

# Protective Effect of *Pluchea lanceolata* against Aluminum Chloride-induced Neurotoxicity in Swiss Albino Mice

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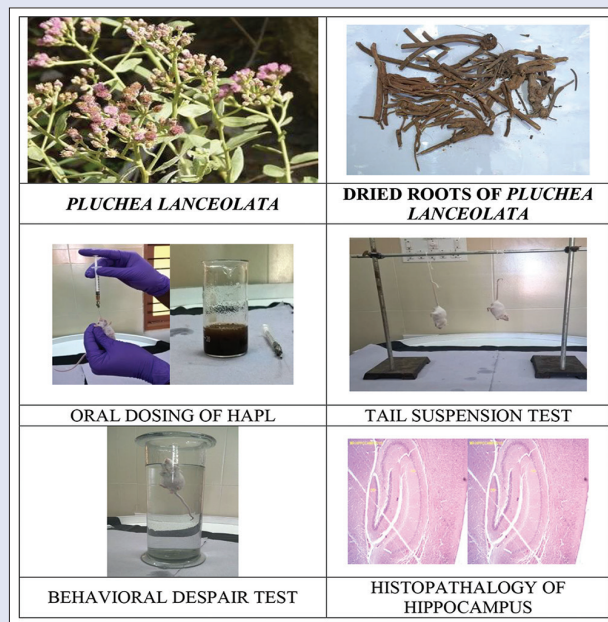
## ABSTRACT

**Background:** Aluminum chloride (AlCl<sub>3</sub>) is a known potent environmental neurotoxin causing progressive neurodegenerative changes in the brain. The herb *Pluchea lanceolata* is commonly known as "Rasana" and used as a nerve tonic in neuroinflammatory conditions in Indian system of medicine. **Objective:** To evaluate the neuroprotective activity of hydroalcoholic extract of *P. lanceolata* in chronic AlCl<sub>3</sub>-induced neurotoxicity in Swiss albino mice. **Materials and Methods:** Albino mice were categorized into four different groups; Group 1 served as vehicle control, Group 2 mice were administered with AlCl<sub>3</sub>, 40 mg/kg body weight by intraperitoneal route for 45 consecutive days. Groups 3 and 4 mice were administered with AlCl<sub>3</sub>, 40 mg/kg body weight intraperitoneal for 45 consecutive days along with hydroalcoholic extract of *P. lanceolata* at 200 and 400 mg/kg body weight. **Results:** Chronic administration of AlCl<sub>3</sub> resulted in behavioral deficits, triggered lipid peroxidation, increased acetylcholinesterase (AChE) activity, and histological alterations. Co-administration of hydroalcoholic extract of *P. lanceolata* attenuated many of the AlCl<sub>3</sub>-induced alterations such as behavioral, lipid peroxidation, AChE, and histological changes of brain tissue. **Conclusion:** The results of the present study have demonstrated the protective role of hydroalcoholic extract of *P. lanceolata* against AlCl<sub>3</sub>-induced neurotoxicity in Swiss albino mice. The neuroprotective efficacy of *P. lanceolata* can help reduce the symptoms caused by toxic protein aggregates in several degenerative diseases.

**Key words:** Acetylcholinesterase, aluminum chloride, lipid peroxidation, neurotoxicity, *Pluchea lanceolata*

## SUMMARY

- The hydro alcoholic extract of *Pluchea lanceolata* showed neuroprotective activity in albino mice against AlCl<sub>3</sub> toxicity
- The benefits of *Pluchea lanceolata* against AlCl<sub>3</sub> toxicity includes reduced lipid peroxidation and acetylcholine esterase activity with improved behavioral functions
- The hydro alcoholic extract of *Pluchea lanceolata* rendered protection against AlCl<sub>3</sub> in forebrain, midbrain, cerebellum and hippocampus
- Therefore *Pluchea lanceolata* holds pharmacological potentials for treating diseases associated with neuronal toxicity.



**Abbreviations used:** HAPL: Hydro alcoholic extract of *Pluchea lanceolata*; CAT: Catalase; GSH-Px: Glutathione peroxidase; SOD: Superoxide dismutase; TBARS: Thio-barbituric acid reactive substances; MDA: Malondialdehyde; AChE: Acetylcholine esterase; AOT: Acute oral toxicity; CNS: Central nervous system; H<sub>2</sub>O<sub>2</sub>: Hydrogen peroxide; ML: molecular layer; GL: granular layer; MC: microcytic changes; BV: blood vessels; DG: dentate gyrus; PC: pyramidal cells; LD: Lethal dose; ANOVA: Analysis of variance; SEM: Standard error of mean; PCL: Pyramidal cell layer; OCL: Outer granular layer; BV: blood vessels; PM: Pia mater.

