

Natural Modulators of Key Signaling Pathways in Skin Inflammageing

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Abstract: Low-grade chronic inflammation without obvious infection is defined as “inflammageing” and a key driver of skin ageing. Although the importance of modulating inflammageing for treating skin diseases and restoring cutaneous homeostasis is increasingly being recognized. However, the mechanisms underlying skin inflammageing, particularly those associated with natural treatments, have not been systematically elucidated. This review explores the signaling pathways associated with skin inflammageing, as well as the natural plants and compounds that directly or indirectly target these pathways. Nine signaling pathways and 60 plants/constituents related to skin anti-inflammageing are discussed, exploring plant mechanisms to mitigate skin inflammageing. Common natural plants with anti-inflammageing activity are detailed by active ingredients, mechanisms, therapeutic potential, and quantitative effects on skin inflammageing modulation. This review strengthens our understanding of these botanical ingredients as natural interventions against skin inflammageing and provides directions for future research.

Keywords: cutaneous homeostasis, inflammageing, plant extract, skin ageing

Introduction

Age-associated sterile, chronic, and low-grade inflammation is commonly termed inflammageing, and is a marker of biological ageing, multimorbidity, and mortality risk.¹ Inflammageing occurs in all organs of our body, and the skin is the most intuitive manifestation. Skin becomes increasingly inflamed with age, with abundant cell inflammation accelerating ageing of the skin, leading to wrinkles, sagging skin, and possibly cancer.² Skin inflammageing is traditionally considered detrimental to human health because of its association with pro-inflammatory factor accumulation and various skin issues. However, from an evolutionary standpoint, inflammageing is increasingly seen as an adaptive or remodeling process.³ Thus, targeting the ageing immune system for immune rejuvenation could be a strategic approach to preserve skin homeostasis and enhance immune-inflammatory functions. Balancing pro-inflammageing and anti-inflammageing states is crucial to effectively address inflammatory skin ageing. As such, understanding the specific mechanisms of skin inflammageing will improve our ability to facilitate healthy skin ageing. External factors such as UV radiation, particulate matter (PM_{2.5}), and seasonal changes all exacerbate inflammageing. Cellular senescence, inflammasome dysregulation, mitochondrial dysfunction, autophagy, ubiquitin-proteasome system impairment, and DNA damage responses also contribute to skin inflammageing.⁴ These processes are governed by several signaling pathways, including NF- κ B/RIG-I, mTOR, TGF- β , AMPK-SIRT6, complement system, NLRP3, Notch, JAK-STAT, and MAPK pathways (Figure 1).^{5–11} However, the pathways that affect skin inflammageing have not yet been systematically summarized.

Botanicals such as quercetin, fisetin, and resveratrol exert anti-inflammatory and anti-ageing benefits.^{12,13} Botanical extract-based skincare is recommended by physicians to combat inflammageing.¹⁴ Thus, natural plant extracts or monomers that target inflammageing pathways show potential as effective preventive strategies.¹⁵ This comprehensive review explores research from the past 13 years available from PubMed and ScienceDirect to update our understanding of skin inflammageing pathways and identify significant medicinal plant extracts or monomers with potential anti-inflammageing effects on the skin.

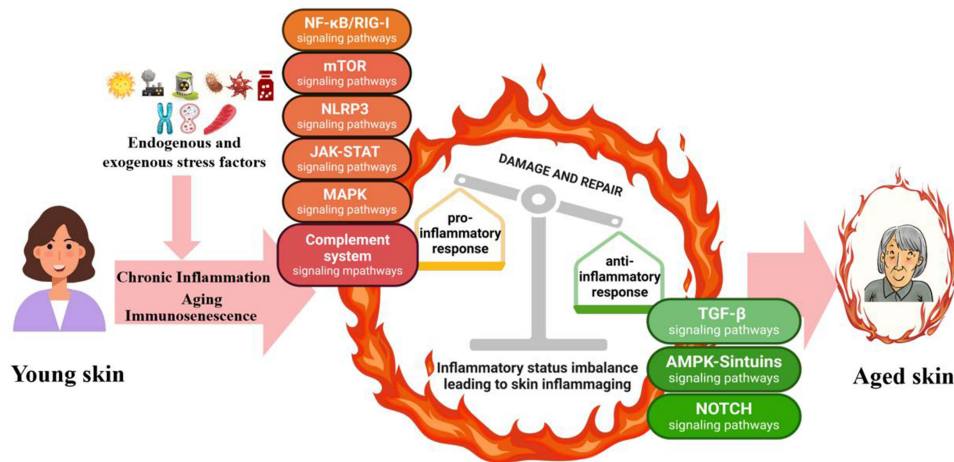


Figure 1 Cellular signaling pathways involved in the regulation of skin inflammaging. Imbalances in skin-related inflammation leads to skin aging.
Notes: Created in BioRender. Ren, Q. (2024) <https://BioRender.com/p661768>.

Despite some plant chemical properties remaining uncharacterized, their pharmacological evaluation has supported the advancement of skincare products sourced from plants with clinically proven anti-inflammaging benefits.

Role of Signaling Pathways in Skin Inflammaging

The main features of senescent cells are manifested in the senescence-associated secretory phenotype (SASP). Skin inflammaging occurs when senescent cells accumulate in the skin and release SASP factors, including IL-(1β, -6, -8, -18), TNF-α and CRP; these cells include keratinocytes, fibroblasts, melanocytes, and immune cells⁴ (Figure 2). Typically, a less than two-fold

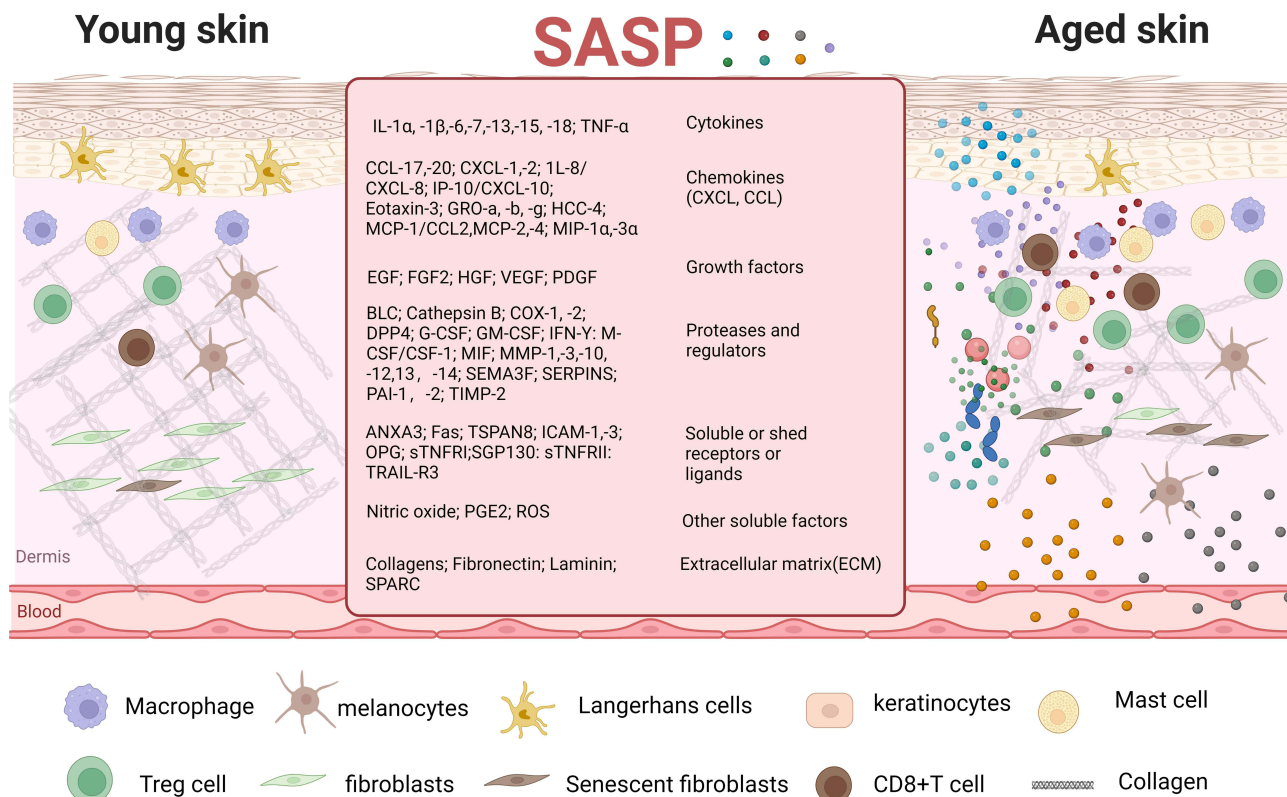


Figure 2 Cells and molecules involved in skin inflammaging. In the process of skin inflammaging, keratinocytes, fibroblasts, melanocytes, and immune cells secrete SASP. SASP further causes aging of surrounding young cells, forming a vicious cycle.
Notes: Created in BioRender. Ren, Q. (2024) <https://BioRender.com/q47v111>.

increase in pro-inflammatory mediators is a positive response by the skin that maintains homeostatic stability, known as cold-inflammageing. However, continuous exposure to external stressors such as UV light and pollution can promote senescence and inflammatory phenotypes in skin.¹⁶ Furthermore, disruption of the skin's homeostatic balance increases the formation of cytokines (two- to four-fold increase).¹⁷ This review involved an extensive literature search, followed by integration of the findings regarding the signaling pathways related to skin inflammageing, including NF- κ B/RIG-I, mTOR, TGF- β , AMPK-SIRT6, Notch, complement system, NLRP3, JAK-STAT, and MAPK, to elucidate the mechanisms of inflammageing (Figure 3).

Role of the NF- κ B/RIG-I Pathway

NF- κ B activation, driven by ageing factors such as ROS, senescent cells, and DNA damage, leads to increased inflammatory cytokine production (TNF- α , IL-1 β , IL-6, and IL-8), further amplifying inflammatory and age-related disturbances through classical pathway activation.¹⁸ The trans-activation motifs of NF- κ B are key regulators associated with ageing across human tissues, and persistent NF- κ B signaling activation fosters cellular senescence.¹⁹ SASP produced by senescent cells necessitates NF- κ B signaling to impact adjacent youthful cells.²⁰ RIG-I, a pattern recognition receptor in the skin, identifies viral pathogen-associated molecular patterns,²¹ and its blockade mitigates senescence-associated inflammation, extending cell growth.²² RIG-I is upregulated in senescent cells via the ATM-IRF1 axis, which mediates IL-6 and IL-8 expression.²³ Klotho, by interacting with RIG-I, inhibits induced IL-6 and IL-8 expression, acting as an anti-ageing agent by dampening RIG-I-mediated inflammation.²⁴ Loss of the ageing-related protein Klotho triggers the NF- κ B/RIG-I pathway, enhancing pro-inflammatory mediator production.²⁵ Furthermore, RIG-I promotes keratinocyte proliferation and wound repair by inducing tissue inhibitor of metalloprotease-1 (TIMP-1) expression through NF- κ B signaling pathway.²³ NF- κ B/RIG-I activation driven by ageing factors therefore increases inflammatory cytokine production and affects skin ageing.¹⁵

Role of the mTOR Signaling Pathway

The mTOR regulates cell growth, metabolism, and nutrient signaling, with hyperactivation often linked to inflammation.²⁶ This pathway comprises two complexes: mTORC1 is vital for regulating cell growth and substrate phosphorylation to increase anabolism (eg, mRNA translation, and lipid synthesis) or inhibit catabolism (eg, autophagy); mTORC2 promotes cell survival and cytoskeletal organization via Akt activation, with both complexes contributing to inflammageing (Figure 3a).²⁷ Overexpression of the upstream mTOR modulators (eg, PI3Ks, Akt, TNF- α , Ras, Raf kinase, MEK, ERK1/2, and Rsk) escalates mTORC1 activity, positioning mTOR at the center of inflammageing processes.²⁶ mTOR overactivation induces cellular senescence, whereas inhibition through rapamycin or genetic interventions halts senescence, attenuates SASP, and prolongs the lifespan of mice.^{28,29} Thus, inhibiting the PI3K/AKT/mTOR or RAS/ERK/mTOR pathways and restoring autophagy may decelerate skin cell senescence. Consequently, the mTOR signaling pathway, and especially its imbalances, affect skin inflammation and cell ageing. Therefore, maintaining balanced mTOR signaling is crucial for skin health.

Role of the TGF- β Signaling Pathway

TGF- β , a multifaceted cytokine, regulates cell growth, differentiation, apoptosis, and cellular homeostasis.³⁰ TGF- β signaling is crucial for modulations to the skin inflammageing process. TGF- β 1 is secreted in a latent form, activated when the LAP is cleaved by mediators such as matrix metalloproteinase (MMP)2/MMP9, ROS, and integrins.³¹ This pathway is divided into classical and non-classical routes. Non-classical signaling pathways include interactions with RAS, MAP3K7, Rho/Rock, and PI3K/AKT.³² Activation of TGF- β /Ras promotes cellular ageing, cell cycle arrest, and increased expression of pro-inflammatory cytokines and chemokines (IL-6, -1 β , -8, -1 α , and MCP).³³ Thus, signaling through TGF- β /Ras and its downstream effectors exacerbates skin inflammageing. Moreover, Ras signaling may be involved in skin inflammageing in animals.³⁴ TGF- β activates MAP3K7 (also known as TAK1) to stimulate the NF- κ B pathway, thereby exacerbating skin inflammageing.³⁵ TGF- β also stimulates the Rho/ROCK pathway to promote pro-inflammatory factor release. Furthermore, TGF- β activates PI3K/AKT to stimulate the mTOR pathway, thereby exacerbating skin inflammageing. Thus, TGF- β non-classical pathway activation primarily affects other factors to subsequently aggravate skin inflammageing. Furthermore, as part of the SASP, TGF- β contributes to maintaining the senescent phenotype and age-related pathologies.³⁰

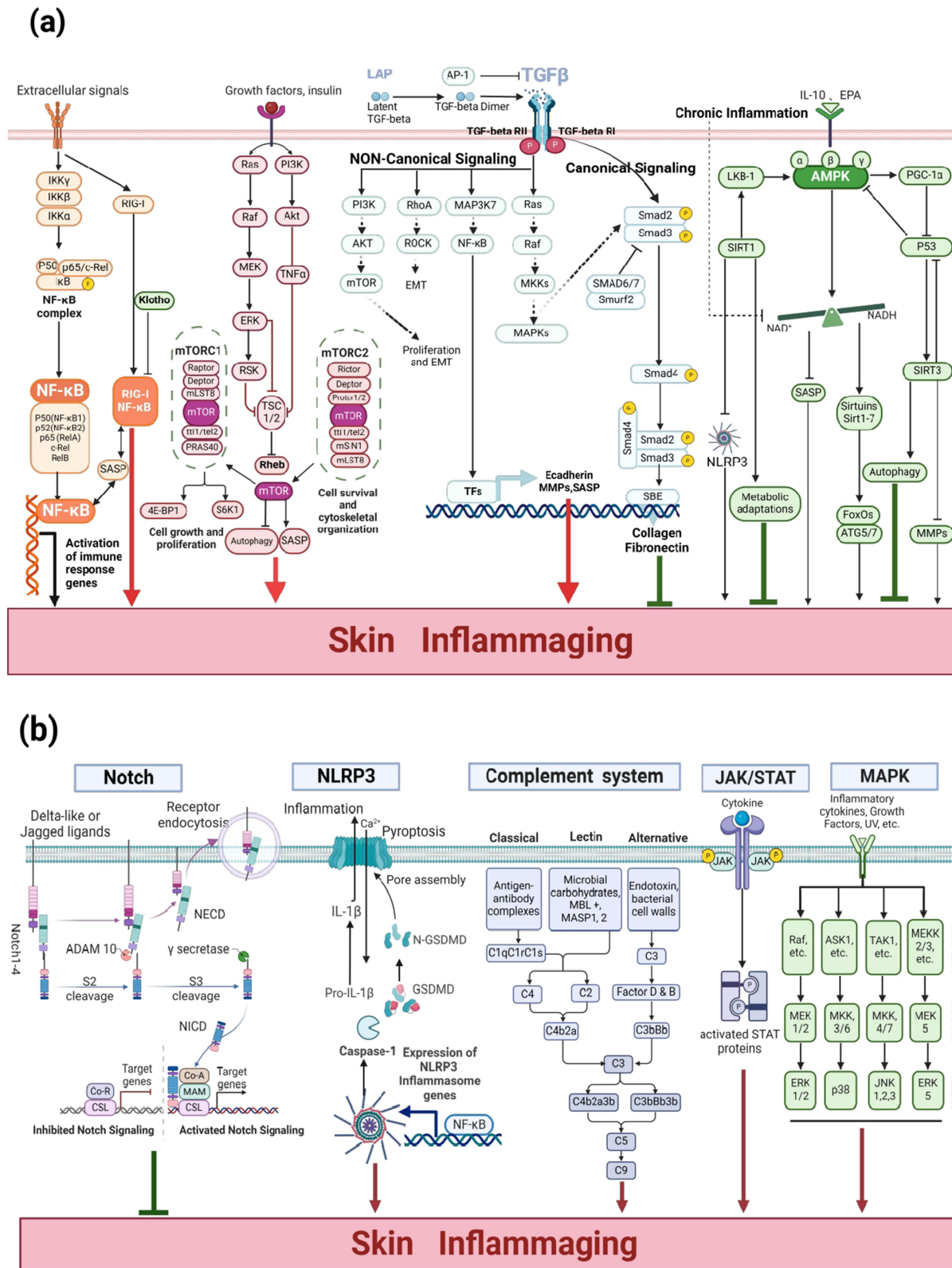


Figure 3 (a) The core regulatory pathways involved skin inflammaging (including NF- κ B/RIG-I, mTOR, TGF- β , AMPK-SIRTs pathways). Created in BioRender. Ren, Q. (2024) <https://BioRender.com/s821596>. (b) The other regulatory pathways of skin inflammaging (including complement system, NLRP3, Notch, JAK-STAT, and MAPK pathways). Created in BioRender. Ren, Q. (2024) <https://BioRender.com/h71i391>.

