#### ORIGINAL RESEARCH ARTICLE



# Pharmacokinetics of the P-gp Inhibitor Tariquidar in Rats After Intravenous, Oral, and Intraperitoneal Administration

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#### Abstract

Background and objective P-glycoprotein (P-gp), a transmembrane transporter expressed at the blood-brain barrier, restricts the distribution of diverse central nervous system-targeted drugs from blood into brain, reducing their therapeutic efficacy. The third-generation P-gp inhibitor tariquidar (XR9576) was shown to enhance brain distribution of P-gp substrate drugs in humans. Oral bioavailability of tariquidar was found to be low in humans requiring the compound to be administered intravenously, which hinders a broader clinical use. The objective of the present study was to investigate the plasma

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pharmacokinetics of tariquidar in rats after single intravenous, oral, and intraperitoneal administration.

Methods Two different tariquidar formulations (A and B) were used, both at a dosage of 15 mg/kg, respectively. Formulation A was a solution and formulation B was a microemulsion which was previously shown to improve the oral bioavailability of the structurally related P-gp inhibitor elacridar in mice.

Results In contrast to human data, the present study found a high bioavailability of tariquidar in rats after oral dosing. Oral bioavailability was significantly higher (p = 0.032) for formulation B (86.3%) than for formulation A (71.6%). After intraperitoneal dosing bioavailability was 91.4% for formulation A and 99.6% for formulation B.

Conclusion The present findings extend the available information on tariquidar and provide a basis for future studies involving oral administration of this compound.

# **Key Points**

This study investigated the pharmacokinetics of tariquidar in rats after oral, intraperitoneal and intravenous administration. Out of two tariquidar formulations used, a microemulsion showed significantly higher oral bioavailability in rats and might be considered for further testing.

Availability of a tariquidar formulation with good oral bioavailability in humans would allow for a broader use of the drug in clinical research as a booster of brain delivery of P-gp substrate drugs.

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#### 1 Introduction

P-glycoprotein (P-gp or ABCB1) is a transmembrane transport protein belonging to the adenosine triphosphate (ATP)-binding cassette (ABC) superfamily, which protects different tissues against potentially toxic xenobiotics [1–3]. It is expressed in cell membranes of various mammalian organ systems including the kidney, liver, intestine, in the luminal side of brain capillary endothelial cells forming the blood–brain barrier (BBB) and in peripheral blood mononuclear cells like lymphocytes and macrophages [1–3]. Accordingly, it has a major impact on the bioavailability, tissue distribution, and excretion of drugs.

Tariquidar (XR9576) is a non-marketed third-generation P-gp inhibitor [4]. Tariquidar's potential to overcome P-gpmediated multidrug resistance (MDR) of tumors has been investigated in human phase I/II clinical studies [5-8]. However, further clinical development of tariquidar as an MDR reversal agent has been halted due to lack of efficacy and side effects in combination with cytotoxic anticancer drugs. More recently, promising data have been obtained with tariquidar in clinical studies in healthy volunteers with respect to its ability to inhibit P-gp at the BBB and to facilitate brain delivery of P-gp substrate drugs [9, 10]. In addition, since P-gp-related transporters in prokaryotic cells have been shown to mediate resistance to antibiotic drugs, tariquidar has also been investigated as a means to reverse bacterial MDR [11-13]. Radiolabeled tariquidar has been used to visualize the function of P-gp and breast cancer-resistance protein (BCRP or ABCG2), another ABC transporter, at the rodent and human BBB [14–17].

In short, despite the fact that tariquidar has never reached marketing authorization, it still attracts a great deal of scientific interest. To date, although a considerable amount of research on and involving tariquidar has been conducted in laboratory animals, most notably rodents, only limited information is available on the pharmacokinetics of the compound in these species.

The main route of excretion of tariquidar is biliary excretion and direct secretion across the gastrointestinal tract mucosa, with urinary excretion constituting only a minor route [18, 19]. From a clinical study involving intravenous administration of ascending doses of tariquidar up to 8 mg/kg body weight to human healthy volunteers, a long elimination half-life, and a large volume of distribution emerged as the drug's key pharmacokinetic features [20]. There is some evidence that tariquidar displays nonlinear pharmacokinetics, which may be related to saturation of transport proteins mediating excretion and/or tissue distribution of tariquidar at higher doses [15, 20].