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## INTRODUCTION

Aluminum is a potent environmental neurotoxin that particularly interferes with several enzymes and proteins related to neurotoxicity leading to many cognitive diseases, especially Alzheimer's disease and Parkinson's disease.<sup>[1]</sup> Chronic aluminum accumulation can induce oxidative stress and pathological changes in vital areas of the brain. There are varieties of aluminum sources from where the human beings

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can expose routinely, such as diet, water purification process, antacids, vaccines, and cosmetic agents. The average dietary intake of aluminum in adults ranges from 3 to 12 mg/day. The population routinely exposed to different sources of aluminum may have higher chances of neurotoxicity.<sup>[2,3]</sup>

The possible mechanism of aluminum chloride ( $\text{AlCl}_3$ )-induced neurotoxicity involves severe oxidative stress followed by inflammatory changes leading to neurodegeneration.  $\text{AlCl}_3$  is a nonredox active metal and capable of increasing the cellular oxidation by potentiating the prooxidant properties. Chronic aluminum exposure generates reactive oxygen species which in turn causes lipid peroxidation and decreased intracellular antioxidants.  $\text{AlCl}_3$  on accumulation leads to affect the slow and fast axonal transport, induces inflammatory responses, and causes synaptic structural abnormalities, which results in progressive neurodegeneration. It is also reported that  $\text{AlCl}_3$  can cause degeneration of cholinergic nerve terminals in cortical and hippocampus areas, leading to cellular depletion and severe learning disability.<sup>[4]</sup>

Plants are considered as rich sources of natural antioxidants, which alleviate the oxidative stress and improve cellular antioxidant status. *Pluchea lanceolata* (family-Asteraceae) popularly known as “Rasana” in Ayurveda is a rapidly spreading perennial rhizomatous weed and distributed throughout the northwestern part of India and neighboring Asian countries.<sup>[5]</sup> In Ayurveda, it is widely used in the treatment of rheumatoid arthritis, fever, pain, and inflammation. It is also used as a nerve tonic in conditions such as neuritis, sciatica, and chronic inflammation of nervous system.<sup>[6-8]</sup> It has been reported to possess anti-inflammatory, analgesic, immunosuppressant, and antimalarial activities.<sup>[9-12]</sup> The phytochemicals such as quercetin and isorhamnetin have been identified in *P. lanceolata*.<sup>[13]</sup> It has been reported that the flavanols derived from *P. lanceolata* possess significant antioxidant properties and well attenuated the cadmium chloride-induced oxidative stress and genotoxicity.<sup>[14]</sup>

Thus, development of potential neuroprotective drugs will be the effective strategy in the management of patients with neurodegenerative cognitive impairment disorders, and hence, there is a great demand for drugs which possess potent anti-inflammatory and antioxidant properties. In the present study, we have evaluated for the first time the neuroprotective activity of hydroalcoholic extract of *P. lanceolata* in  $\text{AlCl}_3$ -induced neurotoxicity in Swiss albino mice.

## MATERIALS AND METHODS

### Chemicals

Sodium dodecyl sulfate, thiobarbituric acid, glutathione standard, n-butanol, and pyridine were purchased from Himedia-Mumbai, India. Aluminum chloride hexahydrate ( $\text{AlCl}_3 \cdot 6\text{H}_2\text{O}$ ) was obtained from Thomas Baker Pvt. Chemicals, Mumbai, India. Acetylcholinesterase (AChE) kit was purchased from Piramal Healthcare Ltd. Thane, Mumbai, India. All other chemicals and reagents used in the present work were of analytical grade.

### Plant material and extract preparation

The rhizome of *P. lanceolata* were procured from Jamnagar, India, during May 2015 and authenticated in Pharmacognosy Laboratory at SDM Centre for Research in Ayurveda and Allied Sciences, Udipi, Karnataka. The voucher specimen no. 15031401-02 has been deposited for future reference. The rhizome was shade dried and powdered at SDM Pharmacy, Udipi, with the help of pulverizer. The hydroalcoholic extract was prepared by soaking 500 g of powdered rhizome of *P. lanceolata* in 2 L of 50% ethanol and 50% cold distilled water for 24 h, filtered, and concentrated by evaporating on water bath till free from water.

### Experimental animals

Male Swiss albino mice weighing 30–40 g body weight were obtained from animal house attached to Pharmacology and Toxicology Laboratory at SDM Centre for research in Ayurveda and Allied Sciences Udipi, India. The animals were maintained at standard laboratory conditions such as temperature at 25°C–27°C, humidity of 50%–55%, and natural light and dark cycles. Animals were fed with commercial pellet diet (*Pranav* Agro Industry, Pune) and water *ad libitum*.

