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Kukoamine B Ameliorate Insulin Resistance, Oxidative Stress, Inflammation and Other Metabolic Abnormalities in High-Fat/High-Fructose-Fed Rats

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submit your manuscript | www.dovepress.com DovePress http://doi.org/10.2147/DMSO.S247844 **Background:** Obesity is characterized by excessive body fat, insulin resistance and dyslipidemia, which increases the chances of developing chronic diseases like type 2 diabetes, cardiovascular diseases, hypertension, nonalcoholic fatty liver diseases, some types of cancers and neurodegenerative diseases. Kukoamine B (Kuk B) is a spermine alkaloid obtained from *Lycium chinense*, and it has been shown to possess antidiabetic, antioxidant and anti-inflammatory properties. In this study, we evaluated the therapeutic effect of Kuk B on high-fat diet/high-fructose (HFDFr)-induced insulin resistance and obesity in experimental rats.

Materials and Methods: Rats were fed with either normal rat diet or HFDFr for 10 consecutive weeks. The groups that were fed with HFDFr received Kuk B (25 and 50 mg/kg) from the beginning of the 6th week to the 10th week. After treatment, the effect of Kuk B on body weight, food, water intake, insulin, blood glucose, serum biochemical parameters, hepatic oxidative stress (malondialdehyde (MDA), superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GSH-Px) and proinflammatory cytokine (interleukin (IL)-6, interleukin (IL)-1 β and tumor necrosis factor alpha (TNF- α)) levels was determined. Histopathological analysis of the liver tissues was also performed.

Results: HFDFr-fed rats showed a significant increase in body weight, fasting blood glucose, insulin, lipid accumulation and liver function enzymes. In addition, HFDFr diet increased hepatic MDA, TNF- α , IL-1 β and IL-6 and decreased hepatic SOD, CAT and GSH-Px activities. On the other hand, Kuk B significantly attenuated body weight, insulin resistance, lipid accumulation, oxidative stress and inflammation.

Conclusion: These results indicated that Kuk B showed protective effect against HFDFrinduced metabolic disorders by downregulating lipid accumulation, oxidative stress and inflammatory factors.

Keywords: kukoamine B, obesity, insulin resistance, high-fat diet, inflammation, oxidative stress

Introduction

The incidence of obesity has increased sporadically in the last two decades and it is gradually becoming a global epidemic that needs urgent attention. In 2014, the World Health Organization estimated that 600 million people were obese, while another 2 billion adults were considered overweight.¹ Obesity is a chronic metabolic disorder that bears the hallmark of excessive body fat accumulation and it is associated with the development of several chronic health conditions such as insulin resistance, type 2 diabetes, cardiovascular diseases, hypertension, nonalcoholic

Diabetes, Metabolic Syndrome and Obesity: Targets and Therapy 2020:13 1843–1853 1843 2020:13 1843–1853 1843 2020:13 1843–1853 1843 2020:13 1843–1853 1843 2020:13 1843–1853 1843 2020:13 1843–1853 1843 2020:13 1843–1853 1843 2020:13 1843–1853 1843 2020:13 1843–1853 1843 2020:13 1843–1853 1843 2020:13 1843–1853 1843 2020:13 1843–1853 1843 2020:13 1843–1853 1843 2020:13 1843–1853 1843 2020:13 1843–1853 1843 2020:13 1843–1853 1843 2020:13 1843–1853 1843 2020:13 1843–1853 1843 2020:13 1843–1853 2020:13 1843–1853 1843 2020:13 1843–1853 1843 2020:13 1843–1853 1843 2020:13 1843–1853 1843 2020:13 1843–1853 1843–1853 2020:13 1843–1853 1843 2020:13 1843–1853 1843–1853 2020:13 1843–1853 1843–1853 2020:14 1843–1853 2020:14 1843–18 fatty liver diseases, some types of cancers and neurodegenerative diseases.^{2,3.} Insulin resistance, a major prevalent factor in obesity, has been closely linked to the accumulation of lipids, oxidative stress and proinflammatory mediators including interleukin-6 and interleukin-1 β , tumor necrosis factor alpha, nuclear factor kappa B, reactive oxygen species and free fatty acids. In addition, anomaly in the metabolism of lipids leading to hyperlipidemia is also a characteristic feature in obesity.^{4–6} Despite the advances in medical sciences, the majority of the treatments currently used for obesity have not achieved absolute success. Therefore, alternative therapies, especially from natural entities that can effectively combat obesity, are now gaining attention.^{7–9}