Preliminary data on oral dosing of tariquidar in human healthy volunteers have been reported, suggesting a low oral bioavailability of the compound [10, 21]. Aim of the present study was to investigate the pharmacokinetic profile of tariquidar in rats after intravenous, oral, and intraperitoneal administration and to test a previously described microemulsion formulation [22] with respect to its ability to enhance the bioavailability of tariquidar. Exhaustive information on oral pharmacokinetics in rodents could be relevant for future studies involving oral administration to humans as well.

## 2 Materials and Methods

## 2.1 Tariquidar Formulations

Two different formulations of tariquidar were tested. Formulation A was obtained by dissolving tariquidar dimesylate (Haoyuan Chemexpress Co., Ltd, Shanghai, PRC) in DMSO and adding heated 5% glucose solution (w/v) to a final DMSO concentration  $\leq 2\%$  (v/v). Formulation B was a microemulsion prepared on the basis of a formulation proposed by Sane et al. [22] which was reported to improve oral bioavailability of the structurally related P-gp/BCRP inhibitor elacridar. In brief, the surfactant Kolliphor® EL (Cremophor EL, Sigma-Aldrich, Austria) and the co-surfactant Carbitol<sup>TM</sup> (diethylene glycol monoethyl ether, Sigma-Aldrich, Austria) were mixed in a 2:1 ratio. After 10 min of heating at 37 °C, the obtained solution was mixed with the medium-chain triglyceride Captex 355 (glyceryl tricaprylate/tricaprate, Abitec, USA) in a 10:1 ratio. Then, tariquidar dimesylate was dissolved in the latter mixture at a concentration of 22.5 mg/mL. This solution was diluted in a 1:3 ratio with deionized water to obtain the desired final tariquidar concentration. Visual inspection of formulation B suggested the formation of a microemulsion, which was not further characterized as this has already been done in detail by Sane et al. [22]. The final concentration of Cremophor EL in formulation B was 0.21 g/mL (w/v).

Both formulations were prepared to give final tariquidar concentrations of 7.5 mg/mL. Aliquots of both tariquidar formulations were snap frozen and stored for a maximum duration of 1 week at  $-80\,^{\circ}$ C immediately after preparation. On the day of scheduled tariquidar administration, the necessary amount of aliquots was thawed and administered after reaching room temperature. Visual appearance of both formulations after thawing was similar to freshly prepared formulations without the presence of precipitates.

#### 2.2 Animals and Study Design

Adult male (10–12 weeks of age and 0.31–0.48 kg of weight, respectively) Sprague–Dawley rats (n = 25,

Charles River, Sulzfeld, Germany) were used. The study protocol was approved by the local Animal Welfare Committee. Experiments were performed at the Institute of Biomedical Research, Medical University of Vienna, in full respect of all applicable institutional, national and international guidelines for the conduct of animal research.

Animals were housed at 22 °C protected from noise and vibration in 12-h day/night cycles and were provided with food and water ad libitum. After 14 days of acclimatization, rats were divided into four groups (n = 5) and treated orally or intraperitoneally with both formulations, respectively. In addition, a fifth group of five animals received one intravenous single dose (formulation A only).

Tariquidar was administered at an individually adapted dosage of 15 mg/kg body weight via gastric gavage, intraperitoneal injection or intravenous bolus injection via the tail vein. Total administered volume did not exceed 0.95, 0.76, and 0.9 mL for intravenous, oral, and intraperitoneal administration, respectively. After each tariquidar administration, sublingual blood draws (blood loss at each timepoint = approximately 120  $\mu$ L) for quantification of tariquidar in plasma were performed at 0.5, 1, 2, 4, 8, and 24-h post-dose in all animals. Blood samples were placed on ice immediately after collection and soon thereafter stored at  $-80\,^{\circ}\text{C}$  until analysis. Intravenous administration of tariquidar and venipuncture for blood sampling was performed under short-duration isoflurane anaesthesia.

