REVIEW Open Access



Cancer metastasis: molecular mechanisms and therapeutic interventions

Xiaofeng Dai^{1*}, Ming Xi¹ and Jitian Li²

Abstract

The metastatic cascade is a complicated process where cancer cells travel across multiple organs distant from their primary site of onset. Despite the wide acceptance of the 'seed and soil' theory, mechanisms driving metastasis organotropism remain mystery. Using breast cancer of different subtypes as the disease model, we characterized the 'metastatic profile of cancer cells' and the 'redox status of the organ microenvironment' as the primary determinants of cancer metastasis organotropism. Mechanically, we identified a positive correlation between cancer metabolic plasticity and stemness, and proposed oxidative stress as the selection power of cancer cells succeeding the metastasis cascade. Therapeutically, we proposed the use of pro-oxidative therapeutics in ablating cancer cells taking advantages of this fragile moment during metastasis. We comprehensively reviewed current pro-oxidative strategies for treating cancers that cover the first line chemo- and radio-therapies, approaches relying on naturally existing power including magnetic field, electric field, light and sound, nanoparticle-based anti-cancer composites obtained through artificial design, as well as cold atmospheric plasma as an innovative pro-oxidative multi-modal modality. We discussed possible combinations of pro-oxidative approaches with existing therapeutics in oncology prior to the forecast of future research directions. This paper identified the fundamental mechanics driving metastasis organotropism and proposed intervention strategies accordingly. Insights provided here may offer clues for the design of innovative solutions that may open a new paradigm for cancer treatment.

Keywords Cancer, Metastasis organotropism, Onco-therapeutics, Oxidative stress, Metabolism, Cold atmospheric plasma

Introduction

Under the long-term synergistic stimulus of various internal and external tumor-causing factors, cells loose the redox homeostasis [1, 2], leading to uncontrollable proliferation and the onset of carcinogenesis. Some transformed cells gain the metastatic features,

Metastasis refers to the process where transformed cells are disseminated from the site of primary initiation to different location(s) of the body. A complicated cascade is involved during metastasis including, e.g., invasion, intravasation, circulation, extravasation, and colonization. The organ to which cancer cells can metastasize is non-random, with the process being known as 'organotropism'. For example, prostate cancers preferably colonize in the bone, uveal melanomas typically relapse to the liver, and breast cancer of different subtypes favor distinct organs to metastasize that include bone, liver, brain, and lung [4]. Transformed cells need to decide 'whether to grow or go' [5, 6] and 'where to go' for sustained survival, as prolifera-

tion and migration represent two mutually exclusive

explaining over 90% cancer-related mortality [3].

xiaofengteam@163.com

National Local Joint Engineering Research Center for Precision Surgery & Regenerative Medicine, Shaanxi Provincial Center for Regenerative Medicine and Surgical Engineering, First Affliated Hospital of Xi'an Jiaotong University, Xi'an 710061, People's Republic of China Molecular Biology Lab, Henan Luoyang Orthopedic Hospital (Henan Provincial Orthopedic Hospital), Henan Province, Zhengzhou 450000, China



^{*}Correspondence: Xiaofeng Dai

Dai et al. Molecular Biomedicine (2025) 6:20 Page 2 of 30

destinations as a result of competitive use of shared cellular resources [7]. Accumulated evidence has suggested that organotropism is regulated by multiple factors such as cancer stemness [8] and tumor matrix stiffness [7], implicating clues for effective cancer metastasis prognosis, prevention and therapeutics.

Breast cancer, remaining as the primary cause of female cancer deaths, is a heterogeneous disease that can be roughly divided into luminal A, luminal B, human epidermal growth factor receptor 2-positive (HER2+), and triple negative breast cancer (TNBC) subtypes according to their biological characteristics and molecular markers [9-11]. Luminal A and B subtypes are over-represented with estrogen receptor (ER); HER2+breast tumors are featured with high levels of HER2; TNBCs lack the expression of ER, progesterone receptor (PR), HER2 and thus are irresponsive to hormonal or targeted therapies such as Tamoxifen [12] and Herceptin [13]. Importantly, TNBCs are known to possess large percentages of cancer stem cells (CSCs), and are therefore more prone to develop metastasis and difficult to treat. Despite such distinct clinical manifestations, breast cancers of different subtypes display different metastatic tendencies. For instance, luminal A/B breast cancers are inclined to develop bone [14] and liver [15] metastasis, and HER2+and TNBCs prefer to metastasize to visceral organs such as brain [16] and lung [8, 17-19]. Such distinct metastatic profiles of tumors originated from the same organ offer us an ideal disease model for investigating factors influencing or driving tumor metastasis organotropism.

The aim of this review is to aid in the effective prognosis, prevention and therapeutics of cancer metastasis for, hopefully, reduced cancer-associated mortality. Using breast cancers of different subtypes as the disease model, this review gives an overview on metastasis organotropism, characterizes critical factors defining cancer metastasis organotropism, identifies primary mechanisms driving the vulnerability of cancer cells to oxidative stress during metastasis, introduces current and emerging pro-oxidative oncotherapeutic modalities, discusses possible strategies for combining pro-oxidative approaches with existing therapeutics, and concludes this paper with future trends forecasted.

Metastasis organotropism

Historical development of the 'seed and soil' theory

The 'seed and soil' theory was proposed initially by Stephen Paget to understand the non-random pattern of metastasis in 1889 [20]. By scrutinizing over 900 autopsy records of patients carrying different types of primary

cancers, Stephen Paget was astonished in finding a discrepant pattern between the relative blood supply and the metastatic frequency of cancer cells to certain organs, i.e., there may exist a non-random distribution of metastatic cells to bones and visceral organs. Therefore, Stephen Paget challenged the prevailing view at that time in proposing the 'seed and soil' theory. This theory stated that the destination of metastatic cancer cells was not by chance but rather determined by cancer cells and the milieu of the organs; by considering tumor cells as the 'seed' and the milieu of certain organs as the 'soil', metastasis could only occur when the seed and the soil were compatible. However, Stephen Paget did not discuss in detail possible factors determining such a compatibility (Fig. 1).

The 'seed and soil' theory was challenged by James Ewing in 1929, who believed that metastatic dissemination of cancer cells, though not completely by chance, was dictated by mechanical factors such as the anatomical structure of the vascular system [21]. This view for the first time substantiated the 'seed and soil' theory in proposing possible influential factors on the compatibility between cancer cells and the colonized organ, but also negated the 'seed and soil' theory in attributing the metastasis organotropism purely to anatomical differences or, in other words, by chance (Fig. 1).

The viewpoint proposed by James Ewing prevailed for several decades until 1970s when Lance Liotta and Jerome Kleinerman characterized the association between cancer metastasis organotropism and the production of proteolytic enzymes by metastatic cancer cells [22]. Also, Ian R. Hart and Isaiah J. Fidler found that besides mechanical attract of traveling tumor cells in the capillary bed of distant organs, specific cells residing the metastatic organs played essential roles in the subsequent cancer cell proliferation in the secondary lesions during the colonization process using B16 melanoma as the cancer model [23]. Such viewpoints were supported by Leonard Weiss [24] and Everett V. Sugarbaker [25], who concluded that distant organ metastasis was site specific for a given type of cancers that could not be solely explained by anatomical or mechanical factors such as efferent venous circulation or lymphatic drainage to regional lymph nodes, after reviewing the clinical data of different human cancers regarding their site preferences of metastasis (Fig. 1).

After a century's struggle and exploration on the possible existence of cancer metastasis organotropism, the 'seed and soil' theory was brought back to our horizon in 1989 when a symposium commemorated the centennial anniversary of Stephen Paget's 'seed and soil' hypothesis. Ever since then, consecutive efforts have been devoted to update and substantiate the content and connotation

Dai et al. Molecular Biomedicine (2025) 6:20 Page 3 of 30

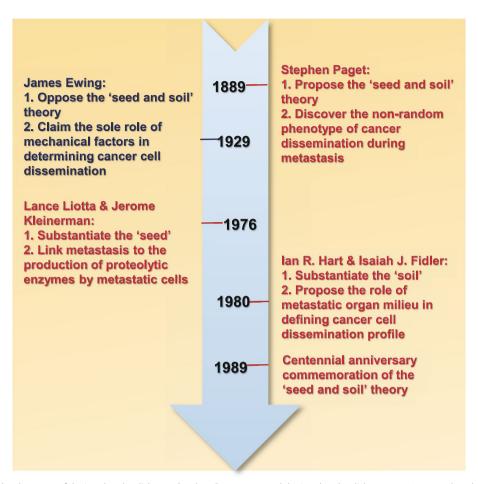


Fig. 1 Historical development of the 'seed and soil' theory. Stephen Paget proposed the 'seed and soil' theory in 1889 to explain the non-random dissemination profile of cancer cells during metastasis he observed. James Ewing challenged this theory by claiming that cancer cell organotropism was solely influenced by mechanical factors. In 1970s, Lance Liotta & Jerome Kleinerman linked metastasis to the metastatic cell produced proteolytic enzymes, and lan R. Hart & Isaiah J. Filder proposed the role of metastatic organ milieu in defining cancer cell dissemination profile that substantiated the 'seed and soil' theory from the 'seed' and 'soil' points respectively. In 1989, the 'seed and soil' theory was commemorated in its centennial anniversary

of the 'seed and soil' theory with our incremental understandings on the pathogenesis and molecular features of cancers [4, 26]. The current beliefs on cancer metastasis organotropism contain the following three principles. First, primary and metastatic tumor lesions both contain cancer cells and host cells which are biologically heterogeneous. Primary components of the host cells include, e.g., epithelial cells, endothelial cells, fibroblasts and infiltrating leukocytes. Neoplasms contain genetically, epigenetically and phenotypically distinct cohorts of transformed cells, each of which have the potential to complete certain but not all steps in the metastatic process. Second, each of the five steps in the metastasis cascade (i.e., invasion, intravasation, circulation, extravasation, colonization) is selective for cells capable of surviving the stress under each particular circumstance, and metastasis can be originated from single cell expansion. To successfully pass through the whole metastasis cascade, metastatic cancer cells (the 'seed') must possess or evolve multiple features to be able to undertake a variety of tasks [27, 28], as the metastasis process may be blocked at diversified stages such as loss of the ability of progressive growth, vascularization and invasion, detachment from the cell cluster, embolization, susceptibility to the attack of the immune system, and the inability of usurping resources from the environment of the secondary loci. Third, a given cluster of travelling cancer cells can only surpass the dormant stage and regain uncontrolled proliferation in specific organs. The biological characteristics of the microenvironments of different metastatic organs (the 'soil') differ that are deterministic on the types of cancer cells capable of establishing Dai et al. Molecular Biomedicine (2025) 6:20 Page 4 of 30

colonization there. In other words, the outcome of metastasis depends on whether a homeostatic crosstalk can be developed between the metastasizing cells and the host environment.

Breast cancer metastasis organotropism

Bone is one of the most common sites for breast cancer metastasis, accounting for approximately 65%-75% of all metastatic cases [29]. Bone metastasis not only leads to serious skeletal complications such as pain, fracture, and hypercalcemia, but also significantly reduces the quality of life and survival time of the cancer patients [30, 31]. It has been documented that ER+breast cancers, i.e., luminal A (largely ER+HER2-) and luminal B (largely ER + HER2 +) subtypes [32–34], more easily develop bone metastasis as compared with ER- subtypes, i.e., HER2+(ER-HER2+) and TN (ER-HER2-) breast cancers (Fig. 2) [14]. It has been reported that 66.8% luminal A patients developed bone metastasis, whereas 38.9% TN patients having cancer cells metastasized to the bone [14]. In another study, the odds between luminal and TN breast cancers in developing bone metastasis was 8.2 [35].

Being one common site for breast cancers metastasis, 40%–50% breast tumor carriers develop liver cancer metastasis with the mortality rate being 50%-62% and the average patient survival time being 31 months [36]. Breast cancers, once metastasized to the liver, exhibit clinical symptoms such as pain in the liver region, hepatomegaly, nausea, vomiting, jaundice, and ascites. Etiology studies have indicated that breast cancers carrying HER2 positivity, i.e., luminal B and HER2+[32-34]breast cancer carriers, are more prone to develop liver metastasis as compared with the other breast cancer subtypes (Fig. 2) [16, 37]. For example, 46.6% HER2+breast cancer patients developed liver metastases, whereas 33% luminal A patients developed metastasis in this organ [14]; and the chance of HER2 + breast cancers in developing liver metastasis versus that of luminal A patients was 4.2/0.8 in another study [35].

Cancers, once metastasized to the brain, are lethal, leaving the percentage of patients capable of surviving for 1-year only appropriately 20% [38]. It has been estimated

that 15-25% of breast cancer patients develop metastasis to the central nervous system [39]. Manifestations of brain metastasis from breast cancers include, e.g., headaches, nausea, vomiting, dizziness, seizures and increased intracranial pressure as a result of tumor formation in the brain and associated tissue compression [40]. It has been reported that ER-PR- patients, i.e., HER2+and TN breast cancers, are more inclined for brain metastasis than luminal A/B breast cancer patients [16], with approximately 50% of HER2+breast cancer carriers and about 1/3 of TNBC patients having gained brain metastasis, respectively, in the end (Fig. 2) [41]. It has also been reported that the rate between HER2+tumors and the luminal A subtype in establishing brain metastasis was 1.1/0.2, and that between TN and luminal A tumors was 0.7/0.2 [35].

Lung metastasis typically does not elicit obvious symptoms until vast replacement of metastatic tumor mass occurs [42], with the primary manifestations including cough, haemoptysis, dyspnoea, malaise and weight loss [43]. Breast cancers especially ER-PR- breast cancer subtypes (i.e., HER2+, TN subtypes) also easily develop lung metastasis, with the incidence ranging from 21 to 32% among breast cancer carriers (Fig. 2) [35]. Specifically, HER2 + breast cancer patients having received anti-HER2 targeted therapies exhibited a high risk of developing lung metastasis [44, 45], and TN breast cancers have high propensities to metastasize to the lung [42, 46]. In one study, 50.4% TN breast cancer patients developed lung metastasis whereas only 21% luminal A patients gained metastasis to the lung [14]. In another report, the ratio between HER2+and luminal subtypes in developing lung metastasis was 3.4/1.2, and that between TN and luminal tumors was 2.5/1.2 [35].

Critical factors defining metastasis organotropism

In the 'seed and soil' theory, cancer cells and the metastasized sites are referred to as 'seeds' and 'soil', respectively; while metastatic seeds can travel to any organs, they can thrive only in the congenial soil [47]. This requires organ-specific metabolic adaptation to help metastasized malignant cells overcome obstacles faced when establishing organotropism phenotypes during distant organ

(See figure on next page.)

Fig. 2 Illustrative diagram on breast cancer metastasis organotropism. Among bone, liver, brain and lung, the four organs commonly developing breast cancer metastasis, the luminal A and B subtypes are inclined to metastasize to the bone, and luminal B breast cancers may also develop liver metastasis, where both bone and liver are characteristic of hypoxia; the HER2+ and TNBC subtypes prefer metastasizing to lung and brain where oxygen supply is sufficient, yet some HER2+ tumors may also metastasize to the liver. Breast cancer stem cells (BCSCs) residing in these breast cancer subtypes differ. While luminal A, luminal B and HER2+ cancers possess BCSCs over-expressing ALDH, TNBC cells have BCSCs characteristic of CD24-CD44+. Importantly, luminal A and luminal B cancers have low percentage of BCSCs, HER2+ and TNBC cells have high percentages of BCSCs

Dai et al. Molecular Biomedicine (2025) 6:20 Page 5 of 30

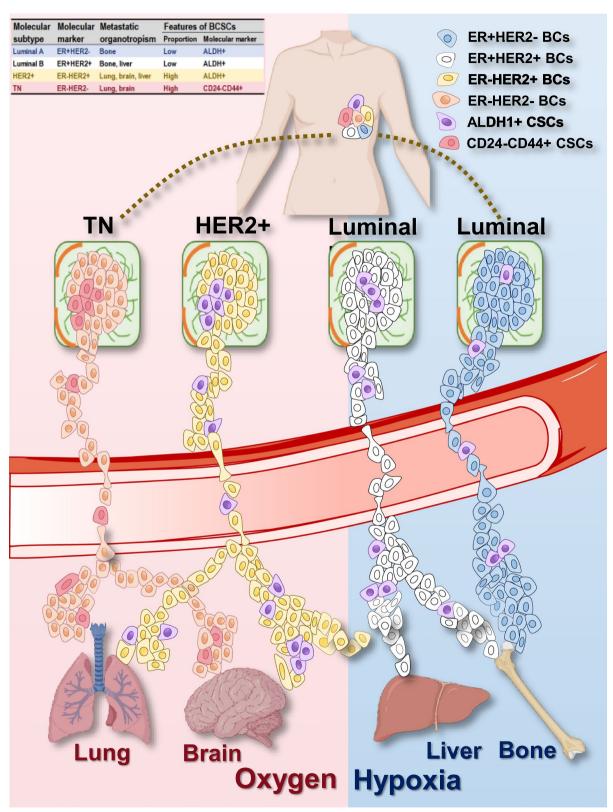


Fig. 2 (See legend on previous page.)

Dai et al. Molecular Biomedicine (2025) 6:20 Page 6 of 30

colonization [48], the process of which is dynamic and heterogeneous [49–51]. A variety of factors have been identified capable of, at least partially, influencing the metastasis organotropism of different breast cancer subtypes during the complicated metastatic cascade, which can be roughly categorized into cancer-intrinsic features and organ-specific microenvironment.

Cancer-intrinsic features

Breast cancer stem cells (BCSCs) are a small cohort of breast cancer cells with the self-renewal, multipotent differentiation and tumor-initiating abilities [52]. The metastasis cascade is initiated by breaking the surrounding basement membrane and invading into the extracellular matrix (ECM), the process of which is energy-intensive [53, 54]. After departure from the primary loci, cancer cells travel in the circulation and evolve various strategies to survive in the bloodstream such as evading the immunosurveillance [55, 56]. On arrival at the sites of tropism, cancer cells may adjust to the new niche and re-initiate tumor proliferation in distant organs. Thus, disseminated cancer cells are considered to retain cancer stemness [55, 57]. Indeed, an increasing number of evidence has indicated the potential relationship between BCSCs and distant breast cancer metastasis, as well as the regulatory role of BCSCs on breast cancer metastatic organotropism. For instance, the plasticity of BCSCs allowed them to transit between the mesenchymal and epithelial states during cancer metastasis [26]. Thus, the properties of CSCs may define the metastasis organotropism from the perspective of cell-intrinsic features.

BCSCs can be grouped into three types, i.e., CD24-CD44 + BCSCs, aldehyde dehydrogenase (ALDH) + BCSCs and CD24-CD44 + ALDH + BCSCs based on the canonical CSC biomarkers used for BCSC characterization. While CD24 (a glycosylated protein linked to the cell membrane) and CD44 (a transmembrane glycoprotein interacting with various components in the ECM) play essential roles in cell adhesion and migration [58, 59], ALDH1 (a member of the aldehyde dedydrogenase family) participates in the regulation of BCSC self-renewal ability [60]. Different BCSCs are located at different positions of the invading cell collection with distinct roles. Specifically, CD24-CD44+BCSCs are localized at the front edge of the tumor collection possessing highly invasive features, ALDH+BCSCs are located at the center of the invading population exhibiting uncontrollable proliferative properties, and CD24-CD44+ALDH+BCSCs represent the CSC subset showing the greatest tumor-initiating capacity [61]. The stemness of these BCSCs increases from ALDH+to CD24-CD44+to those harboring both,

as only 20 BCSCs expressing both CD24-CD44+and ALDH+effectively generated tumors whereas 100 CD24-CD44+BCSCs and 500 ALDH+BCSCs were needed, respectively, to produce the bulk tumor cells [62, 63].

In addition, it has been estimated that up to approximately 50% of ATP was used to support the actin cytoskeleton during cancer metastasis [64, 65], leading to a positive correlation between the intracellular ATP:ADP ratio and migration potential [53]. Several cellular forces associated with bioenergetics during cancer cell migration have been identified including, e.g., integrin activation, cytoskeletal remodeling, and AMPK activation. Differentially expressed genes (DEGs) of CD24-CD44+BCSCs and ALDH+BCSCs have been characterized, which were enriched in focal adhesion and oxidative phosphorylation, respectively [66]. These evidence have implicated the involvement of distinct molecular mechanisms in driving the mobility of different BCSCs during the onset of cancer metastasis that is consistent with the highly invasive nature of CD24-CD44+BCSCs and the elevated glycolysis in ALDH + BCSCs [67].

Breast cancer subtypes contain different amounts and types of BCSCs, i.e. ER+(i.e., luminal A/B) cells contain relative low levels of ALDH+BCSCs [68], HER2+cells are enriched with ALDH+BCSCs, and TNBC cells have high proportion of CD24-CD44+BCSCs [8]. This makes TNBCs that are highly invasive exhibiting the mesenchymal-like properties, and empowers HER2+breast cancers with the aggressive epithelial-like features [8, 69]. The exact amount of CSCs in each breast cancer subtype varies considerably among studies, possibly due to the heterogeneity of cell lines as well as the detection/ quantification approaches used in each investigation. For example, ALDH1+CSCs were reported to be 8.6%, 50%, 72.1%, 26.7% in luminal A, luminal B, HER2+, TNBC subtypes in one study [70], and was estimated to represent approximately 52% HER2+tumors in another study [71]. Similarly, the CD24-CD44+CSC phenotype was manifested in 76.5% TNBCs in one study [72], and in all TNBC cells in another report [71].

Organ-specific microenvironment

Incremental evidence has implicated that the microenvironment of the organ plays essential roles in defining the metastatic profiles of cancer cells. One reason empowering transformed cells with the invasive features is their inefficiency in getting adapted to the microenvironment of the initial organ for survival. Thus, the milieu of the secondary organ selected by cancer cells for colonization may have more favorable features for transformed cells with the corresponding tropism to survive. For instance, cancer cells initiated from an organ enriched with oxygen

Dai et al. Molecular Biomedicine (2025) 6:20 Page 7 of 30

such as the lung may metastasize to hypoxic tissues [73], and malignant cells with a vigorous mitochondrial metabolism may leave the hypoxic liver and opt for the lung [74].