The core function of classical TGF- β signaling in the skin is the induction of type I pre-collagen synthesis and connective tissue growth factor secretion.³⁶ Classically, TGF- β initiates signaling by binding to TGF- β RI and TGF- β RII, leading to Smad2 and Smad3 phosphorylation.³⁷ These phosphorylated Smads form a complex with Smad4, which then regulates gene expression in the nucleus through Smad binding elements. Ageing impacts TGF- β signaling, with decreased TGF- β RII expression in aged dermal fibroblasts disrupting the TGF- β pathway, thereby reducing collagen synthesis and increasing degradation, ultimately leading to skin thinning.^{32,36} TGF- β signaling impairment and TGF- β ligand upregulation results in cellular inflammation.³⁰ Thus, classical TGF- β pathway activation promotes type I collagen production via the Smad pathway, thereby inhibiting skin inflammaging (Figure 3a). In summary, TGF- β is involved in skin inflammaging through classical and non-canonical pathways, and targeting the TGF- β signaling pathway could help modulate the inflammaging phenotype.

Role of the AMPK–SIRT1 Pathway

AMPK is crucial for maintaining cellular energy homeostasis and sensing metabolic stress, it regulates inflammation, oxidative stress, autophagy, and other pathways.³⁸ AMPK enzyme activity declines in aged skin³⁹ and is lost in keratinocytes after acute injury or UVB exposure, which leads to the hyperproliferation and activation of mTOR signaling.⁴⁰ Metformin, a diabetes medication and anti-ageing agent, utilizes AMPK to curb stress-induced cellular proliferation⁴⁰ and modulates mitochondrial function, reducing age-related inflammation.⁴¹ AMPK can regulate the AMPK/SIRT1 and AMPK/mTOR pathways, which relieve neuropathic pain and inflammatory pain.⁴² Furthermore, activation of AMPK enhances the biosynthesis and/or stability of NAD⁺. This augmentation in NAD⁺ concentration subsequently serves as a critical signal to induce the expression of Sirt1.⁴³ Sirt1, in turn, deacetylates LKB-1, an AMPK activator, forming a positive feedback loop.⁴⁴ In terms of functionality, activation of AMPK can suppress mTORC1 through activating TSC1/2, and it also prompts Sirtuins to catalyze the deacetylation of FoxO proteins or autophagy-associated proteins, namely ATG5 and ATG7,⁴⁵ resulting in an upregulation of PGC-1 α expression which improves mitochondrial biogenesis, and cytoplasmic p53 inhibition, which facilitates autophagy.⁴³ Sirtuins, comprising seven proteins (SIRT1–SIRT7), play a pivotal role in postponing cellular ageing and augmenting organismal lifespan through the regulation of various cellular mechanisms.⁴⁶ Age-related metabolic inflammation depletes NAD⁺, which subsequently reduces Sirt activity. The absence of Sirt1 diminishes mitochondrial biogenesis, while the depletion of mitochondrial Sirt3 weakens mitochondrial antioxidant defenses and repair mechanisms. Additionally, a reduction in NAD⁺ levels can hinder the function of Sirt2 and elevate the activity of the NLRP3 inflammasome.⁴⁷ Thus, inflammation may induce a positive feedback loop by altering the Sirtuin function, which promotes inflammaging. SIRT1 deacetylase-activated keratinocytes counteract the inflammatory response induced by LPS⁴⁸ (Figure 3a). Thus, AMPK–SIRT1 pathway activation could help resist inflammaging damage to the skin.

Role of Other Pathways

The mechanisms underlying skin inflammaging are intricate and involve activating NF- κ B/RIG-I, mTOR, and TGF- β non-classical signaling pathways, which exacerbate inflammation and accelerate skin ageing. Conversely, AMPK–SIRT1 and TGF- β classical signaling pathway activation could inhibit inflammaging damage. Several additional signaling pathways have been identified, including Notch, the complement system, and the NLRP3, JAK–STAT, and MAPK signaling pathways; however, relatively little evidence exists for their direct regulation of skin inflammaging and fewer modulators appear to act primarily via these pathways.

The process of epidermal differentiation and proliferation is orchestrated by Notch signaling.⁴⁹ The Notch pathway may regulate various age-related skin diseases characterized by chronic inflammation.⁵⁰ In healthy human skin, Notch receptors and ligands are abundantly expressed in epidermal keratinocytes. Notch signaling has the potential to eliminate senescent cells within the epidermis. Furthermore, a decline in JAG1 expression in the basal layer as a result of ageing may contribute to the accumulation of senescent cells due to weakened Notch signaling activity.⁵¹ Therefore, enhanced JAG1/Notch signaling may help clear epidermal senescent cells and alleviate skin inflammation and ageing (Figure 3b).

Keratinocytes express a variety of genes associated with inflammasomes, providing evidence for the notion that inflammasomes are involved in the process of skin inflammaging.⁸ NLRP3 is involved in pyroptosis. When NLRP3 is activated, the NLRP3 inflammasome assembles to form a complex, which then activates pro-Caspase-1 to form active cleaved Caspase-1 and induces the thermogenic executive protein GSDMD to form cytotoxic GSDMD-N. This is then recruited to the cell membrane to form thermogenic pores, promoting IL-1 β and IL-18 expression.⁵² Skin inflammaging is accompanied by NLRP3 inflammasome activation. The depletion of NAD⁺ serves as a non-transcriptional initiating cue for the activation of NLRP3, facilitated by the clustering of mitochondria around the nucleus. Furthermore, the reduction of NAD⁺ levels associated with ageing can prompt the activation of the NLRP3 inflammasome in environments abundant in ATP.⁵³ Moreover, NLRP3 inflammasome inhibition improved the lifespan of the murine model for Hutchinson–Gilford Progeria,⁵⁴ indicating that the NLRP3 pathway is pivotal for managing skin inflammaging, and that suppression of this pathway can effectively alleviate inflammaging symptoms (Figure 3b).

The skin's complement system is activated via three pathways: lectin, classical, and alternative (Figure 3b).⁹ Keratinocytes produce complement proteins such as C3, Factor B, and C4, which are regulated and promoted by TNF- α , IL-1 α , and IFN- γ .⁵⁵ Inhibiting the C5aR1 signaling cascade within the complement system leads to a decrease in skin microbial diversity and also dampens the expression of cytokines, pattern recognition receptors, and antimicrobial peptides, thereby compromising the skin's defense against pathogenic microorganisms.⁵⁶ With ageing, the skin's complement system increases and activates, potentially contributing to the process of inflammaging.⁵⁷ For example, UV light stimulates epidermal keratinocytes to produce C3 and completion factor B via the alternative pathway.⁵⁸ An overactive complement system can damage the dermoepidermal junction, leading to the release of MMPs and ROS that degrade the extracellular matrix ECM.¹⁶ The complement system also affects melanocyte function and may be associated with age spot development.¹⁶ Complement activation-mediated inflammaging may contribute to enlarged facial pores, a prominent feature of ageing skin.⁵⁹ Understanding how the complement system drives skin inflammaging and ageing-related skin diseases could provide valuable insights for dermatologists and researchers related to the development of improved therapeutics.

The JAK–STAT signaling pathway has been implicated in the pathogenesis of inflammatory and autoimmune diseases of the skin, including vitiligo, psoriasis, and atopic dermatitis.¹⁰ JAKs are a family of four proteins, JAK1, JAK2, JAK3, and TYK2, that regulate inflammation by activating intracytoplasmic transcription factors called STAT.⁶⁰ Inflammation triggered by the JAK–STAT signaling pathway inhibits epidermal stem cell function, leading to skin ageing.⁶¹

MAPK is a group of serine/threonine protein kinases comprising three families, ERK, JNK, and p38. Among them, ERK consists of ERK1/2 and ERK5, p38 MAPK consists of p38 α , p38 β , p38 γ , and p38 δ isoforms, and JNK consists of JNK1, JNK2, and JNK3 isoforms. The MAPK pathway has three levels of signaling: the MAPK, the MAPK kinase (MEK or MKK), and the kinase of the MAPK kinase (MEKK or MKKK). These three kinases can be sequentially activated.⁶² The MAPK pathway promotes the production of MMPs and inflammatory cytokines through the regulation of SASP, which leads to cellular senescence.¹¹ TNF- α rapidly activates the p38 MAPK signaling pathway, causing the premature senescence of human skin fibroblasts.⁶³

Regulatory Roles of Natural Plants in Skin Inflammaging

Scientific advances increasingly facilitate research into the superior efficacy of natural plants as modulators of skin inflammaging. This section underscores the potential of natural plants as adjunctive treatments to combat skin inflammaging by modulating inflammatory and ageing-related pathways. Tables 1–4 detail 60 plant extracts and monomers that influence skin health through the key identified signaling pathways. The aim is to correlate these natural extracts and monomers with observed anti-inflammatory and anti-ageing effects, providing insights into their specific actions on skin health.

Inhibitor of the NF- κ B/RIG-I Pathway

Centella asiatica (L.) Urb. (CA) (Asian pennywort or Gotu kola) is widely used in Ayurvedic and traditional Chinese medicine. This plant contains alkaloids, flavonoids, phenols, tannins, and terpenoids, which exhibit anti-cancer, wound-healing, anti-bacterial, anti-diabetic, anti-inflammatory, and antioxidant effects.¹⁹⁷ The CA phytosome inhibits NF- κ B activities, halts I κ B α degradation, and reduces p65 and p50 nuclear translocation.⁶⁴ CA extract in lotion form protects

Table 1 Natural Plants That Regulate Skin Inflammation via the NF- κ B/RIG-I Pathway

| No. | Natural Plant/Compound | Plant Sources/Main Compounds | Mechanism of Action | Model of Evaluation | References |
|-----|---|--|---|--|------------|
| 1 | <i>Centella asiatica</i> extract | Alkaloids, flavonoids, phenols, tannins, and terpenoids | NF- κ B pathway; iNOS; COX-2; TNF- α ; IL-1 β , -1 α ; IgE; p65; p50; DNA | Clinical trial; animal models and in vitro experiments | [64–67] |
| 2 | Pomegranate | Anthocyanins, catechin, ellagic acid and gallic acids, and other polyphenolic compounds | NF- κ B pathway; MMP-1, -3; Col1A1; Timp3 | Clinical trial; animal models and in vitro experiments | [68–72] |
| 3 | Quercetin | Various natural plants such as <i>Sophora japonica</i> L., <i>Psidium guajava</i> L. | NF- κ B and Src/Syk/NF- κ B pathways; MMP1, -2, -3, -9; COX-2; SOD1, -2; IL-1 β , -6, -8, -17, -10; PKC δ ; JAK2; GSH; CAT; MDA; TRAF3; α -MSH; TNF- α | Clinical trial; animal models and in vitro experiments | [12,73–75] |
| 4 | Quercetin 3-O- β -D-glucuronide | Wine, <i>Hypericum hirsutum</i> , <i>Nelumbo nucifera</i> , <i>Oenothera biennis</i> , and green beans | NF- κ B pathways; AP-1; α -MSH; TNF- α ; COX-2 | Human keratinocytes and melanoma cells | [76] |
| 5 | EGCG | Tea | NF- κ B/RIG-I pathway; AP-1; IL-1, -6, -8, -17; PCNA; MMP-1; collagenase mRNA; TRAF6; SOD; CAT; MDA; JAK2; STAT1; STAT3 | Mouse model and in vitro experiments | [77–79] |
| 6 | Luteolin | Leaves, stems, and branches of <i>Reseda odorata</i> L. and a variety of natural herbs, vegetables, and fruits | NF- κ B pathways; NO; MDA; pro-inflammatory mediators (eg, IL-1 β , IL-6, IL-8, -17, -22, TNF- α , and COX-2) | Clinical trial; mouse models and in vitro experiments | [80] |
| 7 | Carotenoids | β -carotene, astaxanthin, lycopene, lutein, zeaxanthin, canthaxanthin, capsaicin, and astaxanthin | NF- κ B, AKT/mTOR and Bcl-2 pathways | Clinical trial; animal models and in vitro experiments | [81,82] |
| 8 | Pterostilbene | Blueberries | NF- κ B pathway; ROS; COX-2; MMP-9; AQP-3 | Mice model and HaCaTs | [83] |
| 9 | Myricetin | Natural foods such as berries, vegetables, teas, wine, and herbs | NF- κ B pathway; pro-inflammatory mediators (eg, IL-1 β , IL-6, IL-13, IL-8 and CCL17); MMP-9 | Cell experiments | [84] |
| 10 | Artemisinin | <i>Artemisia annua</i> L. | NF- κ B pathway; IL-1 β , -6; TNF α ; TLR2; chemokines (CXCL10, CCL20, CCL2, and CXCL2); ROS; p16INK4a; SOD; β -catenin; apoptosis; proliferation | Mouse models and cell experiments | [85] |
| 11 | Sulforaphane | Eg, broccoli, brussels sprouts, cabbage | NF- κ B pathway; COX-2; IL-6; TNF- α ; IL-1 β ; AP-1; STAT1; MITF; ET-1; PGE2; TYRPI; tyrosinase | Mouse models and cell experiments | [86,87] |
| 12 | Lithospermum erythrorhizon (Gromwell) root extract | Shikonin | NF- κ B pathways; Nrf2/ARE; inhibiting glycation; TNF- α | Cell experiments | [88,89] |
| 13 | <i>Kaempferia parviflora</i> Wall. ex Baker extract | Polymethoxyflavones; methoxyflavones | NF- κ B pathway; IL-6, -8; COX-2; collagen and elastin fibers | Mice models and cell experiments | [90] |
| 14 | Soy isoflavones | Daidzein and genistein | NF- κ B pathway; COX-2; IL-6, -1 α ; antioxidant enzymes | Clinical Trial; animal models and in vitro experiments | [91,201] |