### Acute oral toxicity test

It was carried out as per OECD guidelines 425, using AOT software. The hydroalcoholic extract of rhizome of *Pluchea lanceolata* [HAPL] was made into a suspension in 0.5% carboxymethyl cellulose (CMC) and dosed in the following order: 175, 550, and 2000 mg/kg body weight. The animals were observed for 14 days for mortality. The  $\text{LD}_{50}$  was determined by AOT 425 software.

### Experimental design

The mice were randomized into four different groups, each comprising six animals. Group 1 mice (vehicle control) were given 0.5% CMC orally for 45 consecutive days. Group 2 ( $\text{AlCl}_3$  control) were treated with 40 mg/kg  $\text{AlCl}_3$  (pH 7) intraperitoneally for 45 consecutive days.<sup>[15]</sup> Group 3 and 4 mice were treated with HAPL, 200 and 400 mg/kg body weight, respectively, for 45 consecutive days. The test drug was made as suspension in 0.5% CMC and administered at 1 ml/100 g body weight orally using gavage attached with syringe. The  $\text{AlCl}_3$  (40 mg/kg body weight, pH 7) was administered intraperitoneally for Groups 3 and 4 for 45 consecutive days an hour after the HAPL treatment. At the end of the experimental period, animal behavior was evaluated. Animals were anesthetized, sacrificed, and serum separated from collected blood. The brain tissues were collected from each group. Three brain samples were stored in 10% formalin and used for histopathological studies, whereas remaining three brain samples were homogenized and used for biochemical investigations.

### Behavioral assessment tests

#### Forced swim test

Animals were subjected to forced swim test on the last day of experimentation. An hour after giving the last dose of group-specific treatment, individual mouse was put into water filled (30 cm) glass cylinder measuring about 40 cm in height and 18 cm diameter, and observations were made for 6 min. First 2 min were not considered for recording the drug effect and were taken as stabilizing time. The limb movements and the effort of the mice to get out of the cylinder in the next 4 min were noted and subtracted later from total time (6 min) to find the time of immobility. This was considered as the index of depression.<sup>[16]</sup>

#### Tail suspension test

The total duration of immobility induced by tail suspension was measured according to a standard method. Mice isolated both acoustically and visually were suspended 50 cm above the floor by adhesive tape placed approximately 1 cm from the tip of the tail. Immobility time was recorded during a 6-min test and the immobility time considered as an index of CNS depression.<sup>[17]</sup>

### Estimation of serum acetylcholinesterase

The AChE was estimated by the method described by recommendation of the German society of clinical chemistry.<sup>[18]</sup> The AChE hydrolyses butyrylthiocholine under release of butyric acid and thiocholine. Thiocholine reduces yellow potassium hexacyanoferrate (III) to colorless

potassium hexacyanoferrate (II). The decrease in the absorbance was measured at 405 nm.

### Preparation of brain homogenate

Brain was excised and cleaned with ice cold saline and stored at  $-20^{\circ}\text{C}$  in freezer. Tissues were thawed and homogenized in phosphate buffered saline pH 7.4, centrifuged at 4000 rpm, and supernatant was stored at  $-20^{\circ}\text{C}$ . The homogenate was subjected to determination of catalase, glutathione peroxidase activity, and lipid peroxidation.

### Determination of catalase activity

The brain tissue homogenate (1 ml) was mixed with 5 ml of phosphate buffer and 4 ml of 0.2 M  $\text{H}_2\text{O}_2$  in phosphate buffer and time was noted. Exactly after 180 s after adding  $\text{H}_2\text{O}_2$ , a set of 1 ml of reaction mixture from the above was taken in 2 ml dichromate acetic acid. It was kept in boiling water bath for 10 min, cooled all the tubes under running tap water, and finally noted the reading at 570 nm against reagent blank. Catalase activity in the tissue was expressed as micromoles  $\text{H}_2\text{O}_2$  consumed/mg protein/min.<sup>[19]</sup>

### Determination of lipid peroxidation

Lipid peroxidation activity was determined by measuring the content of the thiobarbituric acid reactive substances. The level of lipid peroxidation was expressed as millimoles of malondialdehyde formed/g wet tissue.<sup>[20]</sup>

### Determination of glutathione peroxidase

Glutathione peroxidase was estimated using a standard protocol, and the glutathione peroxidase activity was expressed as micromolar glutathione utilized per mg protein per minute at  $37^{\circ}\text{C}$ .<sup>[21]</sup>

### Brain histopathology

Three brain samples from each group were used, and ten slices per sample were examined for histopathological study. Immediately after the excision from mice, the brain tissue was transferred into 10% formalin. Sections of 5  $\mu\text{m}$  thickness of brain tissue were prepared using microtome and stained with hematoxylin and eosin for microscopic observations.<sup>[22]</sup> All slides were then evaluated under light microscope (ZEISS Axio Lab A1 India).