Medicinal plants play an integral role in traditional medicinal system across the world, especially in nations where the availability and affordability of allopathic medicine are out of the reach of common people. The presence of an array of bioactive phyto-constituents in many of these plants has afforded them as promising valuable assets for the treatment of several diseases.¹⁰ Lycium chinense is a famous traditional Chinese medicinal plant in the family Solanaceae, and it is widely consumed as a functional food as well as medicine.^{11,12} The dried root bark of L. chinense is frequently used in TCM for the treatment of metabolic diseases, notably diabetes and hypertension,^{13,14} and it has been proven to show potent antioxidant, anti-inflammatory and neuroprotective properties.^{15,16} One of the bioactive constituents isolated from L. chinense is kukoamine B (Figure 1), a spermine alkaloid known for its wide range of therapeutic actions including antioxidant, antiinflammatory and antidiabetic effects.¹⁷ Although kukoamine B has many pharmacological effects associated with it, there is no report on its effect on obesity and insulin resistance. Thus, this study investigated the anti-obesity effect of kukoamine B in high-fat diet/fructose-fed obese rats.



Figure I Chemical structure of kukoamine B (Kuk B).

Materials and Methods Animals and Experimental Design

Adult male Wistar albino rats (150–180 g) were used for the study. The rats were housed under controlled conditions of temperature of 22 ± 2 °C, relative humidity of 55 \pm 10% and a 12 h/12 h light–dark cycle. After 7 days of adaptation, the rats were fed with either normal rat diet with normal drinking water or high-fat diet with 15% fructose solution. Each group of rats except for the normal control group was fed with HFDFr for 5 weeks, and at the beginning of the 6th week, they were administered with kukoamine B (Kuk B) together with the HFDFr for an additional 5 weeks. The composition of the HDF was based on the previous report¹⁸ and is shown in Table 1.

The treatment groups were normal control group, HFDFr control group and Kuk B groups (gavage with 25 and 50 mg/kg). The dose of Kuk B used was according to previous studies.¹⁷ After 10 weeks of treatment, the rats were fasted overnight and blood glucose levels were determined with the aid of a glucometer (Accu-Check Performa, Roche, Mannheim, Germany). The experimental procedure was conducted in line with the National Institute of Health guide for the care and use of laboratory animals (NIH Publication No. 80-23; revised 1978) and was reviewed and approved by the ethics committee of The First People's Hospital of Yunnan Province (ethic number: 20181229).

Intra-Peritoneal Glucose Tolerance Test (IPGTT) and Insulin Tolerance Test (ITT)

After the 10th week of treatment, the animals were fasted for 12 h and intraperitoneally injected with 2.0 g/kg of glucose solution for the determination of IPGTT. Blood glucose levels were estimated at 0, 30, 60, 90 and 120 min after the injection with the aid of a blood glucose meter. For the ITT, the rats were fasted for 6 h and intraperitoneally injected with 0.75 U insulin/kg. Blood samples

Table I Composition of High-Fat Diet

Composition	Amount (g/kg)	
Normal pelletized diet	580.0	
Butter	300.0	
Mineral mix	28.7	
Casein	73.2	
Methionine	1.8	
Coconut oil	16.3	

were taken at 0, 30, 60, 90 and 120 min for the determination of blood glucose level. The concentration of glucose in a time point plot was used to calculate the area under curves of IPGTT and ITT.

Sample Collection

After the experiments, the rats were fasted for 12 h and euthanized with sodium thiopental (150 mg/kg) and sacrificed by cervical dislocation. Blood was collected by cardiac puncture and centrifuged at 2500 rmp for 10 min (4 °C) to obtain the plasma. The liver, epididymal and retroperitoneal fat pads were carefully excised, washed with phosphate buffer and weighed. A part of the liver was preserved in 10% buffered formalin solution for histological analysis while 10% homogenate was prepared from the other part in phosphate buffer and centrifuged at 2500 rmp, 4°C for 10 min. The supernatant obtained was used for further biochemical assays.