Table 2 Natural Plants That Regulate Skin Inflammation via the mTOR Signaling Pathway

| No. | Natural Plant/Compound | Plant Sources/Main Compounds | Mechanism of Action | Model of Evaluation | References |
|-----|--|--|---|--|------------|
| 15 | Cordyceps | <i>Cordyceps sinensis</i> ; <i>Cordyceps cicadae</i> (Miq.) | ROS/PI3K/AKT/mTOR pathway; HAS2; MMP12 collagen and elastin synthesis | Cell experiments | [92–97] |
| 16 | Curcumin | <i>Curcuma longa</i> L. | mTOR pathway; IL-17, -6, -1 β ; TNF- α ; IFN- γ ; involucrin; filaggrin; PPAR- γ ; TLR4-MD2; mRNA; involucrin; filaggrin | Clinical trial; animal models and in vitro experiments | [98–102] |
| 17 | Fisetin | Found in fruits and vegetables, including strawberries, apples, and cucumbers. | PI3K/Akt/mTOR pathway; mTOR/IL-17A; IL-6 β , -53; autophagy; MMPs; COX-2; Nrf2 | Clinical trial; animal models and in vitro experiments | [103–109] |
| 18 | Anthocyanin | Purple sweet potato, pomegranates, and grapes. | PI3K/AKT/mTOR, Ras/MEK/ERK/mTOR pathway; 4E-BP; MMP-9, -2 | Mice models and cell experiments | [110–115] |
| 19 | <i>Myrothamnus flabellifolia</i> extract | Phenolic compounds | mTOR pathway; Park2; Atg7; LC3B; NLRP3; ROS, NF- κ B, IL-1 β , IL-6, MMPs; caspase-1; HMGB1; RAGE; filaggrin; claudin-1 | Clinical Trial; animal models and in vitro experiments | [116] |
| 20 | Novel phytopeptides | – | mTOR pathway | Vitro experiments | [117] |
| 21 | Chrysanthemum boreale flowers extract | Handelin | mTOR/AMPK pathway | Cell experiments | [118] |
| 22 | Latifolin | <i>Dalbergia odorifera</i> T. | mTOR pathway; Nrf2; SIRT1; IL-6; IL-8; RANTES; MDC; ICAM-1; p38; JNK | Cell experiments | [119] |
| 23 | Stigmasterol | Vegetable fats or oils from many plants | AKT/mTOR pathway; TNF- α , IL-1 β , -6; iNOS; COX-2; caspase-3; JAK–STAT pathway | Mice models and cell experiments | [120] |
| 24 | Mangosteen | α -, β -, γ -mangostins, isogarcinol, and gartanin | PI3K/Akt/mTOR pathway; MMP-1, -9; IL-23/Th17; TNF- α ; IL-2, -10; IFN- γ | Mice models and cell experiments | [121–123] |
| 25 | <i>Momordica charantia</i> L. extract | Terpenoids, saponins, flavonoids, sterols, glycosides, phenols, etc. | PI3K/AKT/mTOR pathway | Mice model | [124] |
| 26 | <i>Isatis tinctoria</i> L. leaf extract | 4-hydroxybenzoic acid, anthranilic acid, kynurenic acid, and linarin | mTOR/NF- κ B/SASP, MAPK/NF- κ B pathway; SA- β -GAL | Cell experiment | [125] |
| 27 | <i>Amaranthus cruentus</i> L. seed oil | Palmitic acid; oleic acid; linoleic acid; stearic acid | AKT/mTOR pathway | Cell experiment | [126] |
| 28 | <i>Mikania micrantha</i> L. extract | Flavonoids | FAK/Akt/mTOR pathway; MMP-2, -9 | Animal models and in vitro experiments | [127] |
| 29 | <i>Nypa fruticans</i> Wurmb extract | Polyphenols and flavonoids | PI3K/AKT/mTOR/CREB and MAPK signaling pathways | Cell experiment | [202] |

Table 3 Natural Plants That Regulate Skin Inflammation via the TGF- β Signaling Pathway

| No. | Natural Plant/Compound | Plant Sources/Main Compounds | Mechanism of Action | Model of Evaluation | References |
|-----|---|---|---|-------------------------------------|------------|
| 30 | <i>Prunus yeonensis</i> Matsum. extract | Genistein, naringenin, sakuranetin, prunetin, and amygdalin | TGF- β 1/Smad signaling pathway; MMP-1, -3; NO | Clinical trial and cell experiments | [129–132] |
| 31 | <i>Eucalyptus globulus</i> L. extract | Phenolic compounds | TGF- β /Smad pathway; MMPs; β -galactosidase; IL-6, NO and other pro-inflammatory mediators | Mice model and cell experiments | [128,133] |
| 32 | Ellagic acid | Berries and pomegranates | TGF- β /Smad pathway; IL-1 β , -6; elastin; collagen; Col1A1; TERT; Timp3; MMP3 | Mice model and cell experiments | [134–138] |
| 33 | <i>Terminalia arjuna</i> extract | Pentacyclic triterpenoids | TGF- β signaling pathway; TEWL; IL-6, COX-2, inos and ODC | Clinical Trial and mice model | [139,140] |
| 34 | <i>Portulaca oleracea</i> L. extract | Oleracone C; portulacanone A; portulacanone D | TGF- β /Smad Pathway; MMP-1; type I. procollagen; IL-1 β , -8; TNF- α ; NF- κ B pathway | Clinical Trial and cell experiments | [141,142] |
| 35 | Seaweed extract | Fucosterol; low-molecular-weight fucoidan | TGF- β 1/AP-1 signaling; MMPs; FGF-2; IL-1 β , -6; p-c-Jun, -Fos; NF- κ B; p38-MAPK; Erk1/2; JNK | Animal models and cell experiments | [143] |

(Continued)

Table 3 (Continued).

| No. | Natural Plant/Compound | Plant Sources/Main Compounds | Mechanism of Action | Model of Evaluation | References |
|-----|---|---|---|------------------------------------|------------|
| 36 | <i>Limonium tetragonum</i> (Thunb.) Bullock extract | Myricetin 3-O-β-d-galacto-pyranoside | TGFβ/Smad pathway; pro-inflammatory cytokines; MMP-1; collagen | Cell experiments | [144] |
| 37 | <i>Sacha inchi</i> albumin extract | – | TGF/Smad pathway; TNF-α; IL-6; MDA | Mice model | [145] |
| 38 | Red bean extract | Caffeic acid, rutin, and rosmarinic acid | TGF-β1; MMP-1; IL-1β, -6, -8, CCL17/TARC and CCL22/MDC | Cell experiments | [146] |
| 39 | <i>Prunella vulgaris</i> L. extract | Polyphenols, vitamins, polysaccharides, amino acids, and alkaloids | TGF-β/Smad signaling pathway; MMPs, TNF-α and IL-6 | Cell experiments | [19] |
| 40 | <i>Phyllanthus emblica</i> L. fruit extract | homoplantagin | ERK/TGF-β/Smad pathway; MAPK/AP-1 pathway; IL-1α,-6; PGE2; COX-2 | Cell experiments | [147] |
| 41 | <i>Salvia plebeian</i> extract | Rutin, glycerocolipids, kaempferolrutinoside, etc. | TGF-β/Smad pathway; MMP-1; IL-6 | Animal models and cell experiments | [148] |
| 42 | <i>Panax notoginseng</i> extract | Ginsenoside Rb1, Rb2, Rb3, Rc, Rd, Re, Rg1, and notoginsenoside R1 | TGF-β/Smad signaling pathway; MMP-1; IL-6; COX-2 | Cell experiments | [149,150] |
| 43 | <i>Acer tataricum</i> subsp. Ginnala extract | Ginnalin A | TGFβ/Smad pathway; MMP1; TNF-α, IL-1β, -6 | Cell experiments | [151] |
| 44 | <i>Astragalus</i> and <i>Radix Rehmanniae</i> extract | Saponins, flavonoids and polysaccharides; catalpol, inosine, and echinacoside | TGFβ/Smad pathway; Ras/MAPK; collagens; TIMP-3; MMP-3 | Cell experiments | [37] |
| 45 | <i>Rubus idaeus</i> extract | Anthocyanins, flavonoids, and phenolic compounds | TGFβ/Smad pathway; MMPs; collagens; SOD; Nrf2; NF-κB; COX-2; MAPK | Cell experiments | [203] |

DNA from UV-induced damage, reducing thymine photodimerization by over 28% and downregulating IL-1α expression.⁶⁵ A moisturizing lotion containing CA stem cell extract significantly improved skin moisture and barrier function for up to 24 h.⁶⁶ Moreover, oral supplementation with CA (Centellicum[®]) effectively ameliorated cutaneous stretch marks over a relatively short period.⁶⁷ These results indicate that CA extract is a potent modulator of skin inflammation.

Punica granatum L. (pomegranate) extract, derived from a deciduous tree in the Lythraceae family, is recognized for its anti-inflammatory and anti-senescence properties, primarily through NF-κB pathway inhibition.⁶⁸ Rich in compounds such as anthocyanins, catechin, ellagic acid, gallic acids, and other polyphenols,⁶⁹ punicalagin from the extract mitigates UV-induced growth arrest in the ageing human fibroblast cell line HFB4, enhances Col1A1 and Timp3 gene expression, preserves collagen levels, and reduces MMP3.⁷⁰ The primary dye in pomegranate arils, anthocyanins, decreased facial wrinkles in volunteers using a cold cream containing these compounds.⁷¹ Fermented pomegranate extracts have demonstrated improvements in skin moisture, brightness, elasticity, and collagen density after eight weeks, effectively preventing skin ageing.⁷² Thus, daily consumption of pomegranate extracts supports skin health by combating inflammation and decelerating the ageing process.

Quercetin is a promising anti-inflammatory agent, offering anti-inflammatory, anti-ageing, and antioxidant effects via the NF-κB pathway.¹² Quercetin significantly reduces inflammatory cytokine production in primary human keratinocytes,⁷³ decreases MDA levels in the skin tissues of psoriasis-like mice via NF-κB pathway inhibition,⁷⁴ and directly targets PKCδ and JAK2, protecting against UV-induced skin ageing and inflammation.⁷⁵ Quercetin 3-O-β-D-glucuronide (Q-3-G), a glucuronide conjugate of quercetin, has anti-inflammatory, antioxidant, moisturizing, and anti-melanogenesis effects in human keratinocytes and melanoma cells because of NF-κB pathway modulations.⁷⁶ Quercetin also downregulates pro-inflammatory genes and cytokines such as COX-2 and TNF-α in HaCaT cells and reduces melanin production in B16F10 cells,⁷⁶ highlighting its potential as an effective anti-inflammatory compound.

Green tea's primary polyphenolic catechin, Epigallocatechin gallate (EGCG), along with its structural isomer GCG and a compound containing digallate (specifically, theaflavin-3,3'-digallate), exhibit robust inhibitory effects on RIG-I. At low micromolar concentrations, EGCG interacts with RIG-I in HEK293T cells, thereby suppressing RIG-I signaling.

Table 4 Natural Plants That Regulate Skin Inflammation via the AMPK–Sirtuins and Other Signaling Pathways

| No. | Natural Plant/ Compound | Plant Sources/Main Compounds | Mechanism of Action | Model of Evaluation | References |
|-----|--|--|--|--|--------------|
| 46 | <i>Salvia miltiorrhiza</i> Radix et Rhizoma extract | Cryptotanshinone, tanshinone and salvianolic acid B | AMPK/sirt1, AMPK/ULK1 signaling pathway; MMPs; ROS; IL-4, -10, -2, -6; IFN- γ ; NF- κ B p65; autophagy; MDA; CAT, Nrf2; NQO1; HO-1 | Mouse models and in vitro experiments | [152–158] |
| 47 | <i>Youngia denticulata</i> extract | Phenolic compounds | AMPK–sirtuins, AMPK/Nrf pathway; MMPs; TNF- α ; COX-2; IL-6; p65; p50; autophagic | Cell experiments | [159–164] |
| 48 | Ginseng | Ginsenosides | LKB1/ AMPK/SIRT1 pathway; inflammatory factor; type I collagen; MMPs | Clinical Trial; animal models and in vitro experiments | [46,165–170] |
| 49 | Almond <i>Prunus dulcis</i> (Mill.) D.A. Webb extract | Fatty acids, phytochemical polyphenols, VE and chlorogenic acid | AMPK–sirtuin pathway; COL1A1; COL1A2; TNF- α ; hyaluronic acid synthase mRNA; GR signaling; MCP-1; CCL-5 | Clinical trial and cell experiments | [171–178] |
| 50 | Icariin | Epimedium | SIRT6; NF- κ B | Mice model | [179] |
| 51 | <i>Aronia melanocarpa</i> extract | Phenolic compounds | AMPK/SIRT1/NF- κ B, PI3K/AKT/mTOR pathway; MMP-1, 3; immunoreactivity of collagen types I and III; pro-inflammatory mediators | Mice model and in vitro experiments | [177,180] |
| 52 | <i>Crocus sativus</i> extract | Polyphenols | SIRT1 | Clinical trial | [181] |
| 53 | <i>Tinospora cordifolia</i> extract | Alkaloids, terpenoids, sitosterols, flavonoids, and phenolic acids | AMPK–sirtuins signaling pathway; PI3K/AKT; IL-1 α , -1 β , -6, -8; IFN- γ ; PGE-2; TNF- α | Mouse model | [182] |
| 54 | Peanut skin extract | Oligosaccharides and flavonoids | AMPK; MMP-3, -9; TRP-1; DPPH; MITF; Tyrosinase; Collagenase | Mice model and cell experiments | [183,184] |
| 55 | <i>Hypericum perforatum</i> extract | Hyperforin | AMPK mRNA; III1; II6; II23; III17a; II22; antimicrobial peptides (AMPs); IL-6; IL-17A; TNF- α ; MAPK/STAT3 | Mice model and cell experiments | [185,186] |
| 56 | Resveratrol | The skin of the grape, blueberry, raspberry, mulberry, and peanut. | AMPK; NF- κ B, PI3K/AKT, TLR4/NF- κ B/STAT3, NOTCH; Wnt signaling pathway; p53, p16, p19, NLRP3 and Cas1 p20; (MMP)-1, MMP-9, MMP3 and interleukin-8; CAT, GSH, SOD, MAPK-AP-1/NF- κ B-TNF- α /IL-6, iNOS, COX-2-mediated inflammation-induced ageing; and p53-Bax-cleaved caspase-3-cytochrome C-mediated apoptosis induced ageing | Clinical trial; animal models and in vitro experiments | [39,187–189] |
| 57 | <i>Echinacea purpurea</i> L. Moench extract | Phenolic compounds | Complement system pathway; C3a/C3aR pathway; anti-collagenase, anti-elastase, anti- hyaluronidase activity | Animal models and in vitro experiments | [58,190] |
| 58 | <i>Impatiens textori</i> Miq. extract | Palmitoleic acid, palmitelaidic acid and methyl undecanoate | NLRP3 pathway; IL-1 β ; caspase-1 | Cell experiments | [191] |
| 59 | Piceatannol | Peanuts and grapes | JAK1/STAT3 pathway; Notch pathway; COX-2, iNOS, AP-1, MMP-1, IKK β , MAPK, mTOR | Cell experiments | [192–194] |
| 60 | <i>Houttuynia cordata</i> Thunb. | Volatile oils, polysaccharides, and flavonoids | MAPK pathway | Cell experiments | [195,196] |

Furthermore, it dampens IL-6 secretion and IFN- β mRNA production in BEAS-2B cells.⁷⁷ EGCG also mitigates UV-induced MMP-1 expression, exhibiting anti-ageing effects. EGCG treatment reduced UVA-induced skin damage, such as roughness and sagginess, and protected against decreases in dermal collagen in hairless mouse skin.⁷⁸ Additionally, EGCG obstructed the UV-triggered escalation of collagen secretion and collagenase mRNA levels in fibroblast cultures.⁷⁹ Potential plant-derived RIG-I pathway inhibitors include β -sitosterol,⁶⁰ *Abelmoschus manihot* total flavones,¹⁹⁸ *Asphodelus microcarpus* leaf extract,¹⁹⁹ and berberine.²⁰⁰ Although direct evidence for improving skin inflammation is lacking, these inhibitors show potential.

Phytochemicals such as luteolin,⁸⁰ carotenoids,^{81,82} pterostilbene,⁸³ myricetin,⁸⁴ artemisinin,⁸⁵ sulforaphane,^{86,87} *Lithospermum erythrorhizon* (Gromwell) root extract,^{88,89} *Kaempferia parviflora* Wall. ex Baker,⁹⁰ and soy isoflavones^{91,201} are also beneficial for managing skin inflammation. These compounds contribute to skin health by inhibiting the NF- κ B/RIG-I signaling pathway, indicating their potential as therapeutic agents for promoting skin health and anti-ageing.