### Statistical analysis

The obtained data were expressed as mean  $\pm$  standard error of mean and analyzed by one-way ANOVA, followed by Dunnett's multiple comparison *t*-test using GraphPad Prism 3.  $P < 0.05$  was considered as statistically significant.

## RESULTS

The acute toxicity study revealed no mortality with HAPL in any dose up to 2000 mg/kg body weight. The  $\text{LD}_{50}$  of HAPL is more than 2000 mg/kg. The dose taken for neuroprotective study was  $1/10^{\text{th}}$  and  $1/5^{\text{th}}$  of the higher dose of the study and found to be safe.

### Effect of hydroalcoholic extract of *Pluchea lanceolata* on aluminum chloride-induced behavioral changes in behavioral despair test

In behavioral despair test, the duration of freezing time was significantly increased in  $\text{AlCl}_3$  alone group as compared to vehicle control ( $P < 0.01$ ). Co-administration of HAPL has significantly attenuated the freezing time at both dose levels as compared to  $\text{AlCl}_3$  alone group ( $P < 0.01$ ) [Table 1].

**Table 1:** Effect of hydroalcoholic extract of *Pluchea lanceolata* in behavioral despair test

Group	Duration of freezing time (s)
Vehicle control	23.5 $\pm$ 3.81
$\text{AlCl}_3$ control	42.75 $\pm$ 1.49**
HAPL (200 mg/kg)	10.6 $\pm$ 1.9**
HAPL (400 mg/kg)	19.6 $\pm$ 1.65**

Data expressed as mean $\pm$ SEM, \*\* $P < 0.01$  in comparison to vehicle control group, \*\* $P < 0.01$  in comparison to  $\text{AlCl}_3$  group. HAPL: Hydroalcoholic extract of *Pluchea lanceolata*;  $\text{AlCl}_3$ : Aluminum chloride; SEM: Standard error of mean

**Table 2:** Effect of hydroalcoholic extract of *Pluchea lanceolata* in tail suspension test

Group	Duration of freezing time (s)
Vehicle control	49.8 $\pm$ 6.16
$\text{AlCl}_3$ control	157.25 $\pm$ 38.09*
HAPL (200 mg/kg)	93.2 $\pm$ 16.2
HAPL (400 mg/kg)	113.2 $\pm$ 41.93

Data expressed as mean $\pm$ SEM, \* $P < 0.05$  in comparison to vehicle control group. HAPL: Hydroalcoholic extract of *Pluchea lanceolata*;  $\text{AlCl}_3$ : Aluminum chloride; SEM: Standard error of mean

**Table 3:** Effect of hydroalcoholic extract of *Pluchea lanceolata* on serum acetylcholinesterase activity

Group	Acetylcholinesterase (IU/L)
Vehicle control	3813.3 $\pm$ 168.16
$\text{AlCl}_3$ control	5081.9 $\pm$ 270.56**
HAPL (200 mg/kg)	4225.28 $\pm$ 292.76
HAPL (400 mg/kg)	4945.82 $\pm$ 226.18

Data expressed as mean $\pm$ SEM, \*\* $P < 0.01$  in comparison to vehicle control group. HAPL: Hydroalcoholic extract of *Pluchea lanceolata*;  $\text{AlCl}_3$ : Aluminum chloride; SEM: Standard error of mean

### Effect of hydroalcoholic extract of *Pluchea lanceolata* on aluminum chloride-induced behavioral changes in tail suspension test

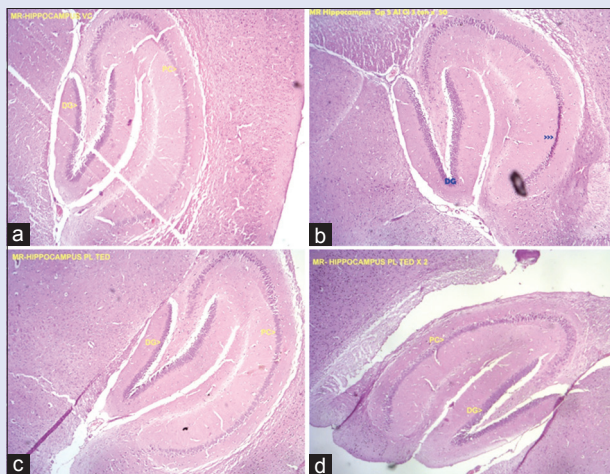
The duration of immobility time was significantly increased in  $\text{AlCl}_3$ -treated mice as compared to vehicle control. Whereas co-administration of HAPL considerably attenuated the freezing time, however, the observed changes were not statistically significant as compared to  $\text{AlCl}_3$  alone group [Table 2].