Bone microenvironment

Bone and bone marrow are comprised of osteocytes, osteoblasts, and osteoclasts. The bone micromilieu contains diversified cell types such as osteoblasts, osteoclasts, adipocytes, haematopoietic stem cells, and infiltrated immune cells [31]. Osteoblasts and osteoclasts play opposite roles. While osteoblasts facilitate new bone deposition to either form osteocytes or mature into lining cells, osteoclasts resorb the bone matrix. Under normal conditions, osteoblasts and osteoclasts establish a dynamic homeostasis between bone formation and decomposition [31]. Cancer cells can hijack the activities of osteoblasts and promote osteoclastogenesis to boost their proliferation and establish a vicious cycle among osteoblasts, osteoclasts, and transformed cells, since such a bone resorption process is accompanied with the release of numerous factors from the bone matrix such as cytokines, calcium, collagens, growth factors, glycoproteins, hyaluronans, proteoglycans, and proteinases [75, 76]. The inorganic part of the bone is primarily comprised of mineral hydroxyapatite crystals, and the bone matrix is enriched with bone sialoprotein, type I collagen, osteopontin, as well as a variety of growth factors and cytokines [77].

The bone microenvironment is characteristic of hypoxia [78] and acidity [79] that are interconnected. Hypoxia triggers hypoxia-inducible factor-1 (HIF-1) expression that can facilitate cancer bone metastasis (i.e., osteolytic metastasis in the case of breast cancers [31]) by promoting osteoclast generation and inhibiting osteoblast differentiation [80]. Hypoxia in the bone metastatic site can reduce pH and thus form an acidic microenvironment due to the large amount of lactic acids produced by bone metastatic cancer cells and osteoclasts [81]. Also, lactates released by breast cancer cells via glycolysis can be used by osteoclasts as the energy source [82]. Thus, breast cancers with high levels of glycolysis may be prone to bone metastasis. It has been reported that estrogenrelated receptors can bind to the cis-regulatory elements of the promoters of glycolytic genes to affect cells' energy metabolism [83]. Thus, breast caner cells over-expressing ER or PR (i.e., luminal A/B) may favor glycolysis and thus bone metastasis.

On the onset of bone resorption, the metabolism of cancer cells needs to undergo a drastic alteration from osteogenic [84, 85] to osteolytic to escape from the metastatic dormancy [4], the process of which is nourished by nutrients released from the bone matrix such as glucose,

glycine, serine and glycerol [86]. The levels of several enzymes controlling de novo serine synthesis such as phosphoserine phosphatase (PSPH), phosphoglycerate dehydrogenase (PHGDH), phosphoserine aminotransferase 1 (PSAT1) and have been identified differentially elevated in breast cancer cells with high propensity for bone metastasis as compared with those without metastatic potency [87], implicating the stimulatory role of serine on osteoclast formation and osteoclastogenesis [87, 88]. On the other hand, serine is required for sustained ER signaling through regulating acetyl-CoA metabolism for histone acetylation [89]. Thus, luminal breast cancer cells harboring overt ER expression can better survive the dormancy period by satisfying the requirement on serine production for transiting cells from the osteogenic to the osteolytic state.

Another character of the bone microenvironment is high calcification deposition. Microcalcifcation in the primary site of breast cancer has been associated with matrix degradation, inflammation, and cell proliferation and migration [90]. Breast microcalcifcation, being subgrouped into type I and II that are composed of calcium oxalate crystals and hyaluronan respectively [90], has been commonly used for breast cancer screening and diagnosis. Thus, breast cancer cells capable of increasing the intracellular concentration of calcium ion through phagocytosis and/or degrading intracellular crystals may better survive the bone microenvironment and colonize in the bone [91]. Importantly, estrogen can regulate intestinal calcium absorption via differential roles of ERα and ERB on duodenal epithelial cellular plasma membrane calcium ATPase (PMCA) and transient receptor potential cation channel subfamily V member 6 (TRPV6) [92]. The effects of estrogenic agonists on stimulating acute calcium signaling have been previously reviewed in the cardiovascular system [93]. In addition, breast cancer cells preferring bone metastasis can sense the extracellular calcium concentration via calcium-sensitive receptors for promoted migration [94–96]. These have, collectively, implicated and explained the propensity of luminal breast cancers towards bone metastasis.

Liver microenvironment

The liver microenvironment is primarily composed of hepatic stellate cells (HSCs), hepatocytes, sinusoidal endothelial cells and various types of immune cells including metastasis-associated macrophages [97]. In particular, metastasis-associated macrophages can secrete granulin to activate HSCs that, once activated, sustain tumor growth by generating a fibrotic and immune-permissive milieu, and facilitate ECM degradation by secreting a panel of growth factors and cytokines [98–100].

Dai et al. Molecular Biomedicine (2025) 6:20 Page 8 of 30

One critical characteristic of the liver microenvironment is hypoxia [101]. This makes cancer cells translocated to the liver exhibiting a similar hypoxic glycolytic metabolic profile with the local hepatic cells, i.e., elevated glycolytic activity and reduced mitochondrial metabolism. A study comparing the metabolisms of breast cancer cells metastasized to different organs reported that transformed cells migrated to the liver displayed enhanced glycolysis, among other features, which can be attributed to the increased expression of HIF-1α [101]. Besides, oxidation phosphorylation and glutamine metabolism are weaker in cells colonized in the liver, further facilitating the adaptation of cancer cells to the hypoxic milieu in the liver [101]. Thus, the highly hypoxic liver microenvironment forces cells capable of surviving the liver milieu to evolve the feature of favoring anaerobic glycolysis, i.e., the Warburg effect. On the other hand, it is known that ALDH1A3 promoted pancreatic cancer metastasis by enhancing cellular glycolysis [102], implicating that breast cancer cells harboring ALDH1 overexpression (i.e., the HER2+subtype) are easier to adapt to the microenvironment of the liver.

Another feature of the liver microenvironment is the production of ketone bodies that cannot be consumed by normal adult hepatocytes [103]. Ketone bodies can be catalyzed to acetyl-CoA through ketolysis for energy production under glucose shortage [104]. It has been underscored that simultaneously targeting fatty acid oxidation and HER2 is feasible for treating HER2+breast cancers or, in other words, blocking ketolysis can sensitize HER2+tumors to the corresponding targeted therapies [105]. This implicates the metabolic plasticity of HER2+breast cancer cells that can be fueled by both glucose and ketone bodies, and also explains the organotropism of the HER2+subtype towards liver.

The third unique feature of the liver microenvironment is its high accessibility due to its dual blood supply system. In particular, liver receives the blood supply from both hepatic arteries and hepatic portal vein, and thus has a much lower sinusoid blood pressure gradient [106, 107]. This unique architecture of liver makes traveling cancer cells easier to attach to the sinusoidal endothelium for seeding. It has been observed that HER2 status is the only breast cancer receptor correlated with the presence of circulating tumor cells [108], suggesting that HER2+breast tumor cells are the easiest, among other subtypes, to gain the migration ability. Thus, it is no wonder that HER2+breast cancers prefer metastasizing to the liver given their abundance in the blood stream.

Brain microenvironment

The brain parenchyma is composed largely of neurons and glial cells (i.e., astrocytes, microglia,

oligodendrocytes) [109]. Among these cells, the roles of astrocytes during brain metastasis are the best-understood. For instance, astrocytes secrete growth factors and cytokines to support neurons, the ability of which can be hijacked by TNBC (MDAMB231) cells to favor metastasis [110, 111]. Specifically, presynaptic neurons release glutamates that are rapidly absorbed by astrocytes and postsynaptic neurons during glutaminergic synapses [112], and metastatic TNBC cells (MDAMB231) can use glutamates released by neuronal cells to activate N-methyl-D-aspartate (NMDA) receptors and form pseudo-tripartite synapses with glutamatergic neuron cells for promoted brain colonization [113]. Oligodendrocytescan can interact with neurons to drive myelin plasticity and participate in the crosstalk between neurons and tumor cells for promoted tumor cell survival and colonization [114]. Microglia, also known as macrophages residing in the brain, facilitate breast cancer cell colonization in the brain via serving as active transporters and guiding rails in a Wnt-dependent manner [115].

One critical characteristic of the brain microenvironment is high oxygen supply and oxidative stress as a result of a high oxygen consumption to satisfy its high demand on energy turnover [116]. Thus, breast cancer cells colonized in the brain displayed a high plasticity regarding pathways used for energy supply that include, e.g., glycolysis, oxidative phosphorylation, pentose phosphate pathway, gluconeogenesis, branched chain amino acid oxidation [117], and acetate metabolism [118]. In consistent with this, cancer cells capable of surviving in the brain utilize fuels from both glucose and/or ketone bodies [116], with glucose being the primary source for generating the energy and nutrients including lactic acids and nonessential amino acids such as alanines, gamma aminobutyric acid (GABA), glutamic acids, glutamines and glycines [119]. Evidence supporting the essential role of lipid metabolism in brain metastasis include, e.g., the aid of FABP7 (a member of the FABP family actively participating in fatty acid metabolism) in the glycolysis and lipid droplet storage process of HER2+breast cancer cells (BT474) for surviving the brain microenvironment [120]. Besides satisfying the requirement of a constant high energy supply, cancer cells capable of colonizing in the brain exhibited improved glutathione (GSH) system to respond to the high oxidative stress [121]. For instance, GABA catabolism has been reported to be utilized by HER2+(SKBR3) and TNBC (MDAMB231) cells metastasized to the brain for enhanced nicotinamide adenine dinucleotide (NADH) generation [122].

One unique feature of the brain is its blood-brain barrier (BBB) that is a non-fenestrated endothelium stitched together by tight junctions and supported by astrocytes, pericytes and a basement membrane [123]. The BBB

Dai et al. Molecular Biomedicine (2025) 6:20 Page 9 of 30

protected the brain from being invaded by cancer cells. Thus, malignant cells capable of metastasizing to the brain need to evolve abilities of transmigrating through the BBB, with several strategies destroying the BBB being identified. For instance, TNBC (MDAMB231) cells produced cathepsin S to proteolyze the junctional adhesion molecule JAM-B [124], and secreted miRNA-181c-enriched extracellular vesicles to down-regulate phosphorylated cofilin for interrupted actin dynamics [125], which ultimately led to BBB breakdown.

Lung microenvironment

The lung microenvironment is composed of varied types of cells including, e.g., alveolar epithelial cells that can be subgrouped into type I and type II, type II alveolar epithelial cells, endothelial cells, fibroblasts, and various kinds of immune cells including alveolar macrophages [126]. Resident cells in the lung can establish a pro-metastatic niche by, e.g., secreting abundant chemokines such as CXCL12 and CCL21 to attract breast cancer cells over-expressing their receptors CXCR4 [127] and CCR7 [128] to the lung [129]. Alveolar macrophages, of note, facilitate lung metastasis by suppressing T cell responses and producing the pro-inflammatory mediator leukotriene B4 (LTB4) that is a potent lipid chemoattractant driving neutrophil swarming and leukocyte migration [130, 131].

One critical characteristic of the lung microenvironment is the oxidative stress. In contrast to the micro milieus of the bone and liver, lung is the respiratory organ responsible for oxygen inhale and carbon dioxide exhale and thus contains high oxygen levels. Therefore, transformed cells need to overcome the oxidative stress before colonizing in the lung [132–134]. Various strategies have been evolved by metastatic cancer cells to adapt to the lung microenvironment. For instance, TNBC (MDAMB231) cells can counteract electron leakage and ROS generation by up-regulating peroxisome proliferator-activated receptor gamma coactivator 1 alpha (PGC-1 α) for improved mitochondrial biogenesis [132], and elevating the expression of antioxidant proteins such as peroxiredoxin 2 (PRDX2) [135]. Alternatively, reprogramming the metabolism to favor abundant oxygen supply represents another strategy of cancer cells to get immune to the oxidative stress. For example, increased activity of pyruvate carboxylase (PC), an enzyme catalyzing pyruvate to produce oxaloacetate for gluconeogenesis and TCA cycle replenishment, has been identified for lung metastatic TNBC-like (i.e., 4T1) cells towards increased glycolysis and oxygen consumption [136].

Another feature of the lung microenvironment is its tightly connected capillary endothelial cell wall. For effective oxygen inhale and exhale, the lung has developed an abundant capillary network. Traversing such a capillary barrier is required for cancer cells to colonize in the lung [126]. One strategy evolved by transformed cells (3LL-LLC, a mouse Lewis lung carcinoma cell line) is to induce increased expression of matrix metalloprotein 9 (MMP9, a protein responsible for ECM breakdown) in lung endothelial cells and macrophages via the VEGFR-1/Flt-1 tyrosine kinase (TK) axis [137].

The third characteristic property of the lung microenvironment is its metastasis-suppressive niches capable of inhibiting cancer cell growth [126]. In particular, a perivascular niche was identified capable of inducing sustained quiescence among breast cancer cells metastasized to the lung, bone marrow and brain [138]. The inhibitory effect of this metastasis-suppressive niche is attributable to endothelial-derived thrombosondin-1 (TSP1) [138]. In addition, CX3C-chemokine receptor 1 (CX3CR1)+monocytes were recruited by the ligand of CX3CR1 derived from lung endothelial cells (i.e., CX3CL1) to activate natural killer (NK) cells for prevented lung metastasis [139]. Also, bone morphogenetic proteins (BMPs) originated from lung fibroblasts had an inhibitory effect on the self-renewal abilities of CSCs [140, 141]. Tumor cells have evolved various approaches to overcome such a growth-suppressive environment for survival. One strategy widely adopted by tumor cells is to augment the cancer stemness through secreting relevant proteins. For instance, TNBC (MDAMB231) cells have been shown capable of producing polypeptide N-acetylgalactosaminyltransferase 14 (GALNT14) that augments the self-renewal properties of breast cancer cells via exploiting macrophage-derived fibroblast growth factors (FGFs) and inducing macrophage infiltration [142]; and TNBC (MDAMB231) cells metastasized to the lung could secrete tenascin C (TNC), an protein residing in the stem cell niche and sites of epithelial-mesenchymal interactions [143, 144], to amplify stem cell-related signalings such as the Wnt and Notch pathways [145]. Another strategy used by TNBC (MDAMB231) cells is to secrete extracellular vesicles and transform the residing cells in the milieu to foster a metastatic-promotive niche. For example, TNBC-derived exosomes (using MDAMB231 as the modeling tumor cell line) expressing integrins α6β4 and α6β1 promoted the expression of the pro-inflammatory gene S100A4 and activated the oncogenic protein Src in resident cells [146]; and chemotherapy-induced extracellular vesicles secreted by TNBC cells (i.e., MDAMB231, 4T1) were enriched with annexin A6 (ANXA6), which is a Ca²⁺-dependent protein capable of activating NFkB-dependent endothelial cells and expanding Ly6C + CCR2 + monocytes [147].

Dai et al. Molecular Biomedicine (2025) 6:20 Page 10 of 30

Organ redox milieu and cancer metabolic profile define metastasis organotropism

From the perspective of the 'soil' or organs frequently being metastasized by breast cancer cells (i.e., bone, liver, brain, lung), bone and liver are relatively hypoxic, brain and lung are enriched with oxygen. Accordingly, breast cancer cells favoring bone and liver metastasis are largely luminal A and B subtypes, and those preferring brain and lung are primarily HER2+and TNBC cells. In particular, the oxidative status increases from bone, to liver, to brain, and to lung, which gradually corresponds to the organotropism of luminal A, luminal B, HER2+and TNBC cells (Fig. 2).

From the perspective of the 'seed' or breast cancer cells, the cancer stemness and metabolic plasticity of these cells increase from luminal A and luminal B, to HER2+and TNBC cells, with a giant discrepancy being occurred between the luminal (luminal A/B) and nonluminal (HER2+and TNBC) subtypes. In other words, while luminal A and B cells can solely resort to the Warburg effect or anaerobic glycolysis to fuel their energy production, HER2+ and TNBC cells can effectively adapt to the oxidative stress imposed by high oxygen exposure and switch to aerobic glycolysis. Indeed, CSCs are known to favor glycolysis instead of oxidative phosphorylation for energy production [148], and aerobic glycolysis is metabolically more efficient than anaerobic glycolysis in producing ATP [149]. On the other hand, as CSCs have a more robust anti-oxidation machinery than the bulk tumor cells that render them easier to survive the oxidative stress, breast cancer cells possessing higher percentages of BCSCs are more easily to colonize in oxygen-enriched organs such as the brain and lung. The type of BCSCs harbored by breast cancer cells also plays a role in the organotropism of transformed cells. By comparing HER2+and TNBC cells that are both enriched with BCSCs, HER2+cancer cells possess a high proportion of ALDH+BCSCs and display more epithelial-like features, and TNBC cells harbor a high percentage of CD24-CD44+BCSCs and manifest more mesenchymallike characteristics [8, 66]. Accordingly, besides brain and lung that can be colonized by both HER2+and TNBC cells, HER2+cancers can also easily metastasize to the liver whereas TNBC cells do not [8]. Recall the vital role of epithelial-mesenchymal transition (EMT) during the metastatic process [150, 151], these evidence suggest that ALDH+BCSCs are more tolerant to hypoxia than CD24-CD44+BCSCs for sustained metabolism or, in other words, ALDH+BCSCs have a less tendency than CD24-CD44+BCSCs in getting metastasis. Such a difference is predisposed by the markers characterizing these BCSCs. That is, ALDH+BCSCs can rely on oxidative phosphorylation for sufficient energy supply, since ALDH is an enzyme involved in glycolysis [66]; and CD44 can aid in tumor migration by participating in the crosstalk between cancer cells and their ECM, given that it is a multi-functional surface adhesion receptor [59].

Therefore, the organotropism of cancer cells is collectively dictated by the redox milieu of the organ and the metabolic profile of metastasized cancer cells (Fig. 3). In other words, cancer cell organotropism is a determination of transformed cells possessing a certain level of metabolic plasticity on whether or not to be ready for colonization in response to the redox levels of the organ microenvironment; and a selection of cancer cells harboring the metabolic properties feasible to be adapted to the redox status of the organ. Indeed, altered redox regulation has been viewed as an essential force driving cancer cell metastasis [152]. Provided with the inefficiency of the migration process where the vast majority of the transformed cells are fated to death, cancer cells capable of traveling through diverse redox environments must possess a highly versatile metabolic profile empowering them with the immunity to the oxidative stress. Cancer cells with the metastatic potential must have higher percentages of CSCs than the bulk tumor cells given the pluripotency of this small cell cohort that can generate tumor cells with diverse metabolic patterns to meet the

(See figure on next page.)

Fig. 3 Factors determining cancer cell metastasis organotropism. The metabolic profile of cancer cells and the redox status of the organ microenvironment dynamically interact during the metastasis cascade to collectively determine cancer metastasis organotropism. Specifically, cancer cells possessing high percentages of CSCs have high metabolic plasticities, since CSCs are able to produce cancer cells relying on different metabolic programs for energy production. Thus, breast cancer subtypes containing high levels of BCSCs (i.e., HER2+, TNBC) have high metabolic plasticities and can metastasize to organs with sufficient oxygen supply (brain, lung); and subtypes harboring low levels of BCSCs (i.e., luminal A, luminal B) are more inclined to develop bone and liver metastasis that are characteristic of hypoxia. In addition to the redox status of the organ microenvironment, other features of the organ milieu may also affect cancer metastasis organotropism. Besides being hypoxic, bone is featured by acidity that is accompanied with hypoxia, osteolysis that provides nutrients for cancer cell survival, and Ca²⁺ deposition that can be regulated by estrogen; liver is characteristic of ketone body production that supply nutrients for cancer cell survival, and dual blood supply system that favors cancer cells with high abundance in the blood (i.e., HER2+). Besides having sufficient oxygen supply that may impose the oxidative stress, brain and lung have characteristic barriers that allow only cancer cells that have evolved the corresponding strategies to colonize, i.e., the unique blood-brain barrier (BBB) in the brain and the capillary endothelial cell wall and metastasis-suppressive niche in the lung

Dai et al. Molecular Biomedicine (2025) 6:20 Page 11 of 30

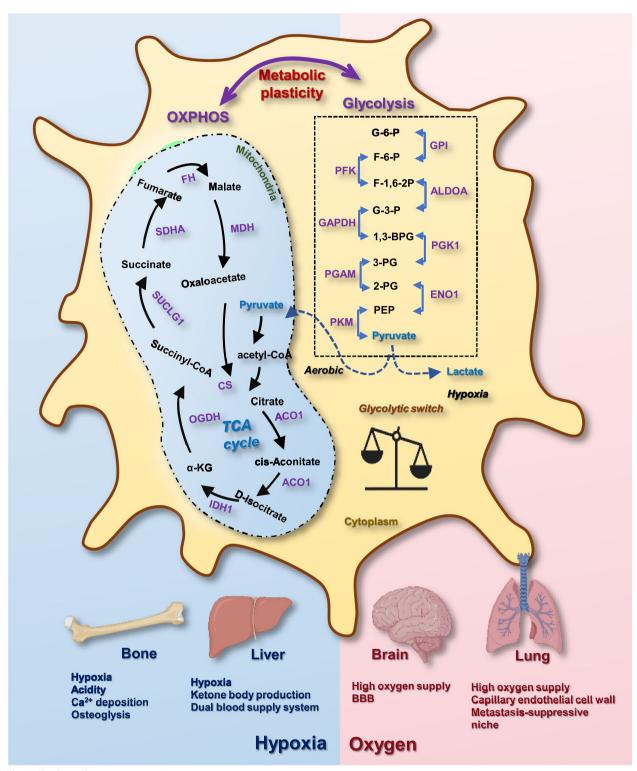


Fig. 3 (See legend on previous page.)

requirement of a new environment. It is thus no wonder that cancer cells with metastatic potential possess higher cancer stemness, and cancer cells with the organotropism for highly oxidative organs such as brain and lung are more stem-like and more malignant as compared with their peers containing more bulk tumor cells. Dai et al. Molecular Biomedicine (2025) 6:20 Page 12 of 30

Insights gained from breast cancer organotropism apply to other types of cancers. That is, the higher metabolic plasticity that cancer cells possess the higher stemness these transformed cells harbor. For example, the metabolic signature of hepatocellular carcinoma cells with stable knockdown of EphA2 and Acly (two genes with demonstrated effects in promoting the aggressiveness and self-renewal ability of cancer cells) was significantly enriched with fatty acid, cholesterol and TCA metabolite profiles and accordingly, Ephrin-A3/EphA2 signaling was identified as an important molecular axis in regulating the metabolic plasticity of hepatocellular carcinoma cells to enhance cancer stemness [153]. As another example, β-hydroxybutyrate significantly enhanced the self-renewal and migration potential of colon cancer cells, the process of which involved decreased extracellular acidification rate (ECAR) and expression of genes related to glycolysis, as well as increased oxygen consumption rate (OCR) and expression of genes associated with mitochondrial biogenesis and cancer stemness [154]. Similarly, metabolic plasticity has been considered as one primary source driving the stemness of pancreatic cancer cells, with the loss of mitochondrial ISGylation (a post-translational modification process regulated by interferon-stimulated gene 15) being a concomitant event occurring along with reduced oxidative phosphorylation (OXPHOS) and decreased amount of cancer stem cells [155]. Consistent with this, another study reported that bioenergetic modulators such as 2-deoxyglucose, dichloroacetate and phenformin decreased the CSC percentage of pancreatic ductal adenocarcinoma cells by switching its metabolic mode from glycolysis to glutamine metabolism [156].