Determination of Serum Biochemical Parameters

Serum total cholesterol (TC), triglycerides (TG), lowdensity lipoprotein cholesterol (LDL-C), high-density lipoprotein cholesterol (HDL-C), alanine transaminase (ALT), and aspartate transaminase (AST) were determined with the aid of an automatic biochemical analyzer. Serum insulin and free fatty acid (FFA) concentrations were measured by using a commercially available ELISA kit (Mlbio, Shanghai Enzyme-linked Biotechnology Co., Ltd, Shanghai, China) according to the manufacturer's instructions. Homeostatic model assessment index (HOMA-IR) was calculated according to the following formula:¹⁹

 $HOMA - IR = \frac{insulin(\mu U/mL) \times blood \ glucose(mg/dL)}{405}$

Determination of Hepatic Lipids

Hepatic lipids were extracted using the previously described protocol.²⁰ Briefly, the liver lipids were extracted by homogenizing liver tissues in chloroform/ methanol solution (ratio 2:1) on an orbital shaker for 25 mins. Thereafter, the liver homogenate was centrifuged at 2500 rmp for 10 min and the supernatants obtained were dried with nitrogen gas. The extracted lipids were dissolved in 2-propanol containing 1% Triton X100 and used for the determination of TC and TG contents.

Determination of Oxidative Stress Parameters

The excised liver samples were homogenized in phosphate buffer (10% homogenate) and centrifuged at 2500 rpm for 10 min at 4 °C. The supernatant obtained from the liver homogenate was used for estimating hepatic oxidative stress marker (SOD, GSH-Px, CAT and MDA) levels using commercial assay kits (Nanjing Jiancheng Bioengineering Institute, Nanjing, China) based on the manufacturer's directives.

Determination of Pro-Inflammatory Mediators

The serum levels of pro-inflammatory cytokines (IL-1 β , TNF- α , IL-6 and NF-kB) were analyzed with the aid of commercially available ELISA kits (Mlbio, Shanghai Enzyme-linked Biotechnology Co., Ltd, Shanghai, China, and Elabscience Biotechnology, Wuhan, China).

Histopathological Analysis of Liver Tissues

The liver tissues were fixed in 10% buffered formalin solution, dehydrated with graded alcohol solution, embedded in paraffin and sliced and into 5- μ m sections. The sections were further stained with hematoxylin and eosin (H&E), and histopathologic changes of the liver tissues were observed under an Olympus light microscope (Tokyo, Japan).

Statistical Analysis

The results are expressed as the mean \pm standard deviation (SD). Differences among groups were analyzed by oneway ANOVA and Tukey's multiple comparison tests using Prism 7.0 software (GraphPad Software Inc., San Diego, CA). Statistically significant was set at P < 0.05 in all cases.

Results

Effect of Kuk B on Food, Water Consumption and Organ Weight in HFDFr-Fed Rats

As shown in Table 2, compared with the normal control group there was a significant reduction in food intake and water intake of the HFDFr group (P< 0.05). On the other hand, there were no significant changes in the food and water intake between the Kuk B-treated groups and

Parameters	Normal Control	HFDFr	Kuk B 25	Kuk B 50
Food intake (g/day)	22.10 ± 1.05	17.14 ± 0.89**	17.80 ± 0.72	18.10 ± 1.22
Water intake (mL/day)	39.63 ± 1.41	25.25 ± 0.72**	26.03 ± 1.21	27.94 ± 0.88
Liver weight (g)	10.00 ± 1.11	15.39 ± 1.41**	12.57 ± 0.7***	11.70 ± 0.7***
Epididymal fat (g/100 g body weight)	0.94 ± 0.11	2.38 ± 0.32**	1.93 ± 0.18***	1.70 ± 0.11***
Retroperitoneal fat (g/100 g body weight)	1.27 ± 0.10	2.66 ± 0.25**	2.27 ± 0.19***	2.01 ± 0.16***

 Table 2 Effect of Kuk B on Food Intake, Water Intake and Organ Weight in Obese Rats

Notes: Data are expressed as the mean ± SD (n = 6). One-way ANOVA with Tukey's post hoc test was used for statistical analysis. **P < 0.05 vs normal control, ***P < 0.05 vs HFDFr.

Abbreviations: Kuk B, kukoamine B; SD, standard deviation; HFDFr, high-fat diet/high fructose.

HFDFr groups. Furthermore, there was a significant increase in the weight of liver, epididymal fat and retroperitoneal fat in the HFDFr group compared with the normal control group. However, treatment with Kuk B reduced the weight of liver and fat compared with the HFDFr group (Table 2).

