activation of poly(ADP-ribose)-dependent apoptosis cascade, as well as on inhibition of myosin light chain kinase that, in turn, enhances microvascular flow, thus improving drug delivery and tumor oxygenation [97–100].

6. Niacin in the Central Nervous System

Besides dermatitis and diarrhea, niacin/tryptophan deficit symptoms also include several nervous system pathologies, such as dementia and depression, as well as other symptoms resembling those observed in neurodegenerative diseases. This evidence, together with accumulating in vitro and in vivo studies, has underlined the importance of niacin (particularly of nicotinamide) in growth and maintenance of the central nervous system (CNS) [101,102].

Nicotinamide biosynthesis actively occurs in the mammalian brain, which contains nanomolar-low micromolar concentrations of nicotinamide precursors derived from the KP [103–105]. Among them, quinolinic acid (unevenly present in different brain regions and, unlike nicotinamide, unable to cross the blood-brain barrier) displays evident neuroactivity [106]: it acts as a *N*-methyl-D-aspartate (NMDA) receptor agonist, thus causing excitotoxic neuronal lesions and oxidative stress [107]. In addition, quinolinic acid concentrations in the brain (particularly in the cortex) positively correlate with age, thus contributing to neuron synapsis dropout occurring during aging [108]. Finally, neuroinflammation, neurodegeneration and mood disturbs are accompanied by increased quinolinic acid levels in plasma and/or cerebrospinal fluid [10,109,110].

Among KP enzymes, TDO activity is rather low in a healthy human healthy brain [111], where it controls neurogenesis with implications in pre- and post-natal development, as well as in anxiety-related behavior [112]. TDO activity is enhanced under pathological conditions: high activity, indeed, has been found in neurodegenerative diseases and during tumor progression [113,114]. Also indolamine-pyrrole 2-3 dioxygenase (IDO) is expressed in the brain and its activity is increased upon pathological conditions, especially in depression, aging and neuroinflammatory diseases [115–117].

Like other vitamins (ascorbic acid, calcitriol and retinoic acid) [118–122], nicotinamide affects neurogenesis by accelerating differentiation of embryonic stem cells or neural progenitors into post-mitotic neurons [123,124]. In vitro vitamin supplementation promotes progression of undifferentiated stem cells to neural progenitors, which further mature into efficient GABAergic neurons; the pro-inducing action is time-dependent as the effects are more pronounced when the vitamin is early received early (day 0) [124]. Accordingly, decreased activity of NNMT (and, therefore, low levels of its metabolic product, N^1 -methylnicotinamide) is required for regulating pluripotency in stem cells: accumulation of NNMT's substrates SAM and nicotinamide, indeed, promotes naïve to primed stem cell transition, by making SAM available for histone methylation and regulation of epigenetic events that control the metabolic changes occurring in early human development [125].

Beside the pro-differentiating action, nicotinamide also promotes neuronal survival, especially during oxidative stress conditions, and this effect is achieved via multiple mechanisms, including: (i) prevention of cytochrome c release and caspase 3- and 9-like activities, (ii) inhibition of caspase-3-mediated degradation of forkhead transcription factor (FOXO3a) and (iii) maintenance of protein kinase B (Akt)-dependent phosphorylation of FOXO3a [126].

CNS vascular integrity also positively correlates with NAD levels in brain, where a fine-tuned control of its metabolism occurs. As an example, heterozygous deletion of nicotinamide phosphoribosyltransferase (NAMPT) in the brain exacerbates focal ischemic stroke-induced neuronal death and brain damage [127], while its selective knock down in projection neurons of adult mice leads to motor dysfunction, neurodegeneration and death [128].

Finally, alterations of NAD metabolism are key features of Wallerian degeneration, a process occurring in crushed nerve fibers and leading to degeneration of the axon distal to the injury, representing an early event of age-related neurodegenerative disorders, as well as of chemotherapy-induced peripheral neuropathy [129]. By inducing intra-axonal Ca²⁺ increase through a pathway requiring the action of the pro-axon death protein SARM1, accumulation of nicotinamide mononucleotide is, indeed, responsible for loss of axonal integrity [130]. The pro-degenerative action of nicotinamide mononucleotide has also been documented during vincristine-induced degeneration in dorsal root ganglion axons [131]. Accordingly, increased activity of nicotinamide/nicotinic acid-mononucleotide-adenylyltransferase (NMNAT) 1–3 protects axons from degeneration, by either limiting nicotinamide mononucleotide levels or activating SIRT1 [132,133].

7. Alzheimer's Disease

Alzheimer's disease (AD) is a neurodegenerative disease affecting about 30 million people worldwide, whose main hallmarks are the presence of amyloid β (A β) plaques and neurofibrillary tangles [134].

Even if tryptophan/niacin deficiency leads to neurological symptoms that cause neurodegenerative decline [135–137], a cause-effect relationship between niacin and AD pathogenesis has not been established (Table 1).

Dietary niacin may protect against AD and age-related cognitive decline, as suggested by a prospective population-based study: the Chicago Health and Aging Project (CHAP) study, considering a geographically defined community of 6158 residents aged 65 years and older, found an inverse association between AD and niacin intakes, after correction for several dietary (antioxidant nutrients, fats, folate, and vitamins B_6 , B_{12} , B_1 and B_2) and non-dietary (age, education, race, ApoE ϵ 4) risk factors for dementia [135].

	Effector	Main Findings	Ref.				
	Niacin	Inverse association between AD and dietary niacin intakes	[135]				
	NAD ⁺	High brain levels restore mitochondrial function and antagonize cognitive decline	[138,139]				
-	Nam/Nam mononucleotide	Protect against Aβ-induced neurotoxicity via reduction of <i>APP</i> and <i>PSEN-1</i> expression and ROS levels	[140,141]				
Alzheimer's disease	Nam riboside	Reduces DNA damage, neuroinflammation and cell death of hippocampal neurons					
	SIRT1	Supports the non-amyloidogenic pathway of AD Lessens AD neuroinflammation, oxidative stress and mitochondrial dysfunction	[143] [144,145]				
	NMNAT1-3	Protects against axon degeneration via reduction of nicotinamide mononucleotide levels and SIRT1 activation	[132,133]				
-	NMNAT2	Activity downregulated prior to neurodegeneration; restoration of activity is neuroprotective against tauopathy Low gene expression in AD patients	[146] [147]				
	Niacin	Increased intake enhances striatal dopamine synthesis and restores optimal NAD ⁺ /NADH ratio High levels sequester transition metal ions Low doses impact macrophage polarization from M1 (pro-inflammatory) to M2 (anti-inflammatory) profile	[148] [149,150] [151]				
Daultin con/c	NAD ⁺	Decreased levels in PD patients	[148]				
Parkinson's disease	NADPH	Inhibits MPTP ⁺ -induced oxidative stress and glia-mediated neuroinflammation	[152]				
	NNMT	High levels in the cerebrospinal fluid and midbrain dopamine neurons of PD patients High activity associated with low activity of mitochondrial complex 1; it counteracts the MPP ⁺ -dependent toxicity on mitochondrial complex 1 and activates neuronal autophagy Induces neurite branching, synaptophysin expression and dopamine release	[153,154] [154,155] [156]				
Huntington's disease	NAD	Low levels correlate with disease progression in Drosophila HD model	[157]				
	Nam	Protects against the toxicity of polyQ proteins in <i>Drosophila</i> HD models Restores BDNF protein levels, increases acetylated PGC-1α, improves motor deficits Prevents motor abnormality via PARP-1-dependent inhibition of neuronal death and oxidative stress	[158] [159] [160–162]				
	SIRT1	Rescues neurons from mutant huntingtin toxicity Ameliorates pathological mechanisms underlying disease onset	[163,164]				

. 1 1	-		•	<i>c</i> •	1.		.1	1	6		•			1			
lahle		- 1//	am	tine	linge	on	the	role	Ot.	n120	cin.	1n	neuroc	loge	mora	t10	m
lavic		1.4.1	am	IIII	Jungo	on	unc	TOIC	O1	Ina	ciri		neuroc	iczi	, i i c i a	uu	л

AD: Alzheimer's disease; APP: amyloid precursor protein; BDNF: brain-derived neurotrophic factor; HD: Huntington's disease; MPTP⁺: *N*-methy-l-4-phenylpyridinium; Nam: nicotinamide; NMNAT: nicotinamide/nicotinic acid-mononucleotide-adenylyltransferases; NNMT: Nicotinamide *N*-Methyltransferase; PARP-1: poly(ADP-ribose) polymerase-1; PD: Parkinson's disease; PGC-1 α : peroxisome proliferator-activated receptor gamma coactivator 1 α ; PSEN-1: presenilin-1; ROS: reactive oxygen species; SIRT1: sirtuin1.

Although the existing epidemiologic evidence remains limited and inconclusive, niacin (especially nicotinamide) may be relevant for AD, especially keeping in mind that, by mediating key biological processes (such as energy metabolism, mitochondrial functions, calcium homeostasis, survival and cell death), NAD has lifespan-extending effects; this is particularly important in brain functions, including neurotransmission, learning and memory. NAD⁺ depletion and mitochondrial dysfunction, fundamental for synaptic plasticity, have usually been found in aging and AD onset [138,165]; accordingly, in mice models of AD, increasing NAD⁺ brain concentrations can restore mitochondrial function and antagonize cognitive decline [138,139]. Nicotinamide and/or nicotinamide mononucleotide also counteract amyloid toxicity, by reducing expression of AD-related genes (amyloid precursor protein and presenilin 1) and reactive oxygen species (ROS) generation, and by improving neuron survival: both in vitro (organotypic hippocampal slice cultures) and in vivo (AD model rats) studies have indeed underlined the protective effects of vitamin B₃ against Aβ-induced neurotoxicity [140,141]. Moreover, the vitamin is able to lessen phosphorylated-Tau

pathology in a novel AD mouse model with introduced DNA repair deficiency: nicotinamide riboside treatment significantly reduces DNA damage, neuroinflammation and cell death of hippocampal neurons, thus suggesting a therapeutic potential of NAD⁺ supplementation for AD [142]. Accordingly, the expression of *Nmnat2*, encoding for the enzyme catalyzing the conversion of nicotinamide to NAD⁺, is downregulated prior to neurodegeneration in a mouse model of dementia, and restoration of enzymatic activity has been shown to be neuroprotective against tauopathy [146]. Low levels of *Nmnat2* have also been found in AD patients and its enzymatic activity is related to clearance of tau protein [147].

Lastly, fluctuations in NAD⁺ availability can reduce AD pathology, also by modulating SIRT1 activity and slowing aging and age-associated diseases [166,167]. Several studies have underlined the key role of SIRTs in AD prevention: in particular, deacetylase activity of SIRT1 has been shown to support the non-amyloidogenic pathway of AD [143], and to counteract phenomena, like neuroinflammation, oxidative stress and mitochondrial dysfunction, contributing to, and aggravating, AD [144,145].

8. Parkinson's Disease

Parkinson's disease (PD) is a progressive disorder characterized by degeneration of dopaminergic neurons within the substantia nigra, whose main hallmarks are abnormal aggregation of the α -synuclein protein, inhibition of mitochondrial respiratory complex 1, oxidative stress and neuroinflammation. Because only 5–10% of PD cases can be ascribed to genetic predisposition, several environmental factors may play a role in sporadic forms of PD [149]. Among them, vitamin B₃ is a promising preventive and therapeutic factor (Table 1), as it can alleviate certain types of early-onset PD symptoms. NAD⁺ levels, indeed, fall in patients with PD and, conversely, increasing niacin intake can increase dopamine synthesis in the striatum and restore optimal NAD⁺/NADH ratio needed for the activity of mitochondrial complex 1 [148]. High niacin levels can also sequester transition metal ions (including iron) that usually accumulate together with the occurrence of aggregated misfolded proteins [149,150]. Furthermore, optimal levels of vitamin B₃ are needed for reducing oxidative stress and neuroinflammation, also implicated in PD pathogenesis: low doses of niacin alter macrophage polarization from M1 (pro-inflammatory) to M2 (anti-inflammatory) phenothype, while exogenous NADPH suppresses oxidative stress and glia-mediated neuroinflammation [151,152].

Neurons are the only cells of the brain expressing NNMT that seems to play an important role in sustaining neuron homeostasis [153]. Despite numerous investigations, the exact cause-effect relationship between NNMT and PD neuropathogenesis remains unclear. Some authors refer to NNMT as a risk factor for PD, since its levels are elevated in the cerebrospinal fluid and midbrain dopamine neurons of PD patients [153,154]. High NNMT activity is associated with low activity of mitochondrial complex 1, thus providing a link with mitochondrial dysfunction triggering neurodegeneration [154,155]. It is noteworthy that N^1 -methylnicotinamide (the metabolite generated by the action of NNMT) is structurally similar to N-methy-l-4-phenylpyridinium (MPP⁺), a toxin damaging dopamine neurons [168]. Conversely, other studies have demonstrated that the enzyme is able to (i) counteract the MPP+-mediated toxicity on mitochondrial complex 1, (ii) activate neuronal autophagy for balancing energy sources and cell homeostasis, and (iii) modulate neuron morphology and differentiation, by inducing neurite branching, synaptophysin expression and dopamine accumulation and release [156]. Likewise, NAD supplementation or inactivation of NAD-consuming enzymes [like PARP-1, a poly(ADP-ribose) polymerase involved in DNA repair] rescue mitochondrial defects and protect neurons against degeneration, in familial forms of PD characterized by mutations in the *pink1* gene; this finding suggests that neurotoxicity associated with mitochondrial defects may be prevented by modulating NAD⁺ salvage metabolism in order to enhance NAD availability [169].