Inhibition of the mTOR Pathway

Cordyceps exert anti-ageing, anti-inflammation, and anti-oxidative stress effects, and their efficacy is attributed to their rich composition of active compounds, including cordyceps, nucleosides, polysaccharides, sterols, proteins, amino acids, polyphenols, and peptides.⁹³ *Cordyceps sinensis* counters cigarette smoke extract-induced cellular senescence by inhibiting the ROS/PI3K/AKT/mTOR signaling pathway;⁹⁴ it also shields human skin cells from UV-induced oxidative stress and DNA damage.^{93,95} *Cordyceps cicadae* (Miq.) aqueous extract enhances HAS2 expression and reduces MMP-12 expression in human skin fibroblasts, demonstrating moisturizing and anti-ageing capabilities.⁹⁶ *Cordyceps* medium-loaded nanoparticles prompted a significant 2.76-fold increase in skin regeneration in human dermal fibroblasts (HDFs) by upregulating ECM production (collagen and elastin synthesis) and repairing H₂O₂-induced cell damage.⁹² *Cordyceps* extract is an ideal topical solution for skin inflammation as it mitigates inflammatory factor secretion, reduces cellular senescence, and boosts collagen synthesis.⁹⁷

Curcumin, a phytochemical in turmeric, inhibits the phosphorylation of S6K1 and 4E-BP1, which are downstream targets of mTOR, in Rh1 and Rh30 cells. It mechanistically blocks mTOR by destabilizing mTORC1 through raptor dissociation at low concentrations and disrupts mTORC2 by detaching Rictor at higher concentrations.⁹⁸ Curcumin and its metabolites have extended the lifespan of several ageing models, including *C. elegans*, *D. melanogaster*, yeast, and mice.^{99–101} Nano-formulated curcumin has a potential for anti-ageing and wound healing.⁹⁹ Curcumin also inhibits the proliferation of differentiated HaCaT cells by reducing IL-17, TNF- α , IFN- γ , and IL-6 levels.¹⁰⁰ A clinical study of 28 women in their 30s who used a turmeric-containing gel (Tricutan[®]) for four weeks reported improved skin firmness according to self-assessments.¹⁰¹ Turmeric hot water extract countered elevated TNF- α and IL-1 β mRNA and protein levels induced by UVB in the skin of healthy subjects and led to increased hyaluronic acid production by keratinocytes, as well as enhanced facial skin hydration.¹⁰² Thus, curcumin has emerged as a potent anti-inflammatory agent and moisturizer.

Fisetin, a flavonol found in various fruits and vegetables such as strawberries, apples, persimmons, and onions, has also emerged as a potent anti-inflammatory agent. Recognized for its senolytic, antioxidant, and anti-proliferative properties, fisetin acts as a natural source of senolytics, offering a novel treatment for ageing.¹⁰³ Fisetin also effectively addresses psoriasis and other inflammatory skin conditions by modulating the mTOR/IL-17A, PI3K/Akt/mTOR, and autophagy pathways across several models.^{104,105} Moreover, fisetin suppresses cutaneous melanocytic lesions by targeting the Akt/mTOR signaling axis¹⁰⁶ and inhibits α -MSH- and IBMX-induced melanosis in B16F10 melanoma cells, simultaneously upregulating mRNA for skin fibril-related genes.¹⁰⁷ Furthermore, fisetin reduces MMP, ROS, PGE1, TNF- α , IL-6 β , IL-53, and cyclooxygenase-2 expression, mitigating UVB-induced skin inflammation, DNA damage, wrinkles, and sunspots.¹⁰⁸ Finally, fisetin enhances filaggrin expression, protecting against the UVB-induced disruption of barrier function.¹⁰⁹

Anthocyanins, water-soluble flavonoid pigments from *Sambucus canadensis*, exhibit anti-inflammatory and antioxidant properties.¹¹⁰ These compounds significantly mitigate cell senescence and lens ageing by inhibiting the PI3K/AKT/mTOR signaling pathway. This inhibition promotes the apoptosis of senescent cells, increases autophagic and mitophagic flux, and enhances mitochondrial renewal to maintain cellular homeostasis.¹¹¹ Purple sweet potato (PSP) anthocyanins reduce oxidative stress and inflammation in rats with neuropathic pain induced by chronic constriction injury,¹¹² and extend the lifespan of *D. melanogaster* by activating autophagy via AKT-1, PI3K, and mTOR gene downregulation and

4E-BP upregulation, while increasing lysosome production and autophagy activation. Thus, PSP extract can be used to delay skin inflammaging.¹¹³ Furthermore, anthocyanins from pomegranates and grapes, as well as caffeic acid from various natural sources, inhibit mTORC1 by preventing the phosphorylation of S6K1 and AKT at the Ser473 residue mediated by mTORC1.¹¹⁴ Furthermore, mulberry anthocyanins reduce MMP-2 and MMP-9 expression in the ECM of melanoma B16-F1 cells by targeting the Ras/MEK/ERK/mTOR pathway.¹¹⁵

A clinical trial showed that a phytocosmetic formulation containing *Myrothamnus flabellifolia* leaf extracts significantly improved skin ageing markers, including spot length, wrinkle area and volume, skin lightening, and homogeneity, by downregulating mTOR genes.¹¹⁶ Additionally, novel phytopeptides have shown efficacy in restoring cellular homeostasis, modulating oxidation, and hyperproliferation, thereby impacting the mTOR pathway.¹¹⁷ Peptides are favored over small-molecule drugs given their higher specificity and lower cytotoxicity, with plant-derived natural peptides having a long history of human consumption, indicating minimal potential adverse effects compared with synthetic or nonspecific therapeutics.¹¹⁷ Various natural plants and compounds, including Handelin from *Chrysanthemum boreale* flowers,¹¹⁸ Latifolin,¹¹⁹ Stigmasterol,¹²⁰ α - β - γ -mangostins, isogarcinol, and gartanin from *Garcinia mangostana* L. (mangosteen),^{121–123} *Momordica charantia* L.,¹²⁴ *Isatis tinctoria* L. leaf,¹²⁵ *Amaranthus cruentus* L. seed oil,¹²⁶ *Mikania micrantha* L.,¹²⁷ and *Nypa fruticans* Wurmb,²⁰² are potential modulators of skin inflammaging via the mTOR signaling pathway. However, further in vitro and in vivo studies are required to verify their effects.

Modulation of the TGF- β Signaling Pathway

Prunus yeonensis Matsum. (Rosaceae) is recognized for its anti-allergy, anti-ageing, and anti-inflammatory properties. The bark extract was traditionally used for treating coughs, urticaria, pruritus, dermatitis, asthma, and measles.¹²⁹ Prunetin from *P. yeonensis* extract (PYE) exhibited anti-inflammatory effects on LPS-stimulated RAW 264.7 macrophages and an LPS-induced septic shock model.¹³⁰ Polyphenols from the bark show high antioxidant activity, contributing to anti-ageing effects.¹³¹ Moreover, PYE at 1, 10, and 100 $\mu\text{g/mL}$ enhanced type I procollagen production in UVB-irradiated normal HDFs via the TGF- β 1/Smad signaling pathway. This involved the upregulation of TGF- β 1 by Smads and the downregulation of transcription activator protein-1 and MAP kinase, resulted in reduced MMP-1 and MMP-3 production.¹³¹ PYE also reduced nitric oxide secretion in LPS-treated RAW 264.7 cells. In an SLS-stimulated patch test, PYE showed lower visual and erythema scores compared with a placebo on days 5 and 9, indicating potent anti-inflammaging effects in vitro and in vivo.¹³² These qualities position PYE as a promising ingredient for skincare products.

Eucalyptus globulus L. (EG) (myrtle family) contains an essential oil in the leaves that is rich in compounds such as thymol, 1,8-cineole, linalool, limonene, and pinenes, and acclaimed for its antioxidant and anti-inflammatory properties. EG extract promotes collagen synthesis by modulating the TGF- β /Smad signaling pathway, including the inhibition of negative Smad regulators. Treatment of UV-irradiated HDFs with 1 to 100 $\mu\text{g/mL}$ EG extract resulted in decreased MMP and IL-6 expression, alongside increased TGF- β 1 and type 1 procollagen levels. Additionally, this treatment prevented activation of the AP-1 transcription factor.¹²⁸ Furthermore, EG essential oil and hydrodistillation residual water minimized the activation of senescence markers, such as β -galactosidase and MMPs, while increasing collagen type I expression and exerting strong anti-inflammatory effects by reducing nitric oxide and pro-inflammatory mediators.¹³³ Therefore, EG extract shows potential as an anti-inflammaging substance.

Ellagic acid (EA), a polyphenolic compound found in numerous fruits (eg, raspberries, pomegranates, and plums), seeds (eg, walnuts and almonds), and vegetables, either exists in free form or as part of ellagitannins. EA exerts antioxidant, anti-inflammatory, and anti-ageing effects.¹³⁴ Specifically, EA shields HaCaT cells and mice from photo-ageing by modulating the TGF- β /Smad signaling pathway (TGF- β 1 \rightarrow pSmad3 \rightarrow Smad7).¹³⁵ The topical application of 10 $\mu\text{mol/L}$ EA to SKH-1 hairless mice, exposed to UV-B for eight weeks, significantly reduced skin wrinkles and UVB-induced inflammation, decreased IL-1 β and IL-6 production, prevented inflammatory macrophage infiltration, and reduced ICAM-1 expression in UVB-irradiated keratinocytes and mouse epidermis.¹³⁶ Moreover, EA boosts elastin and collagen production in HDFs.¹³⁷ Chitosan-coated niosomes containing EA-modulated skin ageing-related genes (upregulating *Coll1A1*, *TERT*, *Timp3*, and downregulating *MMP3*) in HFB4 cells, thereby protecting collagen levels and decelerating skin ageing.¹³⁸

Pentacyclic triterpenoids from *Terminalia arjuna* may enhance in vitro TGF- β expression and vascular endothelial growth factor secretion, significantly increasing skin hydration, reducing transepidermal water loss, and improving skin elasticity.^{139,140} Oleracone C, derived from *Portulaca oleracea* mitigated UVB-induced MMP changes and type I procollagen production in human keratinocytes through the TGF- β /Smad pathway.¹⁴¹ Additionally, two compounds from *P. oleracea* extract, portulacanone A and portulacanone D, inhibited MMP-1 secretion and boosted type I procollagen production in UVB-stressed human keratinocytes, demonstrating potent anti-ageing effects.¹⁴² Fucosterol,¹⁴³ Myricetin 3-O- β -D-galactopyranoside from extracts of *Limonium tetragonum* (Thunb.) Bullock,¹⁴⁴ *Sacha inchi* albumin,¹⁴⁵ red bean (*Vigna angularis*),¹⁴⁶ *Prunella vulgaris* L.,¹⁹ *Phyllanthus emblica* L. fruit,¹⁴⁷ *Salvia plebeian*,¹⁴⁸ *Panax notoginseng*,^{149,150} *Acer tataricum* subsp. Ginnala,¹⁵¹ *Astragalus* and *Radix Rehmanniae*,³⁷ and *Rubus idaeus*²⁰³ all showed efficacy in mitigating inflammaging-induced skin damage. These effects are mediated by modulation of the TGF- β signaling pathway, highlighting the potential of these compounds as therapeutic agents for skin health and anti-ageing.

Regulation of the AMPK–SIRT1 Signaling Pathway

Danshen (*Salviae miltiorrhizae* Radix et Rhizoma) is widely used in traditional Chinese medicine and exhibits extensive pharmacological benefits, including anti-inflammatory, antioxidant, anti-ageing vasodilatory, lipid-lowering, and improved energy metabolism effects.¹⁵³ The active compounds of danshen are primarily fat-soluble tanshinones, such as cryptotanshinone, and water-soluble tanshinone, notably salvianolic acid B.¹⁵⁴ Cryptotanshinone enhances mitochondrial biosynthesis in skin cells via the AMPK/SIRT1/PGC-1 α pathway, reducing skin ageing and extending the lifespan of budding yeast *Saccharomyces cerevisiae*, indicating its potential or delaying senescence.¹⁵⁵ Cryptotanshinone also shows promise in dermatology, including psoriasis treatment,¹⁵⁶ UV-induced melanoma mitigation,¹⁵² and scar formation reduction.¹⁵⁷ Salvianolic acid B activates the AMPK/ULK1 pathway, enhancing the autophagic degradation of pS757-ULK1 and autophagic activity in astrocytes.¹⁵² Salvianolic acid B boosts autophagy, significantly increases Beclin1 expression, a marker of autophagy, and increases the microtubule-associated protein light chain 3 B (LC3B) II/LC3B I ratio. Additionally, salvianolic acid B extends cell survival, enhances the release of anti-inflammatory factors (IL-4, IL-10, IL-2, IL-6, and IFN- γ), and reduces apoptosis; it can also reduce inflammation and oxidative stress, which is attributed to activation of the AMPK/SIRT1 pathway.¹⁵⁸ Thus, danshen extract represents a potential skin anti-inflammaging agent.

Youngia denticulata (YD, Asteraceae; synonym: *Crepidiastrum denticulatum*) is a traditional vegetable in Korea. YD extract contains phenolic compounds such as dicaffeoylquinic acid, chicoric acid, youngiasides A, B, and C, showing anti-inflammatory, anti-ageing and antioxidant activity.^{159,160} These compounds decrease MMPs, including MMP-1, -2, -3, and -9, in HaCaT cells and HDFs, increase collagen production via the AMPK/Nrf2 pathway in HDFs, and inhibit NF- κ B-mediated inflammation.¹⁶¹ Treatment with YD extract results in significant autophagic vesicle formation within the non-cytotoxic range and induces AMPK phosphorylation without mTOR inhibition.¹⁶² The role of autophagic signaling pathway in skin homeostasis has been thoroughly investigated.¹⁶³ Besides its critical role in epidermal differentiation, autophagy contributes to skin barrier integrity, inflammation reduction, and ageing mitigation.¹⁶⁴ Anti-pollution efficacy was evaluated using benzo[a]pyrene (BaP) and cadmium chloride (CdCl₂) as model compounds; the YD extract protected against cytotoxicity and effectively reduced pro-inflammatory cytokines,¹⁶² indicating its potential as an anti-inflammaging cosmetic ingredient.

Ginseng is a perennial herb of the Araliaceae family that is widely used in Asia. Its efficacy is attributed to triterpenoid saponins called ginsenosides, which are categorized into three groups, with microbial transformation producing their active metabolites. Notably, ginsenosides with two glucopyranosyl groups at the C-3 position are recognized as SIRT1 activators. These compounds target the SIRT1 signaling pathway, offering therapeutic potential against oxidative stress, inflammation, ageing, and depression.¹⁶⁵ The ginsenoside metabolite 20-O- β -D-glucopyranosyl-20(S)-protopanaxadiol mitigates UV-induced MMP-1 expression in HDFs by enhancing liver kinase B1 (LKB1) and AMPK phosphorylation.¹⁶⁶ Ginsenoside Rb1, by activating the SIRT1/AMPK pathway, diminishes endothelial cell senescence.⁴⁶ Comprehensive clinical investigations have validated the anti-inflammaging benefits of ginseng extract on skin and the regulation of MMPs and type I collagen in human fibroblasts to augment skin elasticity and hydration.¹⁶⁷ Additionally, ginsenoside Rb1 decreases IL-1 β release at injury sites in mice, which mitigates inflammation.¹⁶⁸ Ginseng extracts, as anti-ageing components, also enhance immunity and modulate inflammation.¹⁶⁹ Herbal formulations with ginseng extract are leading natural candidates for innovative dermocosmetics exhibiting anti-ageing benefits for the skin.¹⁷⁰

Almond (*Prunus dulcis* (Mill.) D.A. Webb), a leading nut crop globally, is a rich source of fatty acids, phytochemical polyphenols, and antioxidants, including vitamin E and chlorogenic acid. Almond bark polyphenol extract notably enhances AMPK phosphorylation and mitigates TNF- α -induced cellular inflammation by reducing the secretion of monocyte chemoattractant protein-1 and chemokine ligand 5.¹⁷¹ This extract also prolongs the lifespan of yeast, possibly because it preserves mitochondrial function with age.¹⁷² β -Violanone from almond oil counters the suppression of dexamethasone-induced collagen and hyaluronic acid synthesis in HDFs.¹⁷³ A study on skin massage with almond oil-infused essential oils indicated significant alleviation of itchiness and mental stress in 64 women (≥ 65 years old).¹⁷⁴ Moreover, almond oil exhibits anti-inflammatory, antioxidant, anti-ageing, wound-healing, and skin barrier-restoration effects.^{183,184} Topical almond oil and extract application diminishes UVB-induced photoageing both in vitro and in vivo.¹⁷⁷ Furthermore, regular almond consumption can reduce facial wrinkles and pigment intensity in postmenopausal women.¹⁷⁸

Natural compounds such as icariin¹⁷⁹ and extracts of *Aronia melanocarpa*,^{117,180} *Crocus sativus*,¹⁸¹ *Tinospora cordifolia*,¹⁸² peanut skin,^{183,184} and *Hypericum perforatum*^{185,186} modulate AMPK signaling. The plant extracts and compounds reviewed in this study attenuate inflammatory senescence by regulating the AMPK–SIRT6 pathway. SIRT6 enzymes have drawn significant interest for their pivotal role in anti-ageing. Thus, comprehensive research into the AMPK–SIRT6 pathway and the molecular underpinnings of inflammaging highlights phytotherapies targeting this pathway as novel interventions for managing skin inflammaging.²⁰⁴

Regulation of Other Pathways

Resveratrol (3,5,4'-trihydroxy-trans-stilbene) is a natural phenol found in grape, blueberry, raspberry, mulberry, and peanut skins, which exhibits anti-inflammatory, antioxidant, and anti-ageing properties.³⁹ As the most potent Notch activator, resveratrol is thus capable of alleviating skin inflammaging damage,¹⁸⁷ potentially by inducing cell death via the ROS-dependent inactivation of the Notch/PTEN/AKT cascade reaction.¹⁸⁸ Resveratrol also inhibits mast cells, sphingosine kinase-1, STAT3, expression of senescence markers (p53, p16, and p19), inflammasome markers (NLRP3 and Cas1 p20), nuclear translocation of NF- κ B, IL-1 β , IL-6, TNF- α , and IL-18, and activation of the NF- κ B p65 signaling pathway.¹⁸⁹

Echinacea purpurea (L.) Moench extract suppresses inflammation by inhibiting the C3a/C3aR signaling pathway in the complement system in TNBS-induced ulcerative colitis rats.⁵⁸ *E. purpurea* extract exhibits significant anti-collagenase (78.5 \pm 0.0%), anti-elastase (69.0 \pm 1.4%), and anti-hyaluronidase activity (64.2 \pm 0.3%).¹⁹⁰ This extract may impede skin inflammaging by inhibiting the complement system.

