Association of *HOTAIR* expression in gastric carcinoma with invasion and distant metastasis

Elaheh Emadi-Andani[§], Parvaneh Nikpour^{1,§}, Modjtaba Emadi-Baygi², Ali Bidmeshkipour

Department of Biology, Faculty of Sciences, Razi University, Kermanshah, ¹Department of Genetics and Molecular Biology, Faculty of Medicine, Pediatric Inherited Diseases Research Center, Child Growth and Development Research Center, Isfahan University of Medical Sciences, Isfahan, ²Department of Genetics, Faculty of Basic Sciences, Research Institute of Biotechnology, Shahrekord University, Shahrekord, Iran

[§]These two authors contributed equally to this work.

Abstract

Background: Gastric cancer is the second and fourth most common cancer in Iranian men and women, respectively, but it is the first leading cause of cancer deaths in Iran. Most Iranian patients with gastric cancer are diagnosed at an advanced stage of disease when the conventional treatments have no effect on improving the survival. So, early gastric cancer detection is of high priority in order to decrease its high mortality rate in Iran. *HOTAIR* is a long non-coding RNA which its overexpression has been documented in different types of human cancer and can be considered as a potential cancer biomarker. The aim of this study was to evaluate the clinicopathological relevance of the expression of *HOTAIR* gene in gastric carcinoma.

Materials and Methods: A total of 60 tumoral and non-tumoral gastric specimens were evaluated for *HOTAIR* gene expression using quantitative real-time PCR.

Results: The expression of *HOTAIR* was markedly increased in gastric cancer tissues compared with adjacent non-tumoral tissues. We further showed that there was a positive significant correlation between the *HOTAIR* gene expression, TNM staging, perineural invasion, and distant metastasis, but not with other clinicopathological features of gastric tumors.

Conclusions: These results suggest that *HOTAIR* expression is modulated during gastric cancer progression and therefore may participate in molecular processes relevant to malignant transformation and metastasis in gastric carcinoma.

Key Words: Gastric carcinoma, gene expression, HOTAIR, invasion, metastasis

Address for correspondence:

Dr. Parvaneh Nikpour, Department of Genetics and Molecular Biology, Faculty of Medicine, Isfahan University of Medical Sciences, Isfahan, Iran. E-mail: pnikpour@med.mui.ac.ir Received: 13.08.2013, Accepted: 14.09.2013

Access this article online			
Quick Response Code:	Website: www.advbiores.net		
	DOI: 10.4103/2277-9175.133278		

INTRODUCTION

Gastric cancer is the second most common cancer worldwide and the second leading cause of cancer mortality.^[1] Gastric cancer is the second and fourth most common cancer in Iranian men and women, respectively, but it is the first leading cause of cancer deaths in Iran.^[2] The main environmental factors

Copyright: © 2014 Emadi-Andani. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

How to cite this article: Emadi-Andani E, Nikpour P, Emadi-Baygi M, Bidmeshkipour A. Association of HOTAIR expression in gastric carcinoma with invasion and distant metastasis. Adv Biomed Res 2014;3:135.

for the high incidence of gastric cancer in Iran are *H. pylori* infection, high intake of salt, and smoking. Most Iranian patients with gastric cancer are diagnosed at an advanced stage of disease when the conventional treatments have no effect on improving the survival. So, early gastric cancer detection is of high priority in order to decrease its high mortality rate in Iran.^[3]

A large part of the genome is transcribed for non-coding RNAs (ncRNAs) with various biological functions.^[4] Long non-coding RNAs (lncRNAs) have a length of >200 nucleotides, with direct effect on the transcription or employing histone modification complexes regulating expression of different genes.^[5,6] Hox transcript antisense intergenic RNA (HOTAIR) is a lncRNA with 2158 nucleotides in length which is expressed from HOXC locus on 12 chromosome.^[7] It interacts with Polycomb Repressive Complex 2 (PRC2) and causes targeting it to the HOXD locus.^[7,8] Several studies display an active role for *HOTAIR* in cancers of the breast,^[9,10] colorectal,^[11] hepatocellular,^[12-14] pancreatic,^[15] gastrointestinal stromal tumors,^[16] nasopharyngeal carcinoma (NPC),^[17] laryngeal squamous cell carcinoma (LSCC),^[18] and sarcoma.^[19] In all studies, increased expression of HOTAIR is associated with malignant progression and poor survival. Hence, HOTAIR may be considered as a potential target for diagnosis and treatment of various cancer types.^[9-19]

