

TP53 Mutations Identified Using NGS Comprise the Overwhelming Majority of TP53 Disruptions in CLL: Results From a Multicentre Study

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*Correspondence:

Mark A. Catherwood mark.catherwood@belfasttrust.hscni.net

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Mark A. Catherwood ^{1*}, Dorte Wren², Laura Chiecchio³, Doriane Cavalieri⁴, David Donaldson¹, Sarah Lawless¹, Ezzat ElHassadi⁵, Amjad Hayat⁶, Mary R. Cahill⁷, Derville O'Shea⁷, Jeremy Sargent⁸, Peter Stewart⁹, Manisha Maurya⁹, John Quinn¹⁰, Philip Murphy¹⁰, David Gonzalez de Castro⁹, Ken Mills⁹, Nicholas C. P. Cross^{3,11}, Francesco Forconi¹¹, Sunil Iyengar², Anna Schuh⁴ and Patrick Thornton¹⁰

¹ Haematology Department, Belfast Health and Social Care Trust, Belfast, United Kingdom, ² The Royal Marsden Hospital and the Institute of Cancer Research, Biomedical Research Centre, London, United Kingdom, ³ Wessex Regional Genetics Laboratory, Salisbury National Health Service (NHS) Foundation Trust, Salisbury, United Kingdom, ⁴ Oxford Molecular Diagnostics Centre, Oxford University Hospitals, Oxford, United Kingdom, ⁵ Department of Haematology, University Hospital Waterford, Ireland, ⁶ Department of Haematology, University Hospital Galway, Galway, Ireland, ⁷ Department of Haematology, Cork University Hospital, Cork, Ireland, ⁸ Department of Haematology, Our Lady of Lourdes Hospital, Queens University Belfast, Drogheda, Ireland, ⁹ Centre for Cancer Research and Cell Biology (CCRCB), Queen's University Belfast, Belfast, United Kingdom, ¹⁰ Department of Haematology, Beaumont Hospital, Dublin, Ireland, ¹¹ Faculty of Medicine, University of Southampton, Southampton, United Kingdom

Limited data exists to show the correlation of (tumour protein 53) TP53 mutation detected by Next generation sequencing (NGS) and the presence/absence of deletions of 17p13 detected by FISH. The study which is the largest series to date includes 2332 CLL patients referred for analysis of del(17p) by FISH and TP53 mutations by NGS before treatment. Using a 10% variant allele frequency (VAF) threshold, cases were segregated into high burden mutations (\geq 10%) and low burden mutations (<10%). TP53 aberrations (17p [del (17p)] and/or TP53 mutation) were detected in 320/2332 patients (13.7%). Using NGS analysis, 429 TP53 mutations were identified in 303 patients (13%). Of these 238 (79%) and 65 (21%) were cases with high burden and low burden mutations respectively. In our cohort, 2012 cases did not demonstrate a TP53 aberration (86.3%). A total of 159 cases showed TP53 mutations in the absence of del(17p) (49/159 with low burden TP53 mutations) and 144 cases had both TP53 mutation and del(17p) (16/144 with low burden mutations). Only 17/2332 (0.7%) cases demonstrated del(17p) with no TP53 mutation. Validated NGS protocols should be used in clinical decision making to avoid missing low-burden TP53 mutations and can detect the vast majority of TP53 aberrations.

Keywords: chronic lymphocytic leukaemia, p53, deletion 17p, prognosis, next generation sequencing

INTRODUCTION

Deletion of chromosome 17p [del(17p)] and TP53 mutation (TP53 mut) referred to as TP53 aberrations can be found in 8%–10% of previously untreated chronic lymphocytic leukaemia (CLL) patients and in up to 30%–40% of relapsed/refractory cases. TP53 aberrations represent the most relevant risk factors for both progression free and overall survival following chemoimmunotherapy (1, 2). The introduction of small molecule inhibitors has led to enhanced response rates in patients with TP53 aberrations (3–5). Therefore, the identification of TP53 aberrations is essential for determining treatment decisions in CLL (6, 7). Historical data using Sanger sequencing suggests that approximately 80% of patients with del (17p) also carry a mutation in the second allele (8). A subset of patients also exhibits TP53 mut without del(17p) (8).

The assessment of del(17p) is routinely performed by Fluorescence *in situ* hybridization (FISH). The cut-off for a positive result varies within laboratories with the threshold >20% of cells with del(17p) deemed to be a clinically relevant clone (9). However, it is recognized that a subset of patients with del(17p) have stable disease without the need for treatment (10).

Sanger sequencing is widely used for TP53 mutational analysis, however it may misclassify cases of TP53 mutations as wildtype when variants with allelic frequencies below the detection limit of Sanger sequencing are present. Recent studies using next generation sequencing (NGS) have shown that TP53 mutations can be present at low clonal abundance in tumour cell populations, termed low-burden and have in certain studies the same detrimental effect on disease course (11-13). Therefore updated guidelines from the TP53 network of ERIC (European Research Initiative on CLL- www.ericcll.org) suggest a threshold of 10% allelic burden for reporting mutations detected by NGS segregating these into high burden (≥10 variant allele frequency (VAF)) and low burden (<10% VAF) mutation (6). In the literature, contradictory results exist regarding the biological relevance of low burden mutations in CLL. This in part may be due to various sequencing strategies.