Impatiens textori Miq. (Balsaminaceae) is a traditional medicinal herb used to treat inflammation-related skin infections and allergic disorders. The extracts demonstrate inhibitory effects on interleukin 1 β secretion and reduce ASC oligomerization and caspase-1 maturation by attenuating NLRP3 inflammasome activation, which were observed at concentrations of 25, 50, and 100 μ g/mL.¹⁹¹ Additionally, the extracts exhibited stimulatory effects on HaCaT cell proliferation, migration, sprout outgrowth, and the synthesis of types I and IV collagen.¹⁹¹ These extracts also alleviate the dulling effect of skin color that is characteristic of skin inflammaging.¹⁹¹ Furthermore, *Myrothamnus flabellifolia* leaf extracts and resveratrol can attenuate inflammaging damage to the skin by inhibiting NLRP3, as outlined in Table 1 for natural plants 19 and 56.

Piceatannol is a resveratrol metabolite commonly found in red wine grapes. It reduces atopic dermatitis by inhibiting JAK1 and decreasing JAK–STAT protein phosphorylation.¹⁹² It also inhibits MMP-1 expression via the JAK1/STAT3 pathway in keratinocytes. Piceatannol exhibits various effects on the skin, including promoting collagen production, inhibiting melanin synthesis, inducing the antioxidant glutathione, and eliminating ROS.¹⁹³ A clinical trial showed that passion fruit seed extract, when taken orally, is rich in piceatannol and can improve the moisture levels of dry skin and reduce fatigue,¹⁹⁴ implying that the compound has a skin inflammaging-prevention effect. Furthermore, EGCG, sulforaphane, stigmasterol, *Hypericum perforatum* extracts, and resveratrol can attenuate inflammaging damage to the skin by inhibiting STAT, as outlined in Tables 1, 2, and 4 for natural plants 5, 11, 23, 55, and 56.

Houttuynia cordata Thunb. (Saururaceae), a native perennial herb, contains many active compounds, including volatile oils, water-soluble polysaccharides, and flavonoids, and exhibits antioxidant, anti-ageing, and anti-inflammatory activities.¹⁹⁵ The hyperoside-enriched fraction obtained from *H. cordata* inhibits skin ageing by

regulating the MAPK signaling pathway and attenuating JNK/ERK/c-Jun activation in HDFs.¹⁹⁶ Furthermore, extracts of *Isatis tinctoria* L. leaves, *Nypa fruticans* Wurmb, seaweed, *Phyllanthus emblica* L. fruit, *Astragalus and Radix Rehmanniae*, *Rubus idaeus*, *Hypericum perforatum*, and piceatannol can attenuate skin inflammaging damage by inhibiting MAPK, as outlined in Tables 2–4 for natural plants 26, 29, 35, 40, 44, 45, 55, and 59.

Conclusion

Chronic low-level inflammation contributes to ageing and age-related degenerative diseases, which manifest outwardly as skin ageing, including wrinkles, sagging, thinning, and pigmentation changes. This review explores the mechanisms of skin inflammaging, focusing on nine critical signaling pathways: NF- κ B/RIG-I, mTOR, TGF- β , AMPK–SIRT6, complement system, NLRP3, Notch, JAK–STAT, and MAPK. In the natural environment, the process of skin inflammaging is marked by complexity, involving intricate interactions among multiple signaling pathways. Notably, the NF- κ B signaling pathway intersects with MAPK, PI3K, and TGF- β pathways to collectively regulate this inflammaging process in the skin. AMPK can regulate AMPK/mTOR pathways, which relieve neuropathic pain and inflammatory pain.⁴² Additionally, this review highlights how various natural plant extracts and monomers impact skin inflammaging through these pathways. Due to their diverse composition, plant extracts exhibit unparalleled advantages in addressing multiple interventions concurrently. This research therefore strengthens our understanding of skin inflammaging mechanisms and identifies promising natural interventions against skin inflammaging, as well as key directions for future research. Unfortunately, few natural plants modulate Notch, the complement system, NLRP3, JAK–STAT, or MAPK pathways. This requires more attention from relevant researchers on the botanical regulators of these pathways, to determine their effectiveness and safety in more skin inflammaging related in vitro and animal models, detailed, well-controlled and larger scale dose-response randomized clinical trials. Limited in-depth animal and clinical studies exist on the regulatory effects of plants extracts, such as *Echinacea purpurea* (L.) Moench, on skin inflammaging via the complement system or other signaling pathways. Further investigation by relevant researchers is warranted. Therefore, future research should explore active substances that affect inflammaging in the identified plant species to promote further development in this field. Given the complex compositions and mechanisms of action of natural plants, clinical trials are required to evaluate their actual effects, safety, and effectiveness. Additionally, the use of natural molecules derived from plant extracts to modulate skin inflammaging is a promising area for future exploration. Elucidating the effects of these natural plant extracts and active chemical monomers on the prevention of skin inflammaging will also enrich the library of raw materials available for treating skin inflammaging. Future research should also aim to discover new natural plants that offer more possibilities for counteracting skin inflammaging damage.

Abbreviations

NF- κ B/RIG-I, nuclear factor kappa-light-chain-enhancer of activated B cells/retinoic acid-inducible gene I; mTOR, mammalian target of rapamycin; TGF- β , transforming growth factor beta; AMPK, AMP-activated protein kinase; NLRP3, NLR family pyrin domain containing 3; JAK, janus kinase; STAT, signal transducers and activators of transcription; MAPK, mitogen-activated protein kinase; TNF- α , tumor necrosis factor- α ; IL, interleukin; ROS, reactive oxygen species; ATM-IRF1, Ataxia telangiectasia-mutated-interferon regulatory factor 1; PI3K, phosphoinositide 3-Kinase; Akt, protein kinase B; Ras, the renin-angiotensin system; Raf, serine/threonine kinase; MEK, mitogen-activated extracellular signal-regulated kinase; ERK, extracellular regulated protein kinases; Rsk, ribosomal S6 kinase; MMP, matrix metalloproteinase; RhoA, ras homolog family member A gene; Rock, Rho-associated coiled-coil-containing protein kinase; LAP, latency associated peptide; AP-1, activator protein-1; MCP, membrane cofactor protein; NADH, nicotinamide adenine dinucleotide; LKB-1, liver kinase B1; FoxO, forkhead Box O; PGC-1 α , peroxisome proliferator-activated receptor-gamma coactivator 1 alpha; P53, tumor protein P53; NICD, Notch Intracellular Domain; GSDMD, gasdermin D; IFN- γ , interferon- γ ; ECM, extracellular matrix; ECM, extracellular matrix; HDF, human dermal fibroblast; LPS, lipopolysaccharides; TIMP, tissue inhibitor of metalloprotease.

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Disclosure

The authors declare no conflicts of interest in this work.

References

- Baechle JJ, Chen N, Makhijani P, Winer S, Furman D, Winer DA. Chronic inflammation and the hallmarks of ageing. *Mol Metabol.* 2023;74:101755. doi:10.1016/j.molmet.2023.101755
- Chung HY, Kim DH, Lee EK, et al. Redefining chronic inflammation in ageing and age-related diseases: proposal of the senoinflammation concept. *Ageing Dis.* 2019;10(2):367–382. doi:10.14336/AD.2018.0324
- Franceschi C, Capri M, Monti D, et al. Inflammageing and anti-inflammageing: a systemic perspective on ageing and longevity emerged from studies in humans. *Mech Ageing Dev.* 2007;128(1):92–105. doi:10.1016/j.mad.2006.11.016
- Pilkington SM, Bulfone-Paus S, Griffiths CEM, Watson REB. Inflammageing and the Skin. *J Invest Dermatol.* 2021;141(4s):1087–1095. doi:10.1016/j.jid.2020.11.006
- Grabowska K, Nowacka-Chmielewska M, Barski J, Liskiewicz D. Inflammageing - contributing mechanisms and cellular signaling pathways. *Postepy Biochem.* 2021;67(2):177–192. doi:10.18388/pb.2021_375
- Pająk J, Nowicka D, Szepletowski JC. Inflammageing and immunosenescence as part of skin ageing-A narrative review. *Int J Mol Sci.* 2023;24(9):1–11. doi:10.3390/ijms24097784
- Quillard T, Charreau B. Impact of notch signaling on inflammatory responses in cardiovascular disorders. *Int J Mol Sci.* 2013;14(4):6863–6888. doi:10.3390/ijms14046863
- Awad F, Assrawi E, Louvrier C, et al. Photoaging and skin cancer: is the inflammasome the missing link? *Mech Ageing Dev.* 2018;172:131–137. doi:10.1016/j.mad.2018.03.003
- de Boer EC, Thielen AJ, Langereis JD, et al. The contribution of the alternative pathway in complement activation on cell surfaces depends on the strength of classical pathway initiation. *Clin Transl Immunol.* 2023;12(1):e1436. doi:10.1002/cti2.1436
- Solimani F, Meier K, Ghoreschi K. Emerging topical and systemic JAK inhibitors in dermatology. *Front Immunol.* 2019;10:2847. doi:10.3389/fimmu.2019.02847
- Liu HM, Cheng MY, Xun MH, et al. Possible mechanisms of oxidative stress-induced skin cellular senescence, inflammation, and cancer and the therapeutic potential of plant polyphenols. *Int J Mol Sci.* 2023;24(4).
- Li X, Li C, Zhang W, Wang Y, Qian P, Huang H. Inflammation and ageing: signaling pathways and intervention therapies. *Signal Transduct Target Ther.* 2023;8(1):239. doi:10.1038/s41392-023-01502-8
- Abbas H, Kamel R, El-Sayed N. Dermal anti-oxidant, anti-inflammatory and anti-ageing effects of compritol ATO-based resveratrol colloidal carriers prepared using mixed surfactants. *Int J Pharm.* 2018;541(1–2):37–47. doi:10.1016/j.ijpharm.2018.01.054
- Fougère B, Boulanger E, Nourhashémi F, Guyonnet S, Cesari M. Chronic inflammation: accelerator of biological ageing. *J Gerontol A Biol Sci Med Sci.* 2017;72(9):1218–1225. doi:10.1093/gerona/glw240
- Kim JE, Lee KW. Molecular targets of phytochemicals for skin inflammation. *Curr Pharm Des.* 2018;24(14):1533–1550. doi:10.2174/1381612824666180426113247
- Zhuang Y, Lyga J, Targets AD. Inflammageing in skin and other tissues - The roles of complement system and macrophage. *Inflamm Allergy Drug Targets.* 2014;13:153–161. doi:10.2174/1871528113666140522112003
- Giunta S, Wei Y, Xu K, Xia S. Cold-inflammageing: when a state of homeostatic-imbalance associated with ageing precedes the low-grade pro-inflammatory-state (inflammageing): meaning, evolution, inflammageing phenotypes. *Clin Exp Pharmacol Physiol.* 2022;49(9):925–934. doi:10.1111/1440-1681.13686
- Bektas A, Schurman SH, Sen R, Ferrucci L. Human T cell immunosenescence and inflammation in ageing. *J Leukoc Biol.* 2017;102(4):977–988. doi:10.1189/jlb.3RI0716-335R
- Zhang M, Hwang E, Lin P, Gao W, Ngo HTT, Yi TH. *Prunella vulgaris* L. exerts a protective effect against extrinsic ageing through NF- κ B, MAPKs, AP-1, and TGF- β /Smad signaling pathways in UVB-aged normal human dermal fibroblasts. *Rejuvenation Res.* 2018;21(4):313–322. doi:10.1089/rej.2017.1971
- Rovillain E, Mansfield L, Caetano C, et al. Activation of nuclear factor-kappa B signalling promotes cellular senescence. *Oncogene.* 2011;30(20):2356–2366. doi:10.1038/onc.2010.611
- Kawamura T, Ogawa Y, Aoki R, Shimada S. Innate and intrinsic antiviral immunity in skin. *J Dermatological Sci.* 2014;75(3):159–166. doi:10.1016/j.jdermsci.2014.05.004
- Liu F, Gu J. Retinoic acid inducible gene-I, more than a virus sensor. *Protein Cell.* 2011;2(5):351–357. doi:10.1007/s13238-011-1045-y
- Zhu H, Li Q, Huang Q, et al. RIG-I contributes to keratinocyte proliferation and wound repair by inducing TIMP-1 expression through NF- κ B signaling pathway. *J Cell Physiol.* 2023;238(8):1876–1890. doi:10.1002/jcp.31049
- Liu F, Wu S, Ren H, Gu J. Klotho suppresses RIG-I-mediated senescence-associated inflammation. *Nat Cell Biol.* 2011;13(3):254–262. doi:10.1038/ncb2167
- Zeng Y, Wang PH, Zhang M, Du JR. ageing-related renal injury and inflammation are associated with downregulation of Klotho and induction of RIG-I/NF- κ B signaling pathway in senescence-accelerated mice. *Ageing Clin Exp Res.* 2016;28(1):69–76. doi:10.1007/s40520-015-0371-y
- Wang J, Eming SA, Ding X. Role of mTOR signaling cascade in epidermal morphogenesis and skin barrier formation. *Biology.* 2022;11(6):931. doi:10.3390/biology11060931
- Leo M, Sivamani RK. Phytochemical modulation of the Akt/mTOR pathway and its potential use in cutaneous disease. *Arch Dermatol Res.* 2014;306(10):861–871. doi:10.1007/s00403-014-1480-8
- Laberge RM, Sun Y, Orjalo AV, et al. MTOR regulates the pro-tumorigenic senescence-associated secretory phenotype by promoting IL1A translation. *Nat Cell Biol.* 2015;17(8):1049–1061. doi:10.1038/ncb3195
- Fok WC, Zhang Y, Salmon AB, et al. Short-term treatment with rapamycin and dietary restriction have overlapping and distinctive effects in young mice. *J Gerontol A Biol Sci Med Sci.* 2013;68(2):108–116. doi:10.1093/gerona/gls127