# 2.3 Bioanalysis, Pharmacokinetic, and Statistical Analysis

The concentrations of tariquidar in rat plasma were measured with high-performance liquid chromatography-mass spectrometry (LC-MS/MS) using a Dionex "UltiMate 3000" system (Dionex Corp., Sunnyvale, CA, USA) connected with an API 4000 triple-quadrupole mass spectrometer (Applied Biosystems, Concord, Ontario, Canada) according to the literature [23]. Briefly, after the addition of 200 μL of methanol to 100 μL of plasma, the samples were centrifuged (5000×g for 5 min at 4 °C) and 80 µL of the clear supernatant was injected onto the HPLC column. Separation of tariquidar was carried out at 35 °C using a Hypersil BDS-C18 column (5  $\mu$ m, 250  $\times$  4.6 mm I.D., Thermo Fisher Scientific, Inc, Waltham, MA, USA), preceded by a Hypersil BDS-C18 precolumn (5 µm,  $10 \times 4.6$  mm I.D.). The mobile phase consisted of a continuous linear gradient, mixed from 10 mM ammonium acetate/acetic acid buffer, pH 5.0 (mobile phase A) and methanol (mobile phase B). Linear calibration curves were generated by spiking drug-free rat plasma with standard solutions of tariquidar (final concentrations ranging from 0.005 µg to 10 µg/mL; average correlation coefficient:

> 0.999). For this method, the lower limit of detection for tariquidar in plasma was 3 ng/mL and the lower limit of quantification 5 ng/mL. The upper limit of quantification was 5  $\mu$ g/mL. Coefficients of accuracy and precision for this compound were < 11%.

Pharmacokinetic parameters were calculated by means of standard noncompartmental analysis using a commercially available computer program (Kinetica 3.0, Innaphase Corp., Philadelphia, PA, USA). Maximum plasma concentration  $(C_{\text{max}})$ , time to maximum plasma concentration  $(T_{\rm max})$ , and area under the concentration–time curve from 0 to last measured concentration (AUC<sub>0-24</sub>) were calculated from non-fitted data. For AUC calculation, the trapezoidal rule was employed. For accurate description of the pharmacokinetic profile of drugs with long half-lives, the US Food and Drug Administration recommends a sampling period covering 3-5 half-lives of the investigated drug [24]. For practical reasons, sampling in the present study was restricted to 24 h post-dose (i.e., approximately one half-life of tariquidar). Therefore, pharmacokinetic parameters reported here are limited to  $C_{\text{max}}$ ,  $T_{\text{max}}$  and  $AUC_{0-24}$ .

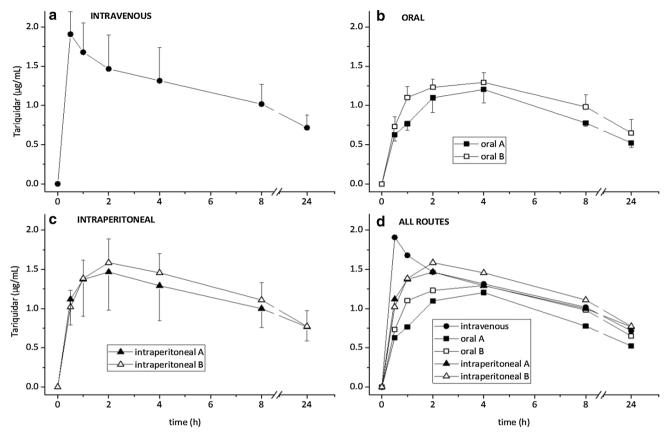
Absolute bioavailability of orally and intraperitoneally administered tariquidar formulations (formulations A and B) was calculated as ratio of the  $AUC_{0-24}$  of the respective formulation/route to the tariquidar  $AUC_{0-24}$  after intravenous administration of formulation A. Statistical analysis was performed using a commercially available program (IBM SPSS Statistics 22; IBM Corp., Armonk, NY, USA). Independent samples Mann–Whitney U test was used to compare key pharmacokinetic parameters between groups. A two-sided p value of 0.05 was considered as threshold for statistical significance. All values are shown as mean  $\pm$  standard deviation unless otherwise stated.

#### 3 Results

The median (range) weight of the animals immediately prior to dosing was 0.35 (0.31–0.38) kg, 0.36 (0.31–0.45) kg, and 0.43 (0.36–0.48) kg for the oral, intraperitoneal, and intravenous route, respectively. Tariquidar was well tolerated by all animals. No immediate drug- or procedure-related complications occurred during intravenous, oral, or intraperitoneal administration of the study drug. Concentration—time curves of both tariquidar formulations in plasma of rats for the three administration routes are shown in Fig. 1.

Thirty minutes after intravenous injection, tariquidar plasma concentration was  $1.91 \pm 0.29 \,\mu\text{g/mL}$  (Fig. 1a).

After oral administration, drug concentrations in plasma reached their maximum at 4 h after dosing for both formulations (Fig. 1b). Overall, tariquidar plasma P. Matzneller et al.