Huntington's disease (HD) is an autosomal dominant neurodegenerative disease characterized by typical progressive motor disturbances (involuntary movements of face and body, abnormalities in gait, posture and balance), psychiatric disorders (dementia) and other cognitive impairments [170]. HD is caused by a CAG expansion in the gene encoding for huntingtin (htt), located on chromosome 4; normally, the *htt* gene contains up to 35 CAG repeats, while in HD it has more than 36 CAG repeats that produce a mutant protein, with an abnormally long polyglutamine repeat (polyQ), responsible for the selective striatal degeneration of medium-sized spiny neurons and cerebral cortex [170]. In neurons, mutant htt protein aggregates, thus critically damaging cellular integrity by impairing proteostasis network, mitochondrial function and energy balance, transcriptional regulation, synaptic function and axonal transport [171].

From metabolomic studies, it has emerged that the metabolite (e.g., Trp, kynurenine, quinolinic acid and 3-hydroxykynurenine) content and activity of KP enzymes are pathologically altered in experimental HD models and human patients [109,110]. Moreover, in a *Drosophila* model of HD, disease progression has been found to be associated with a reduction in NAD levels, suggesting that dietetic or pharmacological supplementation of niacin (or its derivatives) may be useful in HD patients [157]. Several studies, indeed, have put forward a beneficial effect of vitamin B₃ in HD (Table 1): for example, nicotinamide is protective against toxicity of polyQ proteins in *Drosophila* HD models [158], while, in transgenic mouse models, it restores brain-derived neurotrophic factor (BDNF) protein levels, increases acetylated peroxisome proliferator-activated receptor gamma coactivator 1α (PGC- 1α), a master regulator of mitochondrial biogenesis, and improves motor deficits [159]. Nicotinamide effects do not depend on inhibition of mutant htt aggregation, but rather on replenishment of NAD levels required to restore energy balance dysregulation occurring in HD.

Further insights into the neuroprotective action of nicotinamide derive from a recent study showing how nicotinamide dose-dependently prevents motor abnormality in 3-nitropropionic acid-induced rat model of HD. Such an effect seems to be linked to prevention of oxidative stress (i.e., decrease in malondialdehyde and nitrites, increase in glutathione), as well as to inhibition of neuronal death in the striatum, most likely through a PARP-1-dependent mechanism [160]. Accordingly, PARP-1 is activated in response to 3-nitropropionic acid-induced neurotoxicity [161] and PARP-1-triggered astrocyte death is prevented by nicotinamide [162]. Like PARP-1, SIRTs are involved in HD neurodegeneration. In particular, SIRT1 is impaired, most likely because of the ability of mutant htt to directly bind and inhibit it; subsequent hyperacetylation and inactivation of specific genes lead to abrogation of the deacetylase pro-survival action [172]. Accordingly, increased SIRT1 activity rescues neurons from mutant htt toxicity and ameliorates pathological mechanisms underlying HD onset [163,164].

All these findings are somewhat controversial, since other studies reported opposite effects. By using the YAC128 transgenic model (expressing the full-length human mutant *htt* gene), Naia and co-workers [173] compared the effects of nicotinamide (a SIRT1 inhibitor) and resveratrol (a SIRT1 activator), both in vitro and in vivo. Both compounds were able to modify histone H3 acetylation and counteract mitochondrial dysfunction in striatal and cortical neurons isolated from YAC128 embryos; nonetheless, only resveratrol ameliorated energy homeostasis and mitochondrial function, as well as motor coordination, in in vivo HD models. Counterintuitively, in vivo nicotinamide supplementation (especially at high concentrations) did not cause any improvement in motor behavior and, furthermore, it worsened motor performance in wild-type control mice. The harmful action has further been documented in other neurodegenerative pathologies: in lactacystin-lesioned rats (an in vivo model of PD), one-month nicotinamide supplementation leads to SIRT1 inhibition and over-expression of neurotrophic and anti-apoptotic factors, nonetheless it exacerbated degeneration of dopaminergic neurons [174].

Therefore, these data underscore the need of full understanding the pathogenetic mechanisms of neurodegeneration, before suggesting any therapeutic challenge to slow down the progression of symptoms.

10. Other Neurological Diseases

Besides neurodegeneration, the impact of vitamin B_3 on CNS has also been investigated in other neuropathological conditions, among which (i) ischemic and traumatic injuries, (ii) headache and (iii) psychiatric disorders (Table 2).

	Effector	Main Findings	Ref.
	Niacin	Diminishes TBI-dependent behavioral deficits and improves functional recovery	[175–180]
	Nam	Reduces neurologic deficits, hippocampal apoptosis, axonal injury and microglial activation in corpus callosum and oxidative stress; restores NAD(P) content; represses MAPK signaling and caspase 3 cleavage	[181]
Ischemic and	Nam mononucleotide	Ameliorates hippocampal injury and improves neurological outcome, by decreasing poly-ADP-ribosylated proteins and NAD ⁺ catabolism	[182]
traumatic injuries	Nam/PARP-1 antagonists	Pre-treatment improves ATP content and neuronal recovery during re-oxygenation	[183]
	Niaspan (niacin)	Increases local cerebral blood flow; promotes angiogenesis via angpt/Tie2, Akt and eNOS pathways; promotes arteriogenesis via TACE and Notch signaling; ameliorates functional deficits	[184,185]
	Niacin plus selenium	Attenuate cortical cell injury, via an increase in Akt phosphorylation and expression of Nrf2; reduce oxidative stress.	[186]
	Nam plus progesterone	Increase function recovery; reduce lesion cavitation and tissue loss; modulate expression of inflammatory and immune genes	[187,188]
	NAMPT	Decreased activity exacerbates post-ischemic brain damage Heterozygous gene deletion aggravates brain damage following photothrombosis-induced focal ischemia Gene over-expression reduces infarct size	[189,190] [190] [191]
Headaches	Niacin	Restores mitochondrial energy metabolism Ameliorates blood flow and oxygenation in contracted skeletal muscle	[192,193]
	Nicotinic acid	Dilates intracranial vessels and contracts extracranial vessels; increases skin biosynthesis of prostaglandin D2; rises plasma content of 9a,11b-prostaglandin F2	[194–196]
	Niacin	Low dietary intakes in neuropsychiatric patients	[197]
Psychiatric disorders	Nam	Positive correlation between vitamin levels and schizophrenia Chronic supplementation effective in maintaining a bipolar type II patient stable and calm	[198] [199]

Table 2. Main findings on the role of niacin in other neurological diseases.

Akt: protein kinase B; Angpt: angiopoietin1; eNOS: endothelial Nitric oxide synthase; MAPK: mitogen-activated protein kinase; Nam: nicotinamide; NAMPT: nicotinamide phosphoribosyltransferase; Nrf2: Nuclear factor (erythroid-derived 2)-like 2; PARP-1: poly(ADP-ribose) polymerase-1; TACE: tumor necrosis factor-alpha-converting enzyme; TBI: traumatic brain injury.

10.1. Ischemic and Traumatic Injuries

When brain cells are deprived of oxygen for more than a few seconds, severe damage occurs, culminating in cell death, through cerebral infarction or ischemic stroke. During reperfusion following a transient ischemic episode, other significant harm (including oxidative stress, leukocyte infiltration, mitochondrial dysfunction, platelet activation and aggregation, complement activation, and blood-brain-barrier disruption) also occur, contributing to neurological dysfunction [200].

Re-oxygenation of neural tissue dramatically impairs $NAD^+/NADH$ recycling, an event known as NADH hyperoxidation [201]. Over the years, the potential neuroprotective and neurorestorative role of vitamin B₃ in ischemic brain injury has extensively been demonstrated in in vitro and in vivo models. By using hippocampal slices, Shetty and co-workers [183] demonstrated that NADH

hyperoxidation is correlated with diminished neuronal recovery that can be rescued by enhancing NAD⁺ levels. Pre-treatment of brain tissue with nicotinamide (to enhance NAD⁺ availability) or PARP-1 antagonists (to lessen NAD⁺ consumption), indeed, prevents mitochondrial dysfunction, improves ATP content and stimulates neuronal recovery, during re-oxygenation [183]. Nicotinamide seems to be efficacious also when provided after ischemia-reperfusion injury. For example, rats receiving a single high dose or repeated low doses of vitamin B₃ after cardiac arrest show reduced neurologic deficits, hippocampal apoptosis, axonal injury and microglial activation in corpus callosum [181]. Nicotinamide-dependent mechanisms underlying these effects include restoration of NAD(P) content and decrease in oxidative stress, along with repression of mitogen-activated protein kinase signaling and caspase 3 cleavage in brain tissue [181]. These data are in agreement with previous reports showing how nicotinamide significantly reduces brain infarct size and improves neurological deficits in different rat strains [202–206]. Interestingly, neurorestoring effects are also present when niacin is provided several hours after ischemic damage: when administrated 24 h after a middle cerebral artery occlusion, Niaspan (a FDA-approved prolonged release formulation of niacin) increases local cerebral blood flow, promotes angiogenesis (via angiopoietin1/Tie2, Akt and endothelial NOS pathways) and arteriogenesis (via tumor necrosis factor-alpha-converting enzyme and Notch signaling), and ameliorates functional deficits [184,185].

NAMPT is critically involved in vitamin B_3 effects. Proof of its key role include: (i) decreased NAMPT activity significantly worsens post-ischemic brain damage [189,190]; (ii) heterozygous *Nampt* deletion aggravates brain damage following photothrombosis-induced focal ischemia [190], (iii) *Nampt* over-expression reduces infarct size [191]. Accordingly, when intraventricularly injected, the NAMPT substrate nicotinamide mononucleotide reverts the detrimental effects of FK866 (a NAMPT inhibitor) [189], ameliorates hippocampal injury and improves neurological outcome, by decreasing poly-ADP-ribosylated proteins and NAD⁺ catabolism [182].

The evidence of niacin efficacy against ischemic insult strongly prompted researchers to investigate its validity in other brain injuries, including traumatic brain injury (TBI). Rats receiving niacin following a cortical contusion injury (an experimental model of TBI) show reduced behavioral deficits and improved long-lasting functional recovery [175–180].

Regardless the type of brain injury, greater beneficial effects have been observed when vitamin B₃ was administrated in combination with other "natural compounds". Co-administration of nicotinamide and progesterone not only increases function recovery, reduces lesion cavitation and tissue loss in both injured cortex and reactive astrocytes, but also modulates expression of genes involved in inflammatory and immune responses, including *Ccr1* (chemokine (C-C motif) receptor 1), *Clec4e* (C-type lectin domain family 4, member 3), *Fn1* (fibronectin 1), *Hmox1* (heme-oxygenase 1), *Hspb1* (heat shock protein b1), *Igf1* and 2 (insulin like growth factor 1 and 2), *Il1* β (interleukin 1 β), *Il16* and 18 (interleukin 16 and 18), *Mmp8* and 9 (matrix metallopeptidase 8 and 9), *Niacr1* (niacin receptor 1) and *Ptgs2* (prostaglandin-endoperoxide synthase 2) [187,188]. In an in vitro model of ischemia-reperfusion injury, combination of niacin and selenium (at clinically relevant doses) synergistically attenuates cortical cell injury, by increasing Akt phosphorylation and expression of nuclear factor erythroid 2-related factor 2, stimulating glutathione redox cycle and reducing hydrogen peroxide levels [186].

10.2. Headache

Affecting more than fifty percent of adult population, headache represents one of the most widespread causes of disability worldwide. Pathogenic mechanisms underlying migraine and tension-type headache (the most common primary headache types) are mostly superimposable: headache, indeed, is triggered by trigeminovascular complex activation that leads to intracranial vessel vasoconstriction followed by extracranial vessel vasodilation and perivascular nociceptive nerve activation. Pressure changes in cerebrospinal fluid and/or intracranial veins are also involved [207,208].

Some nutrients, such as magnesium, carnitine, coenzyme Q10, vitamins (B₂, B₁₂, D) and alpha lipoic acid, can be used as preventive compounds able to counteract headache migraine attacks [209].

When orally, intramuscularly or intravenously administrated, vitamin B₃ (especially, nicotinic acid) has therapeutic effects in headache management [210–215]. It has been proposed that niacin might exert beneficial effects by acting at both central and peripheral levels; in particular, it efficaciously dilates intracranial vessels and subsequently contracts extracranial vessels, favoring, in parallel, the release of compounds leading to peripheral vasodilation and cutaneous flushing. Taking into account that plasma content of serotonin inversely correlates with headache onset, niacin acts, at the central level, by increasing Trp-dependent synthesis of serotonin, via feedback inhibition of the KP [194]. At the peripheral level, pharmacological doses of nicotinic acid increase skin biosynthesis of prostaglandin D2 [195] and the plasma content of its by-product 9a,11b-PGF2, in healthy volunteers [196].