Effect of Kuk B on Body Weight and Insulin Tolerance in HFDFr-Fed Rats

The effect of Kuk B on body weight is shown in Figure 2A. The body weight of HFDFr-fed rat group rapidly and significantly increased compared to the

normal control group (P < 0.05). In Kuk B-treated groups, the results showed a significant reduction in the body weight of rats as compared with the HFDFr group (P < 0.05). The effect of Kuk B on fasting blood glucose, insulin, and insulin resistance index (HOMA-IR) is shown in Figure 2B–D. The rats in the HFDFr group showed significantly increased fasting blood glucose (Figure 2B) and insulin levels (Figure 2C) when juxtaposed with the normal diet-fed rats (P < 0.05). On the other hand, insulin and fasting blood glucose levels in the Kuk B-treated groups (25 and 50 mg/kg) were significantly lowered compared to the levels in





Α

Blood glucose (mg/dL)

1600

1400

1200

L1000 800

600 400

200

Normal

NC

500

400

300

200

100

0

0

30

60

Time (mins)

90

120

the HFDFr group. In addition, treating with Kuk B significantly improved HOMA-IR compared to HFDFr control group (Figure 2D)

Effect of Kuk B on Glucose and Insulin **Tolerance in HFDFr-Fed Rats**

As indicated in Figure 3A, the HFDFr group showed significantly higher fasting blood glucose levels at all time points measured (0, 30, 60, 90 and 120 min) when compared to the normal control group (P < 0.05) in the IPGTT. On the other hand, the blood glucose levels of Kuk B-treated groups were significantly decreased at 30, 60, 90 and 120 min after the administration of glucose. Furthermore, the AUC in the Kuk B-treated groups was significant lower as compared to the HFDFr group (Figure 3B). In the ITT, the blood glucose levels in the normal control and Kuk B-treated groups (50 mg/ kg) were markedly lower than the blood glucose levels in the HFDFr group (Figure 3C), suggesting that insulin sensitivity in the HFDFr group was reduced. Kuk B-treated group (25 mg/kg) had a lower level of blood glucose; however, the effect was not significant when

compared to the HFDFr group (Figure 3C). Analysis of AUC summarized in Figure 3D indicated that the area under the curve for the ITT in the HFDFr group was significantly increased compared with the normal control group. However, treatment with Kuk B significantly reduced blood glucose levels.

Effect of Kuk B on Biochemical Parameters in HFDFr-Fed Rats

The effect of Kuk B on serum lipid levels in HFDFr-fed rats is shown in Figure 4. Marked increase in the serum levels of TG, TC (Figure 4A), LDL-C (Figure 4B) and FFA (Figure 4E) was noticed in the HFDFr group compared to the normal control group (P < 0.05). On the other hand, after 5 weeks of treatment with Kuk B, the lipid profiles were significantly improved compared to the HFDFr group (P < 0.05). HDL-C levels were significantly lowered in the HFDFr group compared with the normal control group, Kuk B insignificantly increased HDL-C level compared to the HFDFr group (Figure 4B). In addition, high levels of hepatic TG and



Normal

HFDFr

В

vs HFDFr

Abbreviations: Kuk B, kukoamine B; IPGTT, intraperitoneal glucose tolerance test; ITT, intraperitoneal insulin tolerance test; SD, standard deviation; HFDFr, high-fat diet/ high fructose



100

50

0

1000

800

0



60

Time (mins)

90

120

D

30





Figure 4 Effect of Kuk B on (A) serum TG and TC (B) serum LDL-C and HDL-C (C) hepatic TG and TC (D) serum AST and ALT (E) serum FFA in obese rats. Notes: Data are expressed as the mean ± SD (n = 6). One-way ANOVA with Tukey's post hoc test was used for statistical analysis. **P < 0.05 vs normal control, ***P < 0.05 vs HFDFr.

Abbreviations: Kuk B, kukoamine B; TG, triglycerides; TC, total cholesterol; LDL-C, low density lipoprotein cholesterol; HDL-C, high density lipoprotein cholesterol; AST, aspartate transaminase; ALT, alanine transaminase; FFA, free fatty acid; SD, standard deviation; HFDFr, high-fat diet/high fructose.

Kuk B remarkably decreased TG and TC levels in the liver tissues of treated rats (P < 0.05). As shown in Figure 4D, compared to the HFDFr group, treatment with Kuk B significantly decreased the serum levels of AST and ALT (P < 0.05). These results indicated that Kuk B attenuated anomalies of the liver function and lipid metabolism caused by HFDFr.

Effect of Kuk B on Inflammatory Cytokines in HFDFr-Fed Rats

Serum inflammatory cytokines (TNF- α , IL-1 β , IL-6 and NF-kB) concentrations were elevated in HFDFr rats when compared to normal control. However, these parameters were significantly reduced in the Kuk B-treated group (P < 0.05) compared to the HFDFr group (Figure 5A–D).