**Fig. 1** Concentration-time profiles (mean  $\pm$  standard deviation) of two tariquidar formulations (A = solution, closed symbols vs. B = microemulsion, open symbols) in plasma of male Sprague-

Dawley rats after intravenous (a), oral (b) and intraperitoneal (c) single doses of 15 mg/kg, respectively. For comparison, panel d shows all routes of administration plotted in one single graph

concentrations were higher for formulation B (the microemulsion). The difference in  $AUC_{0-24}$  between the two formulations was statistically significant, reflecting a marked increase in bioavailability elicited by the microemulsion (Table 1).

After intraperitoneal injection, peak plasma concentrations were reached more rapidly than after oral administration ( $T_{\rm max}=2~{\rm h}$  for both formulations). Plasma concentrations were numerically higher than after oral dosing, but then showed an elimination pattern very much

**Table 1** Key pharmacokinetic parameters of tariquidar in plasma of Sprague–Dawley rats after intravenous (IV), oral (PO) and intraperitoneal (IP) single doses of 15 mg/kg body weight, respectively

Parameter	IV	PO		IP	
		$\overline{A}$	В	A	В
$C_{\text{max}} (\mu \text{g/mL})$	1.9 (0.3)	1.2 (0.2)	1.3 (0.1)	1.5 (0.5)	1.6 (0.3)
$T_{\rm max}$ (h)	0.5 (0.0)	4.0 (0.0)	3.6 (0.9)	$2.0^{b} (0.0)$	$2.0^{\circ} (0.0)$
$AUC_{0-24} (\mu g \cdot h/mL)$	25.2 (5.5)	18.1 (0.8)	21.9 <sup>a</sup> (3.1)	23.8 (6.2)	25.6 (5.1)
F (%)		71.6 (71.3–71.8)	86.3 <sup>a</sup> (85.3–87.2)	91.4 (89.5–93.3)	99.6 (98.0–101.1)

Values shown as means (standard deviation) except for F shown as geometric means (90% confidence interval) of the PO/IP to IV ratios of the tariquidar AUC<sub>0-24</sub>. Formulation A, solution; formulation B, microemulsion

 $C_{max}$  peak plasma concentration,  $T_{max}$  time-to-peak plasma concentration,  $AUC_{0-24}$  area under the concentration-time curve from timepoint zero to timepoint 24 h. F, absolute bioavailability

<sup>&</sup>lt;sup>a</sup>Statistically significant difference to formulation A

<sup>&</sup>lt;sup>b</sup>Statistically significant difference to formulation A oral

<sup>&</sup>lt;sup>c</sup>Statistically significant difference to formulation B oral

comparable to intravenous and oral administration (Fig. 1c). Again, albeit to a minor extent compared to oral dosing, formulation B showed slightly increased exposure compared to formulation A. Pooling both formulations within each route,  $C_{\text{max}}$  and  $AUC_{0-24}$  mean values after intraperitoneal dosing reached approximately 80 and 97% of values after intravenous dosing, respectively. Overall oral bioavailability of tariquidar was slightly poorer, with mean  $C_{\text{max}}$  and AUC<sub>0-24</sub> values following oral dosing (irrespective of formulation) amounting to approximately 65 and 79% of values after intravenous exposure, respectively. However, with the exception of  $AUC_{0-24}$  after oral dosing, none of the differences between the two formulations were statistically significant. Key pharmacokinetic parameters of tariquidar determined in the present study are summarized in Table 1.

#### 4 Discussion

Over the past years, several P-gp inhibitors, which were originally developed as MDR reversal agents, have been characterized with respect to their ability to inhibit P-gp at the BBB. Among the few compounds tested so far in humans, tariquidar was shown to achieve up to fivefold increases in brain distribution of the model P-gp substrates (R)- $[^{11}C]$ verapamil and  $[^{11}C]N$ -desmethyl-loperamide as measured with positron emission tomography (PET) imaging [9, 10]. Such an increase in brain distribution of P-gp substrates can be expected to lead to significant improvements in efficacy of CNS-targeted drugs. However, in these previous studies, tariquidar was administered to humans as a continuous intravenous infusion [9, 10]. For a widespread applicability of tariquidar as a booster of brain distribution of drugs, an oral administration mode would be clearly preferred. One study failed to reveal an effect of orally administered tariquidar on brain distribution of [11C]N-desmethyl-loperamide, which has been attributed to the low oral bioavailability of tariquidar in humans (approximately 12%) [10]. Similarly, while intravenous administration of the structurally related P-gp/BCRP inhibitor elacridar was found to lead to significant P-gp/BCRP inhibition at the non-human primate BBB resulting in a 3.5-fold increase in brain distribution of the dual P-gp/ BCRP substrate [11C]erlotinib [25], oral dosing of elacridar in humans failed to increase brain uptake of [11C]erlotinib [26]. Approaches to increase the oral bioavailability of elacridar have been investigated [22, 27] and have shown promising results in rodents. In the present study, we characterized the pharmacokinetic profile of tariquidar in plasma of rats after intravenous, oral and intraperitoneal administration and tested a microemulsion formulation,

which has been previously applied to enhance the oral bioavailability of elacridar in rodents [22].