It should also be mentioned that alterations of mitochondrial regulatory networks play a key role in migraine pathophysiology [192,193]. Therefore, by enhancing substrate availability for complex I and reducing lactate concentration, niacin might restore mitochondrial energy metabolism and ameliorate blood flow and oxygenation in sore skeletal muscle, especially in tension-type headache.

10.3. Psychiatric Disorders

A large number of mental disorders have been shown to be influenced by dietary habits, leading to the development of nutritional guidelines for prevention and/or treatment of psychological disorders, including depression, anxiety, schizophrenia, bipolar disorders and psychological distress. In particular, vitamin B₃ dysmetabolism may be linked with some of these neuropsychiatric disorders, although the literature reports conflicting data: as an example, an epidemiologic study conducted on 140 subjects (73 controls and 67 patients with schizophrenia) has revealed that affected individuals show significantly lower dietary intakes of specific nutrients, including niacin [197], whereas a 1-year case-control study performed on 101 controls and 128 cases of schizophrenia found a direct relationship between the disease and nicotinamide levels [198].

The main etiological factors involved in mood disorders appear to be metabolites produced in the KP, as a consequence of the shunt of Trp from serotonin synthesis to kynurenine formation [216]. Serotonergic neurotransmission, indeed, is compromised in the brain of depressed individuals, as a result of activated KP. Since IDO activity is induced under inflammatory and oxidative conditions, and KP is mostly active in astrocytes and microglia (also responsible for production of pro-inflammatory mediators), it has been proposed that unbalanced KP leads to impaired glial-neuronal network, thus priming the CNS against psychological stress [217]. In human postmortem studies, high levels of kynurenic acid (deriving from transamination of kynurenine instead of hydroxylation, see Figure 2) have been found in the prefrontal cortex of schizophrenic individuals; this finding may have clinical relevance, as kynurenic acid is an antagonist of both NMDA and nicotinic acetylcholine receptors, whose blockade is involved in cognitive deficits associated with the disease [218]. Like schizophrenia, alterations in kynurenine precursor have also been observed in bipolar disorder, although, in this case, nicotinamide levels represent a better prognostic factor; indeed, higher nicotinamide levels are correlated with suicide as a cause of death in bipolar patients (1.3-fold increase with respect to bipolar individuals who died from other causes) [219].

The immune-related imbalance of KP can also be responsible for dendritic atrophy and anhedonia associated with major depressive disorder (MDD): comparison between controls (20 healthy subjects) and patients (29 unmedicated individuals who met the Diagnostic and Statistical Manual of Mental Disorders-IV criteria for MDD) revealed, in the MDD group, a lower neuroprotective index [ratio between kynurenic acid (neuroprotective) and quinolinic acid (neurotoxic)], which was negatively correlated with anhedonia and positively correlated with hippocampal and amygdala volume [220]. According to these data, *tdo* knock-out mice show, if compared to wild-type littermates, higher levels of Trp and serotonin in the hippocampus and midbrain, which are connected to increased neurogenesis and amelioration of anxiety-related behavior [221].

Together, such findings suggest a potential antidepressant effect of vitamin B3 or its related products. In a patient with bipolar type II disorder, nicotinamide supplementation (1 g three times daily) for over 11 years has proven effective in maintaining the patient stable and calm [199]. Although a single case report is weak and does not allow us to generalize the results, it may aid in the understanding the potential additional mechanisms accounting for mental disorders.

11. Conclusions

A growing body of evidence highlights the key role of vitamin B3 in neuronal health. What is emerging is that niacin bioavailability is crucial for neuronsurvival and functions: indeed, vitamin deficiency has been recognized as a pathogenic factor for neurological deficits and dementia, as well as for neuronal injury and psychiatric disorders.

Several molecular mechanisms are influenced by vitamin B3 (Figure 4), often strictly linked each other, thus making it difficult to define the precise mechanisms of action of this dietary metabolite. Although further research is needed, it may be speculated that optimal dietary intake of the vitamin will support neuronal health and delay neurodegeneration.



Figure 4. Main molecular mechanisms underlying beneficial effects of niacin in the CNS under physio-pathological conditions. See text for further details. Akt: protein kinase B; Angpt: angiopoietin1; eNOS: endothelial nitric oxide synthase; MAPK: mitogen-activated protein kinase; PARP-1: poly(ADP-ribose) polymerase-1; SIRT1: sirtuin-1; TACE: tumor necrosis factor-alpha-converting enzyme.

Funding: This research received no external funding.

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

AD	Alzheimer's disease
Akt	protein kinase B
ARTC	ADP-ribosyltransferases
ARTD	diphtheria toxin-like ADP-ribosyltransferases
CNS	central nervous system
FOXO3a	forkhead transcription factor
HD	Huntington's disease
HDL	high density lipoprotein
hOAT-10	human organic anion transporter-10
Htt	huntingtin
IDO	indolamine-pyrrole 2-3 dioxygenase
KP	kynurenine pathway
LDL	low density lipoprotein
MDD	major depressive disorder
MPP ⁺	N-methy-l-4-phenylpyridinium
NAD(P)	nicotinamide adenine dinucleotide (phosphate)
NAMPT	nicotinamide phosphoribosyltransferase
NE	niacin equivalents
NMDA	N-methyl-D-aspartate
NNMT	N-methyltransferase
PARP	poly(ADP-ribose) polymerase
PD	Parkinson's disease
polyQ	polyglutamine repeat
ROS	reactive oxygen species
SAM	S-adenosyl-methionine
SIRT	sirtuin
SMCT1/SLC5A8	sodium-coupled monocarboxylate transporter
TBI	traumatic brain injury
TDO	tryptophan 2,3 dioxygenase
Trp	tryptophan
VLDL	very low density lipoprotein

References

- Spies, T.D.; Bean, W.B.; Stone, R.E. The Treatment of Subclinical and Classic Pellagra Use of Nicotinic Acid, Nicotinic Acid Amide and Sodium Nicotinate, with Special Reference to the Vasodilator Action and the Effect on Mental Symptoms. *JAMA* 1938, 111, 584–592. [CrossRef]
- 2. Magni, G.; Amici, A.; Emanuelli, M.; Orsomando, G.; Raffaelli, N.; Ruggieri, S. Enzymology of NAD+ Homeostasis in Man. *Cell. Mol. Life Sci.* **2004**, *61*, 19–34. [CrossRef] [PubMed]
- 3. Goodman, R.P.; Calvo, S.E.; Mootha, V.K. Spatiotemporal Compartmentalization of Hepatic NADH and NADPH Metabolism. *J. Biol. Chem.* **2018**, *293*, 7508–7516. [CrossRef] [PubMed]
- 4. Kirkland, J.B. Niacin Status, NAD Distribution and ADP-Ribose Metabolism. *Curr. Pharm. Des.* **2009**, *15*, 3–11. [CrossRef] [PubMed]
- 5. Nikiforov, A.; Kulikova, V.; Ziegler, M. The Human NAD Metabolome: Functions, Metabolism and Compartmentalization. *Crit. Rev. Biochem. Mol. Biol.* **2015**, *50*, 284–297. [CrossRef] [PubMed]
- Anderson, K.A.; Madsen, A.S.; Olsen, C.A.; Hirschey, M.D. Metabolic Control by Sirtuins and Other Enzymes that Sense NAD(+), NADH, or Their Ratio. *Biochim. Biophys. Acta Bioenerg.* 2017, 1858, 991–998. [CrossRef] [PubMed]
- Mutafova-Yambolieva, V.N.; Hwang, S.J.; Hao, X.; Chen, H.; Zhu, M.X.; Wood, J.D.; Ward, S.M.; Sanders, K.M. Beta-Nicotinamide Adenine Dinucleotide is an Inhibitory Neurotransmitter in Visceral Smooth Muscle. *Proc. Natl. Acad. Sci. USA* 2007, 104, 16359–16364. [CrossRef] [PubMed]

- 8. Moreschi, I.; Bruzzone, S.; Nicholas, R.A.; Fruscione, F.; Sturla, L.; Benvenuto, F.; Usai, C.; Meis, S.; Kassack, M.U.; Zocchi, E.; De Flora, A. Extracellular NAD+ Is an Agonist of the Human P2Y11 Purinergic Receptor in Human Granulocytes. *J. Biol. Chem.* **2008**, *281*, 31419–31429. [CrossRef] [PubMed]
- Klein, C.; Grahnert, A.; Abdelrahman, A.; Muller, C.E.; Hauschildt, S. Extracellular NAD(+) Induces a Rise in [Ca(2+)](i) in Activated Human Monocytes via Engagement of P2Y(1) and P2Y(11) Receptors. *Cell Calcium* 2009, 46, 263–272. [CrossRef] [PubMed]
- 10. Schwarcz, R.; Stone, T.W. The Kynurenine Pathway and the Brain: Challenges, Controversies and Promises. *Neuropharmacology* **2016**, *112*, 237–247. [CrossRef] [PubMed]
- Prousky, J.; Millman, C.; Kirkland, J. Pharmacologic Use of Niacin. J. Evid.-Based Complement. Altern. Med. 2011, 16, 91–101. [CrossRef]
- Bahn, A.; Hagos, Y.; Reuter, S.; Balen, D.; Brzica, H.; Krick, W.; Burckhardt, B.C.; Sabolic, I.; Burckhardt, G. Identification of a New Urate and High Affinity Nicotinate Transporter, hOAT10 (SLC22A13). *J. Biol. Chem.* 2008, 283, 16332–16341. [CrossRef] [PubMed]
- 13. Kumar, J.S.; Subramanian, V.S.; Kapadia, R.; Kashyap, M.L.; Said, H.M. Mammalian Colonocytes Possess a Carrier-Mediated Mechanism for Uptake of Vitamin B3 (niacin): Studies Utilizing Human and Mouse Colonic Preparations. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2013**, *305*, 207–213. [CrossRef] [PubMed]
- 14. Gopal, E.; Miyauchi, S.; Martin, P.M.; Ananth, S.; Roon, P.; Smith, S.B.; Ganapathy, V. Transport of Nicotinate and Structurally Related Compounds by Human SMCT1 (SLC5A8) and Its Relevance to Drug Transport in the Mammalian Intestinal Tract. *Pharm. Res.* **2007**, *24*, 575–584. [CrossRef] [PubMed]
- 15. Gopal, E.; Fei, Y.J.; Miyauchi, S.; Zhuang, L.; Prasad, P.D.; Ganapathy, V. Sodium-Coupled and Electrogenic Transport of B-Complex Vitamin Nicotinic Acid by slc5a8, a Member of the Na/glucose Co-transporter Gene Family. *Biochem. J.* **2005**, *388*, 309–316. [CrossRef] [PubMed]
- 16. Spector, R. Niacinamide Transport Through the Blood-Brain Barrier. *Neurochem. Res.* **1987**, *12*, 27–31. [CrossRef] [PubMed]
- 17. Badawy, A.A. Tryptophan Metabolism in Alcoholism. Nutr. Res. Rev. 2002, 15, 123–152. [CrossRef] [PubMed]
- 18. Badawy, A.A. Kynurenine Pathway of Tryptophan Metabolism: Regulatory and Functional Aspects. *Int. J. Tryptophan Res.* **2017**, *10*. [CrossRef] [PubMed]
- 19. Müller, F. Flavin-Dependent Hydroxylases. Biochem. Soc. Trans. 1985, 13, 443-447. [CrossRef] [PubMed]
- Shibata, K.; Kobayashi, R.; Fukuwatari, T. Vitamin B1 Deficiency Inhibits the Increased Conversion of Tryptophan to Nicotinamide in Severe Food-Restricted Rats. *Biosci. Biotechnol. Biochem.* 2015, 79, 103–108. [CrossRef] [PubMed]
- 21. Shibata, K.; Onodera, M. Comparison of Tryptophan-Niacin Conversion in Rats Fed with a Nicotinic Acid-Free Diet Containing Egg White, Egg White Proteolysate, or Mixtures of Amino Acids. *Agric. Biol. Chem.* **1991**, *55*, 1291–1298. [CrossRef]
- 22. Shibata, K.; Nomamoto, R.; Iwai, K. Effect of Dietary Protein levels on the Urinary Excretion of Nicotinamide and Its Metabolites in Rats. *Agric. Biol. Chem.* **1988**, *53*, 1765–1769. [CrossRef]
- 23. Shibata, K. Nutritional Factors that Regulate on the Conversion of L-Tryptophan to Niacin. *Adv. Exp. Med. Biol.* **1999**, 467, 711–716. [CrossRef] [PubMed]
- 24. Shibata, K. Organ Co-Relationship in Tryptophan Metabolism and Factors That Govern the Biosynthesis of Nicotinamide from Tryptophan. *J. Nutr. Sci. Vitaminol.* **2018**, *64*, 90–98. [CrossRef] [PubMed]
- 25. Shibata, K.; Nakata, C.; Fukuwatari, T. Moderate Food Restriction Suppresses the Conversion of L-tryptophan to Nicotinamide in Weaning Rats. *Biosci. Biotechnol. Biochem.* **2014**, *78*, 478–481. [CrossRef] [PubMed]
- 26. Badawy, A.A. Pellagra and Alcoholism: A Biochemical Perspective. *Alcohol Alcohol.* **2014**, *49*, 238–250. [CrossRef] [PubMed]
- 27. Bender, D.A. Inhibition in Vitro of the Enzymes of the Oxidative Pathway of Tryptophan Metabolism and of Nicotinamide Nucleotide Synthesis by Benserazide, Carbidopa and Isoniazid. *Biochem. Pharmacol.* **1980**, *29*, 707–712. [CrossRef]
- 28. Braidman, I.P.; Rose, D.P. The Effect of Sex Hormones on the Activity of Tryptophan Oxygenase and Other Corticosteroid-Inducible Enzymes in Rat Liver. *Biochem. J.* **1971**, 122, 28P. [CrossRef] [PubMed]
- 29. Nakamura, T.; Shinno, H.; Ichihara, A. Insulin and Glucagon as a New Regulator System for Tryptophan Oxygenase Activity Demonstrated in Primary Cultured Rat Hepatocytes. *J. Biol. Chem.* **1980**, 255, 7533–7535. [PubMed]