Therefore, the aim of this study, which is the largest cohort to date, was to investigate the presence of low and high burden *TP53* mutations in a "real-world" cohort of 2332 CLL cases using sensitive NGS and to correlate results with FISH data.

METHODS

Pretreatment peripheral blood samples from 2332 CLL patients referred for analysis of del(17p13) by FISH and *TP53* mutations by NGS were available for the present study diagnosed between 2015-2019. A retrospective audit of *TP53* status was undertaken. Best practice in the UK follows established guidelines, meaning that *TP53* testing is recommended prior to each line of treatment but not at diagnosis. As participants are part of Specialized HaemOnc Diagnostics services, requests for *TP53* testing in newly diagnosed patients would automatically be rejected.

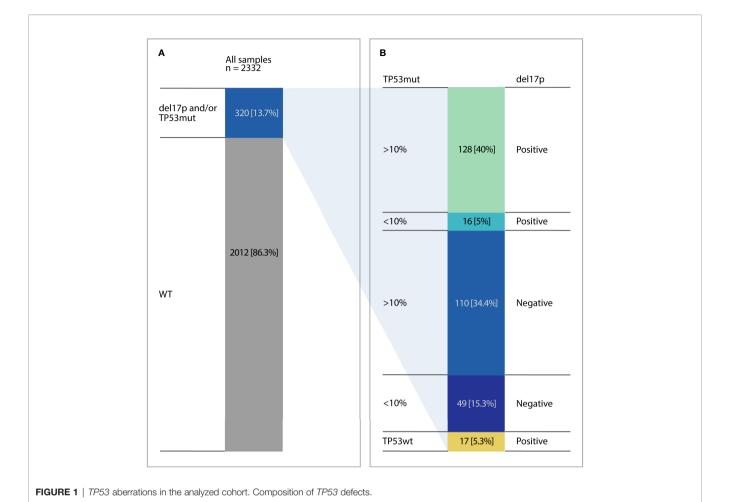
The study was conducted according to the Declaration of Helsinki. Patients were diagnosed according to iwCLL guidelines (14). In all cases, analysis was performed on DNA obtained from >50% tumour cells. FISH analysis for del(17p13) was performed using Vysis Probes with a 10% cut-off for a positive result. TP53 mutation screening was performed by NGS with a panel covering exons 2-11 as previously described (15) or by an Illumina amplicon-based strategy. Briefly, the amplicon-based panel was a bespoke assay and amplicon libraries for are generated by Reverse Complement PCR (RC-PCR) technology. The technique permits both the amplification and the ability to append sequences or functional domains of choice independently to either end of the generated amplicons in a single closed tube reaction. Primers for the TP53 assay were designed in house and sequencing was performed on the Miseq using Illumina chemistry. Raw data was aligned using GATK. Indels are realigned using GeminiMulti indel realigner and Pisces is used for variant calling (both Illumina). Normally analyses with read depths below 5000 are failed.

A VAF cut-off of 1% was used to exclude false positive variants within the cohorts. Pathogenicity assessment of all variants was performed according to ERIC guidelines (6).

RESULTS

Altogether 2332 patients entering first line treatment were included in this study with TP53 aberrations detected in 320/2332 patients (**Figures 1A, B**). Using NGS analysis, 429 TP53 mutations were identified in 303 patients (13%). More than one TP53 mutation was detected in 76 patients (2-8 mutations per patient, **Supplementary Table 1**). When considering all 429 TP53 mutations in the cohort the VAF ranged from 1-97%; mean 28%. Using the 10% VAF threshold, cases were segregated into high burden mutations ($\geq 10\%$) and low burden mutations (<10%). The high and low burden separation was based on the VAF of the most prevalent TP53 mutation. 271 (63%) were classified as high burden mutations (VAF range: 10-97%; mean 42%). 158 were classified as low burden mutations (VAF range: 1-9%; mean 5%) (**Figure 2**).

This translated into 238 patients classified as high burden cases and 65 identified as low burden cases (**Supplementary Table 1**). The needle plot graphs demonstrated no differences in *TP53* coding mutations between high and low burden cases (**Figures 3A, B**). The mutation profile revealed that the majority of mutations were missense mutations followed by frameshift, splicing and nonsense mutations and is in keeping with previous reports (**Figure 3C**) (16, 17). No significant difference within mutation type existed between the low and high burden groups (P=0.5). The amino acid most frequently mutated were at positions 175, 209, 234, 248 and 273 indicating the classical hot spot mutations in CLL. Codons 175, 209, 234, 248 and 273 represented 110/429 (25%) mutations in the total cohort and showed similar allocation in low and high burden case (**Figures 3A, B**).