Cancer cell vulnerability during metastasis Metabolic plasticity and cancer stemness

The high metabolic plasticity and oxidative damage resistance of CSCs can be attributed to the evolutionary selection during metastasis that is fated to happen when the primary tumor site could not meet the nutritional requirement of the ever-growing tumor body (Fig. 4). Oxidative stress functions as the selection power to limit the amount the cancer cells during metastasis [134]. It has been reported that antioxidants increased the number of circulating tumor cells in the blood and metastatic tumor sites [134, 157–159], and metastatic cancer cells undergoing metabolic alterations have been perceived with reduced levels of ROS generation [121, 160–164].

Oxidative stress as a selection pressure

The selection force imposed by the oxidative stress is likely to happen via reducing the ability of metastatic cells to fully reactivate the anabolic pathways required for tumor growth (Fig. 4). For instance, nicotinamide adenine dinucleotide phosphate (NADPH) and its reducing equivalents are needed for cancer cells to combat with the oxidative stress; and this leads to inhibited acetyl-CoA carboxylase and thus decreased fatty acid synthesis (an anabolic process) for reduced NADPH consumption [165]. Nonetheless, once metastatic tumor cells have adapted to the oxidative stress and grown beyond a few millimeters in diameter (i.e., having survived the quiescence stage), they regain the ability of rapid growth that requires a broad activation of the anabolic processes.

The proposed insights here underpin the imprinting role of cell metabolism in empowering cells with discrepant manifestations, some of which may ultimately drive carcinogenesis. Thus, gaining mechanistic insights into the dynamic process of oxidative stress in stratifying and editing the metabolic plasticity of transformed cells may help identify critical clues for the establishment of oncotherapeutics with improved efficacy and safety.

Pro-oxidative approaches in cancer control Existing pro-oxidative onco-therapeutics

Antioxidant supplementation has been proposed to provide health benefits via reducing ROS levels [166]. Despite consecutive clinical efforts examining the efficacy of antioxidants in halting carcinogenesis, no supportive evidence has been reported regarding the hypothesized roles of antioxidants in reducing cancer incidence or cancer-related deaths [167]. Instead, both pre-clinical and clinical data have suggested that antioxidants tend to promote cancer development and increase cancer-related deaths [168-170]. This leaves us the therapeutic opportunity to target cancer cells via imposing a high selection force during cell metastasis, especially before they have successfully colonized and adapted to the redox environment of the targeted sites. In principle, when the selection pressure is sufficiently high, cancer cells fail to metastasize that are eventually restricted to their primary site and become benign.

It has been reported that pro-oxidant therapies can arrest cancer progression via exacerbating the oxidative stress in cancer cells and/or blocking the metabolic adaptation process to confer oxidative stress sensitivity [171]. Many existing onco-therapeutic strategies can be considered fell into this kind, including the primary options for cancer treatment, i.e., chemotherapy and radiotherapy [172–178] (Fig. 5). Specifically, chemo-therapies can all impose oxidative stress to cancer cells [175–177]. For example, paclitaxel (a herbal-derived agent used for treating a plethora of cancers such as breast cancer, ovarian cancer, cervical carcinoma, endometrial cancer, advanced prostate cancer, bladder cancer, and nonsmall cell lung carcinoma [179]) can activate phosphatase

Dai et al. Molecular Biomedicine (2025) 6:20 Page 13 of 30

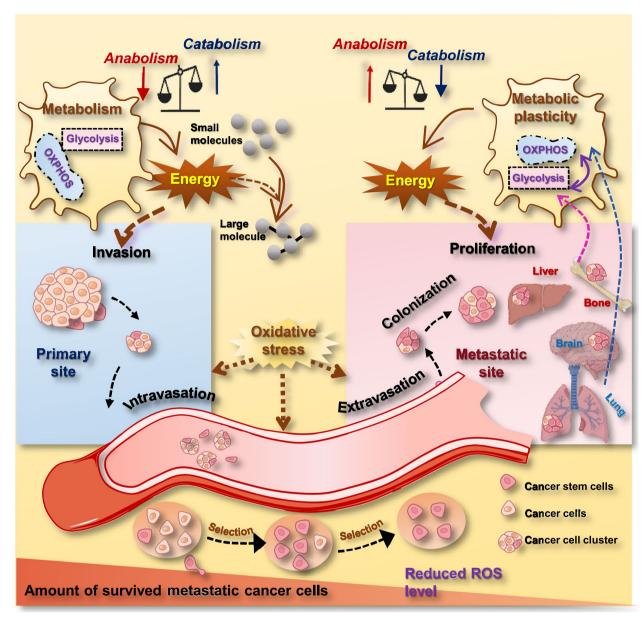


Fig. 4 Metastatic cells are subjected to oxidative stress selection and are vulnerable to pro-oxidant therapeutics. During the metastasis cascade (i.e., invastion, intravasation, circulation, extravasation, colonization), cancer cells are subjected to the selection of the oxidative stress, resulting in declined number of cancer cells and enrichment of cancer stem cells. These cancer stem cells are featured with reduced level of ROS and enhanced metabolic plasticity. Under the selection power of oxidative stress, cells with reduced intracellular ROS generation and reduced basal redox level easily survive. When arriving at the secondary metastatic site, cells with high metabolic plasticity can easily get adapted to the microenvironment of the metastatic site, i.e., cells rely more on glycolysis when the microenvironment is hypoxic, and rely more on oxidative phosphorylation when the microenvironment is oxidative. In the case of breast cancers, TN and HER2 + breast cancer cells that have high metabolic plasticity can easily get adapted to the oxidative microenvironment of brain and lung and thus rely more on OXPHOS for energy production after colonization, and luminal A/B breast cancer cells more easily develop liver and bone metastasis and favor glycolysis for energy production once colonized in the secondary lesions. Also worth mentioning is that successful metastasis often occurs in the form of cell clusters and is accompanied with suppressed anabolism in the beginning that regains the activity after colonization. In the beginning of the metastasis cascade, cells need lots of energy to accomplish invasion, intravasation, circulation and extravasation, leading to reduced amount of energy used for anabolism. Once metastatic cancer cells have colonized to the new environment and grown beyond a few millimeters in diameter, they regain the ability of rapid growth as a result of fully activated anabolic processes

Dai et al. Molecular Biomedicine (2025) 6:20 Page 14 of 30

and tensin homolog deleted on chromosome ten (Pten) and thus suppress the phosphoinositide 3-kinase (PI3K) axis via producing overt amount of ROS [179]; doxorubicin, an anthracycline type of chemotherapeutic agent canonically used for treating cancers such as breast carcinoma, lymphoma and sarcoma, can produce excessive ROS to induce lipid peroxidation as well as damage DNA and protein [180]; imexon, a small molecule capable of depleting GSH and increasing ROS levels via binding to thiols, has been examined for activity against non-Hodgkin lymphoma [181]; arsenic trioxide, capable of causing electron leakage and superoxide anion radical $(O^{2^{\bullet}})$ generation by impairing the function of the electron transport chain, has been used for treating acute promyelocytic leukemia [182]. Radio-therapies are known capable of generating O2.-, hydroxyl radicals (OH.) and hydrogen peroxide (H2O2) [178, 183] to take on their cytotoxic effect [184], and inducing endogenous ROS production in mitochondria to trigger intracellular redox imbalance [185]. However, these commonly seen prooxidative onco-therapeutic approaches are characteristic of unavoidable severe side effects and drug resistance [186], rendering the development of innovative therapies capable of alleviating these disadvantages an ever-lasting focus in the field of oncology.

Some antioxidants such as vitamin C, once given at a dose far exceeding that is recommended for daily intake, turn to be pro-oxidative. Vitamin C (also named ascorbate) is canonically considered as an antioxidant supplement for the benefits of health; yet it becomes prooxidative once infused intravenously at a high dose [187]. This is because that superphysiologic levels of vitamin C can be uptaken by cancer cells in its fully oxidized form, dehydroascorbate, via the glucose transporter 1 (GLUT1) expressed on the membrane of cancer cells; after entering the cancer cells, dehydroascorbate is reduced back to ascorbate, the process of which consumes the reducing equivalents and induces cellular oxidative stress [188, 189]. Given the little toxicity reported, high dose vitamin C seems to be promising in treating cancers as an adjuvant therapy. However, its additive anticancer impact still remains debatable that requires rigorous clinical validations [190]. In this context, increasing attention has been paid to naturally existing matters such as magnetic field, electrical field, light, and ultrasound in treating cancers that are, in principle, of high efficiency and low low cost.

Magnetic fields can be categorized into static magnetic field (also named constant magnetic field) or dynamic magnetic field (Fig. 5). Static magnetic fields can be produced by permanent magnets or solenoidal coils with unidirectional currents, and be classified into weak (<1 mT), medium (1 mT to 1 T), strong (1 to 5 T), and ultra-strong (>5 T) magnetic fields [191]. Dynamic

magnetic fields vary with time and can be grouped as an alternating magnetic field, pulsed magnetic field, pulsating magnetic field, and geomagnetic field. While alternating magnetic fields can be produced by an electromagnetic coil with a current of a certain frequency or through a magnet with regular motion, pulsed magnetic field, pulsating magnetic field and geomagnetic field can be generated by an electromagnetic coil with a pulsed current, alternating current power supply, and Earth and ionosphere, respectively [191]. Magnetic fields at frequencies below 100 kHz could non-invasively induce the death of cancer cells without causing the thermal effect [186, 192]. Thus, static magnetic fields and low-frequency magnetic fields (frequency < 300 Hz) have been shown promising for treating a variety of cancers pre- or clinically including, e.g., breast cancer [193], colon carcinoma [194], Lewis lung carcinoma [195], gastric cancer [196], Ehrlich ascites carcinoma [197], and fibrosarcoma [198]. Magnetic therapies take actions via promoting the generation of ROS [199, 200]. For instance, treating human prostate cancer cells with a 60 Hz sinusoidal magnetic field for 48 h resulted in ROS accumulation that ultimately led to the apoptosis of transformed cells [201]. Despite the increasing evidence supporting the inhibitory effect of magnetic fields on tumor progression, contradictory reports exist. For example, long-term exposure of 7,12-dimethylbenz(a)anthracene (DMBA)-treated female rats to an alternating magnetic field of low flux density promoted the growth and increased the incidence of mammary tumors [202]; and there was some evidence for elevated risk of developing leukemia among children aged above 5 years old living in the proximity to transformer stations [203-205]. These underpin the complicated mechanisms driving the impact of magnetic fields on cancer cells that are far from being completely understood, and such uncertainties regarding the treatment outcome have substantially hindered the clinical translation and wide application of magnetic therapies.

High-voltage electrical pulse (HVEP), taking advantages of concentrated energy induced by electric pulses, has been shown promising in ablating cancers especially those difficult-to-treat solid tumors (Fig. 5). By varying the electrical field strength and treatment duration, electroporation can be either reversible or irreversible. In general, the energy used for generating electrical pulses of irreversible HVEP is much more intensive than that is required for producing reversible HVEP. That is, at least an amplitude up to 3000 V and 80–100 pulses are required for irreversible HVEP to be effective in treating cancers, and an amplitude of 100–1000 V and 8 square wave pulses of 100 µsec are typically used in generating reversible electroporation is lethal, and that of

Dai et al. Molecular Biomedicine (2025) 6:20 Page 15 of 30

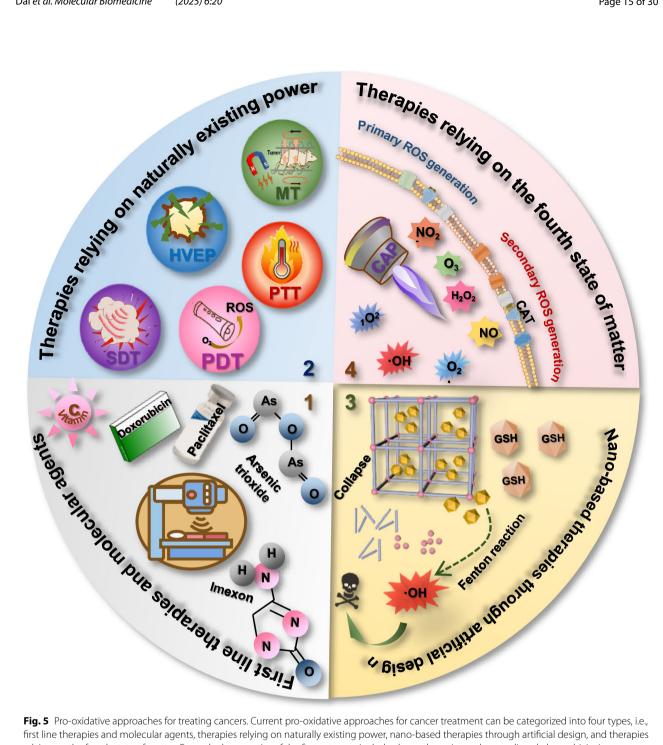


Fig. 5 Pro-oxidative approaches for treating cancers. Current pro-oxidative approaches for cancer treatment can be categorized into four types, i.e., first line therapies and molecular agents, therapies relying on naturally existing power, nano-based therapies through artificial design, and therapies relying on the fourth state of matter. Example therapeutics of the first category include chemotherapies such as paclitaxel, doxorubicin, imexon, arsenic trioxide, radiotherapies, and high dose of vitamin C. Therapeutics of the second category can be further divided into magnetic therapy (MT), high-voltage electrical pulse (HVEP), photodynamic therapy (PDT), photothermal therapy (PTT), and sonodynamic therapy (SDT), which take advantages of the magnetic field, electrical field, light, heat and sound, respectively. One example of the third category is GSH-responsive metal-organic-framework (MOF) that generates ROS via Fenton reaction. Cold atmospheric plasma (CAP) belongs to the fourth state of matter and the fourth category of pro-oxidative therapeutics. CAP is composed of a cocktail of reactive oxygen and nitrogen species such as OH+, O₂+, 1O², H₂O₂, O₃, NO, NO₂. These species take actions via interacting with the cell surface to generate primary and secondary ROS. For example, long-lived species H_2O_2 and NO_2^- from CAP can interact to generate ONOO⁻. In the vicinity to membrane-associated proton pumps, ONOO⁻ is protonated to ONOOH and decomposed into $\cdot NO_2$ and $\cdot OH$. $\cdot OH$ reacts with H_2O_2 to form HO_2 . The subsequent generation of O_2NOOH and O_2NOOT allows for the generation of primary ${}^{1}O_{2}$. Primary ${}^{1}O_{2}$ causes local inactivation of CAT. Surviving $H_{2}O_{2}$ and ONOO at the site of inactivated CAT further generate secondary 10₂. Secondary 10₃ can, in turn, inactivate CAT to trigger 10₃ auto-amplification and promote secondary 10₃ generation

Dai et al. Molecular Biomedicine (2025) 6:20 Page 16 of 30

reversible electroporation is transient that is restricted to the impact on cell membrane permeability. Increased ROS cellular levels has been associated with HVEP. For instance, a statistically significant ROS increase was observed 3 h after electric pulse exposure in glioblastoma cells cultivated using the standard culture condition, and was shown immediately post-exposure in glioblastoma cells cultivated as neurospheres [207]. Lots of clinical evidence has suggested the feasibility of using irreversible HVEP in ablating hepatoma [208-211], kidney carcinoma [212, 213], prostate cancer [214-216] and pancreatic cancer [217-224]; and of using reversible HVEP in treating large size cutaneous metastases of all cancer types [225]. However, HVEP therapies are lack of sufficient targeting ability and may impair the functionality of certain organs receiving the treatment. For example, irreversible electroporation may cause prostate cancer patients loosing up to 41% potency post-treatment [226].

Photodynamic therapy (PDT) is known to rely on the generation of cytotoxic ROS on the ignite of a specific light wavelength in killing transformed cells (Fig. 5). In principle, the photosensitizer administrated to the tumor site can react directly with nucleic acid, protein, or unsaturated lipid to generate ROS such as O²., OH., or hydrogen peroxide (H_2O_2) in the presence of oxygen (O₂) through proton or electron transfer (namely the type I reaction); the excited photosensitizer can react with molecular O₂ to form singlet oxygen (1O²) via energy transfer (called type II reaction), where 1O2 is a leading ROS taking actions in PDT [227, 228]. The anticancer efficiency of PDT largely depends on the properties of photosensitizer that is expected to preferentially accumulate in the tumor tissues. However, as the tumor-targeting feature of a photosensitizer is actualized largely through a passive rout, i.e., via the permeability retention (EPR) effects in tortuous blood vessels and leaky vasculature, one major disadvantage of PDT is the retention of photosensitizers in cells that can last for several weeks [229]. During this period, the skin and eyes can become extremely sensitive to light and quickly become swollen, sunburned, and blistered [230]. Besides, the ROS generation of photosensitizers varies in vivo, rendering it difficult to determine the light dose sufficient to induce PDT in a particular tumor loci without causing photobleaching reactions. Another obstacle faced by PDT is the limited penetration in-depth of light, i.e., approximately 10 mm, restricting its use for treating visceral tumors.

Ultrasound-based sonodynamic therapy (SDT), using sonosensitizers and ultrasound in cancer treatment, also relies on ROS generation in killing cancer cells (Fig. 5). It differs from PDT in having a deeper tissue penetration ability, i.e., 70–100 mm, and thus has become an attractive option for treating solid malignant tumors

[231] and be applied in treating a broader range of cancer types including, e.g., liver cancer [232], breast cancer [233, 234], glioblastoma [235], colon cancer [236], and pancreatic cancer [237]. Similar to PDT, the efficacy and safety of SDT are primarily determined by the properties of sonosensitizers, and appropriate ultrasonic dose setting is vital in reaching the desirable therapeutic outcome [238]. Thus, sonosensitizer screening and state-of-the-art SDT multifunctional equipment establishment are critical issues, among others, to be resolved before SDT can be truly translated into clinical use [238, 239].

Besides, lots of nanoparticles designed for the anticancer purpose such as GSH-responsive metalorganic-framework (MOF) [240] and magnetic-based nanomaterials [241] have been considered capable of imposing the oxidative stress to transformed cells (Fig. 5). For example, the TBD-Pt(IV)@MOF-199 nanocomposite generated O2on light irradiation via consuming intracellular GSH and releasing Pt(IV), leading to the desirable anti-cancer outcome [240]. As another example, the methylene blue immobilized copper ferrite nanocomplex MB-CuFe converted hydrogen peroxide (H₂O₂) to ROS and depleted cervical cancer cells by functioning as the photodynamic therapy under 660 nm laser irradiation [241]. Regardless of the intricate design of these nanoparticles, one unanimous concern is the introduction of nanomaterials into the body, where their distribution and retention should be carefully assessed before nanoparticle-based anti-cancer agents can be considered feasible for clinical use.

Cold atmospheric plasma as an emerging pro-oxidative onco-therapeutics

This section introduces the fourth state of matter, cold atmospheric plasma (CAP), as an innovative type of pro-oxidative strategy in treating cancers (Fig. 5). CAP is composed of a plethora of reactive and oxygen species (RONS) such as short-lived species superoxide anion $(O_2 \bullet)$, OH• and ${}_1O^2$, and long-lived species such as H_2O_2 , ozone (O₃) and nitric oxide (NO), and is typically operated at the atmospheric pressure and the room temperature $(37 \sim 44 \, ^{\circ}\text{C})$ [1, 242–246]. The diversified biomedical applications of CAP has attracted incremental attention in recent years especially in its use as an emerging oncotherapy [1, 242-247]. Lots of pre-clinical studies have confirmed the use of CAP for treating a large spectrum of cancers including, e.g., breast (especially TNBCs) [243, 248–250], prostate [251], bladder [250], brain [252, 253], colon [254], lung [255] and pancreatic [256, 257] cancers as well as melanoma [258, 259], with diversified molecular mechanisms being proposed. Take breast cancers for example, CAP has been shown capable of inducing TNBC apoptosis and synergize with epithelial growth

Dai et al. Molecular Biomedicine (2025) 6:20 Page 17 of 30

factor (EGF) for enhanced cancer cell killing [248], halting the epithelial-to-mesenchymal transition process during TNBC cell metastasis [250], uniquely targeting the BCSC cohort of TNBC cells [249], modulating the activity of immune cells in the microenvironment of breast cancer cell [260], and elevating the sensitivity of breast cancer cells to paclitaxel [261, 262], doxorubicin [263], tamoxifen [264]. In addition, the unique benefits of CAP in functioning an emerging onco-therapeutics [265] have been associated with its roles in targeting other critical hallmarks of cancers such as the induction of immunogenic cell death (ICD) among glioblastoma cells [266] and pancreatic cancer cells [267].

The roles of CAP on cancer cells are multifaceted. Yet, all therapeutic benefits of CAP in treating cancers are originated from its intrinsic oxidative nature. It is worth noting that CAP can effectively and uniquely kill transformed cells without harming their healthy peers under appropriate dosage, preventing issues such as therapeutic resistance and evident toxicity.

In addition, CAP can be administrated directly using devices that can action through direct-discharge and indirect-discharge [268, 269]. The source for direct-discharge is called direct dielectric barrier discharge, and devices for indirect-discharge include plasma jets and plasma torches [265]. In general, direct-discharge produces higher amounts of RONS than indirect-discharge. CAP can also be applied indirectly through activating a solution (cell cultivating medium, Ringer's lactate solution, etc. [270]) to produce plasma-activated medium (PAM) followed by treating cells, animals or patients through injection [271]. Compared with CAP produced from the direct approach that contains ample shortlived species, PAM contain more long-lived species [272, 273]. Such a high plasticity of CAP regarding its production and application makes the clinical translation of the fourth state matter into the field of oncology relatively handy and promising.