- 30. Nakamura, T.; Niimi, S.; Nawa, K.; Noda, C.; Ichihara, A.; Takagi, Y.; Anai, M.; Sakaki, Y. Multihormonal Regulation of Transcription of the Tryptophan 2,3-Dioxygenase Gene in Primary Cultures of Adult Rat Hepatocytes with Special Reference to the Presence of a Transcriptional Protein Mediating the Action of Glucocorticoids. J. Biol. Chem. 1987, 262, 723–732.
- 31. Labrie, F.; Korner, A. Effect of Glucagon, Insulin, and Thyroxine on Tyrosine Transaminase and Tryptophan Pyrrolase of Rat Liver. *Arch. Biochem. Biophys.* **1969**, *129*, 75–78. [CrossRef]
- 32. Chiancone, F.M. Enzyme Activities in the Tryptophan → Nicotinic Acid Path in Physiopathology. *Ital. J. Biochem.* **1964**, *13*, 1–30.
- Ku, Y.; Rogers, Q.R.; Harper, A.E. Effects of Thyroxine and Cortisol on Liver Threonine Dehydratase and Tryptophan Pyrrolase in Rats Fed a High Protein Diet. *Proc. Soc. Exp. Biol. Med.* 1969, 130, 556–563. [CrossRef] [PubMed]
- 34. Shibata, K.; Toda, S. Effect of Thyroxine on the Metabolism of Tryptophan to Nicotinamide in Rats. *Biosci. Biotechnol. Biochem.* **1994**, *58*, 1757–1762. [CrossRef]
- 35. Horwitt, M.K.; Harper, A.E.; Henderson, L.M. Niacin-Tryptophan Relationships for Evaluating Niacin Equivalents. *Am. J. Clin. Nutr.* **1981**, *34*, 423–427. [CrossRef] [PubMed]
- 36. Food and Nutrition Board. *Dietary Reference Intakes for Thiamine, Riboflavin, Niacin, Vitamin B6, Folate, Vitamin B12, Panthotenic Acid, Biotin and Choline;* National Academy Press: Washington, DC, USA, 1998; pp. 374–389.
- 37. Long, A.; Klimova, N.; Kristian, T. Mitochondrial NUDIX hydrolases: A Metabolic Link Between NAD Catabolism, GTP and Mitochondrial Dynamics. *Neurochem. Int.* **2017**, *109*, 193–201. [CrossRef] [PubMed]
- Bogan, K.L.; Evans, C.; Belenky, P.; Song, P.; Burant, C.F.; Kennedy, R.; Brenner, C. Identification of Isn1 and Sdt1 as Glucose- and Vitamin-Regulated Nicotinamide Mononucleotide and Nicotinic Acid Mononucleotide [corrected] 5'-Nucleotidases Responsible for Production of Nicotinamide Riboside and Nicotinic Acid Riboside. J. Biol. Chem. 2009, 284, 34861–34869. [CrossRef]
- Wielgus-Kutrowska, B.; Kulikowska, E.; Wierzchowski, J.; Bzowska, A.; Shugar, D. Nicotinamide Riboside, an Unusual, Non-Typical, Substrate of Purified Purine-Nucleoside Phosphorylases. *Eur. J. Biochem.* 1997, 243, 408–414. [CrossRef] [PubMed]
- 40. Haigis, M.C.; Mostoslavsky, R.; Haigis, K.M.; Fahie, K.; Christodoulou, D.C.; Murphy, A.J.; Valenzuela, D.M.; Yancopoulos, G.D.; Karow, M.; Blander, G.; et al. SIRT4 Inhibits Glutamate Dehydrogenase and Opposes the Effects of Calorie Restriction in Pancreatic Beta Cells. *Cell* **2006**, *126*, 941–954. [CrossRef] [PubMed]
- 41. Mao, Z.; Hine, C.; Tian, X.; Van Meter, M.; Au, M.; Vaidya, A.; Seluanov, A.; Gorbunova, V. SIRT6 Promotes DNA Repair Under Stress by Activating PARP1. *Science* **2011**, *332*, 1443–1446. [CrossRef] [PubMed]
- 42. Wang, Y.; He, J.; Liao, M.; Hu, M.; Li, W.; Ouyang, H.; Wang, X.; Ye, T.; Zhang, Y.; Ouyang, L. An Overview of Sirtuins as Potential Therapeutic Target: Structure, Function and Modulators. *Eur. J. Med. Chem.* **2019**, 161, 48–77. [CrossRef] [PubMed]
- 43. Imai, S.; Guarente, L. Ten Years of NAD-Dependent SIR2 Family Deacetylases: Implications for Metabolic Diseases. *Trends Pharmacol. Sci.* **2010**, *31*, 212–220. [CrossRef] [PubMed]
- 44. Singh, C.K.; Chhabra, G.; Ndiaye, M.A.; Garcia-Peterson, L.M.; Mack, N.J.; Ahmad, N. The Role of Sirtuins in Antioxidant and Redox Signaling. *Antioxid. Redox Signal.* **2018**, *28*, 643–661. [CrossRef] [PubMed]
- 45. Xu, J.; Jackson, C.W.; Khoury, N.; Escobar, I.; Perez-Pinzon, M.A. Brain SIRT1 Mediates Metabolic Homeostasis and Neuroprotection. *Front. Endocrinol.* **2018**, *9*, 702. [CrossRef] [PubMed]
- 46. Cohen, M.S.; Chang, P. Insights Into the Biogenesis, Function, and Regulation of ADP-Ribosylation. *Nat. Chem. Biol.* **2018**, *14*, 236–243. [CrossRef] [PubMed]
- 47. Kunze, F.A.; Hottiger, M.O. Regulating Immunity via ADP-Ribosylation: Therapeutic Implications and Beyond. *Trends Immunol.* **2019**. [CrossRef] [PubMed]
- 48. Crawford, K.; Bonfiglio, J.J.; Mikoč, A.; Matic, I.; Ahel, I. Specificity of Reversible ADP-Ribosylation and Regulation of Cellular Processes. *Crit. Rev. Biochem. Mol. Biol.* **2018**, 53, 64–82. [CrossRef] [PubMed]
- 49. Lee, H.C.; Walseth, T.F.; Bratt, G.T.; Hayes, R.N.; Clapper, D.L. Structural Determination of a Cyclic Metabolite of NAD+ with Intracellular Ca2+-Mobilizing Activity. *J. Biol. Chem.* **1989**, *264*, 1608–1615. [PubMed]
- 50. Swarbrick, J.M.; Graeff, R.; Garnham, C.; Thomas, M.P.; Galione, A.; Potter, B.V. 'Click Cyclic ADP-Ribose': A Neutral Second Messenger Mimic. *Chem. Commun.* **2014**, *50*, 2458–2461. [CrossRef] [PubMed]
- 51. Ferrero, E.; Lo Buono, N.; Horenstein, A.L.; Funaro, A.; Malavasi, F. The ADP-Ribosyl Cyclases–the Current Evolutionary State of the ARCs. *Front. Biosci.* **2014**, *19*, 986–1002. [CrossRef]

- 52. Wei, W.; Graeff, R.; Yue, J. Roles and Mechanisms of the CD38/Cyclic Adenosine Diphosphate Ribose/Ca(2+) Signaling Pathway. *World J. Biol. Chem.* **2014**, *5*, 58–67. [CrossRef] [PubMed]
- 53. Pehar, M.; Harlan, B.A.; Killoy, K.M.; Vargas, M.R. Nicotinamide Adenine Dinucleotide Metabolism and Neurodegeneration. *Antioxid. Redox Signal.* **2018**, *28*, 1652–1668. [CrossRef] [PubMed]
- 54. Chmielewski, J.P.; Bowlby, S.C.; Wheeler, F.B.; Shi, L.; Sui, G.; Davis, A.L.; Howard, T.D.; D'Agostino, R.B., Jr.; Miller, L.D.; Sirintrapun, S.J.; et al. CD38 Inhibits Prostate Cancer Metabolism and Proliferation by Reducing Cellular NAD(+) Pools. *Mol. Cancer Res.* **2018**, *11*, 1687–1700. [CrossRef] [PubMed]
- 55. Morandi, F.; Horenstein, A.L.; Rizzo, R.; Malavasi, F. The Role of Extracellular Adenosine Generation in the Development of Autoimmune Diseases. *Mediators Inflamm.* **2018**, 2018, 7019398. [CrossRef] [PubMed]
- Zhang, M.; Ying, W. NAD(+) Deficiency Is a Common Central Pathological Factor of a Number of Diseases and Aging: Mechanisms and Therapeutic Implications. *Antioxid. Redox Signal.* 2018, 30, 890–905. [CrossRef] [PubMed]
- 57. Komatsu, M.; Kanda, T.; Urai, H.; Kurokochi, A.; Kitahama, R.; Shigaki, S.; Ono, T.; Yukioka, H.; Hasegawa, K.; Tokuyama, H.; et al. NNMT Activation Can Contribute to the Development of Fatty Liver Disease by Modulating the NAD (+) Metabolism. *Sci. Rep.* **2018**, *8*, 8637. [CrossRef] [PubMed]
- 58. Bach, D.H.; Kim, D.; Bae, S.Y.; Kim, W.K.; Hong, J.Y.; Lee, H.J.; Rajasekaran, N.; Kwon, S.; Fan, Y.; Luu, T.T.; et al. Targeting Nicotinamide N-Methyltransferase and miR-449a in EGFR-TKI-Resistant Non-Small-Cell Lung Cancer Cells. *Mol. Ther. Nucleic Acids* 2018, *11*, 455–467. [CrossRef] [PubMed]
- 59. Crujeiras, A.B.; Pissios, P.; Moreno-Navarrete, J.M.; Diaz-Lagares, A.; Sandoval, J.; Gomez, A.; Ricart, W.; Esteller, M.; Casanueva, F.F.; Fernandez-Real, J.M. An Epigenetic Signature in Adipose Tissue Is Linked to Nicotinamide N-Methyltransferase Gene Expression. *Mol. Nutr. Food Res.* 2018, 62, e1700933. [CrossRef] [PubMed]
- 60. Kannt, A.; Rajagopal, S.; Kadnur, S.V.; Suresh, J.; Bhamidipati, R.K.; Swaminathan, S.; Hallur, M.S.; Kristam, R.; Elvert, R.; Czech, J.; et al. A Small Molecule Inhibitor of Nicotinamide N-Methyltransferase for the Treatment of Metabolic Disorders. *Sci. Rep.* **2018**, *8*, 3660. [CrossRef] [PubMed]
- 61. Rudolphi, B.; Zapp, B.; Kraus, N.A.; Ehebauer, F.; Kraus, B.J.; Kraus, D. Body Weight Predicts Nicotinamide N-Methyltransferase Activity in Mouse Fat. *Endocr. Res.* **2018**, *43*, 55–63. [CrossRef] [PubMed]
- 62. Nejabati, H.R.; Mihanfar, A.; Pezeshkian, M.; Fattahi, A.; Latifi, Z.; Safaie, N.; Valiloo, M.; Jodati, A.R.; Nouri, M. N1-Methylnicotinamide (MNAM) as a Guardian of Cardiovascular System. *J. Cell. Physiol.* **2018**, 233, 6386–6394. [CrossRef] [PubMed]
- 63. Schmeisser, K.; Parker, J.A. Nicotinamide-N-Methyltransferase Controls Behavior, Neurodegeneration and Lifespan by Regulating Neuronal Autophagy. *PLoS Genet.* **2018**, *14*, e1007561. [CrossRef] [PubMed]
- 64. Lu, X.M.; Long, H. Nicotinamide N-Methyltransferase as a Potential Marker for Cancer. *Neoplasma* **2018**, *65*, 656–663. [CrossRef] [PubMed]
- 65. Kremer, J.I.; Gömpel, K.; Bakuradze, T.; Eisenbrand, G.; Richling, E. Urinary Excretion of Niacin Metabolites in Humans After Coffee Consumption. *Mol. Nutr. Food Res.* **2018**, *62*, 1700735. [CrossRef] [PubMed]
- 66. Liu, M.; Zhang, D.; Wang, X.; Zhang, L.; Han, J.; Yang, M.; Xiao, X.; Zhang, Y.; Liu, H. Simultaneous Quantification of Niacin and Its Three Main Metabolites in Human Plasma by LC-MS/MS. *J. Chromatogr. B Anal. Technol. Biomed. Life Sci.* **2012**, *904*, 107–114. [CrossRef] [PubMed]
- Zhang, P.; Sun, Y.; Shi, G.; Sui, Y.; Li, Q.; Tang, Y.; Gu, J. Quantification of Niacin and Its Metabolite Nicotinuric Acid in Human Plasma by LC-MS/MS: Application to a Clinical Trial of a Fixed Dose Combination Tablet of Niacin Extended-Release/Simvastatin (500 mg/10 mg) in Healthy Chinese Volunteers. *Int. J. Anal. Chem.* 2015, 2015, 212437. [CrossRef] [PubMed]
- 68. Pittelli, M.; Formentini, L.; Faraco, G.; Lapucci, A.; Rapizzi, E.; Cialdai, F.; Romano, G.; Moneti, G.; Moroni, F.; Chiarugi, A. Inhibition of Nicotinamide Phosphoribosyltransferase: Cellular Bioenergetics Reveals a Mitochondrial Insensitive NAD Pool. *J. Biol. Chem.* **2010**, *285*, 34106–34114. [CrossRef] [PubMed]
- 69. Sydenstricker, V.P. The History of Pellagra, Its Recognition as a Disorder of Nutrition and Its Conquest. *Am. J. Clin. Nutr.* **1958**, *6*, 409–414. [CrossRef] [PubMed]
- 70. Gentilcore, D. Louis Sambon and the Clash of Pellagra Etiologies in Italy and the United States, 1905–1914. *J. Hist. Med. Allied Sci.* **2016**, 19–42. [CrossRef] [PubMed]
- 71. Kirkland, J.B.; Zempleni, J.; Suttie, J.W.; Gregory, J.F., III; Stover, P.J. (Eds.) *Handbook of Vitamins*, 5th ed.; CRC Press: Boca Raton, FL, USA, 2013; pp. 149–190.