Following administration of tariquidar to human healthy volunteers, a second increase of tariquidar plasma concentrations was observed after reaching the initial  $C_{\text{max}}$ . This second peak was ascribed to the enterohepatic circulation of tariquidar [20]. Interestingly, no such phenomenon was observed in any of the concentration-time profiles in the present study in rats, which might point to species-related differences in metabolism and excretion routes. Among other factors, the impact of enterohepatic circulation of a drug on its plasma concentration depends on rate and extent of breakdown of glucuronidated or otherwise conjugated compound by intestinal microbiota [28]. Substantial compositional diversity of the gut microbiota has been reported previously [29]. After all three administration routes, plasma concentrations of tariquidar remained for 24 h above its previously determined in vivo half-maximum effect concentration (EC<sub>50</sub>) to increase brain uptake of (R)-[11C]verapamil in rats  $(545 \pm 30 \text{ ng/mL})$  [30]. This suggests that substantial inhibition of P-gp will occur at the BBB and most likely in other P-gp expressing organs of rats (e.g., liver and kidneys). Interestingly, the previous studies found species differences in P-gp inhibition at the human BBB as compared with the rodent BBB with an approximately threefold higher in vivo EC50 value determined for tariquidar in humans (EC<sub>50</sub> = 1454 ng/mL) [9]. This suggests that higher tariquidar plasma concentrations are required in humans as compared with rats to overcome P-gp-mediated efflux transport at the BBB.

A noteworthy observation made in our study was the good oral bioavailability of tariquidar in rats achieved with both tested formulations (72-87%), which was substantially higher than previously reported in humans (12%) [10]. Absorption of tariquidar from the intestinal lumen may be restricted by efflux transporters expressed in intestinal epithelial cells, i.e., P-gp and BCRP, for which tariquidar was found to be a substrate [15, 17, 31]. However, tariquidar is a potent inhibitor of P-gp and is also reported to inhibit BCRP [31]. The in vitro EC<sub>50</sub> of tariquidar for inhibition of its own transport by human BCRP was approximately 20-fold higher than the EC<sub>50</sub> for inhibition of its own transport by human P-gp (EC<sub>50</sub>: 201 ng/ mL for BCRP vs. 11 ng/mL for P-gp) [15]. Values for total intestinal fluid volume reported in the literature range from approximately 100 to 750 mL in adult humans and from approximately 3 to 8 mL in average-weight rats [32-35]. Accordingly, intestinal concentrations of tariquidar in humans after a 1500 mg oral dose assuming the maximal intestinal fluid volume reported in the literature would be in the range of 2000 ng/mL. Using a median oral dose of 5.3 mg and adjusting volumes from the literature to a

median body weight of 0.35 kg, resulting intestinal concentrations of tariquidar in orally dosed rats in the present study would at best reach values of 1000 ng/mL, i.e., half of those in humans. Both values are several folds above the reported in vitro EC<sub>50</sub> values of tariquidar for inhibition of its own transport by human P-gp and BCRP, which suggests that intestinal transport by P-gp and BCRP most likely did not contribute to the low oral bioavailability of tariquidar observed in humans. If efflux transport is not involved, then species-related differences in intestinal or hepatic first-pass metabolism might play a role. Tariquidar is extensively metabolized in the liver and its metabolites are excreted into bile [18]. In rats and cynomolgus monkeys, phase I metabolism followed three major pathways: O-demethylation, N-dealkylation, and dehydrogenation. In addition, phase II metabolism occurred via O-glucuronidation. The majority of drug-related material recovered from faeces in rats and monkeys after intravenous administration of [14C]tariquidar was in the form of metabolites [18]. The proportion of parent compound to metabolites in faeces was lower in the monkey than in the rat, indicating that tariquidar was more extensively metabolized in the monkey [18]. In contrast, in plasma, most (> 95%) of the tariquidar-related material existed as the unchanged parent compound [18]. This indicates that tariquidar metabolites, whether formed in the liver or in the intestine, do not distribute into blood. In addition, in humans, no tariquidar metabolites were found in plasma after intravenous administration of the compound [14, 20]. However, no information is currently available regarding the proportion of metabolites relative to parent compound excreted in humans into faeces. If intestinal or hepatic firstpass metabolism of tariquidar occurs in both rats and humans, species differences in metabolism may explain differences in oral bioavailabilities between rats and humans.