Pro-oxidative approaches in combination with existing intervention strategies

Pro-oxidative strategies aid in interventional therapy

Tumor resection, utilizing surgical operations to ablate tumors, is one first-line clinical approach for cancer treatment. However, this canonical therapy is challenged by high rates of recurrence that can be primarily attributed to intraoperative cancer cell dissemination and peritumoral invasion [274]. Thus, accurately targeting and eliminating residual tumor cells may prevent metastatic recurrence [275].

Postoperative *in-situ* implantation has been proposed to reduce the likelihood of developing tumor recurrence by removing residual tumor cells. For instance,

an implantable sandwich-structured dual-drug reservoir was fabricated that can concomitantly inhibit neoangiogenesis and induce cancer cell death by releasing combretastatin A4 phosphate and tigecycline [276]. Additionally, pro-oxidative strategies can be used for postoperative *in-situ* implantation. For example, a peroxide copper nanoparticles-loaded hydrogel composite was developed that can eliminate residual lesions via inducing cuproptosis [277].

Intraoperative treatment modalities may also be developed to remove residual tumors taking advantages of pro-oxidative approaches. Given that CAP can be prepared in the liquid form (i.e., PAM), PAM can be used to replace sterile solutions containing antibiotics for rinsing during the surgery. PAM has canonically been used for sterilization [278] and wound healing [279], and was reported capable of inducing immunogenic cell death (ICD) [280-284]. Importantly, PAM has been considered as a mild solution for cancer treatment without harming healthy tissues [285]. Thus, it is expected to achieve desirable therapeutic outcome via intraoperative application of PAM during the surgical process. It is also possible to directly expose post-surgical tissues to CAP to remove residual tumor cells during the operation. Dr. Keith Millikan secured the life of a 75-year advanced pancreatic cancer patient in 2016 through the use of Canady Hybrid Plasma[™] Scalpel that integrates high frequency monopolar current with CAP to simultaneously cut and coagulate biological tissue [286]. Using this tool, the first clinical trial examining the efficacy and safety of CAP as an oncotherapy was approved by the Food and Drug Administration (FDA) in 2019, where 17 out of 20 stage IV solid tumor patents recruited were still alive by the end of this phase I study in 2021 (NCT04267575).

Pro-oxidative strategies mitigate adverse effect of chemotherapy

Chemotherapy has been widely accepted as the primary regimen for treating advanced cancers or unresectable metastatic patients [274]. However, chemotherapy also has metastasis-promoting activities by screening out cells with highly invasive phenotypes, expanding cancer stem cell (CSC) cohorts, and activating the EMT process [287]. For instance, the chemotherapeutic agent paclitaxel increased the number of invadopodia, which is one characteristic feature of cancer cells undergoing systemic dissemination [288, 289]; enhanced the amount of circulating tumor cells by over 1000 folds in breast cancer patients [290], the mechanism of which involves hypoxia-inducible factor-1 (HIF1)-dependent p38 mitogen-activated protein kinase (MAPK) signaling axis [291]; and induced EMT by up-regulating Snail and Twist, which

Dai et al. Molecular Biomedicine (2025) 6:20 Page 18 of 30

are repressive transcription factors of the gene encoding E-cadherin [292].

Chemotherapy functions by killing fast-growing cells without targeting properties. Pro-oxidative strategies may specifically kill cancer cells due to the higher vulnerability of transformed cells to the oxidative stress [1]. This is because that malignant cells have high levels of chaosity that render their anti-oxidative machinery incapable of coping with redox perturbations and thus easily undergo programmed cell death. Therefore, it is plausible to combine chemotherapy with pro-oxidative approaches towards enhanced targeting properties and reduced dosage for minimized adverse effect and low likelihood of promoting metastasis. In addition, pro-oxidative strategies may enhance the therapeutic outcome of chemotherapy. This is because that chemotherapy can enhance cellular oxidative stress [293], rendering its synergies with pro-oxidative approaches naturally compatible. Anthracyclines such as doxorubicin, epirubicin, and daunorubicin, which are widely adopted chemotherapy, have been reported to generate the highest levels of oxidative stress among other agents [294]. It has been reported that anthracycline-based chemotherapy increased levels of oxidative stress and decreased the antioxidant status in small-cell lung cancer patients [295]. Similar results were documented for breast cancer patients after receiving the combined use of doxorubicin, cyclophosphamide and 5-FU [296, 297]; for gastric tumor carriers after receiving adriamyclin, mitomycin and 5-FU; for colon cancer patients having received oxaliplatin, folinic acid and 5-FU; and for prostate cancer patients treated with mitozantrone and prednisolone [298]. A plethora of studies have shown the benefits of combining chemotherapies with pro-oxidative approaches in cancer treatment. For example, CAP, acting as one pro-oxidative tool, synergized with cisplatin in eliminating head and neck cancers [299], and improved the cytotoxic effect of olaparib [300] and paclitaxel [261] in treating TNBC cells.

Pro-oxidative strategies sensitize metastatic tumors to immunotherapy

Immunotherapy has shown a great promise in resolving tumors. The first checkpoint protein blockade shown effective in cancer immunotherapy was antibody-mediated inhibition of cytotoxic T lymphocyte-associated antigen 4 (CTLA-4) that can inhibit T cell expansion and activation via being translocated to the surface of T cells and competing with CD28 for binding to CD80 and CD86 [301]. More interest has been attracted to design therapeutics targeting the programmed cell death protein 1 (PD1)/ programmed cell death—ligand 1 (PD-L1) axis due to its remarkable treatment efficacy, durable response and mild toxicity, which functions by normalizing instead

of purely activating the immune response. Many drugs targeting the PD1/PD-L1 axis have been established and commercialized including anti-PD1 antibodies such as nivolumab (Opdivo), pembrolizumab (Keytruda) and cemiplimab (Libtayo), and anti-PD-L1 antibodies such as avelumab (Bavencio), duravulumab (Imfinzi), and atezolizumab (Tecentriq) [302]. Nivolumab was approved by the FDA for treating advanced melanoma in December 2014, for treating advanced lung cancer in October 2015, for treating unresectable or metastatic melanoma across BRAF status in January 2016, for treating locally advanced or metastatic urothelial carcinoma in February 2017, for treating metastatic colorectal cancer in August 2017, for treating completely resected melanoma gained metastasis in December 2017, in treating metastatic or recurrent non-small cell lung cancer in May 2020, for treating advanced renal cell carcinoma in January 2021, for treating unresectable advanced or metastatic esophageal squamous cell carcinoma in May 2022, and for treating unresectable or metastatic urothelial carcinoma in March 2024. Pembrolizumab was approved by the FDA in the treatment of unresectable advanced or metastatic malignant pleural mesothelioma in September 2024. Cemiplimab received its approval for treating advanced cutaneous squamous cell carcinoma in September 2018, for treating advanced basal cell carcinoma and advanced non-small cell lung cancer in February 2021, for treating advanced metastatic castration-resistant prostate cancer in August 2022, and for treating advanced cervical caner in October 2022. Atezolizumab was approved by the FDA in May 2016 as the first PD-L1 inhibitor of urothelial carcinoma, and for treating metastatic non-squamous non-small cell lung cancer in December 2018. Avelumab was approved by the FDA in March 2017 in the treatment of metastatic Merkel cell carcinoma. Duravulumab was approved by the FDA for treating advanced bladder cancer in May 2017, treating unresectable stage III non-small cell lung cancer in February 2018, treating advanced biliary tract cancer in September 2022, treating metastatic non-small cell lung cancer in November 2022, treating advanced or recurrent endometrial cancer in June 2024, and treating limitedstage small cell lung cancer in December 2024.

Though many immunotherapeutic agents targeting CTLA4 or the PD1/PD-L1 axis have been launched into the clinics for cancer treatment, accumulating clinical evidence has suggested that anti-CTLA4 and anti-PD1 are largely futile in eradicating secondary tumors and extending the lifespan of metastatic cancer patients, due to inherent or acquired drug resistance [303]. Consequently, a number of other T-cell checkpoint inhibitors have been established to treat metastatic tumors over the past years, such as inhibitors targeting lymphocyte

Dai et al. Molecular Biomedicine (2025) 6:20 Page 19 of 30

activation gene 3 (LAG3) and T-cell immunoglobulin and mucin domain 3 (TIM3) [304]. TIM3 expression is associated with advanced cancer stage and lymph node metastasis in lung cancer patients [305], dysfunctional CD8+T cells and NK cells in patients carrying metastatic melanoma [306, 307]. Importantly, TIM3 is implicated in the adapative resistance of cancer cells to PD1 inhibition. Specifically, TIM3 level increased on CD4+and CD8+T cells in metastatic lung cancer patients after anti-PD1 blockage [308]. Indeed, treating lung-tumourbearing CC10-rtTA;Tre-egfr^{T790M/L858R} mice with anti-PD1 and anti-TIM3 extended the lifespan of the diseased animals as compared with applying anti-PD1 blockade alone [308]. In addition, TIM3 inhibition may prime the recruitment of cytotoxic T cells or reinvigoration of exhausted T cells to the tumor site under the metastatic setting. For instance, blocking TIM3 in MMTV-PyMT tumour-carrying mice increased the expression of CXC chemokine ligand 9 (CXCL9) in CD103+dendritic cells, leading to the recruitment of cytotoxic CXC chemokine receptor 3 (CXCR3)+CD8+T cells to tumours [309], which is advantageous in treating immunotherapy-naïve or anti-PD1 refractory cancer patients. Being another checkpoint molecule expressed on NK and T cells, increased LAG3 expression was observed on CD4+and CD8+T cells among mismatch-repairproficient colorectal cancer patients developed liver metastasis [310], and was associated with dysfunctional tumour-infiltrating T cells in human metastatic tumours [310-312]. Combinatorial use of anti-PD1 and anti-LAG3 delayed the growth of fibrosarcoma and colorectal cancer, and attenuated the metastasis of ovarian cancer in vivo [313-315].

Despite these pre- and clinical successes as aforementioned, such immune checkpoint therapy only benefits a fraction of patients. An increasing number of researchers have realized the critical roles played by the tumor microenvironment (TME) in the therapeutic response of these treatment regimens. While an immunogenic (hot) TME contain large amounts of tumor infiltrating cells (TILs), cytokines and high level of PD-L1, non-immunogenic (cold) TME exhibit merely no no T cell infiltration and no PD-L1 expression. Therefore, lots of combinatorial strategies have been established to provide improved clinical results with the aim of creating a hot TME by increasing PD-L1 expression, enhancing TIL infiltration, and targeting other type of cells in the TME.

Besides identifying innovative immune targets and establishing concomitant targeting therapeutics against multiple molecules, combining immunotherapy with pro-oxidative approaches may represent another solution to enhance the sensitivity of metastatic cells to immunotherapy. As aforementioned, metastatic cells especially

those enriched with CSCs are more vulnerable to prooxidative regimen that can easily penetrate through the multi-cell layer within the TME and reach tumor cells. Death of cancer cells in the secondary tumor site elicit signals to the surrounding region to recruit immune cells for enhanced immune cell infiltration and initiate ICD. Indeed, CAP has been shown capable of inducing ICD in a panel of tumor models such as melanoma [280], lung carcinoma [281], nasopharyngeal carinoma [282], and colon cancer [283, 284], restoring skewed macrophage polarization in the TME of TNBC [316], as well as enhancing the therapeutic effect of PD1 blockage therapy in treating head and neck cancer [299] and the treatment outcome of anti-PD-L1 strategy in resolving melanoma [317]. In addition, CAP, being one promising pro-oxidative onco-therapeutic tool, belongs to the naturally existing forms of matter and thus is cheap to produce. It also outweighs other treatment modalities in being specific in killing cancer cells without harming their healthy peers. Thus, combining CAP with immunetherapies may reduce the dosage or treatment time of immunetherapeutic agents, leading to mitigated side effects.

Conclusion

This paper updated the canonical 'seed and soil' theory on metastasis organotropism by viewing the metastatic profile of cancer cells as the 'seed' and the redox status of the organ microenvironment as the 'soil'. Specifically, this paper proposed that cancer cells possessing high plasticity harbor high cancer stemness and can easily colonize in organs with sufficient oxygen supply using breast cancers as the disease model (Fig. 6). From characterizing vital influential factors and the role of redox status in defining organotropism, this paper identified the metastasis process as the most fragile moment sensitive to pro-oxidative perturbations when cancer cells are subjected to the selection of oxidative stress (Fig. 6). Accordingly, this paper reviewed existing pro-oxidative modalities for cancer treatment, and proposed the potential efficacy of CAP, an emerging anti-cancer therapeutics with redox modulatory roles [9, 285], in arresting breast cancer metastasis by intervening with the critical factors identified. In addition to viewing CAP as a redox regulatory tool for interrogating the interactions between cell metabolisms and the oxidative stress, this paper also proposed CAP as a dosage-dependent redox classifier capable of stratifying cancer cells by their metabolic plasticity for specific killing (Fig. 6). These insights may not only advance our understandings on critical factors influencing cancer metastasis organotropism for improved prognosis on cancer metastasis, but also decipher clues why and where pro-oxidative approaches can serve as an

Dai et al. Molecular Biomedicine (2025) 6:20 Page 20 of 30

oncotherapeutic tool and achieve the optimal treatment outcome.

High metabolic plasticity of cancer cells enables them to easily adapt to the redox environment of the secondary organ site, and thus manifests higher malignancy and metastatic potential. CSCs, a small subset of cancer cells characterized by high differentiation plasticity and self-renewal ability, thus more easily develop metastasis than the bulk tumor cells. Given the giant divergence of the redox status between oxygen-enriched organs such as brain and lung and the tumor initiation loci that is typically subjected to hypoxia, cancer cells metastasized to the brain and lung typically contain higher CSC percentages. As a consequence, cancer cells developed brain or lung metastasis are comparably more difficult to treat than those colonized to other organs [318].

By viewing the metastatic process as an opportunity of interrogating the metabolic profiles of transformed cells when they are at the most fragile moment regarding anabolism and redox sensitivity, it is possible to achieve the aim of 'survival with tumors' by killing malignant cells undergoing the metastasis cascade or permanently attracting them to the quiescence state. Yet, cancer cells having survived this selection process may harbor a robust anti-oxidant machinery, resulting in unintended transition of highly-stressed surviving cells into the stemcell-like state. Thus, though many existing pro-oxidative strategies such as chemotherapy, radiotherapy, magnetic therapy, HVEP, PDT, SDT can effectively ablate cancer cells, they may hardly reduce the rate of cancer relapse and metastasis. The proposed innovative type of prooxidative option (i.e., CAP) is unique from this perspective as it can specifically target cancer stem cells [249] and outweighs the other pro-oxidative approaches by, e.g., halting cancer metastasis [250]. These unique traits may be attributed to the multi-modal nature of CAP that modulates the intracellular redox homeostasis instead of simply imposing an external oxidative stress or damaging cells in a brute-force way [319].

CAP is not simply a supplement of current portfolio for cancer therapeutics, but also offers clues for the design of combinatorial anti-cancer strategies by viewing CAP as an adjuvant component. One relevant feature of CAP is to sensitize the TME for enhanced drug sensitivity, rendering it possible to combine CAP with existing therapeutic strategies such as interventional approaches, chemotherapies and immunotherapies to achieve improved treatment response. This may be even more promising for treating cancers with high likelihoods of gaining metastasis or having already gone to the metastasis stage given the essential role of TME and its interplay with metastatic cells in determining cancer cell colonization. Also, with increased research interest in immunotherapies, it is important to resolve issues limiting the clinical efficacy and safety of these treatment approaches. Applying CAP (either directly via ejection or indirectly via administrating its activated liquid) prior to immunotherapies may not only help rewire 'cold' tumors back to the 'hot' state, but also reduce the chance of developing side effects by reducing the load of immuno-agents; and this may be attributed to the priming role of CAP in the immune system besides its TME-sensitizing capacity.

Another future trend sparked by CAP in designing new approaches for treating cancers is to overcome remaining hurdles limiting the clinical translation of CAP taking advantages of nanomaterials. Despite the plethora of unique beneficial features of CAP identified in oncology, short-lived reactive species within CAP such as OH., O_2 • and ${}_1O^2$ play the leading roles in killing transformed cells but have transient life-spans [248, 249]. One strategy would be to preserve the activities of these short-lived species using materials such as covalent organic frameworks, graphene, and hydrogel [320]. Another possibility is to enhance the production of short-lived species by synergizing it with, e.g., magnetic therapies, HVEP, PDT or SDT. For instance, magnetic therapies typically kill cancer cells via the Fenton effect that relies on OH. production, and photosensitizers may serve as the 1O²

(See figure on next page.)

Fig. 6 Intrinsic logic for identifying innovative therapeutic approaches from molecular mechanism for cancer metastasis intervention. ①Using breast cancer as the tumor model, cancer metabolic profile and organ redox milieu are identified as two primary factors influencing tumor organotropism. ②The metabolic plasticity of cancer cells dictates how much they can adapt to the redox milieu of the secondary lesion, and there exists a positive relationship between the metabolic plasticity and cancer stemness. In other words, the higher metabolic plasticity that cancer cells possess, the more easily they can survive the metastatic process and colonize in the new organ, and the higher cancer stem cell percentage that the metastasized cell cluster contains. ③ During metastasis, cancer cells are subjected to the selection pressure that is imposed by the oxidative stress and fated to happen. Cancer cells capable of increasing catabolism to satisfy the constant high demand of energy supply for traveling and reducing intracelluar ROS generation to counteract with the oxidative stress eventually survive. ④Thus, cancer cells during metastasis are vulnerable to oxidative-stress induced cell death. Yet, therapeutics solely imposing external oxidative stress can only enrich the metastatic cluster with cancer stem cells, approaches capable of inducing intracellular ROS generation can eventually sabotage the anti-oxidant machinery of cancer cells and trigger cell death. In this regard, cold atmospheric plasma (CAP) could be considered as one promising intervention strategy to meet this goal

Dai et al. Molecular Biomedicine (2025) 6:20 Page 21 of 30

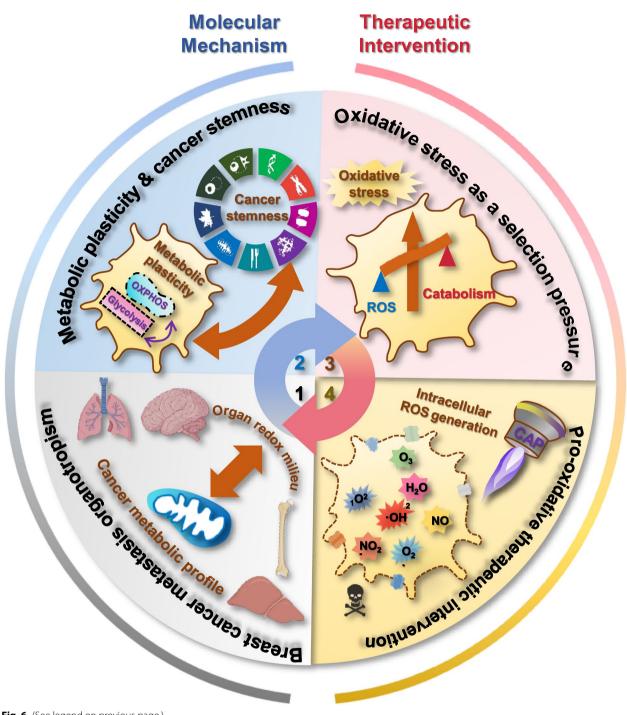


Fig. 6 (See legend on previous page.)

producer on light ignite [321, 322]. These offer a plenty of opportunities for cross-disciplinary therapeutic design that may lead one research direction.

Future endeavors may also be devoted to explore therapeutic regimes beyond CAP that are able to take actions via maintaining cellular redox homeostasis, which may open an innovative paradigm for cancer treatment.

Acknowledgements

Not available.

Authors' contributions

X.F. Dai conceptualized the idea and prepared the initial draft. M. Xi contributed in preparing the figures. J.T. Li agreed to the content of the paper. All authors have read and approved the final manuscript.

Funding

Not available.

Dai et al. Molecular Biomedicine (2025) 6:20 Page 22 of 30

Data availability

Not available.

Declarations

Ethics approval and consent to participate

Not available

Consent for publication

The authors are consent to the submission and publication of this paper.

Competing interests

The authors declare no conflict of interest.