Based on these findings, we tested if *HOTAIR* also shows a similar pattern in gastric cancer. To this aim, we evaluated *HOTAIR* expression in tumoral and non-tumoral gastric tissue samples by using quantitative real-time RT-PCR. Our results demonstrated that the expression of *HOTAIR* was markedly increased in gastric cancer tissues compared with adjacent normal tissues. We further showed that there was a positive correlation between the *HOTAIR* gene expression, TNM (T, N, and M stand for tumor, lymph node, and metastasis, respectively) staging, perineural invasion, and distant metastasis, but not with other clinicopathological features of gastric tumors.

MATERIALS AND METHODS

Tumor and non-tumor tissues

All experimental procedures were approved by the Ethics Committee of Isfahan University of Medical Sciences. A total of 60 specimens of gastric cancer tissues and adjacent benign tissues (paired) were obtained from the Iran Tumoral Bank (Tehran, Iran) as described previously.^[20-22] All patients provided written informed consent to the Iran Tumoral Bank prior to the participation.

Total RNA isolation and cDNA synthesis

Total RNA was extracted from gastric cancer tissues using Qiazol reagent (Qiagen, Hilden, Germany) according to the manufacturer's protocol. The integrity of RNA was verified by electrophoresis on a 1% agarose gel stained with ethidium bromide. The quality and quantity of RNA were determined by ultraviolet spectroscopy. cDNA was synthesized using random hexamer primers and an M-MLV reverse transcriptase (Fermentas, Vilnius, Lithuania) as described elsewhere.^[20-22]

Quantitative real-time PCR

The expression level of HOTAIR gene was determined by quantitative real-time RT-PCR with TBP (TATA box binding protein) as a reference gene.^[23] The primers for the target gene were as follows: 5'-AGACGGCGGCGAGAGGAA-3' and 5 '-CTGAAATGGAGGACCGGCG-3' with an amplicon size of 126 bp. PCR was performed using Maxima[™] SYBR Green/ROX qPCR Master MIX (Fermentas, Vilnius, Lithuania) and an Applied Biosystems StepOnePlus[™] instrument. The PCR amplification conditions consisted of an initial denaturation at 95°C for 10 minutes, 40 cycles of denaturation at 95°C for 15 seconds, annealing at 55°C and 58°C for TBP and HOTAIR genes, respectively, then extension for 15 seconds at 72°C. All samples were measured in triplicate. The 2^{-ΔΔCt} method was used to quantify the relative levels of gene expression.

Statistical analysis

All data are expressed as means \pm standard error of mean (SEM) from at least three separate experiments. Statistical analyses were performed using SPSS version 16.0. Differences between groups were analyzed using a paired t-test or one-way ANOVA with post hoc multiple comparisons. Statistical significance was defined as $P \leq 0.05$.

RESULTS

Optimization of PCR amplification

In order to obtain a specific amplicon for *HOTAIR*, both conventional and real-time PCR was done with a temperature gradient. As it was shown in a previous study that *HOTAIR* is expressed in MCF-7 breast cancer cell line,^[9] we used the cDNA of this cell line for optimization procedures. Gel electrophoresis of amplified product of *HOTAIR* with the designed primers showed a specific band with expected size (126 bp) in 58°C [Figure 1]. Analysis of melting curves of real-time PCR also showed a unique melting curve for this amplicon without primer dimmers (data not shown).



Figure 1: Optimization of PCR conditions for *HOTAIR* gene expression. RT-PCR products were separated on agarose gel electrophoresis as follows: lane 1: DNA size marker, lane 2: Non-template control (adding no cDNA), and lane 3: adding cDNA of MCF-7 cell line

HOTAIR gene expression in tumoral and non-tumoral gastric specimens

To examine the expression of *HOTAIR* in human gastric tissues, cDNA was synthesized for all samples and real-time PCR was performed using specific primers for both *HOTAIR* and *TBP* genes. The relative expression of *HOTAIR* showed statistically significant overexpression in pooled tumoral specimens compared to the paired non-tumoral samples [Figure 2, P = 0.028].