Discrepancies in the bioavailability of drugs between rats and humans have been described before [36]. Of all processes that determine systemic concentrations of a drug, those governing intestinal absorptions seem to correlate reasonably well between the two species [36]. Given the drug's lipophilic character, the ability of tariquidar to penetrate into enterocytes may be similar between rats and humans. However, when administered orally, lipophilic drugs often face the problem of poor water solubility. The latter is determined by various factors including most notably pH and composition of gastrointestinal fluids. While we are not aware of essential differences in physicochemical properties of intestinal fluid between rats and humans, it can be assumed that feeding status at the time of dosing might have an impact on oral bioavailability. Interestingly, the presence of bile salts, which is typically increased in the post-prandial phase [37], is known to enhance the solubilisation and oral absorption of hydrophobic compounds [38]. The animals in the present study had no restrictions regarding intake of food and water during the study, whereas the two studies reporting oral administration of tariquidar in humans do not specify feeding status of the subjects at the time of dosing [10, 21]. Assuming that they were fasted, which seems more likely, one could hypothesize that a poor bile salt content of the intestinal fluid might have determined the poor bioavailability observed in humans opposed to ad libitum-fed rats. Of equal relevance, the impact of the tariquidar formulation should not be ignored. In both studies reporting oral dosing of tariquidar in humans, tariquidar was administered in the form of capsules containing the free base [10, 21], whereas in our study, the dimesylate salt was administered as a solution or microemulsion, which might have facilitated intestinal absorption of the drug.

In the present study, we tested a tariquidar microemulsion for oral and intraperitoneal administration, which has been previously developed and extensively characterized by Sane et al. [22]. The oral bioavailability of elacridar in FVB mice after microemulsion dosing was reported as 47% [22], which was approximately twofold higher as compared with administration of an oral suspension. In addition, the microemulsion formulation was found to retain the P-gp/ Bcrp inhibitory effect of elacridar at the mouse BBB. In our study, the standard formulation already had an oral bioavailability of 71.6% and the microemulsion led to a statistically significant improvement in oral bioavailability to 86.9%. This may indicate that tariquidar has a better solubility and permeability across the intestinal mucosa than elacridar. Despite the small sample size of the animal collectives used in the present study, the good tolerability and at least partially significant evidence pointing to a higher bioavailability might justify consideration of the microemulsion as a promising formulation for further rodent studies involving oral and intraperitoneal administration of tariquidar. Moreover, the reported microemulsion may form the basis for the future development of a tariquidar formulation with good oral bioavailability for human use, which could greatly facilitate the utility of tariquidar as a booster of brain distribution of CNS-targeted P-gp substrate drugs.

### 5 Conclusion

In conclusion, the present project has for the first time described pharmacokinetic profiles of tariquidar in rat plasma after intravenous, oral, and intraperitoneal administration. The good oral bioavailability of tariquidar shown in rats stands in contrast to very low systemic concentrations reported after oral dosing in humans, which

constitutes an obstacle to a broader use of the compound as a booster of brain delivery of drugs which are excluded from the brain by P-gp. Possible reasons for this discrepancy have been discussed here, but remain to be fully elucidated. This study has confirmed that the interplay of sometimes conflicting pharmacokinetic subprocesses makes prediction of oral bioavailability of a drug in humans based on rat data challenging. Out of two formulations tested for oral and intraperitoneal dosing, the microemulsion showed improved bioavailability and might be considered for further investigations. The present findings extend the available information on tariquidar and provide a basis for future studies involving oral administration of this compound.

Data Availability Statement The data sets generated during and/or analysed during the current study are available from the corresponding author on reasonable request.

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#### **Compliance with Ethical Standards**

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**Conflicts of interest** Disclosure of potential conflicts of interest: PM, MK, SE, AMS, WJ, MB, OL, MZ, and WP declare that they have no conflicts of interest.

**Ethics approval** The study protocol was approved by the local Animal Welfare Committee. All applicable international, national, and/or institutional guidelines for the care and use of animals were followed.

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