SHORT COMMUNICATION

Debrisoquine metabolism and genetic predisposition to lung cancer

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It is usually the metabolites of environmental carcinogens that initiate a cancer (Miller & Miller, 1983), and if the metabolism were subject to genetically determined polymorphism, there could be variation between smokers in their susceptibility to smoking-related lung cancer. We report a case-control study using the drug debrisoquine as a potential marker of such genetically determined susceptibility to lung cancer.

Debrisoquine has only one major metabolite, 4-hydroxy debrisoquine (Idle et al., 1979), and the extent of 4hydroxylation before excretion is controlled by a single gene and segregates into two distinct phenotypes - autosomal recessive poor metabolisers (about 9% of white populations, hydroxylating only 1-2% of a 10 mg dose of debrisoquine) and homozygous and heterozygous dominant extensive metabolisers (hydroxylating 10-99%) (Evans et al., 1980; Steiner et al., 1985). Unchanged and 4-hydroxy debrisoquine can be measured easily and with high reproducibility (r=0.88) in urine (Evans et al., 1980). The oxidative metabolism of over 20 drugs and other chemicals is known to be controlled by the same single gene locus as debrisoquine, and exhibits the same polymorphism (Sloan et al., 1978; Eichelbaum, 1984). There is no specific basis for a prior hypothesis, but the metabolism of a carcinogen in tobacco smoke could be subject to the same polymorphism.

Ayesh *et al.* (1984) reported a case-control study showing an association between lung cancer and extensive metabolism of debrisoquine. However D.S. Davies *et al.* (personal communication) have found no such association while Roots *et al.* (1988) considered the association to be at best weak or confined to certain histological types. This prompted us to report the present study to help resolve the uncertainty. It was performed in 1982 as a preliminary study to the larger study of Ayesh *et al.*, but different investigators supervised the tests and measured metabolic ratio, and the subjects were recruited from different sources.

We recruited consecutive caucasian inpatients with newly diagnosed lung cancer from a London hospital. Cigarette smoking histories were documented as accurately as possible and recorded in pack-years (one-twentieth the average daily number of cigarettes smoked multiplied by the number of years of smoking). Subjects whose total cigarette consumption was less than 10 pack years, or who had given up smoking more than five years previously were excluded. We also excluded patients with elevated serum bilirubin $(>25 \text{ mmol } l^{-1})$, and those who were taking drugs known to induce oxidising enzymes (e.g. barbiturates) or to compete with debrisoquine for oxidation (Sloan et al., 1978; Eichelbaum, 1984). A total of 104 cases were included. The histological types were squamous cell (38 cases), large cell cell (31), adenocarcinoma small (22).(11)and undifferentiated (2), diagnosed histologically (92) or

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cytologically (12). The 104 control subjects were residents of three homes for ex-servicemen (70), inpatients with various non-malignant diseases (22) and hospital employees (12). They were matched to the cases by sex (76 men, 28 women), age (mean 62.6 years \pm s.d. 9.2 in cases, 61.4 ± 10.1 in controls) and cigarette consumption (54.5 pack-years \pm 32.0 in cases, 54.0 ± 33.1 in controls).

Cases and controls took a single 10 mg debrisoquine tablet at 7 a.m. and made an 8 h urine collection, from which aliquots were stored at -20° C to await analysis by electroncapture gas-chromatography (Idle *et al.*, 1979). The ratio of unchanged to 4-hydroxy debrisoquine concentration, the metabolic ratio, was used to assign phenotype, the antimode

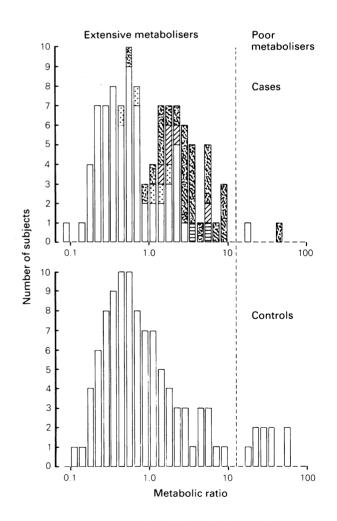


Figure 1 Metabolic ratio for debrisoquine in 104 lung cancer cases and 104 controls. The effect in the cases of certain drugs on metabolic ratio is shown: Distalgesic (irregular shading), cytotoxic drugs (diagonal lines), Moduretic (horizontal lines), diazepam (stippled).