- 72. Jagielska, G.; Tomaszewicz-Libudzic, E.C.; Brzozowska, A. Pellagra: A Rare Complication of Anorexia Nervosa. *Eur. Child. Adolesc. Psychiatry* **2007**, *16*, 417–420. [CrossRef] [PubMed]
- 73. Monteiro, J.P.; da Cunha, D.F.; Filho, D.C.; Silva-Vergara, M.L.; dos Santos, V.M.; da Costa, J.C., Jr.; Etchebehere, R.M.; Gonçalves, J.; de Carvalho da Cunha, S.F.; Jordão, A.A.; et al. Niacin Metabolite Excretion in Alcoholic Pellagra and AIDS Patients with and without Diarrhea. *Nutrition* **2004**, *20*, 778–782. [CrossRef] [PubMed]
- 74. Park, S.M.; Li, T.; Wu, S.; Li, W.Q.; Weinstock, M.; Qureshi, A.A.; Cho, E. Niacin Intake and Risk of Skin Cancer in US Women and Men. *Int. J. Cancer* **2017**, *140*, 2023–2031. [CrossRef] [PubMed]
- 75. Kirkland, J.B. Niacin Status and Treatment-Related Leukemogenesis. *Mol. Cancer Ther.* 2009, *8*, 725–732. [CrossRef] [PubMed]
- 76. Rosmaninho, A.; Sanches, M.; Fernandes, I.C.; Pinto-Almeida, T.; Vilaça, S.; Oliveira, A.; Selores, M. Letter: Pellagra as the Initial Presentation of Crohn Disease. *Dermatol. Online J.* **2012**, *18*, 12. [PubMed]
- 77. Prakash, R.; Gandotra, S.; Singh, L.K.; Das, B.; Lakra, A. Rapid Resolution of Delusional Parasitosis in Pellagra with Niacin Augmentation Therapy. *Gen. Hosp. Psychiatry* **2008**, *30*, 581–584. [CrossRef] [PubMed]
- 78. World Health Organization; United Nations High Commissions for Refugees. Pellagra and Its Prevention and Control in Major Emergencies. World Health Organization. 2000. Available online: http://www.who.int/nutrition/publications/emergencies/WHO_NHD_00.10/en/ (accessed on 1 December 2018).
- 79. Altschul, R.; Hoffer, A.; Stephen, J.D. Influence of Nicotinic Acid on Serum Cholesterol in Man. *Arch. Biochem. Biophys.* **1955**, *54*, 558–559. [CrossRef]
- 80. Zeman, M.; Vecka, M.; Perlík, F.; Staňková, B.; Hromádka, R.; Tvrzická, E.; Širc, J.; Hrib, J.; Žák, A. Pleiotropic Effects of Niacin: Current Possibilities for Its Clinical Use. *Acta Pharm.* **2016**, *66*, 449–469. [CrossRef] [PubMed]
- 81. la Paz, S.M.; Bermudez, B.; Naranjo, M.C.; Lopez, S.; Abia, R.; Muriana, F.J. Pharmacological Effects of Niacin on Acute Hyperlipemia. *Curr. Med. Chem.* **2016**, *23*, 2826–2835. [CrossRef] [PubMed]
- 82. Offermanns, S.; Colletti, S.L.; Lovenberg, T.W.; Semple, G.; Wise, A.; IJzerman, A.P. International Union of Basic and Clinical Pharmacology. LXXXII: Nomenclature and Classification of Hydroxy-carboxylic Acid Receptors (GPR81, GPR109A, andGPR109B). *Pharmacol. Rev.* **2011**, *63*, 269–290. [CrossRef] [PubMed]
- 83. Felts, A.S. Molecule of the Month. TREDAPTIVE (Nicotinic Acid/Laropiprant): A New Lipid-Modifying Therapy for the Treatment of LDL-C, HDL-C and Triglycerides. *Curr. Top. Med. Chem.* **2008**, *8*, 1310. [CrossRef] [PubMed]
- 84. AIM-HIGH Investigators; Boden, W.E.; Probstfield, J.L.; Anderson, T.; Chaitman, B.R.; Desvignes-Nickens, P.; Koprowicz, K.; McBride, R.; Teo, K.; Weintraub, W. Niacin in Patients with Low HDL Cholesterol Levels Receiving Intensive Statin Therapy. *N. Engl. J. Med.* **2011**, *365*, 2255–2267. [CrossRef] [PubMed]
- 85. HPS2-THRIVE Collaborative Group. HPS2-THRIVE Randomized Placebo-Controlled Trial in 25 673 High-Risk Patients of ER Niacin/Laropiprant: Trial Design, Pre-Specified Muscle and Liver Outcomes, and Reasons for Stopping Studytreatment. *Eur. Heart J.* **2013**, *34*, 1279–1291. [CrossRef]
- HPS2-THRIVE Collaborative Group; Landray, M.J.; Haynes, R.; Hopewell, J.C.; Parish, S.; Aung, T.; Tomson, J.; Wallendszus, K.; Craig, M.; Jiang, L.; et al. Effects of Extended-Release Niacin with Laropiprant in High-Risk Patients. *N. Engl. J. Med.* 2014, *371*, 203–212. [CrossRef]
- 87. Graff, E.C.; Fang, H.; Wanders, D.; Judd, R.L. Anti-inflammatory Effects of the Hydroxycarboxylic Acid Receptor 2. *Metabolism* **2016**, *65*, 102–113. [CrossRef] [PubMed]
- Yang, S.; Li, X.; Wang, N.; Yin, G.; Ma, S.; Fu, Y.; Wei, C.; Chen, Y.; Xu, W. GPR109A Expression in the Murine Min6 Pancreatic Beta Cell Line, and Its Relation with Glucose Metabolism and Inflammation. *Ann. Clin. Lab. Sci.* 2015, 45, 315–322. [PubMed]
- Xu, X.; Lin, S.; Chen, Y.; Li, X.; Ma, S.; Fu, Y.; Wei, C.; Wang, C.; Xu, W. The Effect of Metformin on the Expression of GPR109A, NF-κB and IL-1β in Peripheral Blood Leukocytes from Patients with Type 2 Diabetes Mellitus. *Ann. Clin. Lab. Sci.* 2017, 47, 556–562. [PubMed]
- 90. Heemskerk, M.M.; Dharuri, H.K.; van den Berg, S.A.; Jónasdóttir, H.S.; Kloos, D.P.; Giera, M.; van Dijk, K.W.; van Harmelen, V. Prolonged Niacin Treatment Leads to Increased Adipose Tissue PUFA Synthesis and Anti-Inflammatory Lipid and Oxylipin Plasmaprofile. *J. Lipid Res.* **2014**, *55*, 2532–2540. [CrossRef] [PubMed]
- 91. Wanders, D.; Graff, E.C.; White, B.D.; Judd, R.L. Niacin Increases Adiponectin and Decreases Adipose Tissue Inflammation in High Fat Diet-Fed Mice. *PLoS ONE* **2013**, *8*, 71285. [CrossRef] [PubMed]

- Su, G.; Sun, G.; Liu, H.; Shu, L.; Zhang, J.; Guo, L.; Huang, C.; Xu, J. Niacin Suppresses Progression of Atherosclerosis by Inhibiting Vascular Inflammation and Apoptosis of Vascular Smooth Muscle Cells. *Med. Sci. Monit.* 2015, 21, 4081–4089. [CrossRef] [PubMed]
- Zhou, E.; Li, Y.; Yao, M.; Wei, Z.; Fu, Y.; Yang, Z. Niacin Attenuates the Production of Pro-Inflammatory Cytokines in LPS-Induced Mouse Alveolar Macrophages by HCA2 Dependent Mechanisms. *Int. Immunopharmacol.* 2014, 23, 121–126. [CrossRef] [PubMed]
- 94. Cho, K.H.; Kim, H.J.; Rodriguez-Iturbe, B.; Vaziri, N.D. Niacin ameliorates oxidative stress, inflammation, proteinuria, and hypertension in rats with chronic renal failure. *Am. J. Physiol. Ren. Physiol.* **2009**, 297, 106–113. [CrossRef] [PubMed]
- 95. Jeong, K.Y.; Suh, G.J.; Kwon, W.Y.; Kim, K.S.; Jung, Y.S.; Kye, Y.C. The Therapeutic Effect and Mechanism of Niacin on Acute Lung Injury in a Rat Model of Hemorrhagic Shock: Down-Regulation of the Reactive Oxygen Species-Dependent Nuclear Factor κB Pathway. *J. Trauma Acute Care Surg.* 2015, *79*, 247–255. [CrossRef] [PubMed]
- Godin, A.M.; Ferreira, W.C.; Rocha, L.T.; Ferreira, R.G.; Paiva, A.L.; Merlo, L.A.; Nascimento, E.B., Jr.; Bastos, L.F.; Coelho, M.M. Nicotinic Acid Induces Antinociceptive and Anti-Inflammatory Effects in Different Experimental Models. *Pharmacol. BiochemBehav.* 2012, 101, 493–498. [CrossRef] [PubMed]
- Ruddock, M.W.; Hirst, D.G. Nicotinamide Relaxes Vascular Smooth Muscle by Inhibiting Myosin Light Chain Kinase-Dependent Signaling Pathways: Implications for Anticancer Efficacy. Oncol. Res. 2004, 14, 483–489. [CrossRef] [PubMed]
- Ruddock, M.W.; Burns, D.M.; McKeown, S.R.; Murphy, L.; Walsh, I.K.; Keane, P.F.; Hirst, D.G. Contractile Properties of Human Renal Cell Carcinoma Recruited Arteries and Theirresponse to Nicotinamide. *Radiother. Oncol.* 2000, 54, 179–184. [CrossRef]
- Agote, M.; Viaggi, M.; Kreimann, E.; Krawiec, L.; Dagrosa, M.A.; Juvenal, G.J.; Pisarev, M.A. Influence of Nicotinamide on the Radiosensitivity of Normal and Goitrous Thyroid in the Rat. *Thyroid* 2001, *11*, 1003–1007. [CrossRef] [PubMed]
- 100. Hoskin, P.; Rojas, A.; Saunders, M. Accelerated Radiotherapy, Carbogen, and Nicotinamide (ARCON) in the Treatment of Advanced Bladder Cancer: Mature Results of a Phase II Nonrandomized Study. *Int. J. Radiat. Oncol. Biol. Phys.* 2009, 73, 1425–1431. [CrossRef] [PubMed]
- Williams, A.; Ramsden, D. Nicotinamide: A Double Edged Sword. *Parkinsonism Relat. Disord.* 2005, 11, 413–420. [CrossRef] [PubMed]
- 102. Fricker, R.A.; Green, E.L.; Jenkins, S.I.; Griffin, S.M. The Influence of Nicotinamide on Health and Disease in the Central Nervous System. *Int. J. Tryptophan Res.* **2018**, *11*. [CrossRef] [PubMed]
- 103. Turski, W.A.; Nakamura, M.; Todd, W.P.; Carpenter, B.K.; Whetsell, W.O., Jr.; Schwarcz, R. Identification and Quantification of Kynurenic Acid in Human Brain Tissue. *Brain Res.* **1988**, 454, 164–169. [CrossRef]
- 104. Gobaille, S.; Kemmel, V.; Brumaru, D.; Dugave, C.; Aunis, D.; Maitre, M. Xanthurenic Acid Distribution, Transport, Accumulation and Release in the Rat Brain. *J. Neurochem.* **2008**, *105*, 982–993. [CrossRef] [PubMed]
- 105. Baran, H.; Schwarcz, R. Presence of 3-hydroxyanthranilic Acid in Rat Tissues and Evidence for Its Production from Anthranilic Acid in the Brain. *J. Neurochem.* **1990**, *55*, 738–744. [CrossRef] [PubMed]
- 106. Fukui, S.; Schwarcz, R.; Rapoport, S.I.; Takada, Y.; Smith, Q.R. Blood-brain Barrier Transport of Kynurenines: Implications for Brain Synthesis and Metabolism. J. Neurochem. 1991, 56, 2007–2017. [CrossRef] [PubMed]
- Foster, A.C.; Collins, J.F.; Schwarcz, R. On the Excitotoxic Properties of Quinolinic Acid, 2,3-Piperidine Dicarboxylic Acids and Structurally Related Compounds. *Neuropharmacology* 1983, 22, 1331–1342. [CrossRef]
- 108. Moroni, F.; Lombardi, G.; Carlà, V.; Moneti, G. The Excitotoxin Quinolinic Acid is Present and Unevenly Distributed in the Rat Brain. *Brain Res.* **1984**, *19*, 352–355. [CrossRef]
- Bohár, Z.; Toldi, J.; Fülöp, F.; Vécsei, L. Changing the Face of Kynurenines and Neurotoxicity: Therapeutic Considerations. *Int. J. Mol. Sci.* 2015, *16*, 9772–9793. [CrossRef] [PubMed]
- Majewski, M.; Kozlowska, A.; Thoene, M.; Lepiarczyk, E.; Grzegorzewski, W.J. Overview of the Role of Vitamins and Minerals on the Kynurenine Pathway in Health and Disease. *J. Physiol. Pharmacol.* 2016, 67, 3–19. [PubMed]
- 111. Dang, Y.; Dale, W.E.; Brown, O.R. Comparative Effects of Oxygen on Indoleamine 2,3-Dioxygenase and Tryptophan 2,3-Dioxygenase of the Kynurenine Pathway. *Free Radic. Biol. Med.* **2000**, *28*, 615–624. [CrossRef]