### Effect of hydroalcoholic extract of *Pluchea lanceolata* on acetylcholinesterase

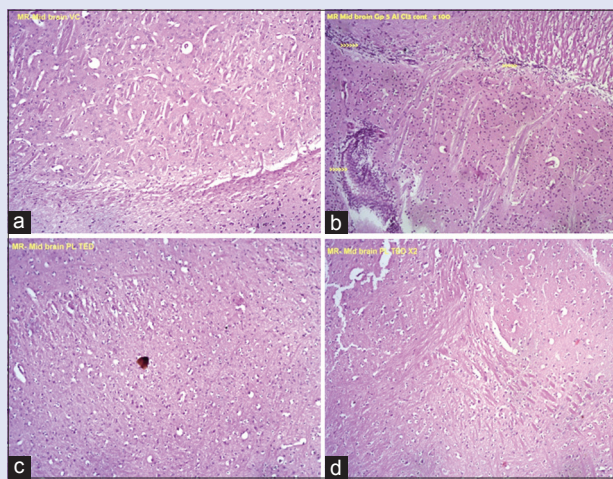
A significant increase in the level of AChE activity was observed in the animals treated with  $\text{AlCl}_3$  as compared to vehicle control ( $P < 0.01$ ). The HAPL co-administration with  $\text{AlCl}_3$  considerably attenuated the  $\text{AlCl}_3$ -induced elevation in the AChE; however, the observed changes were not statistically significant as compared to  $\text{AlCl}_3$  alone group [Table 3].

### Effect of hydroalcoholic extract of *Pluchea lanceolata* on antioxidant parameters

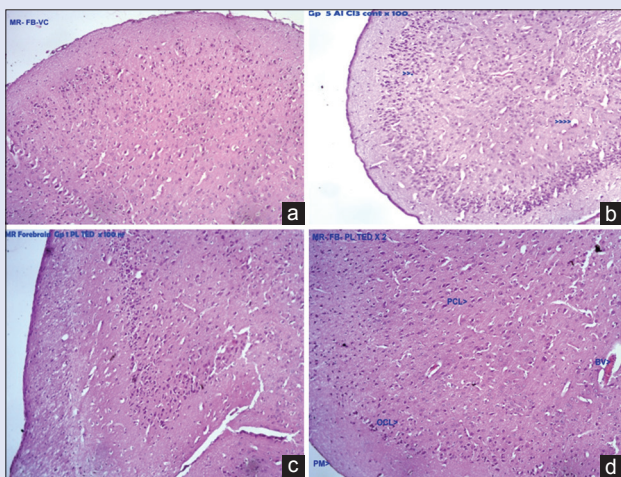
In the present study, repeated administration of  $\text{AlCl}_3$  significantly increased lipid peroxidation as compared to vehicle control ( $P < 0.01$ ). The elevated lipid peroxidation was significantly attenuated at both the dose levels of HAPL as compared to  $\text{AlCl}_3$  alone group. Whereas repeated administration of  $\text{AlCl}_3$  caused marked oxidative stress, which led to decrease in the antioxidant enzymes activities such as catalase and glutathione peroxidase in comparison to control group mice; while



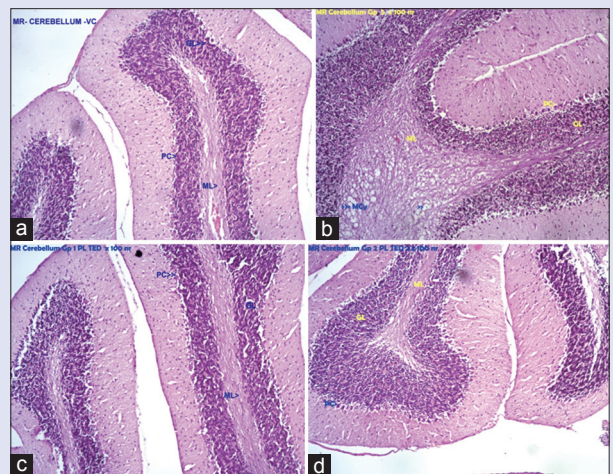
**Figure 1:** Photomicrograph of representative brain sections focused on hippocampus region of vehicle control (a), aluminum chloride group (b) and hydroalcoholic extract of rhizome of *Pluchea lanceolata* administered at 200 and 400 mg/kg + aluminum chloride (c and d). Aluminum chloride-administered group showed decrease in the pyramidal cells population, cellular disorganization, and distorted cells. These changes were considerably reversed by the administration of hydroalcoholic extract of rhizome of *Pluchea lanceolata* at higher dose level. DC: Dentate gyrus; PC: Pyramidal cells