30. Tominaga K, Suzuki HI. TGF- β signaling in cellular senescence and ageing-related pathology. *Int J Mol Sci.* 2019;20(20):5002. doi:10.3390/ijms20205002
31. Bora NS, Mazumder B, Mandal S, et al. Amelioration of UV radiation-induced photoageing by a combinational sunscreen formulation via aversion of oxidative collagen degradation and promotion of TGF- β -Smad-mediated collagen production. *Eur J Pharm Sci.* 2019;127:261–275. doi:10.1016/j.ejps.2018.11.004
32. Ke Y, Wang XJ. TGF β signaling in photoageing and UV-induced skin cancer. *J Invest Dermatol.* 2021;141(4s):1104–1110. doi:10.1016/j.jid.2020.11.007
33. Ikegami K, Yamashita M, Suzuki M, et al. Cellular senescence with SASP in periodontal ligament cells triggers inflammation in ageing periodontal tissue. *Ageing.* 2023;15(5):1279–1305.
34. Mendoza MC, Er EE, Blenis J. The Ras-ERK and PI3K-mTOR pathways: cross-talk and compensation. *Trends Biochem Sci.* 2011;36(6):320–328. doi:10.1016/j.tibs.2011.03.006
35. Yang HW, Lan Y, Li A, et al. Myricetin suppresses TGF- β -induced epithelial-to-mesenchymal transition in ovarian cancer. *Front Pharmacol.* 2023;14:1288883. doi:10.3389/fphar.2023.1288883
36. Quan T, Shao Y, He T, Voorhees JJ, Fisher GJ. Reduced expression of connective tissue growth factor (CTGF/CCN2) mediates collagen loss in chronologically aged human skin. *J Invest Dermatol.* 2010;130(2):415–424. doi:10.1038/jid.2009.224
37. Zhang Q, Fong CC, Yu WK, et al. Herbal formula astragali radix and rehmanniae radix exerted wound healing effect on human skin fibroblast cell line Hs27 via the activation of transformation growth factor (TGF- β) pathway and promoting extracellular matrix (ECM) deposition. *Phytomedicine.* 2012;20(1):9–16. doi:10.1016/j.phymed.2012.09.006
38. Herzig S, Shaw RJ. AMPK: guardian of metabolism and mitochondrial homeostasis. *Nat Rev Mol Cell Biol.* 2018;19(2):121–135. doi:10.1038/nrm.2017.95
39. Ido Y, Duranton A, Lan F, Weikel KA, Breton L, Ruderman NB. Resveratrol prevents oxidative stress-induced senescence and proliferative dysfunction by activating the AMPK-FOXO3 cascade in cultured primary human keratinocytes. *PLoS One.* 2015;10(2):e0115341. doi:10.1371/journal.pone.0115341
40. Crane ED, Wong W, Zhang H, O'Neil G, Crane JD. AMPK Inhibits mTOR-driven keratinocyte proliferation after skin damage and stress. *J Invest Dermatol.* 2021;141(9):2170–2177. doi:10.1016/j.jid.2020.12.036
41. Bharath LP, Agrawal M, McCambridge G, et al. Metformin enhances autophagy and normalizes mitochondrial function to alleviate ageing-associated inflammation. *Cell Metab.* 2020;32(1):44–55. doi:10.1016/j.cmet.2020.04.015
42. Xiang HC, Lin LX, Hu XF, et al. AMPK activation attenuates inflammatory pain through inhibiting NF- κ B activation and IL-1 β expression. *J Neuroinflammation.* 2019;16(1):34. doi:10.1186/s12974-019-1411-x
43. Desjardins EM, Smith BK, Day EA, et al. The phosphorylation of AMPK β 1 is critical for increasing autophagy and maintaining mitochondrial homeostasis in response to fatty acids. *Proc Natl Acad Sci USA.* 2022;119(48):e2119824119. doi:10.1073/pnas.2119824119
44. Chen Y, Liu Y, Jiang K, Wen Z, Cao X, Wu S. Linear ubiquitination of LKB1 activates AMPK pathway to inhibit NLRP3 inflammasome response and reduce chondrocyte pyroptosis in osteoarthritis. *J Orthop Transl.* 2023;39:1–11. doi:10.1016/j.jot.2022.11.002
45. Sadria M, Layton AT. Interactions among mTORC, AMPK and SIRT: a computational model for cell energy balance and metabolism. *Cell Commun Signal.* 2021;19(1):57. doi:10.1186/s12964-021-00706-1
46. Zheng Z, Wang M, Cheng C, et al. Ginsenoside Rb1 reduces H₂O₂-induced HUVEC dysfunction by stimulating the sirtuin-1/AMP-activated protein kinase pathway. *Mol Med Rep.* 2020;22(1):247–256. doi:10.3892/mmr.2020.11096
47. Walker KA, Basisty N, Wilson DM, Ferrucci L. connecting ageing biology and inflammation in the omics era. *J Clin Invest.* 2022;132(14). doi:10.1172/JCI158448
48. Lee JH, Moon JH, Lee YJ, Park SY. SIRT1, a class III histone deacetylase, regulates LPS-induced inflammation in human keratinocytes and mediates the anti-inflammatory effects of hinokitiol. *J Invest Dermatol.* 2017;137(6):1257–1266. doi:10.1016/j.jid.2016.11.044
49. Siebel C, Lendahl U. Notch signaling in development, tissue homeostasis, and disease. *Physiol Rev.* 2017;97(4):1235–1294. doi:10.1152/physrev.00005.2017
50. Wang Y, Li X, Xing X, et al. Notch-Hes1 Signaling regulates IL-17A(+) $\gamma\delta$ (+T) cell expression and IL-17A secretion of mouse psoriasis-like skin inflammation. *Mediators Inflamm.* 2020;2020:8297134. doi:10.1155/2020/8297134
51. Yoshioka H, Yamada T, Hasegawa S, et al. Senescent cell removal via JAG1-NOTCH1 signalling in the epidermis. *Exp Dermatol.* 2021;30(9):1268–1278. doi:10.1111/exd.14361
52. Huang Y, Xu W, Zhou R. NLRP3 inflammasome activation and cell death. *Cell Mol Immunol.* 2021;18(9):2114–2127. doi:10.1038/s41423-021-00740-6
53. Shim DW, Cho HJ, Hwang I, et al. Intracellular NAD(+) depletion confers a priming signal for NLRP3 inflammasome activation. *Front Immunol.* 2021;12:765477. doi:10.3389/fimmu.2021.765477
54. González-Domínguez A, Montañez R, Castejón-Vega B, et al. Inhibition of the NLRP3 inflammasome improves lifespan in animal murine model of Hutchinson-Gilford Progeria. *EMBO Mol Med.* 2021;13(10):e14012. doi:10.15252/emmm.202114012
55. Elieh Ali Komi D, Shafaghat F, Kovanen PT, Meri S. Mast cells and complement system: ancient interactions between components of innate immunity. *Allergy.* 2020;75(11):2818–2828. doi:10.1111/all.14413
56. Chehoud C, Rafail S, Tyldsley AS, Seykora JT, Lambris JD, Grice EA. Complement modulates the cutaneous microbiome and inflammatory milieu. *Proc Natl Acad Sci USA.* 2013;110(37):15061–15066. doi:10.1073/pnas.1307855110
57. Santoro A, Bientinesi E, Monti D. Immunosenescence and inflammaging in the ageing process: age-related diseases or longevity? *Ageing Res Rev.* 2021;71:101422. doi:10.1016/j.arr.2021.101422
58. Gu D, Wang H, Yan M, et al. *Echinacea purpurea* (L.) Moench extract suppresses inflammation by inhibition of C3a/C3aR signaling pathway in TNBS-induced ulcerative colitis rats. *J Ethnopharmacol.* 2023;307:116221. doi:10.1016/j.jep.2023.116221
59. Qiu W, Chen F, Feng X, Shang J, Luo X, Chen Y. Potential role of inflammaging mediated by the complement system in enlarged facial pores. *J Cosmet Dermatol.* 2024;23(1):27–32. doi:10.1111/jocd.15956
60. Zhou BX, Li J, Liang XL, et al. β -sitosterol ameliorates influenza A virus-induced proinflammatory response and acute lung injury in mice by disrupting the cross-talk between RIG-I and IFN/STAT signaling. *Acta Pharmacol Sin.* 2020;41(9):1178–1196. doi:10.1038/s41401-020-0403-9

61. Doles J, Storer M, Cozzuto L, Roma G, Keyes WM. Age-associated inflammation inhibits epidermal stem cell function. *Genes Dev.* 2012;26(19):2144–2153. doi:10.1101/gad.192294.112
62. Cheng Y, Chen J, Shi Y, Fang X, Tang Z. MAPK signaling pathway in oral squamous cell carcinoma: biological function and targeted therapy. *Cancers.* 2022;14(19):4625. doi:10.3390/cancers14194625
63. Mavrogonatu E, Konstantinou A, Kleatsas D. Long-term exposure to TNF- α leads human skin fibroblasts to a p38 MAPK- and ROS-mediated premature senescence. *Biogerontology.* 2018;19(3–4):237–249. doi:10.1007/s10522-018-9753-9
64. Ju Ho P, Jun Sung J, Ki Cheon K, Jin Tae H. Anti-inflammatory effect of *Centella asiatica* phytosome in a mouse model of phthalic anhydride-induced atopic dermatitis. *Phytomedicine.* 2018;43:110–119. doi:10.1016/j.phymed.2018.04.013
65. Maramaldi G, Togni S, Franceschi F, Lati E. Anti-inflammageing and antiglycation activity of a novel botanical ingredient from African biodiversity (Centevita™). *Clin Cosmet Invest Dermatol.* 2014;7:1–9. doi:10.2147/CCID.S49924
66. Milani M, Sparavigna A. The 24-hour skin hydration and barrier function effects of a hyaluronic 1%, glycerin 5%, and *Centella asiatica* stem cells extract moisturizing fluid: an intra-subject, randomized, assessor-blinded study. *Clin Cosmet Invest Dermatol.* 2017;10:311–315. doi:10.2147/CCID.S144180
67. Hu S, Belcaro G, Hosoi M, Feragalli B, Luzzi R, Dugall M. Postpartum stretchmarks: repairing activity of an oral *Centella asiatica* supplementation (Centellicum®). *Minerva Ginecologica.* 2018;70(5):629–634. doi:10.23736/S0026-4784.18.04254-5
68. Rahman MM, Islam MR, Akash S, et al. Pomegranate-specific natural compounds as onco-preventive and onco-therapeutic compounds: comparison with conventional drugs acting on the same molecular mechanisms. *Heliyon.* 2023;9(7):e18090. doi:10.1016/j.heliyon.2023.e18090
69. Saeed M, Naveed M, Bibi J, et al. The promising pharmacological effects and therapeutic/medicinal applications of *Punica Granatum* L. (Pomegranate) as a functional food in humans and animals. *Recent Pat Inflamm Allergy Drug Discov.* 2018;12:24–38. doi:10.2174/1872213X12666180221154713
70. Mohamad EA, Aly AA, Khalaf AA, et al. Evaluation of natural bioactive-derived punicalagin niosomes in skin-ageing processes accelerated by oxidant and ultraviolet radiation. *Drug Des Devel Ther.* 2021;15:3151–3162. doi:10.2147/DDDT.S316247
71. Abdellatif AAH, Alawadh SH, Bouazzaoui A, Alhowail AH, Mohammed HA. Anthocyanins rich pomegranate cream as a topical formulation with anti-ageing activity. *J Dermatol Treat.* 2021;32(8):983–990. doi:10.1080/09546634.2020.1721418
72. Chan LP, Tseng YP, Liu C, Liang CH. Fermented pomegranate extracts protect against oxidative stress and ageing of skin. *J Cosmet Dermatol.* 2022;21(5):2236–2245. doi:10.1111/jocd.14379
73. Vicentini FT, He T, Shao Y, et al. Quercetin inhibits UV irradiation-induced inflammatory cytokine production in primary human keratinocytes by suppressing NF- κ B pathway. *J Dermatological Sci.* 2011;61(3):162–168. doi:10.1016/j.jdermsci.2011.01.002
74. Chen H, Lu C, Liu H, et al. Quercetin ameliorates imiquimod-induced psoriasis-like skin inflammation in mice via the NF- κ B pathway. *Int Immunopharmacol.* 2017;48:110–117. doi:10.1016/j.intimp.2017.04.022
75. Shin EJ, Lee JS, Hong S, Lim TG, Byun S. Quercetin directly targets JAK2 and PKC δ and prevents UV-induced photoageing in human skin. *Int J Mol Sci.* 2019;20(21):1–10. doi:10.3390/ijms20215262
76. Ha AT, Rahmawati L, You L, Hossain MA, Kim JH, Cho JY. Anti-inflammatory, antioxidant, moisturizing, and antimelanogenesis effects of Quercetin 3-O- β -D-Glucuronide in human keratinocytes and melanoma cells via activation of NF- κ B and AP-1 pathways. *Int J Mol Sci.* 2021;23(1). doi:10.3390/ijms23010433
77. Ranjith-Kumar CT, Lai Y, Sarisky RT, Cheng Kao C. Green tea catechin, epigallocatechin gallate, suppresses signaling by the dsRNA innate immune receptor RIG-I. *PLoS One.* 2010;5(9):e12878. doi:10.1371/journal.pone.0012878
78. Zhang S, Liu X, Mei L, Wang H, Fang F. Epigallocatechin-3-gallate (EGCG) inhibits imiquimod-induced psoriasis-like inflammation of BALB/c mice. *BMC Complement Altern Med.* 2016;16(1):334. doi:10.1186/s12906-016-1325-4
79. Prasanth MI, Sivamaruthi BS, Chaiyasut C, Tencomnao T. A review of the role of green tea (*Camellia sinensis*) in anti-photoaging, stress resistance, neuroprotection, and autophagy. *Nutrients.* 2019;11(2):1–24. doi:10.3390/nu11020474
80. Gendrisch F, Esser PR, Schempp CM, Wölfe U. Luteolin as a modulator of skin ageing and inflammation. *BioFactors.* 2021;47(2):170–180. doi:10.1002/biof.1699
81. Kong R, Cui Y, Fisher GJ, et al. A comparative study of the effects of retinol and retinoic acid on histological, molecular, and clinical properties of human skin. *J Cosmet Dermatol.* 2016;15(1):49–57. doi:10.1111/jocd.12193
82. Metibemu DS, Ogungebe IV. Carotenoids in drug discovery and medicine: pathways and molecular targets implicated in human diseases. *Molecules.* 2022;27(18). doi:10.3390/molecules27186005
83. Teng WL, Huang PH, Wang HC, Tseng CH, Yen FL. Pterostilbene attenuates particulate matter-induced oxidative stress, inflammation and ageing in keratinocytes. *Antioxidants.* 2021;10(10):1–11.
84. Lee da H, Lee CS. Flavonoid myricetin inhibits TNF- α -stimulated production of inflammatory mediators by suppressing the Akt, mTOR and NF- κ B pathways in human keratinocytes. *Eur J Pharmacol.* 2016;784:164–172. doi:10.1016/j.ejphar.2016.05.025
85. Tian L, Ke D, Hong Y, et al. Artesunate treatment ameliorates ultraviolet irradiation-driven skin photoageing via increasing β -catenin expression. *Ageing.* 2021;13(23):25325–25341.
86. Jeong SI, Choi BM, Jang SI. Sulforaphane suppresses TARC/CCL17 and MDC/CCL22 expression through heme oxygenase-1 and NF- κ B in human keratinocytes. *Arch Pharmacol Res.* 2010;33(11):1867–1876. doi:10.1007/s12272-010-1120-6
87. Shirasugi I, Kamada M, Matsui T, Sakakibara Y, Liu MC, Suiko M. Sulforaphane inhibited melanin synthesis by regulating tyrosinase gene expression in B16 mouse melanoma cells. *Biosci Biotechnol Biochem.* 2010;74(3):579–582. doi:10.1271/bbb.90778
88. Kim H, Kim J, Park J, et al. Water extract of gromwell (*Lithospermum erythrorhizon*) enhances migration of human keratinocytes and dermal fibroblasts with increased lipid synthesis in an in vitro wound scratch model. *Skin Pharmacol Physiol.* 2012;25(2):57–64. doi:10.1159/000330897
89. Glynn KM, Anderson P, Fast DJ, Koedam J, Rebhun JF, Velliquette RA. Gromwell (*Lithospermum erythrorhizon*) root extract protects against glycation and related inflammatory and oxidative stress while offering UV absorption capability. *Exp Dermatol.* 2018;27(9):1043–1047. doi:10.1111/exd.13706
90. Klinggam W, Rungkamoltip P, Thongin S, et al. Polymethoxyflavones from *Kaempferia parviflora* ameliorate skin ageing in primary human dermal fibroblasts and ex vivo human skin. *Biomed Pharmacother.* 2022;145:112461. doi:10.1016/j.biopha.2021.112461