- Kanai, M.; Nakamura, T.; Funakoshi, H. Identification and Characterization of Novel Variants of the Tryptophan 2,3-Dioxygenase Gene: Differential Regulation in the Mouse Nervous System During Development. *Neurosci. Res.* 2009, 64, 111–117. [CrossRef] [PubMed]
- 113. Opitz, C.A.; Litzenburger, U.M.; Sahm, F.; Ott, M.; Tritschler, I.; Trump, S.; Schumacher, T.; Jestaedt, L.; Schrenk, D.; Weller, M.; et al. An Endogenous Tumour-Promoting Ligand of the Human Aryl Hydrocarbon Receptor. *Nature* 2011, 478, 197–203. [CrossRef] [PubMed]
- 114. Lanz, T.V.; Williams, S.K.; Stojic, A.; Iwantscheff, S.; Sonner, J.K.; Grabitz, C.; Becker, S.; Böhler, L.I.; Mohapatra, S.R.; Sahm, F.; et al. Tryptophan-2,3-Dioxygenase (TDO) Deficiency is Associated with Subclinical Neuroprotection in a Mouse Model of Multiplesclerosis. *Sci. Rep.* 2017, *7*, 41271. [CrossRef] [PubMed]
- 115. O'Connor, J.C.; André, C.; Wang, Y.; Lawson, M.A.; Szegedi, S.S.; Lestage, J.; Castanon, N.; Kelley, K.W.; Dantzer, R. Interferon-Gamma and Tumor Necrosis Factor-Alpha Mediate the Upregulation of Indoleamine 2,3-Dioxygenase and the Induction of Depressive-Like Behavior in Mice in Response to Bacillus Calmette-Guerin. J. Neurosci. 2009, 29, 4200–4209. [CrossRef] [PubMed]
- 116. O'Connor, J.C.; Lawson, M.A.; André, C.; Briley, E.M.; Szegedi, S.S.; Lestage, J.; Castanon, N.; Herkenham, M.; Dantzer, R.; Kelley, K.W. Induction of IDO by Bacille Calmette-Guérin is Responsible for Development of Murine Depressive-Like Behavior. *J. Immunol.* 2009, *182*, 3202–3212. [CrossRef] [PubMed]
- 117. Corona, A.W.; Norden, D.M.; Skendelas, J.P.; Huang, Y.; O'Connor, J.C.; Lawson, M.; Dantzer, R.; Kelley, K.W.; Godbout, J.P. Indoleamine 2,3-Dioxygenase Inhibition Attenuates Lipopolysaccharide Induced Persistent Microglial Activation and Depressive-Like Complications in Fractalkine Receptor (CX(3)CR1)-Deficient Mice. *Brain Behav. Immun.* 2013, *31*, 134–142. [CrossRef] [PubMed]
- 118. Rharass, T.; Lantow, M.; Gbankoto, A.; Weiss, D.G.; Panáková, D.; Lucas, S. Ascorbic Acid Alters Cell Fate Commitment of Human Neural Progenitors in a WNT/β-Catenin/ROS Signaling Dependent Manner. J. Biomed. Sci. 2017, 16, 78. [CrossRef] [PubMed]
- 119. Cataldi, S.; Arcuri, C.; Hunot, S.; Mecca, C.; Codini, M.; Laurenti, M.E.; Ferri, I.; Loreti, E.; Garcia-Gil, M.; Traina, G.; et al. Effect of Vitamin D in HN9.10e Embryonic Hippocampal Cells and in Hippocampus from MPTP-Induced Parkinson's Disease Mouse Model. *Front. Cell. Neurosci.* 2018, 12, 31. [CrossRef] [PubMed]
- 120. Cui, X.; Gooch, H.; Petty, A.; McGrath, J.J.; Eyles, D. Vitamin D and the Brain: Genomicand Non-Genomic Actions. *Mol. Cell. Endocrinol.* **2017**, 453, 131–143. [CrossRef] [PubMed]
- 121. Haushalter, C.; Asselin, L.; Fraulob, V.; Dollé, P.; Rhinn, M. Retinoic Acid Controls Early Neurogenesis in the Developing Mouse Cerebral Cortex. *Dev. Biol.* **2017**, *430*, 129–141. [CrossRef] [PubMed]
- Lin, Y.; Lin, Y.; Lin, H.; Chen, Y.; Wang, H.; Shi, J. Application of Propyl Gallate Alleviates Pericarp Browning in Harvested Longan Fruit by Modulating Metabolisms of Respiration and Energy. *Food Chem.* 2018, 240, 863–869. [CrossRef] [PubMed]
- 123. Griffin, S.M.; Pickard, M.R.; Orme, R.P.; Hawkins, C.P.; Fricker, R.A. Nicotinamide Promotes Neuronal Differentiation of Mouse Embryonic Stem Cells in Vitro. *Neuroreport* 2013, 24, 1041–1046. [CrossRef] [PubMed]
- 124. Griffin, S.M.; Pickard, M.R.; Orme, R.P.; Hawkins, C.P.; Williams, A.C.; Fricker, R.A. Nicotinamide Alone Accelerates the Conversion of Mouse Embryonic Stem Cells into Mature Neuronal Populations. *PLoS ONE* 2017, 12, e0183358. [CrossRef] [PubMed]
- 125. Sperber, H.; Mathieu, J.; Wang, Y.; Ferreccio, A.; Hesson, J.; Xu, Z.; Fischer, K.A.; Devi, A.; Detraux, D.; Gu, H.; et al. The Metabolome Regulates the Epigenetic Landscape During Naive-to-Primed Human Embryonic Stem Cell Transition. *Nat. Cell Biol.* 2015, *17*, 1523–1535. [CrossRef] [PubMed]
- 126. Chong, Z.Z.; Lin, S.H.; Maiese, K. The NAD+ Precursor Nicotinamide Governs Neuronal Survival During Oxidative Stress Through Protein Kinase B Coupled to FOXO3a and Mitochondrial Membrane Potential. *J. Cereb. Blood Flow Metab.* 2004, 24, 728–743. [CrossRef] [PubMed]
- 127. Zhang, W.; Xie, Y.; Wang, T.; Bi, J.; Li, H.; Zhang, L.Q.; Ye, S.Q.; Ding, S. Neuronal Protective Role of PBEF in a Mouse Model of Cerebral Ischemia. *J. Cereb. Blood Flow Metab.* **2010**, *30*, 1962–1971. [CrossRef] [PubMed]
- 128. Wang, X.; Zhang, Q.; Bao, R.; Zhang, N.; Wang, Y.; Polo-Parada, L.; Tarim, A.; Alemifar, A.; Han, X.; Wilkins, H.M.; et al. Deletion of Nampt in Projection Neurons of Adult Mice Leads to Motor Dysfunction, Neurodegeneration, and Death. *Cell Rep.* 2017, 20, 2184–2200. [CrossRef] [PubMed]
- 129. Conforti, L.; Gilley, J.; Coleman, M.P. Wallerian Degeneration: An Emerging Axon Death Pathway Linking Injury and Disease. *Nat. Rev. Neurosci.* **2014**, *15*, 394–409. [CrossRef] [PubMed]

- Loreto, A.; Di Stefano, M.; Gering, M.; Conforti, L. Wallerian Degeneration Is Executed by an NMN-SARM1-Dependent Late Ca2+ Influx butOnly Modestly Influenced by Mitochondria. *Cell Rep.* 2015, 13, 2539–2552. [CrossRef] [PubMed]
- 131. Liu, H.W.; Smith, C.B.; Schmidt, M.S.; Cambronne, X.A.; Cohen, M.S.; Migaud, M.E.; Brenner, C.; Goodman, R.H. Pharmacological Bypass of NAD(+) Salvage Pathway Protects Neurons from Chemotherapy-induced Degeneration. *Proc. Natl. Acad. Sci. USA* 2018, 115, 10654–10659. [CrossRef] [PubMed]
- 132. Araki, T.; Sasaki, Y.; Milbrandt, J. Increased Nuclear NAD Biosynthesis and SIRT1 Activation Prevent Axonal Degeneration. *Science* 2004, *305*, 1010–1013. [CrossRef] [PubMed]
- 133. Gilley, J.; Coleman, M.P. Endogenous Nmnat2 Is an Essential Survival factor for Maintenance of Healthy Axons. *PLoS Biol.* **2010**, *8*, e1000300. [CrossRef] [PubMed]
- 134. Kumar, A.; Singh, A.; Ekavali, E. A Review on Alzheimer's Disease Pathophysiology and Its Management: An Update. *Pharmacol. Rep.* **2015**, *67*, 195–203. [CrossRef] [PubMed]
- 135. Morris, M.C.; Evans, D.A.; Bienias, J.L.; Scherr, P.A.; Tangney, C.C.; Hebert, L.E.; Bennett, D.A.; Wilson, R.S.; Aggarwal, N. Dietary Niacin and Risk of Incident Alzheimer's Disease and of Cognitive Decline. *J. Neurol. Neurosurg. Psychiatry* 2004, 75, 1093–1099. [CrossRef] [PubMed]
- 136. Morris, M.C.; Schneider, J.A.; Tangney, C.C. Thoughts on B-vitamins and Dementia. J. Alzheimers Dis. 2006, 9, 429–433. [CrossRef] [PubMed]
- 137. Williams, A.C.; Hill, L.J.; Ramsden, D.B. Nicotinamide, NAD(P)(H), and Methylgroup Homeostasis Evolved and Became a Determinant of Ageing Diseases: Hypotheses and Lessons from Pellagra. *Curr. Gerontol. Geriatr. Res.* **2012**, 2012, 302875. [CrossRef]
- 138. Kerr, J.S.; Adriaanse, B.A.; Greig, N.H.; Mattson, M.P.; Cader, M.Z.; Bohr, V.A.; Fang, E.F. Mitophagy and Alzheimer's Disease: Cellular and Molecular Mechanisms. *Trends Neurosci.* 2017, 40, 151–166. [CrossRef] [PubMed]
- Liu, D.; Pitta, M.; Jiang, H.; Le, J.H.; Zhang, G.; Chen, X.; Kawamoto, E.M.; Mattson, M.P. Nicotinamide Forestalls Pathology and Cognitive Decline in Alzheimer Mice: Evidence for Improved Neuronal Bioenergetics and Autophagy Procession. *Neurobiol. Aging* 2013, *34*, 1564–1580. [CrossRef] [PubMed]
- 140. Wang, X.; Hu, X.; Yang, Y.; Takata, T.; Sakurai, T. Nicotinamide Mononucleotide Protects Against β-Amyloid Oligomer-Induced Cognitive Impairment and Neuronal Death. *Brain Res.* 2016, 1643, 1–9. [CrossRef] [PubMed]
- 141. Kim, E.J.; Yang, S.J. Nicotinamide Reduces Amyloid Precursor Protein and Presenilin 1 in Brain Tissues of Amyloid Beta-Tail Vein Injected Mice. *Clin. Nutr. Res.* **2017**, *6*, 130–135. [CrossRef] [PubMed]
- 142. Hou, Y.; Lautrup, S.; Cordonnier, S.; Wang, Y.; Croteau, D.L.; Zavala, E.; Zhang, Y.; Moritoh, K.; O'Connell, J.F.; Baptiste, B.A.; et al. NAD+ Supplementation Normalizes Key Alzheimer's Features and DNA Damage Responses in a New AD Mouse Model with Introduced DNA Repair Deficiency. *Proc. Natl. Acad. Sci. USA* 2018, 115, 1876–1885. [CrossRef] [PubMed]
- 143. Wencel, P.L.; Lukiw, W.J.; Strosznajder, J.B.; Strosznajder, R.P. Inhibition of poly(ADP-ribose) Polymerase-1 Enhances Gene Expression of Selected Sirtuins and APP Cleaving Enzymes in Amyloid Beta Cytotoxicity. *Mol. Neurobiol.* 2018, 55, 4612–4623. [CrossRef] [PubMed]
- 144. Cornelius, C.; Trovato Salinaro, A.; Scuto, M.; Fronte, V.; Cambria, M.T.; Pennisi, M.; Bella, R.; Milone, P.; Graziano, A.; Crupi, R.; et al. Cellular Stress Response, Sirtuins and UCP Proteins in Alzheimer Disease: Role of Vitagenes. *Immun. Ageing* **2013**, *10*, 41. [CrossRef] [PubMed]
- 145. Rizzi, L.; Roriz-Cruz, M. Sirtuin 1 and Alzheimer's Disease: An Up-To-Date Review. *Neuropeptides* **2018**, *71*, 54–60. [CrossRef] [PubMed]
- 146. Ljungberg, M.C.; Ali, Y.O.; Zhu, J.; Wu, C.S.; Oka, K.; Zhai, R.G.; Lu, H.C. CREB-Activity and NMNAT2 Transcription are Down-Regulated Prior to Neurodegeneration, while NMNAT2 Overexpression is Neuroprotective, in a Mouse Model of Human Tauopathy. *Hum. Mol. Genet.* 2012, 21, 251–267. [CrossRef] [PubMed]
- 147. Ali, Y.O.; Allen, H.M.; Yu, L.; Li-Kroeger, D.; Bakhshizadehmahmoudi, D.; Hatcher, A.; McCabe, C.; Xu, J.; Bjorklund, N.; Taglialatela, G.; et al. NMNAT2:HSP90 Complex Mediates Proteostasis in Proteinopathies. *PLoS Biol.* **2016**, *14*, e1002472. [CrossRef] [PubMed]
- 148. Wakade, C.; Chong, R.; Bradley, E.; Thomas, B.; Morgan, J. Upregulation of GPR109A in Parkinson's Disease. *PLoS ONE* **2014**, *9*, e109818. [CrossRef] [PubMed]