Received: 13 September 2024 Revised: 7 March 2025 Accepted: 14 March 2025

Published online: 07 April 2025

References

- Dai X, Shen L, Zhang J. Cold atmospheric plasma: redox homeostasis to treat cancers? Trends Biotechnol. 2023;41(1):15–8. https://doi.org/ 10.1016/j.tibtech.2022.07.007.
- Sanmamed MF, Chen L. A paradigm shift in cancer immunotherapy: from enhancement to normalization. Cell. 2018;175(2):313–26. https://doi.org/10.1016/j.cell.2018.09.035.
- Siegel RL, Giaquinto AN, Jemal A. Cancer statistics, 2024. CA Cancer J Clin. 2024;74(1):12–49. https://doi.org/10.3322/caac.21820.
- Gao Y, Bado I, Wang H, Zhang W, Rosen JM, Zhang XH. Metastasis organotropism: redefining the congenial soil. Dev Cell. 2019;49(3):375–91. https://doi.org/10.1016/j.devcel.2019.04.012.
- Giese A, Loo MA, Tran N, Haskett D, Coons SW, Berens ME. Dichotomy of astrocytoma migration and proliferation. Int J Cancer. 1996;67(2):275–82. https://doi.org/10.1002/(SICI)1097-0215(19960717)67:293c275::AID-IJC20%3e3.0.CO;2-9.
- Hecht I, Natan S, Zaritsky A, Levine H, Tsarfaty I, Ben-Jacob E. The motility-proliferation-metabolism interplay during metastatic invasion. Sci Rep. 2015;5: 13538. https://doi.org/10.1038/srep13538.
- Reid SE, Kay EJ, Neilson LJ, Henze AT, Serneels J, McGhee EJ, et al. Tumor matrix stiffness promotes metastatic cancer cell interaction with the endothelium. EMBO J. 2017;36(16):2373–2389. https://doi. org/10.15252/embj.201694912.
- Wang C, Xu K, Wang R, Han X, Tang J, Guan X. Heterogeneity of BCSCs contributes to the metastatic organotropism of breast cancer. J Exp Clin Cancer Res. 2021;40(1):370. https://doi.org/10.1186/ s13046-021-02164-6.
- Murthy SRK, Cheng XQ, Zhuang TS, Ly L, Jones O, Basadonna G, et al. BCL2A1 regulates Canady Helios Cold Plasma-induced cell death in triple-negative breast cancer. Sci Rep. 2022;12(1):ARTN 403810.1038/ s41598-022-07027-4.
- Hu H, Liu YH, Xu L, Zhao JX, Duan XN, Ye JM, et al. Clinicopathological classification and individualized treatment of breast cancer. Chin Med J. 2013;126(20):3921–5. https://doi.org/10.3760/cma.j.issn.0366-6999.20123368.
- Dai X, Li T, Bai Z, Yang Y, Liu X, Zhan J, et al. Breast cancer intrinsic subtype classification, clinical use and future trends. Am J Cancer Res. 2015;5(10):2929–43.
- Ma W, Sun J, Xu J, Luo Z, Diao D, Zhang Z, et al. Sensitizing triple negative breast cancer to tamoxifen chemotherapy via a redox-responsive vorinostat-containing polymeric prodrug nanocarrier. Theranostics. 2020;10(6):2463–78. https://doi.org/10.7150/thno.38973.
- Cameron D, Piccart-Gebhart MJ, Gelber RD, Procter M, Goldhirsch A, de Azambuja E, et al. 11 years' follow-up of trastuzumab after adjuvant chemotherapy in HER2-positive early breast cancer: final analysis of the HERceptin Adjuvant (HERA) trial. Lancet. 2017;389(10075):1195–205. https://doi.org/10.1016/S0140-6736(16)32616-2.
- Bartmann C, Wischnewsky M, Stüber T, Stein R, Krockenberger M, Häusler S, et al. Pattern of metastatic spread and subcategories of breast

- cancer. Arch Gynecol Obstet. 2016;295(1):211–23. https://doi.org/10. 1007/s00404-016-4225-4.
- Kimbung S, Johansson I, Danielsson A, Veerla S, Egyhazi Brage S, Frostvik Stolt M, et al. Transcriptional profiling of breast cancer metastases identifies liver metastasis-selective genes associated with adverse outcome in luminal a primary breast cancer. Clin Cancer Res. 2016;22(1):146–57. https://doi.org/10.1158/1078-0432.CCR-15-0487.
- Kennecke H, Yerushalmi R, Woods R, Cheang MCU, Voduc D, Speers CH, et al. Metastatic behavior of breast cancer subtypes. J Clin Oncol. 2010;28(20):3271–7. https://doi.org/10.1200/jco.2009.25.9820.
- Pourjamal N, Yazdi N, Halme A, Joncour VL, Laakkonen P, Saharinen P, et al. Comparison of trastuzumab emtansine, trastuzumab deruxtecan, and disitamab vedotin in a multiresistant HER2-positive breast cancer lung metastasis model. Clin Exp Metastasis. 2024;41(2):91–102. https:// doi.org/10.1007/s10585-024-10278-2.
- Dang MN, Suri S, Li K, Casas CG, Stigliano G, Riley RS, et al. Antibody and siRNA Nanocarriers to Suppress Wnt Signaling, Tumor Growth, and Lung Metastasis in Triple-Negative Breast Cancer. Adv Ther (Weinh). 2024;7(6). https://doi.org/10.1002/adtp.202300426.
- Lin Y, Huang Y, Zheng Y, Chen W, Zhang Y, Yang Y, et al. Taurine inhibits lung metastasis in triple-negative breast cancer by modulating macrophage polarization through PTEN-PI3K/Akt/mTOR pathway. J Immunother. 2024. https://doi.org/10.1097/CJI.00000000000000518.
- 20. Paget S. The distribution of secondary growths in cancer of the breast. 1889. Cancer Metastasis Rev. 1989;8(2):98–101.
- 21. Ewing J. Neoplastic diseases. 6th ed. Philadelphia: W.B. Saunders; 1928.
- Fidler IJ. The pathogenesis of cancer metastasis: the "seed and soil" hypothesis revisited. Nat Rev Cancer. 2003;3(6):453–8. https://doi.org/ 10.1038/nrc1098.
- 23. Hart IR, Fidler IJ. Role of organ selectivity in the determination of metastatic patterns of B16 melanoma. Cancer Res. 1980;40(7):2281–7.
- Weiss L. Metastasis of cancer: a conceptual history from antiquity to the 1990s. Cancer Metastasis Rev. 2000;19(3–4):I–XI 193–383.
- Sugarbaker EV. Cancer metastasis: a product of tumor-host interactions. Curr Probl Cancer. 1979;3(7):1–59. https://doi.org/10.1016/s0147-0272(79)80008-2.
- Sousa B, Ribeiro AS, Paredes J. Heterogeneity and plasticity of breast cancer stem cells. Adv Exp Med Biol. 2019;1139:83–103. https://doi.org/ 10.1007/978-3-030-14366-4_5.
- Fidler IJ. Critical factors in the biology of human cancer metastasis: twenty-eighth G.H.A. Clowes memorial award lecture. Cancer Res. 1990;50(19):6130–8.
- 28. Fidler IJ, Talmadge JE. Evidence that intravenously derived murine pulmonary melanoma metastases can originate from the expansion of a single tumor cell. Cancer Res. 1986;46(10):5167–71.
- Xu D, Tang M. Advances in the study of biomarkers related to bone metastasis in breast cancer. Br J Radiol. 2023;96(1150): 20230117. https://doi.org/10.1259/bjr.20230117.
- Pang LL, Gan C, Xu J, Jia YX, Chai JY, Huang RZ, et al. Bone metastasis of breast cancer: molecular mechanisms and therapeutic strategies. Cancers. 2022;14(23):ARTN 5727. https://doi.org/10.3390/cancers142 35727.
- Fornetti J, Welm AL, Stewart SA. Understanding the bone in cancer metastasis. J Bone Miner Res. 2018;33(12):2099–113. https://doi.org/10. 1002/jbmr.3618.
- Inic Z, Zegarac M, Inic M, Markovic I, Kozomara Z, Djurisic I, et al. Difference between Luminal A and Luminal B subtypes according to Ki-67, tumor size, and progesterone receptor negativity providing prognostic information. Clin Med Insights Oncol. 2014;8:107–11. https://doi.org/10.4137/CMO.S18006.
- Dai X, Xiang L, Li T, Bai Z. Cancer hallmarks, biomarkers and breast cancer molecular subtypes. J Cancer. 2016;7(10):1281–94. https://doi. org/10.7150/jca.13141.
- Dai X, Cheng H, Bai Z, Li J. Breast cancer cell line classification and its relevance with breast tumor subtyping. J Cancer. 2017;8(16):3131–41. https://doi.org/10.7150/jca.18457.
- Wu Q, Li JJ, Zhu S, Wu J, Chen C, Liu Q, et al. Breast cancer subtypes predict the preferential site of distant metastases: a SEER based study. Oncotarget. 2017;8(17):27990–27996. https://doi.org/10.18632/oncot arget.15856.

- Yan C-Y, Zhao M-L, Wei Y-N, Zhao X-H. Mechanisms of drug resistance in breast cancer liver metastases: dilemmas and opportunities. Mol Ther -Oncolytics. 2023;28:212–29. https://doi.org/10.1016/j.omto.2023.02.001.
- Eroles P, Bosch A, Perez-Fidalgo JA, Lluch A. Molecular biology in breast cancer: intrinsic subtypes and signaling pathways. Cancer Treat Rev. 2012;38(6):698–707. https://doi.org/10.1016/j.ctrv.2011.11.005.
- Lah TT, Novak M, Breznik B. Brain malignancies: glioblastoma and brain metastases. Semin Cancer Biol. 2020;60:262–73. https://doi.org/10. 1016/j.semcancer.2019.10.010.
- Custódio-Santos T, Videira M, Brito MA. Brain metastasization of breast cancer. BBA-Rev Cancer. 2017;1868(1):132–47. https://doi.org/10.1016/j. bbcan.2017.03.004.
- Nagy V, Todor N, Fekete Z, Suteu P. Survival and quality of life after whole brain radiotherapy with 3D conformal boost in the treatment of brain metastases. Med Pharm Rep. 2019;92(1):43–51. https://doi.org/10. 15386/cimed-1040.
- Galloni C, Egnuni T, Zahed Mohajerani S, Ye J, Mittnacht S, Speirs V, et al. Brain endothelial cells promote breast cancer cell extravasation to the brain via EGFR-DOCK4-RAC1 signalling. Commun Biol. 2024;7(1):602. https://doi.org/10.1038/s42003-024-06200-x.
- Yousefi M, Nosrati R, Salmaninejad A, Dehghani S, Shahryari A, Saberi A. Organ-specific metastasis of breast cancer: molecular and cellular mechanisms underlying lung metastasis. Cell Oncol. 2018. https://doi. org/10.1007/s13402-018-0376-6.
- Shim J, Brindle L, Simon M, George S. A systematic review of symptomatic diagnosis of lung cancer. Fam Pract. 2013;31(2):137–48. https://doi.org/10.1093/fampra/cmt076.
- Antonarelli G, Corti C, Tarantino P, Salimbeni BT, Zagami P, Marra A, et al. Management of patients with HER2-positive metastatic breast cancer after trastuzumab deruxtecan failure. ESMO Open. 2023;8(4): 101608. https://doi.org/10.1016/j.esmoop.2023.101608.
- Modi S, Saura C, Yamashita T, Park YH, Kim S-B, Tamura K, et al. Trastuzumab deruxtecan in previously treated HER2-positive breast cancer. N Engl J Med. 2020;382(7):610–21. https://doi.org/10.1056/NEJMoa1914 510.
- Font-Clos F, Zapperi S, La Porta CAM. Classification of triple negative breast cancer by epithelial mesenchymal transition and the tumor immune microenvironment. Sci Rep. 2022. https://doi.org/10.1038/ s41598-022-13428-2.
- Liu Q, Zhang H, Jiang X, Qian C, Liu Z, Luo D. Factors involved in cancer metastasis: a better understanding to "seed and soil" hypothesis. Mol Cancer. 2017;16(1):176. https://doi.org/10.1186/s12943-017-0742-4.
- Schild T, Low V, Blenis J, Gomes AP. Unique metabolic adaptations dictate distal organ-specific metastatic colonization. Cancer Cell. 2018;33(3):347–54. https://doi.org/10.1016/j.ccell.2018.02.001.
- DeBerardinis RJ, Chandel NS. Fundamentals of cancer metabolism. Sci Adv. 2016;2(5):e1600200. https://doi.org/10.1126/sciadv.1600200.
- Bi J, Wu S, Zhang W, Mischel PS. Targeting cancer's metabolic codependencies: a landscape shaped by genotype and tissue context. BBA-Rev Cancer. 2018;1870(1):76–87. https://doi.org/10.1016/j.bbcan. 2018.05.002.
- Kim J, DeBerardinis RJ. Mechanisms and implications of metabolic heterogeneity in cancer. Cell Metab. 2019;30(3):434–46. https://doi.org/ 10.1016/j.cmet.2019.08.013.
- Dittmer J. Breast cancer stem cells: features, key drivers and treatment options. Semin Cancer Biol. 2018;53:59–74. https://doi.org/10.1016/j. semcancer.2018.07.007.
- Zanotelli MR, Goldblatt ZE, Miller JP, Bordeleau F, Li J, VanderBurgh JA, et al. Regulation of ATP utilization during metastatic cell migration by collagen architecture. Mol Biol Cell. 2018;29(1):1–9. https://doi.org/10. 1091/mbc.E17-01-0041.
- Stevenson RP, Veltman D, Machesky LM. Actin-bundling proteins in cancer progression at a glance. J Cell Sci. 2012;125(Pt 5):1073–9. https://doi.org/10.1242/jcs.093799.
- Lambert AW, Pattabiraman DR, Weinberg RA. Emerging biological principles of metastasis. Cell. 2017;168(4):670–91. https://doi.org/10.1016/j.cell.2016.11.037.
- Riggi N, Aguet M, Stamenkovic I. Cancer metastasis: a reappraisal of its underlying mechanisms and their relevance to treatment. Annu Rev Pathol. 2018;13:117–40. https://doi.org/10.1146/annur ev-pathol-020117-044127.

- Oskarsson T, Batlle E, Massague J. Metastatic stem cells: sources, niches, and vital pathways. Cell Stem Cell. 2014;14(3):306–21. https://doi.org/ 10.1016/j.stem.2014.02.002.
- Kristiansen G, Sammar M, Altevogt P. Tumour biological aspects of CD24, a mucin-like adhesion molecule. J Mol Histol. 2004;35(3):255–62. https://doi.org/10.1023/b:hijo.0000032357.16261.c5.
- Senbanjo LT, Chellaiah MA. CD44: a multifunctional cell surface adhesion receptor is a regulator of progression and metastasis of cancer cells. Front Cell Dev Biol. 2017;5: 18. https://doi.org/10.3389/fcell.2017.00018
- Tomita H, Tanaka K, Tanaka T, Hara A. Aldehyde dehydrogenase 1A1 in stem cells and cancer. Oncotarget. 2016;7(10):11018–32. https://doi. org/10.18632/oncotarget.6920.
- 61. Liu S, Cong Y, Wang D, Sun Y, Deng L, Liu Y, et al. Breast cancer stem cells transition between epithelial and mesenchymal states reflective of their normal counterparts. Stem Cell Rep. 2014;2(1):78–91. https://doi.org/10.1016/j.stemcr.2013.11.009.
- Ginestier C, Hur MH, Charafe-Jauffret E, Monville F, Dutcher J, Brown M, et al. ALDH1 is a marker of normal and malignant human mammary stem cells and a predictor of poor clinical outcome. Cell Stem Cell. 2007;1(5):555–67. https://doi.org/10.1016/j.stem.2007.08.014.
- Al-Hajj M, Wicha MS, Benito-Hernandez A, Morrison SJ, Clarke MF. Prospective identification of tumorigenic breast cancer cells. Proc Natl Acad Sci U S A. 2003;100(7):3983–8. https://doi.org/10.1073/pnas.05302 91100.
- 64. Bernstein BW, Bamburg JR. Actin-ATP hydrolysis is a major energy drain for neurons. J Neurosci. 2003;23(1):1–6. https://doi.org/10.1523/JNEUR OSCI.23-01-00002.2003.
- Daniel JL, Molish IR, Robkin L, Holmsen H. Nucleotide exchange between cytosolic ATP and F-actin-bound ADP may be a major energy-utilizing process in unstimulated platelets. Eur J Biochem. 1986;156(3):677–84. https://doi.org/10.1111/j.1432-1033.1986.tb096 31 x.
- Colacino JA, Azizi E, Brooks MD, Harouaka R, Fouladdel S, McDermott SP, et al. Heterogeneity of human breast stem and progenitor cells as revealed by transcriptional profiling. Stem Cell Rep. 2018;10(5):1596– 609. https://doi.org/10.1016/j.stemcr.2018.03.001.
- Mori Y, Yamawaki K, Ishiguro T, Yoshihara K, Ueda H, Sato A, et al. ALDH-dependent glycolytic activation mediates stemness and paclitaxel resistance in patient-derived spheroid models of uterine endometrial cancer. Stem Cell Rep. 2019;13(4):730–46. https://doi.org/10.1016/j.stemcr.2019.08.015.
- Lv X, Wang Y, Song Y, Pang X, Li H. Association between ALDH1+/ CD133+ stem-like cells and tumor angiogenesis in invasive ductal breast carcinoma. Oncol Lett. 2016;11(3):1750–6. https://doi.org/10. 3892/ol.2016.4145.
- Xu H, Tian Y, Yuan X, Liu Y, Wu H, Liu Q, et al. Enrichment of CD44 in basal-type breast cancer correlates with EMT, cancer stem cell gene profile, and prognosis. Onco Targets Ther. 2016;9:431–44. https://doi. org/10.2147/OTT.S97192.
- Tsukabe M, Shimazu K, Morimoto K, Naoi Y, Kagara N, Shimoda M, et al. Clinicopathological analysis of breast ductal carcinoma in situ with ALDH1-positive cancer stem cells. Oncology. 2013;85(4):248–56. https://doi.org/10.1159/000355476.
- Park SY, Lee HE, Li H, Shipitsin M, Gelman R, Polyak K. Heterogeneity for stem cell-related markers according to tumor subtype and histologic stage in breast cancer. Clin Cancer Res. 2010;16(3):876–87. https://doi. org/10.1158/1078-0432.CCR-09-1532.
- Ricardo S, Vieira AF, Gerhard R, Leitao D, Pinto R, Cameselle-Teijeiro
 JF, et al. Breast cancer stem cell markers CD44, CD24 and ALDH1:
 expression distribution within intrinsic molecular subtype. J Clin Pathol.
 2011;64(11):937–46. https://doi.org/10.1136/jcp.2011.090456.
- Hung PF, Hong TM, Chang CC, Hung CL, Hsu YL, Chang YL, et al. Hypoxia-induced Slug SUMOylation enhances lung cancer metastasis. J Exp Clin Cancer Res. 2019;38(1):5. https://doi.org/10.1186/s13046-018-0996-8.
- Zhang Z, Li TE, Chen M, Xu D, Zhu Y, Hu BY, et al. MFN1-dependent alteration of mitochondrial dynamics drives hepatocellular carcinoma metastasis by glucose metabolic reprogramming. Br J Cancer. 2020;122(2):209–20. https://doi.org/10.1038/s41416-019-0658-4.

- Thomas RJ, Guise TA, Yin JJ, Elliott J, Horwood NJ, Martin TJ, et al. Breast cancer cells interact with osteoblasts to support osteoclast formation. Endocrinology. 1999;140(10):4451–8. https://doi.org/10.1210/endo.140. 10.7037.
- 76. Casimiro S, Guise TA, Chirgwin J. The critical role of the bone microenvironment in cancer metastases. Mol Cell Endocrinol. 2009;310(1–2):71–81. https://doi.org/10.1016/j.mce.2009.07.004.
- Zhang W, Bado I, Wang H, Lo HC, Zhang XH. Bone metastasis: find your niche and fit in. Trends Cancer. 2019;5(2):95–110. https://doi.org/10. 1016/j.trecan.2018.12.004.
- Spencer JA, Ferraro F, Roussakis E, Klein A, Wu J, Runnels JM, et al. Direct measurement of local oxygen concentration in the bone marrow of live animals. Nature. 2014;508(7495):269–73. https://doi.org/10.1038/natur e13034.
- Nakazawa MS, Keith B, Simon MC. Oxygen availability and metabolic adaptations. Nat Rev Cancer. 2016;16(10):663–73. https://doi.org/10. 1038/nrc.2016.84.
- Hiraga T, Kizaka-Kondoh S, Hirota K, Hiraoka M, Yoneda T. Hypoxia and hypoxia-inducible factor-1 expression enhance osteolytic bone metastases of breast cancer. Cancer Res. 2007;67(9):4157–63. https://doi.org/ 10.1158/0008-5472.CAN-06-2355.
- Yoneda T, Hiasa M, Nagata Y, Okui T, White F. Contribution of acidic extracellular microenvironment of cancer-colonized bone to bone pain. Biochim Biophys Acta. 2015;1848(10 Pt B):2677–84. https://doi.org/10. 1016/j.bbamem.2015.02.004.
- Lemma S, Di Pompo G, Porporato PE, Sboarina M, Russell S, Gillies RJ, et al. MDA-MB-231 breast cancer cells fuel osteoclast metabolism and activity: A new rationale for the pathogenesis of osteolytic bone metastases. Biochim Biophys Acta Mol Basis Dis. 2017;1863(12):3254–64. https://doi.org/10.1016/j.bbadis.2017.08.030.
- 83. Long W, Wu J, Shen G, Zhang H, Liu H, Xu Y, et al. Estrogen-related receptor participates in regulating glycolysis and influences embryonic development in silkworm Bombyx mori. Insect Mol Biol. 2020;29(2):160–9. https://doi.org/10.1111/imb.12619.
- Zhang L, Zhang S, Yao J, Lowery FJ, Zhang Q, Huang WC, et al. Microenvironment-induced PTEN loss by exosomal microRNA primes brain metastasis outgrowth. Nature. 2015;527(7576):100–4. https://doi. org/10.1038/nature15376.
- Zeng Z, Li Y, Pan Y, Lan X, Song F, Sun J, et al. Cancer-derived exosomal miR-25-3p promotes pre-metastatic niche formation by inducing vascular permeability and angiogenesis. Nat Commun. 2018;9(1):5395. https://doi.org/10.1038/s41467-018-07810-w.
- Shi Z, Mirza M, Wang B, Kennedy MA, Weber GF. Osteopontin-a alters glucose homeostasis in anchorage-independent breast cancer cells. Cancer Lett. 2014;344(1):47–53. https://doi.org/10.1016/j.canlet.2013.10. 008.
- 87. Pollari S, Kakonen SM, Edgren H, Wolf M, Kohonen P, Sara H, et al. Enhanced serine production by bone metastatic breast cancer cells stimulates osteoclastogenesis. Breast Cancer Res Treat. 2011;125(2):421–30. https://doi.org/10.1007/s10549-010-0848-5.
- Ogawa T, Ishida-Kitagawa N, Tanaka A, Matsumoto T, Hirouchi T, Akimaru M, et al. A novel role of L-serine (L-Ser) for the expression of nuclear factor of activated T cells (NFAT)2 in receptor activator of nuclear factor kappa B ligand (RANKL)-induced osteoclastogenesis in vitro. J Bone Miner Metab. 2006;24(5):373–9. https://doi.org/10.1007/ s00774-006-0705-0.
- 89. Li AM, He B, Karagiannis D, Li Y, Jiang H, Srinivasan P, et al. Serine starvation silences estrogen receptor signaling through histone hypoacetylation. Proc Natl Acad Sci U S A. 2023;120(38): e2302489120. https://doi.org/10.1073/pnas.2302489120.
- O'Grady S, Morgan MP. Microcalcifications in breast cancer: from pathophysiology to diagnosis and prognosis. Biochim Biophys Acta Rev Cancer. 2018;1869(2):310–20. https://doi.org/10.1016/j.bbcan.2018.04.
- 91. Cooke MM, McCarthy GM, Sallis JD, Morgan MP. Phosphocitrate inhibits calcium hydroxyapatite induced mitogenesis and upregulation of matrix metalloproteinase-1, interleukin-1beta and cyclooxygenase-2 mRNA in human breast cancer cell lines. Breast Cancer Res Treat. 2003;79(2):253–63. https://doi.org/10.1023/a:1023908307108.
- 92. Nie X, Jin H, Wen G, Xu J, An J, Liu X, et al. Estrogen regulates duodenal calcium absorption through differential role of estrogen receptor on