91. Khan AQ, Khan R, Rehman MU, et al. Soy isoflavones (daidzein & genistein) inhibit 12-O-tetradecanoylphorbol-13-acetate (TPA)-induced cutaneous inflammation via modulation of COX-2 and NF- κ B in Swiss albino mice. *Toxicology*. 2012;302(2–3):266–274. doi:10.1016/j.tox.2012.08.008
92. Upatcha N, Kaokaen P, Sorraksa N, et al. Nanoencapsulated cordyceps extract enhances collagen synthesis and skin cell regeneration through antioxidation and autophagy. *J Microencapsul*. 2023;40(5):303–317. doi:10.1080/02652048.2023.2198008
93. He H, Tang J, Ru D, et al. Protective effects of Cordyceps extract against UVB-induced damage and prediction of application prospects in the topical administration: an experimental validation and network pharmacology study. *Biomed Pharmacother*. 2020;121:109600. doi:10.1016/j.biopha.2019.109600
94. Liu A, Wu J, Li A, et al. The inhibitory mechanism of Cordyceps sinensis on cigarette smoke extract-induced senescence in human bronchial epithelial cells. *Int J Chronic Obstr*. 2016;11:1721–1731. doi:10.2147/COPD.S107396
95. Wong WC, Wu JY, Benzie IFF. Photoprotective potential of Cordyceps polysaccharides against ultraviolet B radiation-induced DNA damage to human skin cells. *Br J Dermatol*. 2011;164(5):980–986. doi:10.1111/j.1365-2133.2010.10201.x
96. Shao L, Jiang S, Li Y, et al. Aqueous extract of Cordyceps cicadae (Miq.) promotes hyaluronan synthesis in human skin fibroblasts: a potential moisturizing and anti-ageing ingredient. *PLoS One*. 2023;18(7):e0274479. doi:10.1371/journal.pone.0274479
97. Prommaban A, Sriyab S, Marsup P, et al. Comparison of chemical profiles, antioxidation, inhibition of skin extracellular matrix degradation, and anti-tyrosinase activity between mycelium and fruiting body of Cordyceps militaris and Isaria tenuipes. *Pharm Biol*. 2022;60(1):225–234. doi:10.1080/13880209.2021.2025255
98. Gong X, Jiang L, Li W, Liang Q, Li Z. Curcumin induces apoptosis and autophagy in human renal cell carcinoma cells via Akt/mTOR suppression. *Bioengineered*. 2021;12(1):5017–5027. doi:10.1080/21655979.2021.1960765
99. Zia A, Farkhondeh T, Pourbagher-Shahri AM, Samarghandian S. The role of curcumin in ageing and senescence: molecular mechanisms. *Biomed Pharmacother*. 2021;134:111119. doi:10.1016/j.biopha.2020.111119
100. Mazzarino L, Silva LF, Curta JC, et al. Curcumin-loaded lipid and polymeric nanocapsules stabilized by nonionic surfactants: an in vitro and In vivo antitumor activity on B16-F10 melanoma and macrophage uptake comparative study. *J Biomed Nanotechnol*. 2011;7(3):406–414. doi:10.1166/jbn.2011.1296
101. Vollono L, Falconi M, Gaziano R, et al. Potential of curcumin in skin disorders. *Nutrients*. 2019;11(9):1–25. doi:10.3390/nu11092169
102. Asada K, Ohara T, Muroyama K, Yamamoto Y, Murosaki S. Effects of hot water extract of Curcuma longa on human epidermal keratinocytes in vitro and skin conditions in healthy participants: a randomized, double-blind, placebo-controlled trial. *J Cosmet Dermatol*. 2019;18(6):1866–1874. doi:10.1111/jocd.12890
103. Roy T, Boateng ST, Banang-Mbeumi S, et al. Synthesis, inverse docking-assisted identification and in vitro biological characterization of Flavonol-based analogs of fisetin as c-Kit, CDK2 and mTOR inhibitors against melanoma and non-melanoma skin cancers. *Bioorg Chem*. 2021;107:104595. doi:10.1016/j.bioorg.2020.104595
104. Roy T, Banang-Mbeumi S, Boateng ST, et al. Dual targeting of mTOR/IL-17A and autophagy by fisetin alleviates psoriasis-like skin inflammation. *Front Immunol*. 2022;13:1075804. doi:10.3389/fimmu.2022.1075804
105. Chamcheu JC, Esnault S, Adhami VM, et al. Fisetin, a 3,7,3',4'-tetrahydroxyflavone inhibits the PI3K/Akt/mTOR and MAPK pathways and ameliorates psoriasis pathology in 2D and 3D organotypic human inflammatory skin models. *Cells*. 2019;8(9):1–27. doi:10.3390/cells8091089
106. Syed DN, Adhami VM, Khan MI, Mukhtar H. Inhibition of Akt/mTOR signaling by the dietary flavonoid fisetin. *Anticancer Agents Med Chem*. 2013;13(7):995–1001. doi:10.2174/18715206113139990129
107. Shon MS, Kim RH, Kwon OJ, Roh SS, Kim GN. Beneficial role and function of fisetin in skin health via regulation of the CCN2/TGF- β signaling pathway. *Food Sci Biotechnol*. 2016;25(Suppl 1):133–141. doi:10.1007/s10068-016-0110-y
108. Chiang H-M, Chan S-Y, Chu Y, Wen K-C. Fisetin ameliorated photodamage by suppressing the mitogen-activated protein Kinase/Matrix metalloproteinase pathway and nuclear factor- κ B pathways. *J Agric Food Chem*. 2015;63(18):4551–4560. doi:10.1021/jf502500t
109. Wu PY, Lyu JL, Liu YJ, et al. Fisetin regulates Nrf2 Expression and the inflammation-related signaling pathway to prevent UVB-induced skin damage in hairless mice. *Int J Mol Sci*. 2017;18(10):1–23. doi:10.3390/ijms18102118
110. Waswa EN, Li J, Mkala EM, et al. Ethnobotany, phytochemistry, pharmacology, and toxicology of the genus *Sambucus* L. (Viburnaceae). *J Ethnopharmacol*. 2022;292:115102. doi:10.1016/j.jep.2022.115102
111. Hu X, Yang Y, Tang S, et al. Anti-ageing effects of anthocyanin extracts of *Sambucus canadensis* caused by targeting mitochondrial-induced oxidative stress. *Int J Mol Sci*. 2023;24(2):1528. doi:10.3390/ijms24021528
112. Widyadharma IP, Purwata T, Suprpta D, Sudewi R. Anthocyanin derived from Purple Sweet Potato water extracts ameliorated oxidative stress, inflammation, mechanical allodynia, and cold allodynia among chronic constriction injury-Induced neuropathic pain in rats. *Open Access Maced J Med Sci*. 2020;8:1–9. doi:10.3889/oamjms.2020.4524
113. Han Y, Guo Y, Cui SW, Li H, Shan Y, Wang H. Purple sweet potato extract extends lifespan by activating autophagy pathway in male Drosophila melanogaster. *Exp Gerontology*. 2021;144:111190. doi:10.1016/j.exger.2020.111190
114. Sajadimajd S, Bahramsoltani R, Iranpanah A, et al. Advances on natural polyphenols as anticancer agents for skin cancer. *Pharmacol Res*. 2020;151:104584. doi:10.1016/j.phrs.2019.104584
115. Hao J, Gao Y, Xue J, et al. Phytochemicals, pharmacological effects and molecular mechanisms of mulberry. *Foods*. 2022;11(8):1–19. doi:10.3390/foods11081170
116. Biscaro RC, Mussi L, Sufi B, et al. Modulation of autophagy by an innovative phytocosmetic preparation (*Myrothamnus flabelifolia* and *Coffea arabica*) in human fibroblasts and its effects in a clinical randomized placebo-controlled trial. *J Cosmet Dermatol*. 2022;21(10):4901–4912. doi:10.1111/jocd.14888
117. Salekeen R, Ahmed A, Islam ME, Billah MM, Rahman H, Islam KMD. In-silico screening of bioactive phytopeptides for novel anti-ageing therapeutics. *J Biomol Struct Dyn*. 2022;40(10):4475–4487. doi:10.1080/07391102.2020.1859411
118. Chu J, Xiang Y, Lin X, et al. Handelin protects human skin keratinocytes against ultraviolet B-induced photodamage via autophagy activation by regulating the AMPK-mTOR signaling pathway. *Arch Biochem Biophys*. 2023;743:109646. doi:10.1016/j.abb.2023.109646
119. Dong L, Lee H, Liu Z, Lee DS. Anti-skin inflammatory and anti-oxidative effects of the neoflavonoid latifolin isolated from *Dalbergia odorifera* in HaCaT and BJ-5ta cells. *Int J Mol Sci*. 2023;24(8):1–10. doi:10.3390/ijms24087371

120. Bakrim S, Benkhaira N, Bourais I, et al. Health benefits and pharmacological properties of stigmasterol. *Antioxidants*. 2022;11(10). doi:10.3390/antiox11101912
121. Im AR, Kim YM, Chin YW, Chae S. Protective effects of compounds from *Garcinia mangostana* L. (mangosteen) against UVB damage in HaCaT cells and hairless mice. *Int J Mol Med*. 2017;40(6):1941–1949. doi:10.3892/ijmm.2017.3188
122. Wang F, Ma H, Liu Z, Huang W, Xu X, Zhang X. α -Mangostin inhibits DMBA/TPA-induced skin cancer through inhibiting inflammation and promoting autophagy and apoptosis by regulating PI3K/Akt/mTOR signaling pathway in mice. *Biomed Pharmacother*. 2017;92:672–680. doi:10.1016/j.biopha.2017.05.129
123. Chen S, Han K, Li H, et al. Isogarcinol extracted from *Garcinia mangostana* L. ameliorates imiquimod-induced psoriasis-like skin lesions in mice. *J Agric Food Chem*. 2017;65(4):846–857. doi:10.1021/acs.jafc.6b05207
124. Wang D, Wang E, Li Y, et al. Anti-ageing effect of *Momordica charantia* L. on d-Galactose-induced subacute ageing in mice by activating PI3K/AKT signaling pathway. *Molecules*. 2022;27(14):1–14.
125. Woo J, Shin S, Ji H, et al. *Isatis tinctoria* L. leaf extract inhibits replicative senescence in dermal fibroblasts by regulating mTOR-NF- κ B-SASP signaling. *Nutrients*. 2022;14(9):1–13. doi:10.3390/nu14091979
126. Wolosik K, Chalecka M, Palka J, Surazynski A. Protective effect of *amaranthus cruentus* L. seed oil on UVA-radiation-induced apoptosis in human skin fibroblasts. *Int J Mol Sci*. 2023;24(13):1–15. doi:10.3390/ijms241310795
127. Das G, Farhan M, Sinha S, Bora HK, Singh WR, Meeran SM. *Mikania micrantha* extract enhances cutaneous wound healing activity through the activation of FAK/Akt/mTOR cell signaling pathway. *Injury*. 2023;54(8):110856. doi:10.1016/j.injury.2023.110856
128. Park B, Hwang E, Seo SA, Cho J-G, Yang J-E, Yi T-H. Eucalyptus globulus extract protects against UVB-induced photoageing by enhancing collagen synthesis via regulation of TGF- β /Smad signals and attenuation of AP-1. *Arch Biochem Biophys*. 2018;637:31–39. doi:10.1016/j.abb.2017.11.007
129. Lee K, Ham I, Yang G, et al. Vasorelaxant effect of *Prunus yedoensis* bark. *BMC Complement Altern Med*. 2013;13:31. doi:10.1186/1472-6882-13-31
130. Lee J, Yang G, Lee K, et al. Anti-inflammatory effect of *Prunus yedoensis* through inhibition of nuclear factor- κ B in macrophages. *BMC Complement Altern Med*. 2013;13:92. doi:10.1186/1472-6882-13-92
131. Li L, Hwang E, Ngo HTT, et al. Antiphotageing effect of *Prunus yeonensis* blossom extract via inhibition of MAPK/AP-1 and regulation of the TGF- β /Smad and Nrf2/ARE signaling pathways. *Photochem Photobiol*. 2018;94(4):725–732. doi:10.1111/php.12894
132. Zhang YQ, Guan L, Zhong ZY, et al. The anti-inflammatory effect of cherry blossom extract (*Prunus yedoensis*) used in soothing skincare product. *Int J Cosmet Sci*. 2014;36(6):527–530. doi:10.1111/ics.12149
133. Moreira P, Sousa FJ, Matos P, et al. Chemical composition and effect against skin alterations of bioactive extracts obtained by the hydrodistillation of *Eucalyptus globulus* leaves. *Pharmaceutics*. 2022;14(3):561. doi:10.3390/pharmaceutics14030561
134. Rios JL, Giner RM, Marín M, Recio MC. A pharmacological update of ellagic acid. *Planta Med*. 2018;84(15):1068–1093. doi:10.1055/a-0633-9492
135. Moon NR, Kang S, Park S. Consumption of ellagic acid and dihydromyricetin synergistically protects against UV-B induced photoageing, possibly by activating both TGF- β 1 and wnt signaling pathways. *J Photochem Photobiol B Biol*. 2018;178:92–100. doi:10.1016/j.jphotobiol.2017.11.004
136. Bae JY, Choi JS, Kang SW, Lee YJ, Park J, Kang YH. Dietary compound ellagic acid alleviates skin wrinkle and inflammation induced by UV-B irradiation. *Exp Dermatol*. 2010;19(8):e182–190. doi:10.1111/j.1600-0625.2009.01044.x
137. Duckworth C, Stutts J, Clatterbuck K, Nosoudi N. Effect of ellagic acid and retinoic acid on collagen and elastin production by human dermal fibroblasts. *Bio-Med Mater Eng*. 2023;34(5):473–480. doi:10.3233/BME-230007
138. Abd-Elghany AA, Mohamad EA. Chitosan-coated niosomes loaded with ellagic acid present antiageing activity in a skin cell line. *ACS Omega*. 2023;8(19):16620–16629. doi:10.1021/acsomega.2c07254
139. Farwick M, Köhler T, Schild J, et al. Pentacyclic triterpenes from *Terminalia arjuna* show multiple benefits on aged and dry skin. *Skin Pharmacol Physiol*. 2014;27(2):71–81. doi:10.1159/000351387
140. Majed F, Nafees S, Rashid S, et al. *Terminalia chebula* attenuates DMBA/Croton oil-induced oxidative stress and inflammation in Swiss albino mouse skin. *Toxicol Int*. 2015;22(1):21–29. doi:10.4103/0971-6580.172252
141. Oh JH, Karadeniz F, Lee JI, Seo Y, Kong CS. *Oleracone C* from *portulaca oleracea* attenuates UVB-induced changes in matrix metalloproteinase and type I procollagen production via MAPK and TGF- β /Smad pathways in human keratinocytes. *Int J Cosmet Sci*. 2023;45(2):166–176. doi:10.1111/ics.12828
142. Oh JH, Seo Y, Kong CS. Anti-photoageing effects of solvent-partitioned fractions from *Portulaca oleracea* L. on UVB-stressed human keratinocytes. *J Food Biochem*. 2019;43(4):e12814. doi:10.1111/jfbc.12814
143. Kim MS, Oh GH, Kim MJ, Hwang JK. Fucosterol inhibits matrix metalloproteinase expression and promotes type-I procollagen production in UVB-induced HaCaT cells. *Photochem Photobiol*. 2013;89(4):911–918. doi:10.1111/php.12061
144. Oh JH, Karadeniz F, Lee JI, Park SY, Seo Y, Kong CS. Anticatabolic and anti-inflammatory effects of myricetin 3-O- β -D-Galactopyranoside in UVA-irradiated dermal cells via repression of MAPK/AP-1 and activation of TGF β /Smad. *Molecules*. 2020;25(6):1–18. doi:10.3390/molecules25061331
145. Zeng J, An M, Tian W, Wang K, Du B, Li P. Sacha inchi albumin delays skin-ageing by alleviating inflammation, oxidative stress and regulating gut microbiota in d-galactose induced-ageing mice. *J Sci Food Agric*. 2023;103(9):4470–4480. doi:10.1002/jsfa.12555
146. Bak SG, Lim HJ, Park EJ, et al. Effects of *Vigna angularis* extract and its active compound hemiphloin against atopic dermatitis-like skin inflammation. *Heliyon*. 2023;9(2):e12994. doi:10.1016/j.heliyon.2023.e12994
147. Qu L, Wang F, Chen Y. Protective effect and mechanism research of *Phyllanthus emblica* Linn. fruit extract on UV-induced photodamage in keratinocytes. *Photochem Photobiol Sci*. 2023;22(8):1945–1959. doi:10.1007/s43630-023-00423-3
148. Guo Y, Zhang Y, Wang YS, Ma L, Liu H, Gao W. Protective effect of *Salvia plebeia* R. Br ethanol extract on UVB-induced skin photoageing in vitro and in vivo. *Photodermatol Photoimmunol Photomed*. 2023;39(5):466–477. doi:10.1111/phpp.12879
149. Liu J, Wu Y, Ma W, et al. Anti-inflammatory activity of *Panax notoginseng* flower saponins quantified using LC/MS/MS. *Molecules*. 2023;28(5):2416. doi:10.3390/molecules28052416
150. Liu XY, Hwang E, Park B, Ngo HTT, Xiao YK, Yi TH. Ginsenoside C-Mx isolated from *Notoginseng* stem-leaf ginsenosides attenuates ultraviolet B-mediated photoageing in human dermal fibroblasts. *Photochem Photobiol*. 2018;94(5):1040–1048. doi:10.1111/php.12940