**Figure 2:** Photomicrograph of representative brain sections focused on midbrain region of vehicle control (a), aluminum chloride group (b) and hydroalcoholic extract of rhizome of *Pluchea lanceolata* administered at 200 and 400 mg/kg body weight + aluminum chloride (c and d). The midbrain sections from aluminum chloride group show edematous changes, cellular disorganization, sparsely cellular light staining areas, and cell distortion. The sections from hydroalcoholic extract of rhizome of *Pluchea lanceolata*-treated groups showed mild-to-moderate edematous changes and less cell infiltration



**Figure 3:** Photomicrograph of representative brain sections focused on forebrain region of vehicle control (a), aluminum chloride group (b), hydroalcoholic extract of rhizome of *Pluchea lanceolata* administered at 200 and 400 mg/kg body weight + aluminum chloride (c and d). The forebrain sections from aluminum chloride group showed edematous changes, cellular disorganization, and cell distortion. The sections from low-dose hydroalcoholic extract of rhizome of *Pluchea lanceolata* (200 mg/kg)-treated group exhibited edematous changes with cellular disorganization and distortion whereas high-dose hydroalcoholic extract of rhizome of *Pluchea lanceolata* (400 mg/kg)-treated group showed almost normal cytoarchitecture



**Figure 4:** Photomicrograph of representative brain sections focused on cerebellum region of vehicle control (a), aluminum chloride group (b), and hydroalcoholic extract of rhizome of *Pluchea lanceolata* administered at 200 and 400 mg/kg body weight + aluminum chloride (c and d). The cerebellum sections from aluminum chloride group exhibited caused microcytic changes in the cellular layer of cerebellum. The sections from hydroalcoholic extract of rhizome of *Pluchea lanceolata* (200 and 400 mg/kg)-treated groups exhibited normal cytoarchitecture. ML: Molecular layer; GL: Granular layer; MC: Microcytic changes; BV: Blood vessels

HAPL co-administered with  $AlCl_3$  has increased the activity of catalase and caused no significant changes in glutathione peroxidase levels as compared to  $AlCl_3$  alone group [Table 4].

### Histopathological changes

Chronic administration of  $AlCl_3$  produced moderate intensity of neurodegeneration in different parts of the brain such as hippocampus,

midbrain, forebrain, and cerebellum. In hippocampus, there is a decrease in the pyramidal cells population, with apparent cellular disorganization and distorted cells as compared to control group. The forebrain and midbrain sections of  $AlCl_3$  alone group have shown edematous changes, cellular disorganization, and cell distortion. There are microcytic changes observed in the cellular layer of cerebellum, and these changes were attenuated to moderate extent by administration of HAPL at the higher dose level [Figures 1-4].

**Table 4:** Effect of hydroalcoholic extract of *Plantago lanceolata* on antioxidant parameters in aluminum chloride-induced neurotoxicity

Group	Catalase (µmoles/min/mg protein)	Glutathione peroxidase (µmoles glutathione/mg protein for 10 min)	Lipid peroxidation (mmoles of MDA formed/g wet tissue)
Vehicle control	6996.96±1078.1	3522.7±6.3	25.7621±318.2
AlCl <sub>3</sub> control	1373.31±10.7**	2325.92±788.1	404.165±39.8**
HAPL (200 mg/kg)	1256.16±129.5	1856.9±267.25	90.36±38.9**
HAPL (400 mg/kg)	8116.99±41.11**	3598.85±14.59	46.84±0.902**

Data expressed as mean±SEM, \*\* $P < 0.01$  in comparison to vehicle control group, \*\* $P < 0.01$  in comparison to AlCl<sub>3</sub> group. HAPL: Hydroalcoholic extract of *Plantago lanceolata*; AlCl<sub>3</sub>: Aluminum chloride; SEM: Standard error of mean; MDA: Malondialdehyde

## DISCUSSION

Aluminum being an important environmental neurotoxin also acts as a prooxidant. On chronic exposure, it can cause severe oxidative stress in brain tissues. The oxidative stress can lead to neuroinflammation followed by neurodegenerative changes. It has been reported that the phytoconstituents such as curcumin, quercetin, naringin, and catechin have potential antioxidant properties and has shown potent neuroprotective activity in AlCl<sub>3</sub>-induced neurotoxicity.<sup>[23,24]</sup>

*P. lanceolata* contains flavonoids such as quercetin and isorhamnetin and also chemicals such as sesquiterpenes, monoterpenes, and triterpenoids. These chemicals possess significant antioxidant and anti-inflammatory activities.<sup>[14,25,26]</sup> It has been reported that the decoction prepared from the *P. lanceolata* was used in pain and inflamed conditions such as arthritis. The taraxasterol derived from *P. lanceolata* has significant role in reducing neuroinflammation in C6 rat glial cells.<sup>[27]</sup> By extraction method, the fractions isolated from *P. lanceolata* were reported to have immunosuppressive properties by inhibiting the cytokines.<sup>[12]</sup> So far, there are no reports to substantiate the neuroprotective effect of *P. lanceolata* in AlCl<sub>3</sub> toxicity.