- 149. Aaseth, J.; Dusek, P.; Roos, P.M. Prevention of Progression in Parkinson's Disease. *Biometals* **2018**, *31*, 737–747. [CrossRef] [PubMed]
- 150. Anderson, D.W.; Bradbury, K.A.; Schneider, J.S. Broad Neuroprotective Profile of Nicotinamide in Different Mouse Models of MPTP-Induced Parkinsonism. *Eur. J. Neurosci.* **2008**, *28*, 610–617. [CrossRef] [PubMed]
- 151. Wakade, C.; Giri, B.; Malik, A.; Khodadadi, H.; Morgan, J.C.; Chong, R.K.; Baban, B. Niacin Modulates Macrophage Polarization in Parkinson's Disease. *J. Neuroimmunol.* **2018**, *320*, 76–79. [CrossRef] [PubMed]
- 152. Zhou, Y.; Wu, J.; Sheng, R.; Li, M.; Wang, Y.; Han, R.; Han, F.; Chen, Z.; Qin, Z.H. Reduced Nicotinamide Adenine Dinucleotide Phosphate Inhibits MPTP-Induced Neuroinflammation and Neurotoxicity. *Neuroscience* **2018**, *391*, 140–153. [CrossRef] [PubMed]
- 153. Parsons, R.B.; Smith, M.L.; Williams, A.C.; Ramsden, D.B. Expression of Nicotinamide N-Methyltransferase (E.C. 2.1.1.1) in the Parkinsonian Brain. *J. Neurol. Exp. Neurol.* **2002**, *61*, 111–124. [CrossRef]
- 154. Parsons, R.B.; Smith, S.W.; Waring, R.H.; Williams, A.C.; Ramsden, D.B. High Expression of Nicotinamide N-Methyltransferase in Patients with Idiopathic Parkinson's Disease. *Neurosci. Lett.* 2003, 342, 13–16. [CrossRef]
- 155. Williams, A.C.; Cartwright, L.S.; Ramsden, D.B. Parkinson's Disease: The First Common Neurological Disease due to Auto-Intoxication? *QJM* **2005**, *98*, 215–226. [CrossRef] [PubMed]
- 156. Thomas, M.G.; Saldanha, M.; Mistry, R.J.; Dexter, D.T.; Ramsden, D.B.; Parsons, R.B. Nicotinamide N-Methyltransferase Expression in SH-SY5Y Neuroblastoma and N27 Mesencephalic Neurones Induces Changes in Cell Morphology via Ephrin-B2 and Aktsignalling. *Cell Death Dis.* 2013, 4, 669. [CrossRef] [PubMed]
- 157. Singh, V.; Sharma, R.K.; Athilingam, T.; Sinha, P.; Sinha, N.; Thakur, A.K. NMR Spectroscopy-based Metabolomics of Drosophila Model of Huntington's Disease Suggests Altered Cell Energetics. J. Proteome Res. 2017, 16, 3863–3872. [CrossRef] [PubMed]
- 158. Ghosh, S.; Feany, M.B. Comparison of Pathways Controlling Toxicity in the Eye and Brain in Drosophila Models of Human Neurodegenerative Diseases. *Hum. Mol. Genet.* **2004**, *13*, 2011–2018. [CrossRef] [PubMed]
- 159. Hathorn, T.; Snyder-Keller, A.; Messer, A. Nicotinamide Improves Motor Deficits and Upregulates PGC-1α and BDNF Gene Expression in a Mouse Model of Huntington's Disease. *Neurobiol. Dis.* 2011, 41, 43–50. [CrossRef] [PubMed]
- 160. Sidhu, A.; Diwan, V.; Kaur, H.; Bhateja, D.; Singh, C.K.; Sharma, S.; Padi, S.S.V. Nicotinamide Reverses Behavioral Impairments and Provides Neuroprotection in 3-Nitropropionic Acid Induced Animal Model of Huntington's Disease: Implication of Oxidative Stress- Poly(ADP- Ribose) Polymerase Pathway. *Metab. Brain Dis.* 2018, 33, 1911–1921. [CrossRef] [PubMed]
- Chidambaram, S.B.; Vijayan, R.; Sekar, S.; Mani, S.; Rajamani, B.; Ganapathy, R. Simultaneous Blockade of NMDA Receptors and PARP-1 Activity Synergistically Alleviate Immunoexcitotoxicity and Bioenergetics in 3-Nitropropionic Acid Intoxicated Mice: Evidences from Memantine and 3-Aminobenzamide Interventions. *Eur. J. Pharmacol.* 2017, *803*, 148–158. [CrossRef] [PubMed]
- 162. Suzuki, E.; Okuda, H.; Nishida, K.; Fujimoto, S.; Nagasawa, K. Protective Effect of Nicotinamide Against Poly(ADP-Ribose) Polymerase-1-Mediated Astrocyte Death Depends on Its Transporter-Mediated Uptake. *Life Sci.* 2010, *86*, 676–682. [CrossRef] [PubMed]
- 163. Sadri-Vakili, G.; Cha, J.H. Histone Deacetylase Inhibitors: A Novel Therapeutic Approach to Huntington's Disease (Complex Mechanism of Neuronal Death). *Curr. Alzheimer Res.* 2006, *3*, 403–408. [CrossRef] [PubMed]
- 164. Pallos, J.; Bodai, L.; Lukacsovich, T.; Purcell, J.M.; Steffan, J.S.; Thompson, L.M.; Marsh, J.L. Inhibition of Specific HDACs and Sirtuins Suppresses Pathogenesis in a Drosophila Model of Huntington's Disease. *Hum. Mol. Genet.* 2008, 17, 3767–3775. [CrossRef] [PubMed]
- Verdin, E. NAD+ in Aging, Metabolism, and Neurodegeneration. *Science* 2015, 350, 1208–1213. [CrossRef]
 [PubMed]
- Bonkowski, M.S.; Sinclair, D.A. Slowing Ageing by Design: The Rise of NAD. *Nat. Rev. Mol. Cell Biol.* 2016, 17, 679–690. [CrossRef] [PubMed]
- 167. Jęśko, H.; Wencel, P.; Strosznajder, R.P.; Strosznajder, J.B. Sirtuins and Their Roles in Brain Aging and Neurodegenerative Disorders. *Neurochem. Res.* **2017**, *42*, 876–890. [CrossRef] [PubMed]
- 168. Williams, A.C.; Ramsden, D.B. Autotoxicity, Methylation and a Road to the Prevention of Parkinson's Disease. *J. Clin. Neurosci.* 2005, *12*, 6–11. [CrossRef] [PubMed]

- Lehmann, S.; Loh, S.H.; Martins, L.M. Enhancing NAD(+) Salvage Metabolism is Neuroprotective in a PINK1 Model of Parkinson's Disease. *Biol. Open* 2017, *6*, 141–147. [CrossRef] [PubMed]
- 170. Testa, C.M.; Jankovic, J. Huntington Disease: A Quarter Century of Progress Since the Gene Discovery. J. Neurol. Sci. 2019, 396, 52–68. [CrossRef] [PubMed]
- 171. McColgan, P.; Tabrizi, S.J. Huntington's Disease: A Clinical Review. *Eur. J. Neurol.* **2018**, 25, 24–34. [CrossRef] [PubMed]
- 172. Tulino, R.; Benjamin, A.C.; Jolinon, N.; Smith, D.L.; Chini, E.N.; Carnemolla, A.; Bates, G.P. SIRT1 Activity Is Linked to Its Brain Region-Specific Phosphorylation and Is Impaired in Huntington's Disease Mice. *PLoS ONE* **2016**, *11*, e0145425. [CrossRef]
- 173. Naia, L.; Rosenstock, T.R.; Oliveira, A.M.; Oliveira-Sousa, S.I.; Caldeira, G.L.; Carmo, C.; Laço, M.N.; Hayden, M.R.; Oliveira, C.R.; Rego, A.C. Comparative Mitochondrial-Based Protective Effects of Resveratrol and Nicotinamide in Huntington's Disease Models. *Mol. Neurobiol.* 2017, 54, 5385–5399. [CrossRef] [PubMed]
- 174. Harrison, I.F.; Powell, N.M.; Dexter, D.T. The Histone Deacetylase Inhibitor Nicotinamide Exacerbates Neurodegeneration in the Lactacystin Rat Model of Parkinson's Disease. J. Neurochem. 2019, 148, 136–156. [CrossRef] [PubMed]
- 175. Hoane, M.R.; Akstulewicz, S.L.; Toppen, J. Treatment with Vitamin B3 Improves Functional Recovery and Reduces GFAP Expression Following Traumatic Brain Injury in Rats. J. Neurotrauma 2003, 20, 1189–1199. [CrossRef] [PubMed]
- 176. Goffus, A.M.; Anderson, G.D.; Hoane, M. Sustained Delivery of Nicotinamide Limits Cortical Injury and Improves Functional Recovery Following Traumatic Brain Injury. Oxid. Med. Cell. Longev. 2010, 3, 145–152. [CrossRef] [PubMed]
- 177. Vonder Haar, C.; Maas, W.R.; Jacobs, E.A.; Hoane, M.R. Deficits in Discrimination after Experimental Frontal Brain Injury Are Mediated by Motivation and Can Be Improved by Nicotinamide Administration. *J. Neurotrauma* 2014, *31*, 1711–1720. [CrossRef] [PubMed]
- 178. Vonder Haar, C.; Anderson, G.D.; Hoane, M.R. Continuous Nicotinamide Administration Improves Behavioral Recovery and Reduces Lesion Size Following Bilateral Frontal Controlled Cortical Impact Injury. *Behav. Brain Res.* 2011, 224, 311–317. [CrossRef] [PubMed]
- Won, S.J.; Choi, B.Y.; Yoo, B.H.; Sohn, M.; Ying, W.; Swanson, R.A.; Suh, S.W. Prevention of Traumatic Brain Injury-Induced Neuron Death by Intranasal Delivery of Nicotinamide Adenine Dinucleotide. *J. Neurotrauma* 2012, 29, 1401–1409. [CrossRef] [PubMed]
- Swan, A.A.; Chandrashekar, R.; Beare, J.; Hoane, M.R. Preclinical Efficacy Testing in Middle-Aged Rats: Nicotinamide, a Novel Neuroprotectant, Demonstrates Diminished Preclinical Efficacy after Controlled Cortical Impact. J. Neurotrauma 2011, 28, 431–440. [CrossRef] [PubMed]
- 181. Kwon, W.Y.; Suh, G.J.; Kim, K.S.; Lee, H.J.; Jeong, K.Y.; Kwak, Y.H.; Kim, K. Niacin Suppresses the Mitogen-Activated Protein Kinase Pathway and Attenuates Brain Injury after Cardiac Arrest in Rats. *Crit. Care Med.* 2013, 41, e223–e232. [CrossRef] [PubMed]
- Park, J.H.; Long, A.; Owens, K.; Kristian, T. Nicotinamide Mononucleotide Inhibits Post-Ischemic NAD(+) Degradation and Dramatically Ameliorates Brain Damage Following Global Cerebral Ischemia. *Neurobiol. Dis.* 2016, 95, 102–110. [CrossRef] [PubMed]
- 183. Shetty, P.K.; Galeffi, F.; Turner, D.A. Nicotinamide Pre-treatment Ameliorates NAD(H) Hyperoxidation and Improves Neuronal Function after Severe Hypoxia. *Neurobiol. Dis.* **2014**, *62*, 469–478. [CrossRef] [PubMed]
- Chen, J.; Cui, X.; Zacharek, A.; Jiang, H.; Roberts, C.; Zhang, C.; Lu, M.; Kapke, A.; Feldkamp, C.S.; Chopp, M. Niaspan Increases Angiogenesis and Improves Functional Recovery after Stroke. *Ann. Neurol.* 2007, 62, 49–58. [CrossRef] [PubMed]
- 185. Chen, J.; Cui, X.; Zacharek, A.; Ding, G.L.; Shehadah, A.; Jiang, Q.; Lu, M.; Chopp, M. Niaspan Treatment Increases Tumor Necrosis Factor-Alpha-Converting Enzyme and Promotes Arteriogenesis after Stroke. *J. Cereb. Blood Flow Metab.* 2009, 29, 911–920. [CrossRef] [PubMed]
- 186. Kwon, W.Y.; Suh, G.J.; Kim, K.S.; Jung, Y.S.; Kim, S.H.; Lee, A.R.; You, K.M.; Park, M.J. Niacin and Selenium Attenuate Brain Injury After Cardiac Arrest in Rats by Up-Regulating DJ-1-Akt Signaling. *Crit. Care Med.* 2018, 46, e788–e796. [CrossRef] [PubMed]
- 187. Peterson, T.C.; Anderson, G.D.; Kantor, E.D.; Hoane, M.R. A Comparison of the Effects of Nicotinamide and Progesterone on Functional Recovery of Cognitive Behavior Following Cortical Contusion Injury in the Rat. *J. Neurotrauma* 2012, 29, 2823–2830. [CrossRef] [PubMed]