Correlation of *HOTAIR* expression with clinicopathological features in gastric carcinoma

In order to examine the clinical importance of the *HOTAIR* overexpression, the correlation between clinicopathological status of gastric tumor samples and level of *HOTAIR* expression was investigated. Analyses showed a significant and positive association between the expression levels of *HOTAIR*, TNM staging, perineural invasion, and distant metastasis. A trend was also evident toward the same pattern for lymph node metastasis (N classification) and invasion depth although not reaching statistical significance. No significant correlation was found for other clinicopathological features of gastric tumors [Table 1].

DISCUSSION

Our study showed for the first time that *HOTAIR* expression is significantly correlated with perineural invasion and distant metastasis in gastric cancer. An increased *HOTAIR* expression in gastric carcinomas compared to their non-tumoral adjacent tissues was also documented.

Until now, altered expression of *HOTAIR* gene has been documented in different types of human cancer. Gupta *et al.*^[9] showed increased expression of *HOTAIR* in primary breast tumors as well as metastases. They showed that high expression of *HOTAIR* is an independent prognostic factor for metastasis and death in breast cancer patients. In another study by Milhem *et al.*,^[19] increased expression of *HOTAIR* Advanced Biomedical Research | 2014



Figure 2: The relative expression levels of *HOTAIR* in tumoral vs non-tumoral gastric samples. Error bars represent standard error of mean (SEM) and the asterisk represents a statistically significant difference ($p \le 0.05$)

Table 1: Relationship between HOTAIR expression le	evels and
clinicopathological parameters of tumoral gastric s	samples

Characteristics	Numbers (%)	HOTAIR relative expression (mean±SEM*)	<i>P</i> value
Sex			0.13
Male	18 (60)	0.15±0.05	
Female	12 (40)	0.51±0.28	
Age (years)			0.31
≥70	15 (50)	0.36±0.23	
<70	15 (50)	0.23±0.09	
Depth of invasion			0.07
T1-T2	3 (10)	0.11±0.06	
T3-T4	27 (90)	0.31±0.13	
N classification			0.06
NO	9 (30)	0.12±0.03	
N1-N3	21 (70)	0.37±0.17	
M classification			0.04
Mx	15 (50)	0.20±0.07	
MO	11 (36.6)	0.19±0.08	
M1	4 (13.3)	0.94±0.36	
TNM stage			0.03
I-III	26 (86.7)	0.19±0.05	
IV	4 (13.3)	0.94±0.36	
Perineural invasion	× ,		0.04
Negative	6 (20)	0.08±0.03	
Positive	24 (80)	0.35±0.11	
Lymphatic invasion	()		0.11
Negative	16 (55.17)	0.14±0.04	
Positive	13 (44.82)	0.50±0.27	
Tumor size (cm)	()		0.45
≥5	20 (68.7)	0.31±0.17	
<5	9 (31. 3)	0.29±0.14	
Tumor grades	()		0.21
I	10 (33.3)	0.47±0.34	
	8 (26.6)	0.21±0.12	
III	12 (40)	0.20±0.09	
Tumor types	· · /		0.16
Diffuse	15 (50)	0.17±0.07	
Intestinal	15 (50)	0.42±0.23	

*SEM: Standard error of mean

was detected in most of the primary and metastatic sarcoma patient tumor samples in such a way that its expression was correlated with the likelihood of metastasis in primary sarcoma tumors. In 2011, Geng *et al.* measured *HOTAIR* gene expression in hepatocellular carcinoma (HCC) tissues and showed that the expression of *HOTAIR* is significantly higher in tumoral vs. non-tumoral HCC tissue samples. Furthermore, they reported a positive correlation between the HOTAIR gene expression and lymph node metastasis. However, they did not observe any significant correlation between HOTAIR expression levels and other clinicopathological features of patients like age, gender, tumor size, tumor number, and portal invasion.^[13] Kogo *et al.* also showed *HOTAIR* overexpression in colorectal cancerous tissues compared to the noncancerous ones. They divided their patients into two groups, one with high and one with the low *HOTAIR* expression. They found a strong correlation between HOTAIR expression and histological grade, depth of tumor and liver metastasis but not with age, gender, lymph node metastasis, lymphatic or venous invasion. The group with higher HOTAIR expression had also poorer prognosis.^[11] In another study of HCC patients, HOTAIR expression level was shown to be higher in tumor tissues than their adjacent non-tumoral ones. Furthermore, the higher level of HOTAIR was linked to shorter survival and more probability of recurrence after liver transplantation.^[12] In contrast to the other previous reports, a study by Lu et al. in 2012 demonstrated that high levels of *HOTAIR* are correlated with the lower chance of recurrence and mortality in 348 examined tissues from patients with breast cancer. They also found no significant correlation between HOTAIR levels and various pathological and clinical features of breast cancer samples like tumor stage, grade, histological type, tumor size, and nodal status.^[24] In 2012, a study by Chisholm et al.^[10] was performed to analyze HOTAIR gene expression in formalin-fixed paraffin-embedded breast cancer tissues. They showed a positive correlation between HOTAIR levels and ER/PR (estrogen receptor/progesterone receptor) positivity and also with worse survival rates. In 2013, overexpression of HOTAIR was also shown in LSCC tissue samples compared to their adjacent non-tumoral tissues. In that study, a positive association between HOTAIR gene expression levels and T classification, neck nodal metastasis, and clinical stage was reported.^[18] Nie et al. examined HOTAIR gene expression using in situ hybridization and real-time PCR in NPC samples. Similar to other studies, they also showed increased expression of HOTAIR in cancerous samples in comparison to non-cancerous tissues. They also reported a positive association of HOTAIR gene expression with tumor

