



Complete Genome Sequence of Solvent-Tolerant *Clostridium* beijerinckii Strain SA-1

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We report the complete genome sequence of *Clostridium beijerinckii* SA-1, derived by directed evolution from *C. beijerinckii* NCIMB 8052, selecting for enhanced solvent tolerance. This sequence allows for accurate placement of SA-1 as *C. beijerinckii*, permits functional analyses of mutant phenotypes, and suggests methods for distinguishing SA-1 from its parent.

Received 7 November 2014 Accepted 10 November 2014 Published 18 December 2014

Citation Noar J, Makwana ST, Bruno-Bárcena JM. 2014. Complete genome sequence of solvent-tolerant *Clostridium beijerinckii* strain SA-1. Genome Announc. 2(6):e01310-14. doi:10.1128/genomeA.01310-14.

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C*lostridium beijerinckii* is a Gram-positive, obligately anaerobic solventogenic organism, of interest primarily for acetone-butanol-ethanol (ABE) fermentations. Taxonomic assignments of solventogenic species of *Clostridium* historically have experienced some turmoil, and strains originally classified as one species have often been reassigned to another (1).

In 1983, using stress-directed evolution with a series of increasing butanol concentrations, Lin and Blaschek isolated an especially butanol-resistant strain of *Clostridium*, originally designated an offspring of *C. acetobutylicum* ATCC 824 (2). This strain was named SA-1, deposited, and catalogued as ATCC 35702; later it was reclassified as a strain of *Clostridium beijerinckii*, an offspring of *C. beijerinckii* NCIMB 8052 (1). SA-1's nutritional requirements were determined previously, and butanol resistance and delayed sporulation phenotypes have been observed (3). Other studies on SA-1 have evaluated its extracellular alpha-amylase and glucoamylase activity and the effects of butanol stress on its membrane fatty acid composition (4, 5). Reportedly, SA-1 is genetically malleable (6).

Sequencing the genome of *C. beijerinckii* SA-1 allows for definitive placement of this strain within the *C. beijerinckii* species. Identification of variations in SA-1's sequence from that of its parent can guide functional physiological studies to ascertain the extent of SA-1's butanol tolerance and solventogenic capacities, as well as other mutant phenotypes (3, 7); alternatively, such a project allows for the identification of errors in the original sequence of *C. beijerinckii* NCIMB 8052 (7). Finally, discovery of large variations between strains suggests a simple molecular method for distinguishing them from each other (7).

The genome sequence of *C. beijerinckii* SA-1 was determined using the Illumina Genome Analyzer IIx platform by the US Department of Energy Joint Genome Institute. Reads were assembled by comparison to the sequence of parent strain *C. beijerinckii* NCIMB 8052 (GenBank accession no. NC_009617), and algorithms MAQ (8) and BreakDancer (9) were used to find small and large variations from the reference sequence. PCR and Sanger dyeterminator sequencing (Eton Bioscience, Research Triangle Park, NC) were used to validate variations found *in silico*. Annotations were copied from the GenBank record of the genome of NCIMB 8052 and modified as necessary.

Nucleotide sequence accession number. The genome sequence of *C. beijerinckii* strain SA-1 (ATCC 35702) has been deposited in DDBJ/EMBL/GenBank under the accession no. CP006777. The version described in this paper is the first version.

ACKNOWLEDGMENTS

This work was partially supported by the College of Life Sciences at NC State University, the North Carolina Agricultural Research Service, and the Biofuels Center of North Carolina. The work conducted by the U.S. DOE JGI was supported by the Office of Science of the U.S. Department of Energy under contract no. DE-AC02-05CH11231. J.N. was supported under an NSF Graduate Research Fellowship.

REFERENCES

- Johnson JL, Toth J, Santiwatanakul S, Chen J-S. 1997. Cultures of "Clostridium acetobutylicum" from various collections comprise Clostridium acetobutylicum, Clostridium beijerinckii, and two other distinct types based on DNA-DNA reassociation. Int. J. Syst. Bacteriol. 47:420–424. http:// dx.doi.org/10.1099/00207713-47-2-420.
- Lin Y-L, Blaschek HP. 1983. Butanol production by a butanol-tolerant strain of *Clostridium acetobutylicum* in extruded corn broth. Appl. Environ. Microbiol. 45:966–973.
- Heluane H, Evans MR, Dagher SF, Bruno-Bárcena JM. 2011. Metaanalysis and functional validation of nutritional requirements of solventogenic clostridia growing under butanol stress conditions and coutilization of D-glucose and D-xylose. Appl. Environ. Microbiol. 77:4473–4485. http:// dx.doi.org/10.1128/AEM.00116-11.
- Matoi S. 1986. Effect of butanol challenge on the lipid composition of butanol-tolerant *Clostridium acetobutylicum* SA-1, poster 86, p 267. Abstr 86th Annu Meet Am Soc Microbiol 1986. American Society for Microbiology, Washington, DC.
- Chojecki A, Blaschek HP. 1986. Effect of carbohydrate source on alphaamylase and glucoamylase formation by *Clostridium acetobutylicum* SA-1. J. Ind. Microbiol. 1:63–67. http://dx.doi.org/10.1007/BF01569418.

- Lin Y-L, Blaschek HP. 1984. Transformation of heat-treated *Clostridium* acetobutylicum protoplasts with pUB110 plasmid DNA. Appl. Environ. Microbiol. 48:737–742.
- Sandoval-Espinola WJ, Makwana ST, Chinn MS, Thon MR, Azcárate-Peril MA, Bruno-Bárcena JM. 2013. Comparative phenotypic analysis and genome sequence of *Clostridium beijerinckii* SA-1, an offspring of NCIMB 8052. Microbiology 159:2558–2570. http://dx.doi.org/10.1099/ mic.0.069534-0.
- 8. Li H, Ruan J, Durbin R. 2008. Mapping short DNA sequencing reads and calling variants using mapping quality scores. Genome Res. 18:1851–1858. http://dx.doi.org/10.1101/gr.078212.108.
- Chen K, Wallis JW, McLellan MD, Larson DE, Kalicki JM, Pohl CS, McGrath SD, Wendl MC, Zhang Q, Locke DP, Shi X, Fulton RS, Ley TJ, Wilson RK, Ding L, Mardis ER. 2009. BreakDancer: an algorithm for high-resolution mapping of genomic structural variation. Nat. Methods 6:677–681. http://dx.doi.org/10.1038/nmeth.1363.