- 188. Peterson, T.C.; Hoane, M.R.; McConomy, K.S.; Farin, F.M.; Bammler, T.K.; MacDonald, J.W.; Kantor, E.D.; Anderson, G.D. A Combination Therapy of Nicotinamide and Progesterone Improves Functional Recovery following Traumatic Brain Injury. J. Neurotrauma 2015, 32, 765–779. [CrossRef] [PubMed]
- 189. Wang, P.; Xu, T.Y.; Guan, Y.F.; Tian, W.W.; Viollet, B.; Rui, Y.C.; Zhai, Q.W.; Su, D.F.; Miao, C.Y. Nicotinamide Phosphoribosyltransferase Protects against Ischemic Stroke through SIRT1-Dependent Adenosine Monophosphate-Activated Kinase Pathway. Ann. Neurol. 2011, 69, 360–374. [CrossRef] [PubMed]
- Zhang, T.; Kraus, W.L. SIRT1-Dependent Regulation of Chromatin and Transcription: Linking NAD(+) Metabolism and Signaling to the Control of Cellular Functions. *Biochim. Biophys. Acta* 2010, 1804, 1666–1675. [CrossRef] [PubMed]
- 191. Jing, Z.; Xing, J.; Chen, X.; Stetler, R.A.; Wen, Z.; Gan, Y.; Zhang, F.; Gao, Y.; Chen, J.; Leak, R.K.; et al. Neuronal NAMPT is Released after Cerebral Ischemia and Protects against White Matter Injury. J. Cereb. Blood Flow Metab. 2014, 34, 1613–1621. [CrossRef] [PubMed]
- 192. Hardison, W.R.; Yorns, H.H., Jr. Mitochondrial Dysfunction in Migraine. *Semin. Pediatr. Neurol.* 2013, 20, 188–193.
- 193. Neri, M.; Frustaci, A.; Milic, M.; Valdiglesias, V.; Fini, M.; Bonassi, S.; Barbanti, P. A Meta-Analysis of Biomarkers Related to Oxidative Stress and Nitric Oxide Pathway in Migraine. *Cephalalgia* 2015, 35, 931–937. [CrossRef] [PubMed]
- Bohár, Z.; Párdutz, Á.; Vécsei, L. Tryptophan Catabolites and Migraine. *Curr. Pharm. Des.* 2016, 22, 1013–1021.
 [CrossRef] [PubMed]
- 195. Morrow, J.D.; Awad, J.A.; Oates, J.A.; Roberts, L.J. Identification of Skin as a Major Site on Prostaglandin D2 Release Following Oral Administration of Niacin in Humans. J. Investig. Dermatol. 1992, 98, 812–815. [CrossRef] [PubMed]
- 196. Morrow, J.D.; Parsons, W.G.; Roberts, L., II. Release of Markedly Increased Quantities of Prostaglandin D2 in Vivo in Humans Following the Administration of Nicotinic Acid. *Prostaglandins* **1989**, *38*, 263–274. [CrossRef]
- 197. Kim, E.J.; Lim, S.Y.; Lee, H.J.; Lee, J.Y.; Choi, S.; Kim, S.Y.; Kim, J.M.; Shin, I.S.; Yoon, J.S.; Yang, S.J.; et al. Low Dietary Intake of n-3 Fatty Acids, Niacin, Folate, and Vitamin C in Korean Patients with Schizophrenia and the Development of Dietary Guidelines for Schizophrenia. *Nutr. Res.* **2017**, *45*, 10–18. [CrossRef] [PubMed]
- 198. Cao, B.; Sun, X.Y.; Zhang, C.B.; Yan, J.J.; Zhao, Q.Q.; Yang, S.Y.; Yan, L.L.; Huang, N.H.; Zeng, J.; Liao, J.Y.; et al. Association Between B Vitamins and Schizophrenia: A Population-Based Case-Control Study. *Psychiatry Res.* 2018, 259, 501–505. [CrossRef] [PubMed]
- 199. Jonsson, B.H. Nicotinic Acid Long-Term Effectiveness in a Patient with Bipolar Type II Disorder: A Case of Vitamin Dependency. *Nutrients* **2018**, *10*, 134. [CrossRef] [PubMed]
- 200. Ryou, M.G.; Mallet, R.T. An In Vitro Oxygen-Glucose Deprivation Model for Studying Ischemia-Reperfusion Injury of Neuronal Cells. *Methods Mol. Biol.* **2018**, 1717, 229–235. [CrossRef]
- 201. Martin, E.; Rosenthal, R.E.; Fiskum, G. Pyruvate Dehydrogenase Complex: Metabolic Link to Ischemic Brain Injury and Target of Oxidative Stress. *J. Neurosci. Res.* **2005**, *79*, 240–247. [CrossRef] [PubMed]
- 202. Mokudai, T.; Ayoub, I.A.; Sakakibara, Y.; Lee, E.J.; Ogilvy, C.S.; Maynard, K.I. Delayed Treatment with Nicotinamide (Vitamin B3) Improves Neurological Outcome and Reduces Infarct Volume after Transient Focal Cerebral Ischemia in Wistar Rats. *Stroke* 2000, *31*, 1679–1685. [CrossRef] [PubMed]
- 203. Ayoub, I.A.; Lee, E.J.; Ogilvy, C.S.; Beal, M.F.; Maynard, K.I. Nicotinamide Reduces Infarction up to Two Hours after the Onset of Permanent Focal Ischemia in Wistar Rats. *Neurosci. Lett.* **1999**, 259, 21–24. [CrossRef]
- 204. Sakakibara, Y.; Mitha, A.P.; Ogilvy, C.S.; Maynard, K.I. Post-Treatment with Nicotinamide (Vitamin B3) Reduces the Infarct Volume Following Permanent Focal Ischemia in Female Sprague–Dawley and Wistar Rats. *Neurosci. Lett.* 2000, 281, 111–114. [CrossRef]
- 205. Sakakibara, Y.; Mitha, A.P.; Ayoub, I.A.; Ogilvy, C.S.; Maynard, K.I. Delayed Treatment with Nicotinamide (Vitamin B3) Reduces the Infarct Volume Following Focal Cerebral Ischemia in Spontaneously Hypertensive Rats, Diabetic and Non-Diabetic Fischer 344 Rats. *Brain Res.* 2002, *931*, 68–73. [CrossRef]
- Feng, Y.; Paul, I.A.; LeBlanc, M.H. Nicotinamide Reduces Hypoxic Ischemic Brain Injury in the Newborn Rat. Brain Res. Bull. 2006, 69, 117–122. [CrossRef] [PubMed]
- 207. Rizzoli, P.; Mullally, W.J. Headache. Am. J. Med. 2018, 131, 17-24. [CrossRef] [PubMed]
- 208. Close Headache Classification Subcommittee of the International Headache Society. The International Classification of Headache Disorders: 3rd Edition. *Cephalalgia* **2013**, *33*, 629–808. [CrossRef] [PubMed]

- Nattagh-Eshtivani, E.; Sani, M.A.; Dahri, M.; Ghalichi, F.; Ghavami, A.; Arjang, P.; Tarighat-Esfanjani, A. The Role of Nutrients in the Pathogenesis and Treatment of Migraine Headaches: Review. *Biomed. Pharmacother.* 2018, 102, 317–325. [CrossRef] [PubMed]
- 210. Goldzieher, J.W.; Popkin, G.L. Treatment of Headache with Intravenous Sodium Nicotinate. J. Am. Med. Assoc. 1946, 131, 103–105. [CrossRef] [PubMed]
- 211. Grenfell, R.F. Treatment of Migraine with Nicotinic Acid. Am. Pract Dig. Treat. 1949, 3, 542–544. [PubMed]
- 212. Grenfell, R.F. Treatment of Tension Headache. Am. Pract. Dig. Treat. 1951, 2, 933–936. [PubMed]
- 213. Morgan, Z.R. Nicotinic Acid Therapy in Vasoconstriction Type of Headache. *Md. State Med. J.* **1953**, 2, 377–382. [PubMed]
- 214. Morgan, Z.R. A Newer Method of Nicotinic Acid Therapy in Headache of the Vasoconstrictive Type. J. Am. *Geriatr. Soc.* **1955**, *3*, 545–551. [CrossRef] [PubMed]
- 215. Prousky, J.; Seely, D. The Treatment of Migraines and Tension-Type Headaches with Intravenous and Oral Niacin (Nicotinic Acid): Systematic Review of the Literature. *Nutr. J.* **2005**, *4*, 3. [CrossRef] [PubMed]
- 216. Oxenkrug, G.F. Tryptophan Kynurenine Metabolism as a Common Mediator of Genetic and Environmental Impacts in Major Depressive Disorder: The Serotonin Hypothesis Revisited 40 Years Later. *Isr. J. Psychiatry Relat. Sci.* **2010**, 47, 56–63. [CrossRef] [PubMed]
- 217. Myint, A.M.; Schwarz, M.J.; Müller, N. The Role of the Kynurenine Metabolism in Major Depression. *J. Neural Transm.* **2012**, *119*, 245–251. [CrossRef] [PubMed]
- 218. Wonodi, I.; Schwarcz, R. Cortical Kynurenine Pathway Metabolism: A Novel Target for Cognitive Enhancement in Schizophrenia. *Schizophr. Bull.* **2010**, *36*, 211–218. [CrossRef] [PubMed]
- 219. Miller, C.L.; Llenos, I.C.; Cwik, M.; Walkup, J.; Weis, S. Alterations in Kynurenine Precursor and Product Levels in Schizophrenia and Bipolar Disorder. *Neurochem. Int.* **2008**, *52*, 1297–1303. [CrossRef] [PubMed]
- 220. Savitz, J.; Drevets, W.C.; Smith, C.M.; Victor, T.A.; Wurfel, B.E.; Bellgowan, P.S.; Bodurka, J.; Teague, T.K.; Dantzer, R. Putative Neuroprotective and Neurotoxic Kynurenine Pathway Metabolites are Associated with Hippocampal and Amygdalar Volumes in Subjects with Major Depressive Disorder. *Neuropsychopharmacology* 2015, 40, 463–471. [CrossRef] [PubMed]
- 221. Kanai, M.; Funakoshi, H.; Takahashi, H.; Hayakawa, T.; Mizuno, S.; Matsumoto, K.; Nakamura, T. Tryptophan 2,3-Dioxygenase is a Key Modulator of Physiological Neurogenesis and Anxiety-Related Behavior in Mice. *Mol. Brain* 2009, 2, 8. [CrossRef] [PubMed]



© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).