- calcium transport proteins. Dig Dis Sci. 2020;65(12):3502–13. https://doi.org/10.1007/s10620-020-06076-x.
- 93. Tran QK. Reciprocality between estrogen biology and calcium signaling in the cardiovascular system. Front Endocrinol (Lausanne). 2020;11: 568203. https://doi.org/10.3389/fendo.2020.568203.
- 94. Liao J, Schneider A, Datta NS, McCauley LK. Extracellular calcium as a candidate mediator of prostate cancer skeletal metastasis. Cancer Res. 2006;66(18):9065–73. https://doi.org/10.1158/0008-5472.CAN-06-0317.
- 95. Saidak Z, Boudot C, Abdoune R, Petit L, Brazier M, Mentaverri R, et al. Extracellular calcium promotes the migration of breast cancer cells through the activation of the calcium sensing receptor. Exp Cell Res. 2009;315(12):2072–80. https://doi.org/10.1016/j.yexcr.2009.03.003.
- Joeckel E, Haber T, Prawitt D, Junker K, Hampel C, Thuroff JW, et al. High calcium concentration in bones promotes bone metastasis in renal cell carcinomas expressing calcium-sensing receptor. Mol Cancer. 2014;13: 42. https://doi.org/10.1186/1476-4598-13-42.
- Brodt P. Role of the microenvironment in liver metastasis: from pre- to prometastatic niches. Clin Cancer Res. 2016;22(24):5971–82. https://doi. org/10.1158/1078-0432.Ccr-16-0460.
- Van den Eynden GG, Majeed AW, Illemann M, Vermeulen PB, Bird NC, Høyer-Hansen G, et al. The multifaceted role of the microenvironment in liver metastasis: biology and clinical implications. Cancer Res. 2013;73(7):2031–43. https://doi.org/10.1158/0008-5472.Can-12-3931.
- Kang N, Gores GJ, Shah VH. Hepatic stellate cells: partners in crime for liver metastases? Hepatology. 2011;54(2):707–13. https://doi.org/10. 1002/hep.24384.
- Nielsen SR, Quaranta V, Linford A, Emeagi P, Rainer C, Santos A, et al. Macrophage-secreted granulin supports pancreatic cancer metastasis by inducing liver fibrosis. Nat Cell Biol. 2016;18(5):549–60. https://doi. org/10.1038/ncb3340.
- Dupuy F, Tabariès S, Andrzejewski S, Dong Z, Blagih J, Annis MG, et al. PDK1-dependent metabolic reprogramming dictates metastatic potential in breast cancer. Cell Metab. 2015;22(4):577–89. https://doi.org/10.1016/j.cmet.2015.08.007.
- McLean ME, MacLean MR, Cahill HF, Arun RP, Walker OL, Wasson MD, et al. The expanding role of cancer stem cell marker ALDH1A3 in cancer and beyond. Cancers (Basel). 2023;15(2). https://doi.org/10.3390/cance rs15020492.
- Newman JC, Verdin E. Ketone bodies as signaling metabolites. Trends Endocrinol Metab. 2014;25(1):42–52. https://doi.org/10.1016/j.tem. 2013.09.002
- Hwang CY, Choe W, Yoon KS, Ha J, Kim SS, Yeo EJ, et al. Molecular mechanisms for ketone body metabolism, signaling functions, and therapeutic potential in cancer. Nutrients. 2022;14(22). https://doi.org/ 10.3390/nu14224932.
- Nandi I, Ji L, Smith HW, Avizonis D, Papavasiliou V, Lavoie C, et al. Targeting fatty acid oxidation enhances response to HER2-targeted therapy. Nat Commun. 2024;15(1):6587. https://doi.org/10.1038/ s41467-024-50998-3.
- Kumar A, Sharma P, Sarin SK. Hepatic venous pressure gradient measurement: time to learn! Indian J Gastroenterol. 2008;27(2):74–80.
- MacPhee PJ, Schmidt EE, Groom AC. Intermittence of blood flow in liver sinusoids, studied by high-resolution in vivo microscopy. Am J Physiol. 1995;269(5 Pt 1):G692–8. https://doi.org/10.1152/ajpgi.1995.269.5.G692.
- Lang JE, Mosalpuria K, Cristofanilli M, Krishnamurthy S, Reuben J, Singh B, et al. HER2 status predicts the presence of circulating tumor cells in patients with operable breast cancer. Breast Cancer Res Treat. 2009;113(3):501–7. https://doi.org/10.1007/s10549-008-9951-2.
- Termini J, Neman J, Jandial R. Role of the neural niche in brain metastatic cancer. Cancer Res. 2014;74(15):4011–5. https://doi.org/10.1158/ 0008-5472 Can-14-1226.
- Valiente M, Ahluwalia MS, Boire A, Brastianos PK, Goldberg SB, Lee EQ, et al. The evolving landscape of brain metastasis. Trends Cancer. 2018;4(3):176–96. https://doi.org/10.1016/j.trecan.2018.01.003.
- Chen Q, Boire A, Jin X, Valiente M, Er EE, Lopez-Soto A, et al. Carcinomaastrocyte gap junctions promote brain metastasis by cGAMP transfer. Nature. 2016;533(7604):493–8. https://doi.org/10.1038/nature18268.
- 112. Danbolt NC. Glutamate uptake. Prog Neurobiol. 2001;65(1):1–105. https://doi.org/10.1016/s0301-0082(00)00067-8.
- 113. Zeng Q, Michael IP, Zhang P, Saghafinia S, Knott G, Jiao W, et al. Synaptic proximity enables NMDAR signalling to promote brain

- metastasis. Nature. 2019;573(7775):526–31. https://doi.org/10.1038/s41586-019-1576-6.
- Taylor KR, Monje M. Neuron–oligodendroglial interactions in health and malignant disease. Nat Rev Neurosci. 2023;24(12):733–46. https://doi. org/10.1038/s41583-023-00744-3.
- Pukrop T, Dehghani F, Chuang HN, Lohaus R, Bayanga K, Heermann S, et al. Microglia promote colonization of brain tissue by breast cancer cells in a Wnt-dependent way. Glia. 2010;58(12):1477–89. https://doi. org/10.1002/glia.21022.
- Boire A, Brastianos PK, Garzia L, Valiente M. Brain metastasis. Nat Rev Cancer. 2020;20(1):4–11. https://doi.org/10.1038/s41568-019-0220-y.
- Chen J, Lee HJ, Wu X, Huo L, Kim SJ, Xu L, et al. Gain of glucose-independent growth upon metastasis of breast cancer cells to the brain. Cancer Res. 2015;75(3):554–65. https://doi.org/10.1158/0008-5472. Can-14-2268
- Mashimo T, Pichumani K, Vemireddy V, Hatanpaa KJ, Singh DK, Sirasanagandla S, et al. Acetate is a bioenergetic substrate for human glioblastoma and brain metastases. Cell. 2014;159(7):1603–14. https://doi.org/10.1016/j.cell.2014.11.025.
- Maher EA, Marin-Valencia I, Bachoo RM, Mashimo T, Raisanen J, Hatanpaa KJ, et al. Metabolism of [U-13 C]glucose in human brain tumors in vivo. NMR Biomed. 2012;25(11):1234–44. https://doi.org/10.1002/ nbm.2794
- 120. Cordero A, Kanojia D, Miska J, Panek WK, Xiao A, Han Y, et al. FABP7 is a key metabolic regulator in HER2+ breast cancer brain metastasis. Oncogene. 2019;38(37):6445–60. https://doi.org/10.1038/s41388-019-0893-4.
- Chen El, Hewel J, Krueger JS, Tiraby C, Weber MR, Kralli A, et al. Adaptation of energy metabolism in breast cancer brain metastases. Cancer Res. 2007;67(4):1472–86. https://doi.org/10.1158/0008-5472. CAN-06-3137.
- 122. Neman J, Termini J, Wilczynski S, Vaidehi N, Choy C, Kowolik CM, et al. Human breast cancer metastases to the brain display GABAergic properties in the neural niche. Proc Natl Acad Sci U S A. 2014;111(3):984–9. https://doi.org/10.1073/pnas.1322098111.
- 123. Weidle UH, Birzele F, Kollmorgen G, Rüger R. Dissection of the process of brain metastasis reveals targets and mechanisms for molecular-based intervention. Cancer Genom Proteom. 2016;13(4):245–58.
- Sevenich L, Bowman RL, Mason SD, Quail DF, Rapaport F, Elie BT, et al. Analysis of tumour- and stroma-supplied proteolytic networks reveals a brain-metastasis-promoting role for cathepsin S. Nat Cell Biol. 2014;16(9):876–88. https://doi.org/10.1038/ncb3011.
- Tominaga N, Kosaka N, Ono M, Katsuda T, Yoshioka Y, Tamura K, et al. Brain metastatic cancer cells release microRNA-181c-containing extracellular vesicles capable of destructing blood-brain barrier. Nat Commun. 2015;6:6716. https://doi.org/10.1038/ncomms7716.
- 126. Altorki NK, Markowitz GJ, Gao D, Port JL, Saxena A, Stiles B, et al. The lung microenvironment: an important regulator of tumour growth and metastasis. Nat Rev Cancer. 2019;19(1):9–31. https://doi.org/10.1038/s41568-018-0081-9.
- Guo F, Wang Y, Liu J, Mok SC, Xue F, Zhang W. CXCL12/CXCR4: a symbiotic bridge linking cancer cells and their stromal neighbors in oncogenic communication networks. Oncogene. 2016;35(7):816–26. https://doi.org/10.1038/onc.2015.139.
- Forster R, Davalos-Misslitz AC, Rot A. CCR7 and its ligands: balancing immunity and tolerance. Nat Rev Immunol. 2008;8(5):362–71. https://doi.org/10.1038/nri2297.
- Müller A, Homey B, Soto H, Ge N, Catron D, Buchanan ME, et al. Involvement of chemokine receptors in breast cancer metastasis. Nature. 2001;410(6824):50–6. https://doi.org/10.1038/35065016.
- Nosaka T, Baba T, Tanabe Y, Sasaki S, Nishimura T, Imamura Y, et al. Alveolar macrophages drive hepatocellular carcinoma lung metastasis by generating leukotriene B(4). J Immunol. 2018;200(5):1839–52. https://doi.org/10.4049/jimmunol.1700544.
- Sharma SK, Chintala NK, Vadrevu SK, Patel J, Karbowniczek M, Markiewski MM. Pulmonary alveolar macrophages contribute to the premetastatic niche by suppressing antitumor T cell responses in the lungs. J Immunol. 2015;194(11):5529–38. https://doi.org/10.4049/jimmunol.1403215.
- 132. LeBleu VS, O'Connell JT, Gonzalez Herrera KN, Wikman H, Pantel K, Haigis MC, et al. PGC-1α mediates mitochondrial biogenesis and oxidative

- phosphorylation in cancer cells to promote metastasis. Nat Cell Biol. 2014;16(10):992–1003. https://doi.org/10.1038/ncb3039. 1-15.
- Schafer ZT, Grassian AR, Song L, Jiang Z, Gerhart-Hines Z, Irie HY, et al. Antioxidant and oncogene rescue of metabolic defects caused by loss of matrix attachment. Nature. 2009;461(7260):109–13. https://doi.org/ 10.1038/nature08268.
- Piskounova E, Agathocleous M, Murphy MM, Hu Z, Huddlestun SE, Zhao Z, et al. Oxidative stress inhibits distant metastasis by human melanoma cells. Nature. 2015;527(7577):186–91. https://doi.org/10.1038/nature.15776
- 135. Stresing V, Baltziskueta E, Rubio N, Blanco J, Arriba MC, Valls J, et al. Peroxiredoxin 2 specifically regulates the oxidative and metabolic stress response of human metastatic breast cancer cells in lungs. Oncogene. 2013;32(6):724–35. https://doi.org/10.1038/onc.2012.93.
- Shinde A, Wilmanski T, Chen H, Teegarden D, Wendt MK. Pyruvate carboxylase supports the pulmonary tropism of metastatic breast cancer. Breast Cancer Res. 2018;20(1):76. https://doi.org/10.1186/ s13058-018-1008-9.
- Hiratsuka S, Nakamura K, Iwai S, Murakami M, Itoh T, Kijima H, et al. MMP9 induction by vascular endothelial growth factor receptor-1 is involved in lung-specific metastasis. Cancer Cell. 2002;2(4):289–300. https://doi.org/10.1016/S1535-6108(02)00153-8.
- Ghajar CM, Peinado H, Mori H, Matei IR, Evason KJ, Brazier H, et al. The perivascular niche regulates breast tumour dormancy. Nat Cell Biol. 2013;15(7):807–17. https://doi.org/10.1038/ncb2767.
- Hanna RN, Cekic C, Sag D, Tacke R, Thomas GD, Nowyhed H, et al. Patrolling monocytes control tumor metastasis to the lung. Science. 2015;350(6263):985–90. https://doi.org/10.1126/science.aac9407.
- Guan R, Yuan L, Li J, Wang J, Li Z, Cai Z, et al. Bone morphogenetic protein 4 inhibits pulmonary fibrosis by modulating cellular senescence and mitophagy in lung fibroblasts. Eur Respir J. 2022;60(6):2102307. https://doi.org/10.1183/13993003.02307-2021.
- 141. Jiang Y, Chen Y, Fu J, Zhao R, Xu J, Liu Y. Bone morphogenetic protein 4 alleviates pulmonary fibrosis by regulating macrophages. Int Immunopharmacol. 2024;139: 112530. https://doi.org/10.1016/j.intimp.2024. 112530.
- 142. Song K-H, Park MS, Nandu TS, Gadad S, Kim S-C, Kim M-Y. GALNT14 promotes lung-specific breast cancer metastasis by modulating self-renewal and interaction with the lung microenvironment. Nat Commun. 2016;7(1): 13796. https://doi.org/10.1038/ncomms13796.
- 143. von Holst A. Tenascin C in stem cell niches: redundant, permissive or instructive? Cells Tissues Organs. 2008;188(1–2):170–7. https://doi.org/10.1159/000112848.
- 144. Orend G, Chiquet-Ehrismann R. Tenascin-C induced signaling in cancer. Cancer Lett. 2006;244(2):143–63. https://doi.org/10.1016/j.canlet.2006.
- 145. Oskarsson T, Acharyya S, Zhang XHF, Vanharanta S, Tavazoie SF, Morris PG, et al. Breast cancer cells produce tenascin C as a metastatic niche component to colonize the lungs. Nat Med. 2011;17(7):867–74. https://doi.org/10.1038/nm.2379.
- Hoshino A, Costa-Silva B, Shen TL, Rodrigues G, Hashimoto A, Tesic Mark M, et al. Tumour exosome integrins determine organotropic metastasis. Nature. 2015;527(7578):329–35. https://doi.org/10.1038/nature15756.
- Keklikoglou I, Cianciaruso C, Güç E, Squadrito ML, Spring LM, Tazzyman S, et al. Chemotherapy elicits pro-metastatic extracellular vesicles in breast cancer models. Nat Cell Biol. 2019;21(2):190–202. https://doi.org/ 10.1038/s41556-018-0256-3.
- 148. Zhu T, Zheng J, Zhuo W, Pan P, Li M, Zhang W, et al. ETV4 promotes breast cancer cell stemness by activating glycolysis and CXCR4-mediated sonic Hedgehog signaling. Cell Death Dis. 2021;7(1):126. https://doi.org/10.1038/s41420-021-00508-x.
- Granchi C, Bertini S, Macchia M, Minutolo F. Inhibitors of lactate dehydrogenase isoforms and their therapeutic potentials. Curr Med Chem. 2010;17(7):672–97. https://doi.org/10.2174/092986710790416263.
- Ma Q, Ye S, Liu H, Zhao Y, Mao Y, Zhang W. HMGA2 promotes cancer metastasis by regulating epithelial-mesenchymal transition. Front Oncol. 2024;14: 1320887. https://doi.org/10.3389/fonc.2024.1320887.
- Hussein MA, Valinezhad K, Adel E, Munirathinam G. MALAT-1 is a key regulator of epithelial-mesenchymal transition in cancer: a potential therapeutic target for metastasis. Cancers (Basel). 2024;16(1). https:// doi.org/10.3390/cancers16010234.

- Tasdogan A, Ubellacker JM, Morrison SJ. Redox regulation in cancer cells during metastasis. Cancer Discov. 2021;11(11):2682–92. https://doi. org/10.1158/2159-8290.CD-21-0558.
- 153. Husain A, Chiu YT, Sze KM, Ho DW, Tsui YM, Suarez EMS, et al. Ephrin-A3/EphA2 axis regulates cellular metabolic plasticity to enhance cancer stemness in hypoxic hepatocellular carcinoma. J Hepatol. 2022;77(2):383–96. https://doi.org/10.1016/j.jhep.2022.02.018.
- 154. Shakery A, Pourvali K, Ghorbani A, Fereidani SS, Zand H. Beta-hydroxy-butyrate promotes proliferation, migration and stemness in a subpopulation of 5FU treated SW480 cells: evidence for metabolic plasticity in colon cancer. Asian Pac J Cancer Prev. 2018;19(11):3287–94. https://doi.org/10.31557/APJCP.2018.19.11.3287.
- Alcala S, Sancho P, Martinelli P, Navarro D, Pedrero C, Martin-Hijano L, et al. ISG15 and ISGylation is required for pancreatic cancer stem cell mitophagy and metabolic plasticity. Nat Commun. 2020;11(1):2682. https://doi.org/10.1038/s41467-020-16395-2.
- 156. Tavares-Valente D, Cannone S, Greco MR, Carvalho TMA, Baltazar F, Queiros O, et al. Extracellular matrix collagen i differentially regulates the metabolic plasticity of pancreatic ductal adenocarcinoma parenchymal cell and cancer stem Cell. Cancers (Basel). 2023;15(15). https:// doi.org/10.3390/cancers15153868.
- Wiel C, Le Gal K, Ibrahim MX, Jahangir CA, Kashif M, Yao H, et al. BACH1 stabilization by antioxidants stimulates lung cancer metastasis. Cell. 2019;178(2):330–345 e22. https://doi.org/10.1016/j.cell.2019.06.005.
- Sayin VI, Ibrahim MX, Larsson E, Nilsson JA, Lindahl P, Bergo MO. Antioxidants accelerate lung cancer progression in mice. Sci Transl Med. 2014;6(221):221ra15. https://doi.org/10.1126/scitranslmed.3007653.
- Wang H, Liu X, Long M, Huang Y, Zhang L, Zhang R, et al. NRF2 activation by antioxidant antidiabetic agents accelerates tumor metastasis.
 Sci Transl Med. 2016;8(334):334ra51. https://doi.org/10.1126/scitranslmed.aad6095.
- Basnet H, Tian L, Ganesh K, Huang YH, Macalinao DG, Brogi E, et al. Flura-seq identifies organ-specific metabolic adaptations during early metastatic colonization. Elife. 2019;8:8. https://doi.org/10.7554/eLife. 43627.
- Dong C, Yuan T, Wu Y, Wang Y, Fan TW, Miriyala S, et al. Loss of FBP1 by Snail-mediated repression provides metabolic advantages in basal-like breast cancer. Cancer Cell. 2013;23(3):316–31. https://doi.org/10.1016/j. ccr.2013.01.022.
- Kamarajugadda S, Cai Q, Chen H, Nayak S, Zhu J, He M, et al. Manganese superoxide dismutase promotes anoikis resistance and tumor metastasis. Cell Death Dis. 2013;4(2): e504. https://doi.org/10.1038/cddis.2013.
- 163. Qu Y, Wang J, Ray PS, Guo H, Huang J, Shin-Sim M, et al. Thioredoxin-like 2 regulates human cancer cell growth and metastasis via redox homeostasis and NF-kappaB signaling. J Clin Invest. 2011;121(1):212–25. https://doi.org/10.1172/JCl43144.
- 164. Lu X, Bennet B, Mu E, Rabinowitz J, Kang Y. Metabolomic changes accompanying transformation and acquisition of metastatic potential in a syngeneic mouse mammary tumor model. J Biol Chem. 2010;285(13):9317–21. https://doi.org/10.1074/jbc.C110.104448.
- Chen L, Xing X, Zhang P, Chen L, Pei H. Homeostatic regulation of NAD(H) and NADP(H) in cells. Genes Dis. 2024;11(5): 101146. https://doi. org/10.1016/j.gendis.2023.101146.
- 166. Goodman M, Bostick RM, Kucuk O, Jones DP. Clinical trials of antioxidants as cancer prevention agents: past, present, and future. Free Radic Biol Med. 2011;51(5):1068–84. https://doi.org/10.1016/j.freeradbiomed. 2011.05.018.
- Chandel NS, Tuveson DA. The promise and perils of antioxidants for cancer patients. N Engl J Med. 2014;371(2):177–8. https://doi.org/10. 1056/NEJMcibr1405701.
- 168. Alpha-Tocopherol BCCPSG. The effect of vitamin E and beta carotene on the incidence of lung cancer and other cancers in male smokers. N Engl J Med. 1994;330(15):1029–35. https://doi.org/10.1056/NEJM1 99404143301501.
- 169. Ebbing M, Bonaa KH, Nygard O, Arnesen E, Ueland PM, Nordrehaug JE, et al. Cancer incidence and mortality after treatment with folic acid and vitamin B12. JAMA. 2009;302(19):2119–26. https://doi.org/10.1001/jama.2009.1622.
- 170. Shen L, Lv X, Li Y, Dai X. Spermidine antagonizes the anti-cancer effect of cold atmospheric plasma and induces transit G(0)/G(1) cell