size, clinical stage, lymph node tumor burden, and distant metastasis.^[17]

Nearly all studies that have examined the HOTAIR expression levels in tumoral and non-tumoral samples of different cancer types have reported HOTAIR elevated levels in cancerous specimens.^[9-19] Taken together our data are in accord with other studies^[9-19] that showed HOTAIR expression increased significantly in cancerous tissues. We also observed a positive significant correlation between HOTAIR expression, staging, invasion, and metastasis; in a same way to the other studies.^[9-11,13,17-19] Moreover, a trend was also evident toward the same pattern for lymph node metastasis (N classification) and invasion depth although not reaching statistical significance. This is consistent with findings of previous studies.^[11,13,17,18] However, we did not observe a significant association between the target gene expression and other clinicopathological parameters, like age, gender, tumor size, grades, histological types, and lymphatic invasion consistent with other studies.^[11-13,17,18,24] Finally, during processing of the manuscript, two relevant studies appeared in the Pubmed in which they also showed that HOTAIR is overexpressed in gastric cancer and is associated with TNM staging, lymph node metastasis, and poor overall survival.^[25,26]

In summary, this is the first study showing a positive and significant correlation between *HOTAIR* gene expression profile, perineural invasion, and distant metastasis in gastric tumor samples. These results suggest that *HOTAIR* expression is modulated during gastric cancer progression and therefore may participate in molecular processes relevant to malignant transformation and metastasis in gastric carcinoma.

REFERENCES

- Gomceli I, Demiriz B, Tez M. Gastric carcinogenesis. World J Gastroenterol 2012;18:5164-70.
- Mousavi SM, Gouya MM, Ramazani R, Davanlou M, Hajsadeghi N, Seddighi Z. Cancer incidence and mortality in Iran. Ann Oncol 2009;20:556-63.
- Malekzadeh R, Derakhshan MH, Malekzadeh Z. Gastric cancer in Iran: Epidemiology and risk factors. Arch Iran Med 2009;12:576-83.
- Pan YF, Feng L, Zhang XQ, Song LJ, Liang HX, Li ZQ, *et al.* Role of long non-coding RNAs in gene regulation and oncogenesis. Chin Med J (Engl) 2011;124:2378-83.
- Kurokawa R. Long noncoding RNA as a regulator for transcription. Prog Mol Subcell Biol 2011;51:29-41.
- De Lucia F, Dean C. Long non-coding RNAs and chromatin regulation. Curr Opin Plant Biol 2011;14:168-73.
- Rinn JL, Kertesz M, Wang JK, Squazzo SL, Xu X, Brugmann SA, et al. Functional demarcation of active and silent chromatin domains in human HOX Loci by Noncoding RNAs. Cell 2007;129:1311-23.
- Tsai MC, Manor O, Wan Y, Mosammaparast N, Wang JK, Lan F, et al. Long noncoding RNA as modular scaffold of histone modification complexes. Science 2010;329:689-93.