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Table I Metabolic ratios among extensive metabolisers

	n	log ₁₀ metabolic ratio (mean (s.d.))	Comparison with 'remaining cases' (Z)
Cases taking:			
Distalgesic	25	+0.47(0.32)	9.2 $(P < 0.001)$
cytotoxic drugs	5	+0.35(0.23)	5.1 (P < 0.001)
Moduretic	2	+0.65 (0.14)	8.1 (P < 0.001)
diazepam	5	+0.03 (0.19)	2.1 (P = 0.03)
Remaining cases	65	-0.30(0.34)	· · · ·
Controls	95	-0.14 (0.43)	-2.5 (P=0.01)

distinguishing poor from extensive metabolisers occurring at a metabolic ratio of 12.6 (Evans *et al.*, 1980).

Results are shown in Figure 1. Two of 104 (1.9%) lung cancer cases and nine of 104 (8.7%) controls were poor metabolisers: P (one-tailed)=0.03 (Fisher's exact test). Statistical significance is greater if the larger series of 258 normal subjects of Evans et al. (1980) is used as controls (2P=0.02). Among extensive metabolisers the distribution of metabolic ratio in controls was approximately log-normal and similar to that in studies of normal volunteers (Evans et al., 1980; Steiner et al., 1985). In the cases it was skewed to the right by the effect of certain drugs, as shown in Figure 1. Metabolic ratios were statistically significantly higher in patients taking four drugs, namely Distalgesic, cytotoxics Moduretic and diazepam, than in the remaining cases (Table I). No drugs were associated with a lower metabolic ratio. In the 65 extensive metaboliser cases not taking any of the above four drugs, mean metabolic ratio was statistically significantly lower than in the 95 extensive metaboliser controls (none of whom were taking any of the four drugs) (Table I).

This study confirms that fewer lung cancer cases than controls were poor metabolisers of debrisoquine (1.9% vs 8.7%). The results of Ayesh *et al.* (1984) were very similar; four of 245 (1.6%) cases and 21 of 234 (9.0%) controls were poor metabolisers (P < 0.01). Poor metabolisers thus have one-fifth the risk of smoking-related lung cancer of extensive metabolisers. Among extensive metabolisers one might expect more lung cancer cases than controls to be homozygous, and in keeping with this metabolic ratio was lower on average (i.e. more debrisoquine was hydroxylated) in lung cancer cases (excluding those taking certain drugs) than in controls.

No apparent sources of bias can explain the association of extensive debrisoquine hydroxylation with lung cancer. Those drugs known to affect metabolic ratio increase it, the bias thus operating against the result of fewer poor metabolisers among lung cancer cases. A biological effect of the cancer is a possible explanation, but in animal studies mono-oxygenase activities appear depressed by the presence of cancer (Rosso et al., 1971), such a bias again operating against the result. Steiner et al. (1985) found that smoking and various other environmental factors such as alcohol, body weight and exercise that might differ between cases and controls did not demonstrably influence metabolic ratio, apart from a modest association with coffee intake. Moreover, as discussed by Nebert (1981), the metabolic effects of environmental factors are generally not powerful enough to mimic genetic effects of the magnitude observed here (extensive metabolisers hydroxylating about 50 times as much debrisoquine as poor metabolisers on average). We conclude that the association observed in two studies, in the absence of any apparent source of bias, constitutes evidence for genetic predisposition to smoking-related lung cancer. The association may be interpreted directly as a shared enzymatic pathway with an unspecified carcinogen, or indirectly as linkage disequilibrium, the gene locus regulating debrisoquine hydroxylation being closely associated on the same chromosome with a locus that independently affects lung cancer risk.

Further progress in elucidating metabolic predisposition to cancer is likely to be dominated by recombinant DNA technology, which is free from the bias associated with metabolic studies. The gene responsible for extensive debrisoquine hydroxylation is a structural gene, its complementary DNA has been cloned and sequenced, and three different mutant genes have been identified that give rise to incorrectly spliced messenger RNAs unable to yield an immunodetectable protein in the liver (Gonzalez *et al.*, 1988). Unfortunately these three mutant genes can account for only about half of all poor metabolisers, so that molecular genetics cannot at present replace the pharmacological measure of debrisoquine metaboliser status.

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