- cycle arrest of triple negative breast cancers. Free Radic Biol Med. 2025;229:30–8. https://doi.org/10.1016/j.freeradbiomed.2025.01.024.
- Fry FH, Jacob C. Sensor/effector drug design with potential relevance to cancer. Curr Pharm Des. 2006;12(34):4479–99. https://doi.org/10. 2174/138161206779010512.
- 172. Wang Y, Qi H, Liu Y, Duan C, Liu X, Xia T, et al. The double-edged roles of ROS in cancer prevention and therapy. Theranostics. 2021;11(10):4839–57. https://doi.org/10.7150/thno.56747.
- 173. Shimada K, Reznik E, Stokes ME, Krishnamoorthy L, Bos PH, Song Y, et al. Copper-binding small molecule induces oxidative stress and cell-cycle arrest in glioblastoma-patient-derived cells. Cell Chem Biol. 2018;25(5):585–594 e7. https://doi.org/10.1016/j.chembiol.2018.02.010.
- 174. Adams DJ, Boskovic ZV, Theriault JR, Wang AJ, Stern AM, Wagner BK, et al. Discovery of small-molecule enhancers of reactive oxygen species that are nontoxic or cause genotype-selective cell death. ACS Chem Biol. 2013;8(5):923–9. https://doi.org/10.1021/cb300653v.
- Cummings J, Anderson L, Willmott N, Smyth JF. The molecular pharmacology of doxorubicin in vivo. Eur J Cancer. 1991;27(5):532–5. https:// doi.org/10.1016/0277-5379(91)90209-v.
- Ramanathan B, Jan KY, Chen CH, Hour TC, Yu HJ, Pu YS. Resistance to paclitaxel is proportional to cellular total antioxidant capacity. Cancer Res. 2005;65(18):8455–60. https://doi.org/10.1158/0008-5472. CAN-05-1162.
- 177. Renschler MF. The emerging role of reactive oxygen species in cancer therapy. Eur J Cancer. 2004;40(13):1934–40. https://doi.org/10.1016/j.ejca.2004.02.031.
- 178. Ward JF. Biochemistry of DNA lesions. Radiat Res Suppl. 1985;8:S103–11.
- 179. Zhao S, Tang Y, Wang R, Najafi M. Mechanisms of cancer cell death induction by paclitaxel: an updated review. Apoptosis. 2022;27(9–10):647–67. https://doi.org/10.1007/s10495-022-01750-z.
- Christidi E, Brunham LR. Regulated cell death pathways in doxorubicininduced cardiotoxicity. Cell Death Dis. 2021;12(4):339. https://doi.org/ 10.1038/s41419-021-03614-x.
- Barr PM, Miller TP, Friedberg JW, Peterson DR, Baran AM, Herr M, et al. Phase 2 study of imexon, a prooxidant molecule, in relapsed and refractory B-cell non-Hodgkin lymphoma. Blood. 2014;124(8):1259–65. https://doi.org/10.1182/blood-2014-04-570044.
- 182. Pelicano H, Feng L, Zhou Y, Carew JS, Hileman EO, Plunkett W, et al. Inhibition of mitochondrial respiration: a novel strategy to enhance drug-induced apoptosis in human leukemia cells by a reactive oxygen species-mediated mechanism. J Biol Chem. 2003;278(39):37832–9. https://doi.org/10.1074/jbc.M301546200.
- Kim W, Lee S, Seo D, Kim D, Kim K, Kim E, et al. Cellular stress responses in radiotherapy. Cells. 2019;8:9. https://doi.org/10.3390/cells8091105.
- Zou Z, Chang H, Li H, Wang S. Induction of reactive oxygen species: an emerging approach for cancer therapy. Apoptosis. 2017;22(11):1321– 35. https://doi.org/10.1007/s10495-017-1424-9.
- 185. Miranda S, Correia M, Dias AG, Pestana A, Soares P, Nunes J, et al. Evaluation of the role of mitochondria in the non-targeted effects of ionizing radiation using cybrid cellular models. Sci Rep. 2020;10(1):6131. https://doi.org/10.1038/s41598-020-63011-w.
- 186. Xu A, Wang Q, Lv X, Lin T. Progressive study on the non-thermal effects of magnetic field therapy in oncology. Front Oncol. 2021;11: 638146. https://doi.org/10.3389/fonc.2021.638146.
- Yun J, Mullarky E, Lu C, Bosch KN, Kavalier A, Rivera K, et al. Vitamin C selectively kills KRAS and BRAF mutant colorectal cancer cells by targeting GAPDH. Science. 2015;350(6266):1391–6. https://doi.org/10.1126/ science.aaa5004.
- Cameron E, Pauling L. Supplemental ascorbate in the supportive treatment of cancer: prolongation of survival times in terminal human cancer. Proc Natl Acad Sci U S A. 1976;73(10):3685–9. https://doi.org/10. 1073/pnas.73.10.3685.
- Ngo B, Van Riper JM, Cantley LC, Yun J. Targeting cancer vulnerabilities with high-dose vitamin C. Nat Rev Cancer. 2019;19(5):271–82. https://doi.org/10.1038/s41568-019-0135-7.
- Abiri B, Vafa M. Vitamin C and cancer: the role of vitamin C in disease progression and quality of life in cancer patients. Nutr Cancer. 2021;73(8):1282–92. https://doi.org/10.1080/01635581.2020.1795692.
- Zhang G, Liu X, Liu Y, Zhang S, Yu T, Chai X, et al. The effect of magnetic fields on tumor occurrence and progression: Recent advances. Prog

- Biophys Mol Biol. 2023;179:38–50. https://doi.org/10.1016/j.pbiomolbio. 2023.04.001.
- Israel M, Zaryabova V, Ivanova M. Electromagnetic field occupational exposure: non-thermal vs. thermal effects. Electromagn Biol Med. 2013;32(2):145–54. https://doi.org/10.3109/15368378.2013.776349.
- Moori M, Norouzian D, Yaghmaei P, Farahmand L. Electromagnetic field as a possible inhibitor of tumor invasion by declining E-cadherin/Ncadherin switching in triple negative breast cancer. Electromagn Biol Med. 2024:1–10. https://doi.org/10.1080/15368378.2024.2381575.
- 194. Ried K, Eng P, Binjemain T. Metastatic colon cancer an effective treatment protocol of integrative therapies including electromagnetic field frequencies: a case report. Case Rep Oncol. 2023;16(1):1324–34. https://doi.org/10.1159/000534628.
- 195. Tofani S, Barone D, Berardelli M, Berno E, Cintorino M, Foglia L, et al. Static and ELF magnetic fields enhance the in vivo anti-tumor efficacy of cis-platin against lewis lung carcinoma, but not of cyclophosphamide against B16 melanotic melanoma. Pharmacol Res. 2003;48(1):83–90.
- Wang J, Hou Q, Qu J, Huo X, Li H, Feng Y, et al. Polyhedral magnetic nanoparticles induce apoptosis in gastric cancer stem cells and suppressing tumor growth through magnetic force generation. J Control Release. 2024;373:370–84. https://doi.org/10.1016/j.jconrel.2024.07.041.
- Sergeeva EY, Titova NM, Sherbinina AS, Sergeev NV, Shirokova AV. Effect of magnetic fields on antioxidant system enzymes in mice with Ehrlich ascites carcinoma. Bull Exp Biol Med. 2011;150(3):365–7. https://doi.org/ 10.1007/s10517-011-1143-0.
- Martino CF, Portelli L, McCabe K, Hernandez M, Barnes F. Reduction of the Earth's magnetic field inhibits growth rates of model cancer cell lines. Bioelectromagnetics. 2010;31(8):649–55. https://doi.org/10.1002/ bem.20606.
- Storch K, Dickreuter E, Artati A, Adamski J, Cordes N. BEMER electromagnetic field therapy reduces cancer cell radioresistance by enhanced ROS formation and induced DNA damage. PLoS ONE. 2016;11(12): e0167931. https://doi.org/10.1371/journal.pone.0167931.
- Kamalipooya S, Abdolmaleki P, Salemi Z, Javani Jouni F, Zafari J, Soleimani H. Simultaneous application of cisplatin and static magnetic field enhances oxidative stress in HeLa cell line. In Vitro Cell Dev Biol Anim. 2017;53(9):783–90. https://doi.org/10.1007/s11626-017-0148-z.
- Koh EK, Ryu BK, Jeong DY, Bang IS, Nam MH, Chae KS. A 60-Hz sinusoidal magnetic field induces apoptosis of prostate cancer cells through reactive oxygen species. Int J Radiat Biol. 2008;84(11):945–55. https:// doi.org/10.1080/09553000802460206.
- Loscher W, Mevissen M, Lehmacher W, Stamm A. Tumor promotion in a breast cancer model by exposure to a weak alternating magnetic field. Cancer Lett. 1993;71(1–3):75–81. https://doi.org/10.1016/0304-3835(93) 90100-n.
- Malavolti M, Malagoli C, Wise LA, Poli M, Notari B, Taddei I, et al. Residential exposure to magnetic fields from transformer stations and risk of childhood leukemia. Environ Res. 2024;245: 118043. https://doi.org/10.1016/j.envres.2023.118043.
- Malagoli C, Malavolti M, Wise LA, Balboni E, Fabbi S, Teggi S, et al. Residential exposure to magnetic fields from high-voltage power lines and risk of childhood leukemia. Environ Res. 2023;232: 116320. https://doi.org/10.1016/j.envres.2023.116320.
- Brabant C, Geerinck A, Beaudart C, Tirelli E, Geuzaine C, Bruyere O. Exposure to magnetic fields and childhood leukemia: a systematic review and meta-analysis of case-control and cohort studies. Rev Environ Health. 2023;38(2):229–53. https://doi.org/10.1515/reveh-2021-0112.
- Yarmush ML, Golberg A, Sersa G, Kotnik T, Miklavcic D. Electroporationbased technologies for medicine: principles, applications, and challenges. Annu Rev Biomed Eng. 2014;16:295–320. https://doi.org/10. 1146/annurev-bioeng-071813-104622.
- Casciati A, Tanori M, Gianlorenzi I, Rampazzo E, Persano L, Viola G, et al. Effects of ultra-short pulsed electric field exposure on glioblastoma cells. Int J Mol Sci. 2022;23(6): 3001. https://doi.org/10.3390/ijms230630
- Thomson KR, Cheung W, Ellis SJ, Federman D, Kavnoudias H, Loader-Oliver D, et al. Investigation of the safety of irreversible electroporation in humans. J Vasc Interv Radiol. 2011;22(5):611–21. https://doi.org/10. 1016/j.jvir.2010.12.014.

- Scheffer HJ, Vroomen LG, Nielsen K, van Tilborg AA, Comans EF, van Kuijk C, et al. Colorectal liver metastatic disease: efficacy of irreversible electroporation—a single-arm phase Il clinical trial (COLDFIRE-2 trial).
 BMC Cancer. 2015;15:772. https://doi.org/10.1186/s12885-015-1736-5.
- 210. Ruarus AH, Vroomen L, Puijk RS, Scheffer HJ, Zonderhuis BM, Kazemier G, et al. Irreversible electroporation in hepatopancreaticobiliary tumours. Can Assoc Radiol J. 2018;69(1):38–50. https://doi.org/10.1016/j.carj.2017.10.005.
- Scheffer HJ, Nielsen K, van Tilborg AA, Vieveen JM, Bouwman RA, Kazemier G, et al. Ablation of colorectal liver metastases by irreversible electroporation: results of the COLDFIRE-I ablate-and-resect study. Eur Radiol. 2014;24(10):2467–75. https://doi.org/10.1007/s00330-014-3259-x.
- 212. Wendler JJ, Pech M, Kollermann J, Friebe B, Siedentopf S, Blaschke S, et al. Upper-urinary-tract effects after Irreversible Electroporation (IRE) of human localised Renal-Cell Carcinoma (RCC) in the IRENE pilot phase 2a ablate-and-resect study. Cardiovasc Intervent Radiol. 2018;41(3):466–76. https://doi.org/10.1007/s00270-017-1795-x.
- Buijs M, Zondervan PJ, de Bruin DM, van Lienden KP, Bex A, van Delden OM. Feasibility and safety of irreversible electroporation (IRE) in patients with small renal masses: results of a prospective study. Urol Oncol. 2019;37(3):183 e1–183 e8. https://doi.org/10.1016/j.urolonc.2018.11. 008
- 214. Onik G, Rubinsky B. Irreversible electroporation: first patient experience focal therapy of prostate cancer. In: B. Rubinsky (Ed.). Irreversible electroporation. Berlin, Germany: Springer; 2010.
- Van den Bos W, Jurhill RR, de Bruin DM, Savci-Heijink CD, Postema AW, Wagstaff PG, et al. Histopathological outcomes after irreversible electroporation for prostate cancer: results of an ablate and resect study. J Urol. 2016;196(2):552–9. https://doi.org/10.1016/j.juro.2016.02.2977.
- Van den Bos W, Scheltema MJ, Siriwardana AR, Kalsbeek AMF, Thompson JE, Ting F, et al. Focal irreversible electroporation as primary treatment for localized prostate cancer. BJU Int. 2018;121(5):716–24. https://doi.org/10.1111/bju.13983.
- Narayanan G, Hosein PJ, Beulaygue IC, Froud T, Scheffer HJ, Venkat SR, et al. Percutaneous image-guided irreversible electroporation for the treatment of unresectable, locally advanced pancreatic adenocarcinoma. J Vasc Interv Radiol. 2017;28(3):342–8. https://doi.org/10.1016/j. ivir.2016.10.023.
- Martin RC 2nd, Kwon D, Chalikonda S, Sellers M, Kotz E, Scoggins C, et al. Treatment of 200 locally advanced (stage III) pancreatic adenocarcinoma patients with irreversible electroporation: safety and efficacy. Ann Surg. 2015;262(3):486–94. https://doi.org/10.1097/SLA.0000000000 001441.
- Leen E, Picard J, Stebbing J, Abel M, Dhillon T, Wasan H. Percutaneous irreversible electroporation with systemic treatment for locally advanced pancreatic adenocarcinoma. J Gastrointest Oncol. 2018;9(2):275–81. https://doi.org/10.21037/jgo.2018.01.14.
- Rombouts SJ, Walma MS, Vogel JA, van Rijssen LB, Wilmink JW, Mohammad NH, et al. Systematic review of resection rates and clinical outcomes after FOLFIRINOX-based treatment in patients with locally advanced pancreatic cancer. Ann Surg Oncol. 2016;23(13):4352–60. https://doi.org/10.1245/s10434-016-5373-2.
- Vincent A, Herman J, Schulick R, Hruban RH, Goggins M. Pancreatic cancer. Lancet. 2011;378(9791):607–20. https://doi.org/10.1016/S0140-6736(10)62307-0.
- 222. Kourounis G, Paul Tabet P, Moris D, Papalambros A, Felekouras E, Georgiades F, et al. Irreversible electroporation (Nanoknife(R) treatment) in the field of hepatobiliary surgery: current status and future perspectives. J BUON. 2017;22(1):141–9.
- 223. Silk MT, Wimmer T, Lee KS, Srimathveeravalli G, Brown KT, Kingham PT, et al. Percutaneous ablation of peribiliary tumors with irreversible electroporation. J Vasc Interv Radiol. 2014;25(1):112–8. https://doi.org/10.1016/j.jvir.2013.10.012.
- Scheffer HJ, Nielsen K, de Jong MC, van Tilborg AA, Vieveen JM, Bouwman AR, et al. Irreversible electroporation for nonthermal tumor ablation in the clinical setting: a systematic review of safety and efficacy. J Vasc Interv Radiol. 2014;25(7):997–1011. https://doi.org/10.1016/j.jvir. 2014.01.028. quiz 1011.
- Gehl J, Sersa G, Matthiessen LW, Muir T, Soden D, Occhini A, et al. Updated standard operating procedures for electrochemotherapy of

- cutaneous tumours and skin metastases. Acta Oncol. 2018;57(7):874–82. https://doi.org/10.1080/0284186X.2018.1454602.
- 226. Faiella E, Santucci D, Vertulli D, Vergantino E, Vaccarino F, Perillo G, et al. Irreversible Electroporation (IRE) for Prostate Cancer (PCa) treatment: the state of the art. J Pers Med. 2024;14(2). https://doi.org/10.3390/jpm14020137.
- Schmidt R. Photosensitized generation of singlet oxygen. Photochem Photobiol. 2006;82(5):1161–77. https://doi.org/10.1562/2006-03-03-IR-833.
- 228. Singh N, Sen Gupta R, Bose S. A comprehensive review on singlet oxygen generation in nanomaterials and conjugated polymers for photodynamic therapy in the treatment of cancer. Nanoscale. 2024;16(7):3243–68. https://doi.org/10.1039/d3nr05801h.
- Nwogu C, Kloc A, Attwood K, Bshara W, Durrani F, Pandey R. Porfimer sodium versus PS785 for Photodynamic Therapy (PDT) of lung cancer xenografts in mice. J Surg Res. 2021;263:245–50. https://doi.org/10. 1016/i.iss.2020.12.067.
- Sanchez-Cruz P, Vazquez K, Lozada EL, Valiyeva F, Sharma R, Vivas PE, et al. Photosensitized co-generation of nitric oxide and singlet oxygen Enhanced toxicity against ovarian cancer cells. J Nanopart Res. 2022;24(4): 82. https://doi.org/10.1007/s11051-022-05463-x.
- 231. Um W, E KP, Lee J, Kim CH, You DG, Park JH. Recent advances in nanomaterial-based augmented sonodynamic therapy of cancer. Chem Commun (Camb). 2021;57(23):2854-2866. https://doi.org/10.1039/d0cc07750j.
- 232. Yin H, Sun L, Pu Y, Yu J, Feng W, Dong C, et al. Ultrasound-controlled CRISPR/Cas9 system augments sonodynamic therapy of hepatocellular carcinoma. ACS Cent Sci. 2021;7(12):2049–62. https://doi.org/10.1021/acscentsci.1c01143.
- Yue W, Chen L, Yu L, Zhou B, Yin H, Ren W, et al. Checkpoint blockade and nanosonosensitizer-augmented noninvasive sonodynamic therapy combination reduces tumour growth and metastases in mice. Nat Commun. 2019;10(1):2025. https://doi.org/10.1038/ \$41467-019-09760-3.
- 234. Dahan M, Cortet M, Lafon C, Padilla F. Combination of focused ultrasound, immunotherapy, and chemotherapy: new perspectives in breast cancer therapy. J Ultrasound Med. 2023;42(3):559–73. https://doi.org/10.1002/jum.16053.
- Wu T, Liu Y, Cao Y, Liu Z. Engineering macrophage exosome disguised biodegradable nanoplatform for enhanced sonodynamic therapy of glioblastoma. Adv Mater. 2022;34(15): e2110364. https://doi.org/10. 1002/adma.202110364.
- 236. Lu D, Wang L, Wang L, An L, Huo M, Xu H, et al. Probiotic engineering and targeted sonoimmuno-therapy augmented by STING agonist. Adv Sci (Weinh). 2022;9(22): e2201711. https://doi.org/10.1002/advs.20220 1711.
- 237. Nesbitt H, Logan K, Thomas K, Callan B, Gao J, McKaig T, et al. Sonodynamic therapy complements PD-L1 immune checkpoint inhibition in a murine model of pancreatic cancer. Cancer Lett. 2021;517:88–95. https://doi.org/10.1016/j.canlet.2021.06.003.
- 238. Gong Z, Dai Z. Design and challenges of sonodynamic therapy system for cancer theranostics: from equipment to sensitizers. Adv Sci (Weinh). 2021;8(10): 2002178. https://doi.org/10.1002/advs.202002178.
- Li X, Gao J, Wu C, Wang C, Zhang R, He J, et al. Precise modulation and use of reactive oxygen species for immunotherapy. Sci Adv. 2024;10(20):eadl0479. https://doi.org/10.1126/sciadv.adl0479.
- 240. Wang YB, Wu WB, Mao D, Teh C, Wang B, Liu B. Metal-organic framework assisted and tumor microenvironment modulated synergistic imageguided photo-chemo therapy. Adv Funct Mater. 2020;30(28):ARTN 2002431. https://doi.org/10.1002/adfm.202002431.
- 241. Kuo SH, Wu PT, Huang JY, Chiu CP, Yu J, Liao MY. Fabrication of Anisotropic Cu Ferrite-Polymer Core-Shell Nanoparticles for Photodynamic Ablation of Cervical Cancer Cells. Nanomaterials (Basel). 2020;10(12). https://doi.org/10.3390/nano10122429.
- 242. Dai X, Bazaka K, Thompson EW, Ostrikov KK. Cold atmospheric plasma: a promising controller of cancer cell states. Cancers (Basel). 2020;12(11): 3360. https://doi.org/10.3390/cancers12113360.
- Xiang L, Xu X, Zhang S, Cai D, Dai X. Cold atmospheric plasma conveys selectivity on triple negative breast cancer cells both in vitro and in vivo. Free Radic Biol Med. 2018;124:205–13. https://doi.org/10.1016/j. freeradbiomed.2018.06.001.