In the present study, two neurobehavioral tests were carried out to explore the degree of cognitive impairment and depressant component of the central nervous system. The chronic exposure to AlCl<sub>3</sub> significantly increased the freezing time in behavioral despair test and increased immobility time in tail suspension test. This indicates on repeated dose of AlCl<sub>3</sub> decreased motor activity, and CNS depression is evident. On the other hand, the co-administration of HAPL significantly attenuated AlCl<sub>3</sub>-induced CNS depression.

Normal brain cells maintain intact antioxidant milieu of enzymatic and nonenzymatic mediators. The enzymatic components such as superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GSH-Px) and the nonenzymatic components such as glutathione, thioredoxin, and thiol-containing molecules play a major role in maintaining cellular integrity. SOD converts superoxide anions into H<sub>2</sub>O<sub>2</sub> and oxygen, and it is neutralized by CAT and GSH-Px. Thus, the enzymes and indigenous antioxidant molecules protect the cells from damaging aggressive hydroxyl radicals.<sup>[28]</sup> In the present study, chronic exposure to AlCl<sub>3</sub> significantly increased the lipid peroxidation levels ( $P < 0.01$ ) and significantly decreased the activity of CAT and moderately the GSH-Px, whereas these adverse effects were significantly ( $P < 0.01$ ) attenuated by co-administration of HAPL, except for GSH-Px.

Acetylcholine is an important neurotransmitter in the cholinergic system, and AChE is an important enzyme involved in the metabolism of acetylcholine neurotransmitter. The increase in the AChE level in turn increases the metabolism of acetylcholine and leads to oxidative stress causing neurobehavioral changes, especially memory and cognitive failure. However, recently, it has been shown that circulating AChE activity reflects inflammatory response since acetylcholine suppresses inflammation. Based on this contention, donepezil, an AChE inhibitor, is being investigated for neuroprotective activity. In the present study, significant elevation in serum AChE activity was observed in AlCl<sub>3</sub>-administered group in

comparison to the control. This elevation was found to be moderately attenuated by HAPL drug treatment. This observation can be considered as an additional evidence for the neuroprotective activity indicating the role of *P. lanceolata* in the regulation of cholinergic function.<sup>[29]</sup>

The histological investigation also supports the pathological changes induced by chronic exposure of AlCl<sub>3</sub>. The high-dose HAPL drug has shown considerable protective effect. AlCl<sub>3</sub>-induced microcytic changes in the cellular layer of cerebellum and in forebrain sections with evidence of edematous changes, cellular disorganization, and cell distortion in few mice could be visualized. In the hippocampus, there was a decrease in the pyramidal cells population, cellular disorganization, and distorted cells. Thus, chronic administration of AlCl<sub>3</sub> produced moderate intensity of neurodegeneration in different parts of the brain such as forebrain, hippocampus, and cerebellum. The test drug administered at higher dose level attenuated the AlCl<sub>3</sub>-induced histopathological changes from mild to moderate extent. These results support the protective role of HAPL in AlCl<sub>3</sub>-induced neurotoxicity and improved functional outcome.

## CONCLUSIONS

Based on the findings of the present work, it can be concluded that the hydroalcoholic extract of *P. lanceolata* exhibited neuroprotection in mice against AlCl<sub>3</sub> neurotoxicity. Our preliminary experiments using histological, behavioral, and biochemical analysis support this proof of concept and warrants deeper investigations in future using HAPL for gaining better pharmacological information and intervention.

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## Conflicts of interest

There are no conflicts of interest.

## REFERENCES

- Yokel RA. Aluminum chelation principles and recent advances. Co-ord. Chem Rev 2002;228:97-113.
- Ribes D, Colomina MT, Vicens P, Domingo JL. Effects of oral aluminum exposure on behavior and neurogenesis in a transgenic mouse model of Alzheimer's disease. Exp Neurol 2008;214:293-300.
- Savory J, Herman MM, Ghribi O. Mechanisms of aluminum-induced neurodegeneration in animals: Implications for Alzheimer's disease. J Alzheimers Dis 2006;10:135-44.
- González MA, Bernal CA, Mahieu S, Carrillo MC. The interactions between the chronic exposure to aluminum and liver regeneration on bile flow and organic anion transport in rats.