Figure 5 Effect of Kuk B on (A) TNF- α , (B) IL-6, (C) ILI- β , (D) NF-kB in obese rats in obese rats. Notes: Data are expressed as the mean ± SD (n = 6). One-way ANOVA with Tukey's post hoc test was used for statistical analysis. **P < 0.05 vs normal control, ***P < 0.05 vs HFDFr: Abbreviations: Kuk B, kukoamine B; TNF- α ,tumor necrosis factor alpha; IL-6, interleukin 6; ILI- β , interleukin 1 beta; NF-kB, nuclear factor kappa B; SD, standard deviation; HFDFr, high-fat diet/high fructose.

Effect of Kuk B on Oxidative Stress Markers in HFDFr-Fed Rats

We further examined the effect of Kuk B on oxidative stress parameters in HFDFr-fed rats by estimating the hepatic levels of MDA, SOD, GSH-Px and CAT. As shown in Figure 6A, MDA level in the HFDFr-fed rats was increased by 1.6-fold compared to the rats that were fed with the normal diet. In contrast, treatment with Kuk B attenuated MDA levels in the treated rats. Furthermore, in the HFDFr-fed group, the activities of SOD, GSH-Px and CAT were reduced; however, a significant increase was observed in the activities of these enzymes after 5 weeks of treatment with Kuk B (Figure 6B–D).

Effect of Kuk B on Pathological Lesions in HFDFr-Fed Rats

The effect of Kuk B on histopathological alterations in HFDFr-induced obesity was evaluated by histological analysis. As shown in Figure 7, the normal control rats

showed no abnormalities, whereas, hepatocyte lipid droplet accumulation as well as inflammatory cells were observed in the liver histology of the HFDFr group (Figure 7B). In the Kuk B-treated groups, there were obvious signs of alleviation of these pathological alterations (Figure 7C–D).

Discussion

Kukoamine B (Kuk B) is a spermine alkaloid obtained from the root bark of *L. chinense*, and several studies have reported it as a potent antioxidant and anti-inflammatory agent, and its ability to inhibit oxidative stress–induced damages has been highlighted.^{21–24} Since oxidative stress and low-grade inflammation have been largely implicated in the pathogenesis of insulin resistance and obesity, natural substances with antioxidant and anti-inflammatory effects may be promising in the treatment of obesity and metabolic disorders. Therefore, this study investigated the effect of Kuk B on high-fat/fructose diet–induced obesity as well as



Figure 6 Effect of Kuk B on (A) MDA, (B) SOD, (C) CAT, (D) GSH-Px in obese rats. Notes: Data are expressed as the mean ± SD (n = 6). One-way ANOVA with Tukey's post hoc test was used for statistical analysis. **P < 0.05 vs normal control, ***P < 0.05 vs HFDFr: Abbreviations: Kuk B, kukoamine B; MDA, malondialdehyde; SOD, superoxide dismutase; CAT, catalase; GSH-Px, glutathione peroxidase; SD,standard deviation; HFDFr, high-fat diet/high fructose.

its role in alleviating oxidative stress, lipid accumulation, inflammation and insulin resistance in obese rats.

The consumption of diets with high quantities of fats and fructose has been widely viewed as one of the key elements associated with obesity, insulin resistance and other metabolic disorders. Excessive lipid accumulation in the adipose tissue can result in the formation of lipid intermediates including fatty acyl-CoA, diacylglycerols and ceramides, which alters several processes in the body, especially in the muscle and liver, resulting in metabolic anomalies such as glucose intolerance and hepatic steatosis.^{25–28} The accumulation of free fatty acids initiates insulin resistance in skeletal muscle, which subsequently activates protein kinase C and inflammatory pathways such as JNK, IKK and NF-k²⁹⁻³¹ Furthermore, high fructose consumption has also been associated with insulin resistance due to its effect on elevating plasma insulin, FFA, fasting glucose and glucose intolerance.^{32,33}

Insulin resistance is a condition that is associated with reduction in insulin sensitivity, utilization of insulin by peripheral tissues and glucose uptake, leading to excessive secretion and circulation of insulin in the blood (hyperinsulinemia). Hyperinsulinemia can result in several metabolic disorders such as type 2 diabetes mellitus, hypertension, coronary heart disease, cerebrovascular disease, obesity and dyslipidemia.^{34,35} Furthermore, insulin resistance is a requisite for the elevation of blood glucose level, serum lipids and cholesterol levels, thus creating a fertile ground for the development of cardiovascular disease and metabolic syndrome.^{36,37} The results of this study clearly demonstrated that Kuk B significantly ameliorated insulin resistance caused by HFDFr. Indeed, the administration of Kuk B reduced levels of blood glucose, insulin, serum and hepatic lipids, as well as liver function enzymes. These results suggest that Kuk B enhances insulin sensitivity, inhibits insulin resistance and alleviates other metabolic anomalies associated with insulin resistance.