- 244. Sapino A, Lehmann BD, Jovanović B, Chen X, Estrada MV, Johnson KN, et al. Refinement of triple-negative breast cancer molecular subtypes: implications for neoadjuvant chemotherapy selection. PLoS One. 2016;11(6). https://doi.org/10.1371/journal.pone.0157368.
- Keidar M, Walk R, Shashurin A, Srinivasan P, Sandler A, Dasgupta S, et al. Cold plasma selectivity and the possibility of a paradigm shift in cancer therapy. Br J Cancer. 2011;105(9):1295–301. https://doi.org/10.1038/bjc. 2011.386.
- 246. Roy S, Ma Y, Ha CS, Hwang SW, Lee HJ, Kim GC, et al. Non-Thermal Atmospheric Pressure Plasma Preferentially Induces Apoptosis in p53-Mutated Cancer Cells by Activating ROS Stress-Response Pathways. PLoS One. 2014;9(4). https://doi.org/10.1371/journal.pone.0091947.
- 247. Weltmann KD, Kindel E, von Woedtke T, Hähnel M, Stieber M, Brandenburg R. Atmospheric-pressure plasma sources: Prospective tools for plasma medicine. Pure Appl Chem. 2010;82(6):1223–37. https://doi.org/10.1351/pac-con-09-10-35.
- Wang P, Zhou R, Zhou R, Feng S, Zhao L, Li W, et al. Epidermal growth factor potentiates EGFR(Y992/1173)-mediated therapeutic response of triple negative breast cancer cells to cold atmospheric plasma-activated medium. Redox Biol. 2024;69: 102976. https://doi.org/10.1016/j. redox 2023 102976
- 249. Dai X, Cai D, Wang P, Nan N, Yu L, Zhang Z, et al. Cold atmospheric plasmas target breast cancer stemness via modulating AQP3-19Y mediated AQP3-5K and FOXO1 K48-ubiquitination. Int J Biol Sci. 2022;18(8):3544–61. https://doi.org/10.7150/ijbs.72296.
- Wang P, Zhou R, Thomas P, Zhao L, Zhou R, Mandal S, et al. Epithelial-to-mesenchymal transition enhances cancer cell sensitivity to cytotoxic effects of cold atmospheric plasmas in breast and bladder cancer systems. Cancers (Basel). 2021;13(12). https://doi.org/10.3390/cancers13122889.
- 251. Hua D, Cai D, Ning M, Yu L, Zhang Z, Han P, et al. Cold atmospheric plasma selectively induces G(0)/G(1) cell cycle arrest and apoptosis in AR-independent prostate cancer cells. J Cancer. 2021;12(19):5977–86. https://doi.org/10.7150/jca.54528.
- Adhikari M, Adhikari B, Adhikari A, Yan D, Soni V, Sherman J, et al. Cold atmospheric plasma as a novel therapeutic tool for the treatment of brain cancer. Curr Pharm Des. 2020;26(19):2195–206. https://doi.org/10. 2174/1381612826666200302105715.
- Cai L, Chen Y, Dai X. CD44 as a prognostic index of brain cancers and therapeutic target of cold atmospheric plasma. Aging. 2024:in print.
- Soulat A, Mohsenpour T, Roshangar L, Naghshara H. A two-stage transferred cold atmospheric plasma as a unique therapeutic strategy for targeting colon cancer stem cells. Adv Pharm Bull. 2024;14(2):400–11. https://doi.org/10.34172/apb.2024.041.
- Wang Y, Mang X, Li X, Cai Z, Tan F. Cold atmospheric plasma induces apoptosis in human colon and lung cancer cells through modulating mitochondrial pathway. Front Cell Dev Biol. 2022;10: 915785. https:// doi.org/10.3389/fcell.2022.915785.
- Privat-Maldonado A, Verloy R, Cardenas Delahoz E, Lin A, Vanlanduit S, Smits E, et al. Cold atmospheric plasma does not affect stellate cells phenotype in pancreatic cancer tissue in Ovo. Int J Mol Sci. 2022;23(4): 1954. https://doi.org/10.3390/ijms23041954.
- Verloy R, Privat-Maldonado A, Smits E, Bogaerts A. Cold Atmospheric Plasma Treatment for Pancreatic Cancer-The Importance of Pancreatic Stellate Cells. Cancers (Basel). 2020;12(10). https://doi.org/10.3390/ cancers12102782.
- Chen C, Zhou S, Yang X, Ren M, Qi Y, Mao Y, et al. In vitro study of cold atmospheric plasma-activated liquids inhibits malignant melanoma by affecting macrophage polarization through the ROS/JAK2/STAT1 pathway. Biomed Pharmacother. 2024;175: 116657. https://doi.org/10. 1016/j.biopha.2024.116657.
- 259. Yazdani Z, Pasandi MS, Golpour M, Eslami M, Rafiei A. Effect of cold atmospheric plasma on changing of biomolecular structures involved in apoptosis pathways of melanoma cancer. Skin Res Technol. 2024;30(1): e13544. https://doi.org/10.1111/srt.13544.
- Aggelopoulos CA, Christodoulou A-M, Tachliabouri M, Meropoulis S, Christopoulou M-E, Karalis TT, et al. Cold atmospheric plasma attenuates breast cancer cell growth through regulation of cell microenvironment effectors. Front Oncol. 2022;11: 826865. https://doi.org/10.3389/fonc.2021.826865.

- Mihai CT, Mihaila I, Pasare MA, Pintilie RM, Ciorpac M, Topala I. Cold atmospheric plasma-activated media improve paclitaxel efficacy on breast cancer cells in a combined treatment model. Curr Issues Mol Biol. 2022;44(5):1995–2014. https://doi.org/10.3390/cimb44050135.
- 262. Park S, Kim H, Ji HW, Kim HW, Yun SH, Choi EH, et al. Cold atmospheric plasma restores paclitaxel sensitivity to paclitaxel-resistant breast cancer cells by reversing expression of resistance-related genes. Cancers (Basel). 2019;11(12). https://doi.org/10.3390/cancers11122011.
- Dezhpour A, Ghafouri H, Jafari S, Nilkar M. Effects of cold atmosphericpressure plasma in combination with doxorubicin drug against breast cancer cells in vitro and invivo. Free Radic Biol Med. 2023;209(Pt 2):202–10. https://doi.org/10.1016/i.freeradbiomed.2023.10.405.
- Lee S, Lee H, Jeong D, Ham J, Park S, Choi EH, et al. Cold atmospheric plasma restores tamoxifen sensitivity in resistant MCF-7 breast cancer cell. Free Radic Biol Med. 2017;110:280–90. https://doi.org/10.1016/j. freeradbiomed.2017.06.017.
- Dai X, Bazaka K, Richard DJ, Thompson ERW, Ostrikov KK. The emerging role of gas plasma in oncotherapy. Trends Biotechnol. 2018;36(11):1183–98. https://doi.org/10.1016/i.tibtech.2018.06.010.
- Van Loenhout J, Freire Boullosa L, Quatannens D, De Waele J, Merlin C, Lambrechts H, et al. Auranofin and cold atmospheric plasma synergize to trigger distinct cell death mechanisms and immunogenic responses in glioblastoma. Cells. 2021;10(11): 2936. https://doi.org/10.3390/cells 10112936.
- Van Loenhout J, Flieswasser T, Freire Boullosa L, De Waele J, Van Audenaerde J, Marcq E, et al. Cold atmospheric plasma-treated PBS eliminates immunosuppressive pancreatic stellate cells and induces immunogenic cell death of pancreatic cancer cells. Cancers (Basel). 2019;11(10). https://doi.org/10.3390/cancers11101597.
- Isbary G, Shimizu T, Li YF, Stolz W, Thomas HM, Morfill GE, et al. Cold atmospheric plasma devices for medical issues. Expert Rev Med Devices. 2013;10(3):367–77. https://doi.org/10.1586/erd.13.4.
- Hoffmann C, Berganza C, Zhang J. Cold Atmospheric Plasma: methods of production and application in dentistry and oncology. Med Gas Res. 2013;3(1): 21. https://doi.org/10.1186/2045-9912-3-21.
- Tanaka H, Nakamura K, Mizuno M, Ishikawa K, Takeda K, Kajiyama H, et al. Non-thermal atmospheric pressure plasma activates lactate in Ringer's solution for anti-tumor effects. Sci Rep. 2016;6: 36282. https://doi.org/10.1038/srep36282.
- Azzariti A, lacobazzi RM, Di Fonte R, Porcelli L, Gristina R, Favia P, et al. Plasma-activated medium triggers cell death and the presentation of immune activating danger signals in melanoma and pancreatic cancer cells. Sci Rep. 2019;9(1):4099. https://doi.org/10.1038/ s41598-019-40637-z.
- Bauer G, Sersenova D, Graves DB, Machala Z. Cold atmospheric plasma and plasma-activated medium trigger RONS-based tumor cell apoptosis. Sci Rep. 2019;9(1):14210. https://doi.org/10.1038/ s41598-019-50291-0.
- Yan D, Talbot A, Nourmohammadi N, Cheng X, Canady J, Sherman J, et al. Principles of using cold atmospheric plasma stimulated media for cancer treatment. Sci Rep. 2015;5: 18339. https://doi.org/10.1038/srep1 8339.
- Liu Z, Chen J, Ren Y, Liu S, Ba Y, Zuo A, et al. Multi-stage mechanisms of tumor metastasis and therapeutic strategies. Signal Transduct Target Ther. 2024;9(1):270. https://doi.org/10.1038/s41392-024-01955-5.
- Cañellas-Socias A, Cortina C, Hernando-Momblona X, Palomo-Ponce S, Mulholland EJ, Turon G, et al. Metastatic recurrence in colorectal cancer arises from residual EMP1(+) cells. Nature. 2022;611(7936):603–13. https://doi.org/10.1038/s41586-022-05402-9.
- Fang Y, Luo X, Xu Y, Liu Z, Mintz RL, Yu H, et al. Sandwich-structured implants to obstruct multipath energy supply and trigger selfenhanced hypoxia-initiated chemotherapy against postsurgical tumor recurrence and metastasis. Adv Sci (Weinh). 2023;10(22): e2300899. https://doi.org/10.1002/advs.202300899.
- Fang Y, Huang S, Hu Q, Zhang J, King JA, Wang Y, et al. Injectable zwitterionic physical hydrogel with enhanced chemodynamic therapy and tumor microenvironment remodeling properties for synergistic anticancer therapy. ACS Nano. 2023;17(24):24883–900. https://doi.org/10.1021/acsnano.3c05898.
- Boekema B, Stoop M, Vlig M, van Liempt J, Sobota A, Ulrich M, et al. Antibacterial and safety tests of a flexible cold atmospheric plasma

- device for the stimulation of wound healing. Appl Microbiol Biotechnol. 2021;105(5):2057–70. https://doi.org/10.1007/s00253-021-11166-5.
- 279. Boeckmann L, Schäfer M, Bernhardt T, Semmler ML, Jung O, Ojak G, et al. Cold atmospheric pressure plasma in wound healing and cancer treatment. Appl Sci. 2020;10(19):6898.
- Lin A, Gorbanev Y, De Backer J, Van Loenhout J, Van Boxem W, Lemiere F, et al. Non-thermal plasma as a unique delivery system of short-lived reactive oxygen and nitrogen species for immunogenic cell death in melanoma cells. Adv Sci (Weinh). 2019;6(6):1802062. https://doi.org/10. 1002/advs.201802062.
- Lin A, Truong B, Pappas A, Kirifides L, Oubarri A, Chen S, et al. Uniform nanosecond pulsed dielectric barrier discharge plasma enhances anti-tumor effects by induction of immunogenic cell death in tumors and stimulation of macrophages. Plasma Processes and Polymers. 2015;12(12):1392–9. https://doi.org/10.1002/ppap.201500139.
- Lin A, Truong B, Patel S, Kaushik N, Choi EH, Fridman G, et al. Nanosecond-pulsed DBD plasma-generated reactive oxygen species trigger immunogenic cell death in A549 lung carcinoma cells through intracellular oxidative stress. Int J Mol Sci. 2017;18(5). https://doi.org/10.3390/iims18050966.
- 283. Lin AG, Xiang B, Merlino DJ, Baybutt TR, Sahu J, Fridman A, et al. Nonthermal plasma induces immunogenic cell death in vivo in murine CT26 colorectal tumors. Oncoimmunology. 2018;7(9): e1484978. https://doi.org/10.1080/2162402X.2018.1484978.
- Bekeschus S, Mueller A, Miller V, Gaipl U, Weltmann KD. Physical Plasma Elicits Immunogenic Cancer Cell Death and Mitochondrial Singlet Oxygen. IEEE Transactions on Radiation and Plasma Medical Sciences. 2018;2(2):138–46. https://doi.org/10.1109/TRPMS.2017.2766027.
- Dai X, Wu J, Lu L, Chen Y. Current status and future trends of cold atmospheric plasma as an oncotherapy. Biomol Ther (Seoul). 2023;31(5):496–514. https://doi.org/10.4062/biomolther.2023.027.
- Canady MDJ, Shashurin A, Wiley K, Fisch N, Keidar M. Characterization of plasma parameters and tissue injury produced by plasma electrosurgical systems. Plasma Med. 2013;3:279–89. https://doi.org/10.1615/Plasm aMed.2014011979.
- 287. Su JX, Li SJ, Zhou XF, Zhang ZJ, Yan Y, Liu SL, et al. Chemotherapyinduced metastasis: molecular mechanisms and clinical therapies. Acta Pharmacol Sin. 2023;44(9):1725–36. https://doi.org/10.1038/ s41401-023-01093-8
- Eddy RJ, Weidmann MD, Sharma VP, Condeelis JS. Tumor cell invadopodia: invasive protrusions that orchestrate metastasis. Trends Cell Biol. 2017;27(8):595–607. https://doi.org/10.1016/j.tcb.2017.03.003.
- 289. Quintavalle M, Elia L, Price JH, Heynen-Genel S, Courtneidge SA. A cell-based high-content screening assay reveals activators and inhibitors of cancer cell invasion. Sci Signal. 2011;4(183):ra49. https://doi.org/10.1126/scisignal.2002032.
- Camara O, Rengsberger M, Egbe A, Koch A, Gajda M, Hammer U, et al. The relevance of circulating epithelial tumor cells (CETC) for therapy monitoring during neoadjuvant (primary systemic) chemotherapy in breast cancer. Ann Oncol. 2007;18(9):1484–92. https://doi.org/10.1093/ annonc/mdm206.
- Lu H, Tran L, Park Y, Chen I, Lan J, Xie Y, et al. Reciprocal regulation of DUSP9 and DUSP16 expression by HIF1 controls ERK and p38 MAP kinase activity and mediates chemotherapy-induced breast cancer stem cell enrichment. Cancer Res. 2018;78(15):4191–202. https://doi. org/10.1158/0008-5472.Can-18-0270.
- 292. Kajiyama H, Shibata K, Terauchi M, Yamashita M, Ino K, Nawa A, et al. Chemoresistance to paclitaxel induces epithelial-mesenchymal transition and enhances metastatic potential for epithelial ovarian carcinoma cells. Int J Oncol. 2007;31(2):277–83.
- 293. Chiang FF, Huang SC, Yu PT, Chao TH, Huang YC. Oxidative stress induced by chemotherapy: evaluation of glutathione and its related antioxidant enzyme dynamics in patients with colorectal cancer. Nutrients. 2023;15(24): 5104. https://doi.org/10.3390/nu15245104.
- 294. Conklin KA. Chemotherapy-associated oxidative stress: impact on chemotherapeutic effectiveness. Integr Cancer Ther. 2004;3(4):294–300. https://doi.org/10.1177/1534735404270335.
- 295. Erhola M, Kellokumpu-Lehtinen P, Metsä-Ketelä T, Alanko K, Nieminen MM. Effects of anthracyclin-based chemotherapy on total plasma antioxidant capacity in small cell lung cancer patients. Free Radic Biol Med. 1996;21(3):383–90. https://doi.org/10.1016/0891-5849(96)00041-x.

Dai et al. Molecular Biomedicine (2025) 6:20 Page 30 of 30

- 296. Kasapović J, Pejić S, Stojiljković V, Todorović A, Radošević-Jelić L, Saičić ZS, et al. Antioxidant status and lipid peroxidation in the blood of breast cancer patients of different ages after chemotherapy with 5-fluorouracil, doxorubicin and cyclophosphamide. Clin Biochem. 2010;43(16–17):1287–93. https://doi.org/10.1016/j.clinbiochem.2010.08.009.
- Hamza TA, Muhsin SA, Khalil TT. Evaluation of malondialdehyde, homocysteine and antioxidant influence chemotherapy in breast cancer patients. Military Med Sci Lett. 2023;92(2):105–11.
- Thanoon IA, Ahmed FA, Jadoaa KR. Lipid peroxidation and antioxidant status in post-operative patients with cancer treated with chemotherapy. Med Sci. 2018;14:64–7.
- 299. Wang Y, Mang X, Li D, Wang Z, Chen Y, Cai Z, et al. Cold atmospheric plasma sensitizes head and neck cancer to chemotherapy and immune checkpoint blockade therapy. Redox Biol. 2024;69: 102991. https://doi.org/10.1016/j.redox.2023.102991.
- Lafontaine J, Boisvert JS, Glory A, Coulombe S, Wong P. Synergy between Non-Thermal Plasma with Radiation Therapy and Olaparib in a Panel of Breast Cancer Cell Lines. Cancers (Basel). 2020;12(2). https://doi. org/10.3390/cancers12020348.
- Buchbinder E, Hodi FS. Cytotoxic T lymphocyte antigen-4 and immune checkpoint blockade. J Clin Invest. 2015;125(9):3377–83. https://doi. org/10.1172/jci80012.
- Jiang Y, Chen M, Nie H, Yuan Y. PD-1 and PD-L1 in cancer immunotherapy: clinical implications and future considerations. Hum Vaccin Immunother. 2019;15(5):1111–22. https://doi.org/10.1080/21645515. 2019.1571892.
- Syn NL, Teng MWL, Mok TSK, Soo RA. De-novo and acquired resistance to immune checkpoint targeting. Lancet Oncol. 2017;18(12):e731–41. https://doi.org/10.1016/s1470-2045(17)30607-1.
- Qin S, Xu L, Yi M, Yu S, Wu K, Luo S. Novel immune checkpoint targets: moving beyond PD-1 and CTLA-4. Mol Cancer. 2019;18(1):155. https://doi.org/10.1186/s12943-019-1091-2.
- Gao X, Zhu Y, Li G, Huang H, Zhang G, Wang F, et al. TIM-3 expression characterizes regulatory T cells in tumor tissues and is associated with lung cancer progression. PLoS ONE. 2012;7(2): e30676. https://doi.org/ 10.1371/journal.pone.0030676.
- Fourcade J, Sun Z, Benallaoua M, Guillaume P, Luescher IF, Sander C, et al. Upregulation of Tim-3 and PD-1 expression is associated with tumor antigen-specific CD8+T cell dysfunction in melanoma patients. J Exp Med. 2010;207(10):2175–86. https://doi.org/10.1084/jem.20100 637.
- Gallois A, Silva I, Osman I, Bhardwaj N. Reversal of natural killer cell exhaustion by TIM-3 blockade. Oncoimmunology. 2014;3(12): e946365. https://doi.org/10.4161/21624011.2014.946365.
- 308. Koyama S, Akbay EA, Li YY, Herter-Sprie GS, Buczkowski KA, Richards WG, et al. Adaptive resistance to therapeutic PD-1 blockade is associated with upregulation of alternative immune checkpoints. Nat Commun. 2016;7: 10501. https://doi.org/10.1038/ncomms10501.
- 309. de Mingo PÁ, Gardner A, Hiebler S, Soliman H, Rugo HS, Krummel MF, et al. TIM-3 regulates CD103(+) dendritic cell function and response to chemotherapy in breast cancer. Cancer Cell. 2018;33(1):60–74.e6. https://doi.org/10.1016/j.ccell.2017.11.019.
- Zhou G, Noordam L, Sprengers D, Doukas M, Boor PPC, van Beek AA, et al. Blockade of LAG3 enhances responses of tumor-infiltrating T cells in mismatch repair-proficient liver metastases of colorectal cancer. Oncoimmunology. 2018;7(7): e1448332. https://doi.org/10.1080/21624 02x.2018.1448332.
- 311. Matsuzaki J, Gnjatic S, Mhawech-Fauceglia P, Beck A, Miller A, Tsuji T, et al. Tumor-infiltrating NY-ESO-1-specific CD8+T cells are negatively regulated by LAG-3 and PD-1 in human ovarian cancer. Proc Natl Acad Sci U S A. 2010;107(17):7875–80. https://doi.org/10.1073/pnas.10033
- 312. Baitsch L, Baumgaertner P, Devêvre E, Raghav SK, Legat A, Barba L, et al. Exhaustion of tumor-specific CD8⁺T cells in metastases from melanoma patients. J Clin Invest. 2011;121(6):2350–60. https://doi.org/10.1172/jci46102.
- Woo SR, Turnis ME, Goldberg MV, Bankoti J, Selby M, Nirschl CJ, et al. Immune inhibitory molecules LAG-3 and PD-1 synergistically regulate T-cell function to promote tumoral immune escape. Cancer Res. 2012;72(4):917–27. https://doi.org/10.1158/0008-5472.Can-11-1620.

- Huang RY, Eppolito C, Lele S, Shrikant P, Matsuzaki J, Odunsi K. LAG3 and PD1 co-inhibitory molecules collaborate to limit CD8+T cell signaling and dampen antitumor immunity in a murine ovarian cancer model. Oncotarget. 2015;6(29):27359–77. https://doi.org/10.18632/ oncotarget.4751.
- Huang RY, Francois A, McGray AR, Milliotto A, Odunsi K. Compensatory upregulation of PD-1, LAG-3, and CTLA-4 limits the efficacy of singleagent checkpoint blockade in metastatic ovarian cancer. Oncoimmunology. 2017;6(1): e1249561. https://doi.org/10.1080/2162402x.2016. 1249561.
- 316. Zhu K, Lv Q, Lu X, Wang Y, Dai X. Cold atmospheric plasma restores skewed macrophage polarization in triple negative breast cancers via enhancing KAT6A acetylation. Free Radic Biol Med. 2024;226:364–73. https://doi.org/10.1016/j.freeradbiomed.2024.11.028.
- Chen G, Chen Z, Wen D, Wang Z, Li H, Zeng Y, et al. Transdermal cold atmospheric plasma-mediated immune checkpoint blockade therapy. Proc Natl Acad Sci U S A. 2020;117(7):3687–92. https://doi.org/10.1073/ pnas.1917891117.
- 318. He DJ, Yu DQ, Wang QM, Yu ZY, Qi YH, Shao QJ, et al. Breast cancer subtypes and mortality of breast cancer patients with brain metastasis at diagnosis: a population-based study. Inquiry. 2021;58: 469580211055636. https://doi.org/10.1177/00469580211055636.
- 319. Dai X, Thompson EW, Ostrikov KK. Receptor-mediated redox imbalance: an emerging clinical avenue against aggressive cancers. Biomolecules. 2022;12(12). https://doi.org/10.3390/biom12121880.
- 320. Yang Y, Dai X. Current status of controlled onco-therapies based on metal organic frameworks. RSC Adv. 2024;14(18):12817–28. https://doi.org/10.1039/d4ra00375f.
- Dai X, Dai Y, Zheng Y, Lv Y. Magnetic nanoparticles and possible synergies with cold atmospheric plasma for cancer treatment. RSC Adv. 2024;14(40):29039–51. https://doi.org/10.1039/d4ra03837a.
- 322. Dai X, Yang Y. Metal-organic frameworks: potential synergies with cold atmospheric plasmas for cancer control. J Mater Chem B. 2024;12(42):10770–85. https://doi.org/10.1039/d4tb00968a.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.