151. Jin YJ, Ji Y, Jang YP, Choung SY. *Acer tataricum* subsp. *ginnala* inhibits skin photoageing via regulating MAPK/AP-1, NF- κ B, and TGF β /Smad signaling in UVB-irradiated human dermal fibroblasts. *Molecules*. 2021;26(3):1–16. doi:10.3390/molecules26030662
152. Ye T, Zhu S, Zhu Y, et al. Cryptotanshinone induces melanoma cancer cells apoptosis via ROS-mitochondrial apoptotic pathway and impairs cell migration and invasion. *Biomed Pharmacother*. 2016;82:319–326. doi:10.1016/j.biopha.2016.05.015
153. Maione F, Mascolo N. Danshen and the Cardiovascular system: new advances for an old remedy. *Semin Thromb Hemost*. 2016;42(3):321–322. doi:10.1055/s-0036-1580086
154. Guo K, Liu R, Jing R, et al. Cryptotanshinone protects skin cells from ultraviolet radiation-induced photoageing via its antioxidant effect and by reducing mitochondrial dysfunction and inhibiting apoptosis. *Front Pharmacol*. 2022;13:1036013. doi:10.3389/fphar.2022.1036013
155. Wu Z, Song L, Liu SQ, Huang D. Tanshinones extend chronological lifespan in budding yeast *Saccharomyces cerevisiae*. *Appl Microbiol Biotechnol*. 2014;98(20):8617–8628. doi:10.1007/s00253-014-5890-5
156. Tang L, He S, Wang X, et al. Cryptotanshinone reduces psoriatic epidermal hyperplasia via inhibiting the activation of STAT3. *Exp Dermatol*. 2018;27(3):268–275. doi:10.1111/exd.13511
157. Li Y, Shi S, Gao J, et al. Cryptotanshinone downregulates the profibrotic activities of hypertrophic scar fibroblasts and accelerates wound healing: a potential therapy for the reduction of skin scarring. *Biomed Pharmacother*. 2016;80:80–86. doi:10.1016/j.biopha.2016.03.006
158. Liao D, Chen Y, Guo Y, et al. Salvianolic acid B improves chronic mild stress-induced depressive behaviors in rats: involvement of AMPK/SIRT1 signaling pathway. *J Inflamm Res*. 2020;13:195–206. doi:10.2147/JIR.S249363
159. Kang K, Jho EH, Lee HJ, et al. *Youngia denticulata* protects against oxidative damage induced by tert-butylhydroperoxide in HepG2 cells. *J Med Food*. 2011;14(10):1198–1207. doi:10.1089/jmf.2010.1557
160. Kim KA, Kang KD, Lee EH, Nho CW, Jung SH. Edible wild vegetable, *Gymnaster koraiensis* protects retinal ganglion cells against oxidative stress. *Food Chem Toxicol*. 2011;49(9):2131–2143. doi:10.1016/j.fct.2011.05.028
161. Kim M, Park YG, Lee HJ, Lim SJ, Nho CW. Youngiasides A and C isolated from *youngia denticulata* inhibit UVB-induced MMP expression and promote type I procollagen production via repression of MAPK/AP-1/NF- κ B and activation of AMPK/Nrf2 in HaCaT cells and human dermal fibroblasts. *J Agric Food Chem*. 2015;63(22):5428–5438. doi:10.1021/acs.jafc.5b00467
162. Yoon SJ, Lim CJ, Chung HJ, et al. Autophagy activation by crepidiastrum denticulatum extract attenuates environmental pollutant-Induced damage in dermal fibroblasts. *Int J Mol Sci*. 2019;20(3):517. doi:10.3390/ijms20030517
163. Akinduro O, Sully K, Patel A, et al. Constitutive autophagy and nucleophagy during Epidermal Differentiation. *J Invest Dermatol*. 2016;136(7):1460–1470. doi:10.1016/j.jid.2016.03.016
164. Kim HS, Park SY, Moon SH, Lee JD, Kim S. Autophagy in human skin fibroblasts: impact of age. *Int J Mol Sci*. 2018;19(8):1–13.
165. Lou T, Huang Q, Su H, Zhao D, Li X. Targeting Sirtuin 1 signaling pathway by ginsenosides. *J Ethnopharmacol*. 2021;268:113657. doi:10.1016/j.jep.2020.113657
166. Shin D, Kim J, Lim T, et al. 20-O- β -D-glucopyranosyl-20(S)-protopanaxadiol suppresses UV-Induced MMP-1 expression through AMPK-mediated mTOR inhibition as a downstream of the PKA-LKB1 pathway. *J Cell Biochem*. 2014;115(10):1702–1711. doi:10.1002/jcb.24833
167. Meng H, Liu XK, Li JR, Bao TY, Yi F. Bibliometric analysis of the effects of ginseng on skin. *J Cosmet Dermatol*. 2022;21(1):99–107. doi:10.1111/jocd.14450
168. Kimura Y, Sumiyoshi M, Sakanaka M. Effects of ginsenoside Rb₁ on skin changes. *J Biomed Biotechnol*. 2012;2012:946242. doi:10.1155/2012/946242
169. Tian T, Ko CN, Luo W, Li D, Yang C. The anti-ageing mechanism of ginsenosides with medicine and food homology. *Food Funct*. 2023;14(20):9123–9136.
170. Costa EF, Magalhães WV, Di Stasi LC. Recent advances in herbal-derived products with skin anti-ageing properties and cosmetic applications. *Molecules*. 2022;27(21):1–29.
171. Huang WC, Chen CY, Wu SJ. Almond skin polyphenol extract inhibits inflammation and promotes lipolysis in differentiated 3T3-L1 adipocytes. *J Med Food*. 2017;20(2):103–109. doi:10.1089/jmf.2016.3806
172. Tungmunthum D, Abid M, Elamrani A, Drouet S, Addi M, Hano C. Almond skin extracts and chlorogenic acid delay chronological ageing and enhanced oxidative stress response in yeast. *Life*. 2020;10(6):1–18. doi:10.3390/life10060080
173. Choi D, Kang W, Park S, Son B, Park T. β -ionone attenuates dexamethasone-induced suppression of collagen and hyaluronic acid synthesis in human dermal fibroblasts. *Biomolecules*. 2021;11(5):1–14. doi:10.3390/biom11050619
174. Shim MS, Je NJ, Lee DY. Aromatherapy massage for relief of pruritus and stress in older women. *Altern Ther Health Med*. 2023;29(2):36–41.
175. Lin TK, Zhong L, Santiago JL. Anti-inflammatory and skin barrier repair effects of topical application of some plant oils. *Int J Mol Sci*. 2017;19(1):70. doi:10.3390/ijms19010070
176. Hajhashemi M, Rafeian M, Rouhi Boroujeni HA, et al. The effect of Aloe vera gel and sweet almond oil on striae gravidarum in nulliparous women. *J Matern Fetal Neonatal Med*. 2018;31(13):1703–1708. doi:10.1080/14767058.2017.1325865
177. Li JN, Henning SM, Thames G, et al. Almond consumption increased UVB resistance in healthy Asian women. *J Cosmet Dermatol*. 2021;20(9):2975–2980. doi:10.1111/jocd.13946
178. Rybak I, Carrington AE, Dhaliwal S, et al. Prospective randomized controlled trial on the effects of almonds on facial wrinkles and pigmentation. *Nutrients*. 2021;13(3):785. doi:10.3390/nu13030785
179. Song L, Chen X, Mi L, et al. Icaritin-induced inhibition of SIRT6/NF- κ B triggers redox mediated apoptosis and enhances anti-tumor immunity in triple-negative breast cancer. *Cancer Sci*. 2020;111(11):4242–4256. doi:10.1111/cas.14648
180. Lee HR, Ryu HG, Lee Y, et al. Effect of aronia extract on collagen synthesis in human skin cell and dermal equivalent. *Oxid Med Cell Longev*. 2022;2022:4392256. doi:10.1155/2022/4392256
181. Akhtar N, Khan HM, Ashraf S, Mohammad IS, Saqib NU, Bashir K. Moisturizing effect of stable cream containing *Crocus sativus* extracts. *Pak J Pharm Sci*. 2014;27(6):1881–1884.
182. Bisht A, Tewari D, Kumar S, Chandra S. Network pharmacology, molecular docking, and molecular dynamics simulation to elucidate the mechanism of anti-ageing action of *Tinospora cordifolia*. *Mol Divers*. 2023;28(3):1743–1763. doi:10.1007/s11030-023-10684-w
183. Gam DH, Hong JW, Kim JH, Kim JW. Skin-whitening and anti-wrinkle effects of bioactive compounds isolated from peanut shell using ultrasound-assisted extraction. *Molecules*. 2021;26(5):1231. doi:10.3390/molecules26051231

184. Yi M, Fasina OB, Li Y, Xiang L, Qi J. Mixture of peanut skin extract, geniposide, and isoquercitrin improves the hepatic lipid accumulation of mice via modification of gut microbiota homeostasis and the TLR4 and AMPK signaling pathways. *Int J Mol Sci.* 2023;24(23):16684. doi:10.3390/ijms242316684
185. Rafailovska E, Tushevski O, Shijakova K, Simic SG, Kjojkarovska SD, Miova B. *Hypericum perforatum* L. extract exerts insulinotropic effects and inhibits gluconeogenesis in diabetic rats by regulating AMPK expression and PKC ϵ concentration. *J Ethnopharmacol.* 2023;302:115899. doi:10.1016/j.jep.2022.115899
186. Zhang S, Zhang J, Yu J, et al. Hyperforin ameliorates imiquimod-induced psoriasis-like murine skin inflammation by modulating IL-17A-producing $\gamma\delta$ T cells. *Front Immunol.* 2021;12:635076. doi:10.3389/fimmu.2021.635076
187. LaFoya B, Munroe JA, Albig AR. A comparison of resveratrol and other polyphenolic compounds on Notch activation and endothelial cell activity. *PLoS One.* 2019;14(1):e0210607. doi:10.1371/journal.pone.0210607
188. Ghafouri-Fard S, Bahroudi Z, Shoorei H, et al. Disease-associated regulation of gene expression by resveratrol: special focus on the PI3K/AKT signaling pathway. *Can Cell Inter.* 2022;22(1):298. doi:10.1186/s12935-022-02719-3
189. Carlucci CD, Hui Y, Chumanevich AP, et al. Resveratrol protects against skin inflammation through inhibition of mast cell, sphingosine Kinase-1, Stat3 and NF- κ B p65 signaling activation in mice. *Int J Mol Sci.* 2023;24(7):1–14. doi:10.3390/ijms24076707
190. Chaiyana W, Charoensup W, Sriyab S, Punyoyai C, Neimkhum W. Herbal extracts as potential antioxidant, anti-ageing, anti-inflammatory, and whitening cosmeceutical ingredients. *Chem Biodivers.* 2021;18(7):e2100245. doi:10.1002/cbdv.202100245
191. Won YR, Won KJ, Kim DY, Kim MJ, Hong BS, Lee HM. Chemical composition of *Impatiens textori* Miq. flower absolute and its potential wound repair and anti-Melanogenesis-promoting activities in skin cells. *Pharmaceuticals.* 2022;15(11):1–15. doi:10.3390/ph15111397
192. Lee CH, Yang H, Park JHY, Kim JE, Lee KW. Piceatannol, a metabolite of resveratrol, attenuates atopic dermatitis by targeting Janus kinase 1. *Phytomedicine.* 2022;99:153981.
193. Maruki-Uchida H, Kurita I, Sugiyama K, Sai M, Maeda K, Ito T. The protective effects of piceatannol from passion fruit (*Passiflora edulis*) seeds in UVB-irradiated keratinocytes. *Biol Pharm Bull.* 2013;36(5):845–849. doi:10.1248/bpb.b12-00708
194. Maruki-Uchida H, Morita M, Yonei Y, Sai M. Effect of passion fruit seed extract rich in piceatannol on the skin of women: a randomized, placebo-controlled, double-blind trial. *J Nutr Sci Vitaminol.* 2018;64(1):75–80. doi:10.3177/jnsv.64.75
195. Kumar M, Prasad SK, Hemalatha S. A current update on the phytopharmacological aspects of *Houttuynia cordata* Thunb. *Pharmacogn Rev.* 2014;8(15):22–35. doi:10.4103/0973-7847.125525
196. Mapoung S, Umsumarn S, Semmarath W, et al. Photoprotective effects of a hyperoside-enriched fraction prepared from *Houttuynia cordata* Thunb. on ultraviolet B-induced skin ageing in human fibroblasts through the MAPK signaling pathway. *Plants.* 2021;10(12):1–20. doi:10.3390/plants10122628
197. Ibrahim N, Nadian I, Noor DR, Fadilah F. Prediction of translational regulation by network interaction in synaptic plasticity induced with *Centella asiatica*. *Sci World J.* 2023;2023:4199614. doi:10.1155/2023/4199614
198. Gao Y, Liang Z, Lv N, et al. Exploring the total flavones of *Abelmoschus manihot* against IAV-induced lung inflammation by network pharmacology. *BMC Complement Med Therap.* 2022;22(1):36. doi:10.1186/s12906-022-03509-0
199. Di Pettillo A, Fais A, Pintus F, et al. Broad-range potential of *Asphodelus microcarpus* leaves extract for drug development. *BMC Microbiol.* 2017;17(1):159. doi:10.1186/s12866-017-1068-5
200. Yang N, Zou L, Wang Y. Label-free quantitative proteomic analysis of reserpine-induced depression in mice intervened by berberine. *Pak J Pharm Sci.* 2022;35(1):151–155.
201. Na Takuathung M, Klinjan P, Sakuludomkan W, et al. Efficacy and safety of the genistein nutraceutical product containing Vitamin E, Vitamin B3, and ceramide on skin health in postmenopausal women: a randomized, double-blind, placebo-controlled clinical trial. *J Clin Med.* 2023;12(4):1–12. doi:10.3390/jcm12041326
202. Han S-Y, Jang TW, Park H-J, et al. *Nyssa fruticans* Wurmb inhibits melanogenesis in isobutylmethylxanthine-treated melanoma via the PI3K/AKT/mTOR/CREB and MAPK signaling pathways. *Exp Ther Med.* 2022;24.
203. Tao Y, Bao J, Zhu F, Pan M, Liu Q, Wang P. Ethnopharmacology of *Rubus idaeus* Linnaeus: a critical review on ethnobotany, processing methods, phytochemicals, pharmacology and quality control. *J Ethnopharmacol.* 2023;302(Pt A):115870. doi:10.1016/j.jep.2022.115870
204. Shen CY, Jiang JG, Yang L, Wang DW, Zhu W. Anti-ageing active ingredients from herbs and nutraceuticals used in traditional Chinese medicine: pharmacological mechanisms and implications for drug discovery. *Br J Pharmacol.* 2017;174(11):1395–1425. doi:10.1111/bph.13631

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