The etiology of insulin resistance is multi-faceted and a full understanding of it is still lacking; however, certain parameters like oxidative stress and inflammation have



Figure 7 The effect of Kuk B on liver pathological lesions.

Notes: Representative images of liver section stained with H&E. Lipid accumulation (black arrow). Inflammatory cells (brown arrow). Abbreviations: Kuk B, kukoamine B; H&E, haemotoxylin and eosin; HFDFr, high-fat diet/high fructose.

been widely implicated in the etiology of insulin resistance and obesity. Numerous reports have shown the existing cordial relationship between oxidative stress, insulin resistance and obesity. The activities of cellular antioxidant enzymes are diminished and the generation of oxidative stress agents such as ROS and other oxidative modification products is increased in the serum, liver, adipose and brain tissues during insulin resistance.^{6,38-40} Accumulation of excess fat, which is often the case in obesity, or HFDFr intake leads to increase in FFA oxidation, which subsequently triggers nicotinamide adenine dinucleotide (NADPH) oxidase to overgenerate ROS. In addition, previous reports have associated oxidative stress agents with high-fat accrual.41-43 Furthermore, it has been widely acknowledged that insulin resistance and oxidative stress can trigger inflammation and the release of cytokines by the liver and adipose tissue. Oxidative stress also has the ability to activate inflammatory response leading to insulin resistance through the upregulation of several inflammatory-related pathway signaling molecules such as serine kinases c-Jun N-terminal kinase (JNK) and I kappa B kinase (IKK-b) in the adipose tissues and liver that inhibit normal insulin signaling. In response to the instigation of chronic inflammation, several inflammatory

mediators such as TNF- α , IL-6, IL-1 β , COX-2 and MCP-1 are released, which can aggravate insulin resistance and further reduce the sensitivity of tissues to insulin, thus limiting glucose uptake.^{44–48} Previous reports have clearly indicated that HFDFr stimulates ROS, oxidative stress and inflammatory mediators.^{4,49} Our study indicated that the HFDFr-fed rats showed increased oxidative stress and inflammatory markers (MDA, TNF- α , IL-6, IL-1 β and NF-kB) and decreased antioxidant enzyme activities (GSH-Px, SOD, and CAT). Kuk B attenuated oxidative stress and inflammation by enhancing antioxidant defense activity and reducing the release of inflammatory-related cytokine.

Numerous studies have clearly illustrated that lipid profiles are significantly altered in diet-induced obesity and metabolic abnormalities. Consumption of HFDFr enhances the accumulation of lipids in the liver and subsequently increased secretion in blood.^{50,51} High levels of TC, TG and LDL-C and reduced levels of HDL-C are vital indicators that have been linked to the incidence of diabetes mellitus, coronary heart diseases and cardiovascular diseases.⁵² The results from this study demonstrated that the administration of Kuk B decreased serum and hepatic TC and TG as well as serum FFA and LDL-C. The Kuk B-treated groups also showed improvement in HDL-C levels.

It was observed that the HFDFr group had high levels of serum AST and ALT, indicating liver damage. The release of these liver function enzymes has been linked to alterations in the plasmatic membrane of the hepatocytes. The decrease in hepatic lipid profiles and liver function enzymes in the Kuk B-treated groups correlates with the histopathology analysis of the liver, where a reduction in the lipid droplets (lipid vacuole content) was obviously seen in the Kuk B-treated groups when compared to the HFDFr group. These results are consistent with those of other studies.^{51,53}

Conclusion

Kuk B protected against HFDFr-induced insulin resistance and metabolic dysfunction in rats through the regulation of dyslipidemia, antioxidant defenses and inflammatory response. Thus, *L. chinense* and its active components especially kukoamine B may have possible applications as dietary supplements in ameliorating metabolic disorders.

Author Contributions

All authors contributed to data analysis, drafting and revising of the article, gave final approval of the version to be published, and agree to be accountable for all aspects of the work.

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Disclosure

The authors report no conflicts of interest in this work.

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