- Biol Trace Elem Res 2009;127:164-76.
5. Anonymous. The Wealth of India: Raw Materials. New Delhi, India: Publications and Information Directorate, Council of Scientific and Industrial Research (CSIR); 1969.
  6. Chopra RN, Chopra IC, Handa KL, Kapur LD. The use of drugs in arthritis and bronchitis. In: Indigenous Drugs of India. Calcutta, India: UN Dhur, Sons Ltd.; 1958. p. 20.
  7. Inderjit S, Foy CL, Dakshini KM. *Pluchea lanceolata*: Anoxious perennial weed. Weed Technol 1998;12:190-3.
  8. Gupta OP. Handbook of Ayurvedic Medicine. In: Chaukhamba Sanskrit Bhawan. Varanasi, India: Chaukhamba Publications; 2006. p. 4.
  9. Srivastava P, Shanker K. *Pluchea lanceolata* (Rasana): Chemical and biological potential of rasayana herb used in traditional system of medicine. Fitoterapia 2012;83:1371-85.
  10. Akihisa T, Yasukawa K, Oinuma H, Kasahara Y, Yamanouchi S, Takido M, et al. Triterpene alcohols from the flowers of compositae and their anti-inflammatory effects. Phytochemistry 1996;43:1255-60.
  11. Kaith BS. Neolupinol and antiinflammatory activity. Int J Pharmacogn 1995;34:73-5.
  12. Bhagwat DP, Kharya MD, Bani S, Kaul A, Kour K, Chauhan PS, et al. Immunosuppressive properties of *Pluchea lanceolata* leaves. Indian J Pharmacol 2010;42:21-6.
  13. Dixit GS, Tewari RP. Chemical constituents of *Pluchea lanceolata*. Sacitra Ayurveda 1991;43:841.
  14. Jahangir T, Khan TH, Prasad L, Sultana S. *Pluchea lanceolata* attenuates cadmium chloride induced oxidative stress and genotoxicity in swiss albino mice. J Pharm Pharmacol 2005;57:1199-204.
  15. Shati AA, Elsaid FG, Hafez EE. Biochemical and molecular aspects of aluminium chloride-induced neurotoxicity in mice and the protective role of *Crocus sativus* L. extraction and honey syrup. Neuroscience 2011;175:66-74.
  16. Porsolt RD, Bertin A, Jalfre M. Behavioral despair in mice: A primary screening test for antidepressants. Arch Int Pharmacodyn Ther 1977;229:327-36.
  17. Steru L, Chermat R, Thierry B, Simon P. The tail suspension test: A new method for screening antidepressants in mice. Psychopharmacology (Berl) 1985;85:367-70.
  18. Recommendations of the German Society of Clinical Chemistry. Standardization of Methods for the Estimation of Enzymes Estimation in Biological Fluids. Standard Method for the Determination of Cholinesterase Activity. J Clin Chem Clin Biochem 1992;30:163-70.
  19. Sinha AK. Colorimetric assay of catalase. Anal Biochem 1972;47:389-94.
  20. Ohkawa H, Ohishi N, Yagi K. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. Anal Biochem 1979;95:351-8.
  21. Rotruck JT, Pope AL, Ganther HE, Swanson AB, Hafeman DG, Hoekstra WG. Selenium: Biochemical role as a component of glutathione peroxidase. Science 1973;179:588-90.
  22. Bancroft JD, Gamble M. Theory and Practice of Histological Techniques. 5<sup>th</sup> ed. London: Churchill Livingstone; 2002.
  23. Rios JL, Recio MC, Giner RM, Máñez S. An update review of saffron and its active constituents. Phytother Res 1996;10:189-93.
  24. Comalada M, Camuesco D, Sierra S, Ballester I, Xaus J, Gálvez J, et al. *In vivo* quercitrin anti-inflammatory effect involves release of quercetin, which inhibits inflammation through down-regulation of the NF-kappaB pathway. Eur J Immunol 2005;35:584-92.
  25. Srivastava V, Verma N, Tandon JS, Srimal RC, Zetlinger LS. Anti-inflammatory activity of *Pluchea lanceolata*: Isolation of an active principle. Int J Crude Drug Res 1990;28:135-7.
  26. Chawla AS, Kaith BS, Handa SS, Kulshreshtha DK, Srimal RC. Chemical investigation and anti-inflammatory activity of *Pluchea lanceolata*. Fitoterapia 1991;62:441-4.
  27. Srivastava P, Mohanti S, Bawankule DU, Khan F, Shanker K. Effect of *Pluchea lanceolata* bioactives in LPS-induced neuroinflammation in C6 rat glial cells. Naunyn Schmiedebergs Arch Pharmacol 2014;387:119-27.
  28. Schrader M, Fahimi HD. Peroxisomes and oxidative stress. Biochim Biophys Acta 2006;1763:1755-66.
  29. Noh YH, Baek JY, Jeong W, Rhee SG, Chang TS. Sulfiredoxin translocation into mitochondria plays a crucial role in reducing hyperoxidized peroxiredoxin III. J Biol Chem 2009;284:8470-7.