- Gupta RA, Shah N, Wang KC, Kim J, Horlings HM, Wong DJ, et al. Long non-coding RNA HOTAIR reprograms chromatin state to promote cancer metastasis. Nature 2010;464:1071-6.
- Chisholm KM, Wan Y, Li R, Montgomery KD, Chang HY, West RB. Detection of long non-coding RNA in archival tissue: Correlation with polycomb protein expression in primary and metastatic breast carcinoma. PLoS One 2012;7:e47998.
- Kogo R, Shimamura T, Mimori K, Kawahara K, Imoto S, Sudo T, *et al.* Long Noncoding RNA HOTAIR Regulates Polycomb-Dependent Chromatin Modification and Is Associated with Poor Prognosis in Colorectal Cancers. Cancer Res 2011;71:6320-6.
- Yang Z, Zhou L, Wu LM, Lai MC, Xie HY, Zhang F, et al. Overexpression of long non-coding RNA HOTAIR predicts tumor recurrence in hepatocellular carcinoma patients following liver transplantation. Ann Surg Oncol 2011;18:1243-50.
- Geng YJ, Xie SL, Li Q, Ma J, Wang GY. Large intervening non-coding RNA HOTAIR is associated with hepatocellular carcinoma progression. J Int Med Res 2011;39:2119-28.
- 14. Ishibashi M, Kogo R, Shibata K, Sawada G, Takahashi Y, Kurashige J, *et al.* Clinical significance of the expression of long non-coding RNA HOTAIR in primary hepatocellular carcinoma. Oncol Rep 2013;29:946-50.
- Kim K, Jutooru I, Chadalapaka G, Johnson G, Frank J, Burghardt R, *et al.* HOTAIR is a negative prognostic factor and exhibits pro-oncogenic activity in pancreatic cancer. Oncogene 2013;32:1616-25.
- Niinuma T, Suzuki H, Nojima M, Nosho K, Yamamoto H, Takamaru H, *et al*. Upregulation of miR-196a and HOTAIR drive malignant character in gastrointestinal stromal tumors. Cancer Res 2012;72:1126-36.
- Nie Y, Liu X, Qu S, Song E, Zou H, Gong C. Long non-coding RNA HOTAIR is an independent prognostic marker for nasopharyngeal carcinoma progression and survival. Cancer Sci 2013;104:458-6.
- Li D, Feng J, Wu T, Wang Y, Sun Y, Ren J, *et al.* Long Intergenic Noncoding RNA HOTAIR Is Overexpressed and Regulates PTEN Methylation in Laryngeal Squamous Cell Carcinoma. Am J Pathol 2013;182:64-70.

- Milhem MM, Knutson T, Yang S, Zhu D, Wang X, Leslie KK, et al. Correlation of MTDH/AEG-1 and HOTAIR Expression with Metastasis and Response to Treatment in Sarcoma Patients. J Cancer Sci Ther 2012;S5. Pii:004.
- Nikpour P, Emadi-Baygi M, Mohhamad-Hashem F, Maracy MR, Haghjooy-Javanmard S. MSI1 overexpression in diffuse type of gastric cancer. Pathol Res Pract 2013;209:10-3.
- Nikpour P, Emadi-Baygi M, Mohammad-Hashem F, Maracy MR, Haghjooy-Javanmard S. Differential expression of ZFX gene in gastric cancer. J Biosci 2012;37:85-90.
- Baygi ME, Nikpour P. Deregulation of MTDH gene expression in gastric cancer. Asian Pac J Cancer Prev 2012;13:2833-6.
- Nikpour P, Baygi ME, Steinhoff C, Hader C, Luca AC, Mowla SJ, et al. The RNA binding protein Musashi1 regulates apoptosis, gene expression and stress granule formation in urothelial carcinoma cells. J Cell Mol Med 2011;15:1210-24.
- Lu L, Zhu G, Zhang C, Deng Q, Katsaros D, Mayne ST, *et al.* Association of large noncoding RNA HOTAIR expression and its downstream intergenic CpG island methylation with survival in breast cancer. Breast Cancer Res Treat 2012;136:875-83.
- Hajjari M, Behmanesh M, Sadeghizadeh M, Zeinoddini M. Up-regulation of HOTAIR long non-coding RNA in human gastric adenocarcinoma tissues. Med Oncol 2013;30:670.
- Xu ZY, Yu QM, Du YA, Yang LT, Dong RZ, Huang L, *et al.* Knockdown of Long Non-coding RNA HOTAIR Suppresses Tumor Invasion and Reverses Epithelial-mesenchymal Transition in Gastric Cancer. Int J Biol Sci 2013;9:587-97.

Source of Support: The study was supported in part by a research grant from Isfahan University of Medical Sciences, Isfahan and Razi University, Kermanshah, Iran, **Conflict of Interest:** None declared.