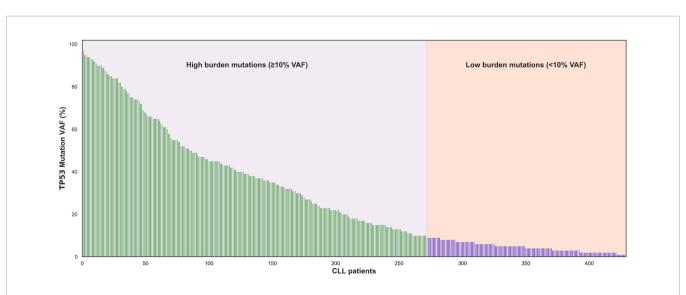


FIGURE 2 | Molecular Profile of TP53 mutations in the cohort. Using a cutoff of 10% VAF 271 TP53 mutations (228 patients) had high burden mutations and 158 TP53 mutations (65 patients) had low burden mutations.

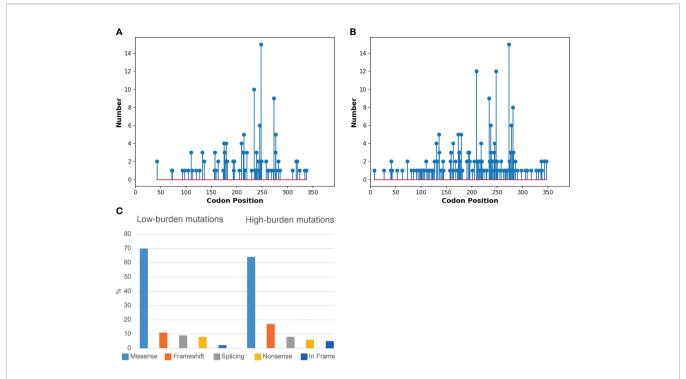


FIGURE 3 | Molecular Profile of *TP53* mutations in low and high burden cohorts. **(A)** Needle plot graph of low burden *TP53* mutations along the *TP53* coding sequence. **(B)** Needle plot graph of high burden *TP53* mutations along the *TP53* coding sequence. **(C)** Bar chart of mutations effect on the p53 protein in terms of amino acid changes in the low and high burden context.

Combining FISH data on del(17p) with *TP53* mutation data in our cohort, 2012 cases did not demonstrate a *TP53* aberration (86.3%). However, 17 cases demonstrated del(17p) only (0.7%). Average del(17p) was 40% (range 10-91%) in del(17p) only cases and was significantly higher in del(17p)/*TP53* mut cases (55% (range 10-100%: p<0.05). One hundred and fifty-nine patients (159) were *TP53* mutated only cases (49/159 with low burden TP53 mutations) and 144 cases with both del(17p) and *TP53* mutation (16/144 with low burden mutations, **Figure 1B**).

DISCUSSION

In this study, which is the largest study to date assessing *TP53* aberrations for both del(17p) and *TP53* mutation by NGS in cases of treatment naïve CLL. Using NGS analysis, 429 *TP53* mutations were identified in 303 patients (13%). Current guidelines from the *TP53* network of ERIC suggest a threshold of 10% allelic burden for reporting mutations detected by NGS (6). An acknowledgement is made in reference to cases with 5-10% VAF. In this study we employed a threshold of 10% VAF separating the cohort into high and low burden subgroups. High burden mutations were evident in 10.2% (238 cases) and low burden mutations in 2.8% (65 cases). This figure is lower than that reported in other studies and is likely due to the threshold of 1% used in this study (12, 13). Even with this threshold, 49 cases [*TP53* mut/del (17p) wt] in this cohort would have been misclassified as *TP53*

proficient cases. This is an important observation given the recent publication that clearly demonstrates a shorter survival in cases with VAFs of 5-10% (13). This study again questions the threshold of 10% VAF and the impact this has in the misclassification of *TP53* aberrations.

Whilst most tumour suppressors are inactivated by frameshift or nonsense mutations, the most frequent mode of inactivation of TP53 in CLL is by missense mutations which is a unique phenomenon. The mutation profile of the cohort did not differ when separated into high and low burden mutations. The vast majority of mutations were missense and no significant differences were observed between the low and high burden cohorts (Figures 3A-C). Unique to CLL is the presence of a specific hot spot variant leading to premature termination [p.(R209Kfs*6)]. This specific variant was demonstrated both in low and high burden cases highlighting the similar mutation profile between the cohorts (Figures 3A, B). The majority of TP53 mutations are located within the DNA binding domain of the gene and hot spot mutations are frequently observed in CLL. This study showed an enrichment of mutations in codons 175, 209, 234, 248 and 273 representing (25%) of all mutations in the total cohort. A similar pattern was evident in both low and high burden subgroups confirming the disease specific TP53 mutation profile in CLL (Figures 3A, B) (16).

Combining FISH data on del(17p) with *TP53* mutation data in our cohort, 2012 cases did not demonstrate a *TP53* aberration (86.3%) whereas *TP53* aberrations were detected in 13.7% of patients. This is in keeping with recent data from independent

groups that utilized various NGS strategies and bioinformatics pipelines (11, 13, 18).

In this study we have demonstrated the existence of del(17p) in the absence of a TP53 mutation in 17/2332 (0.7%) which is in keeping with the literature (1, 13). The average del(17p) clone was 40% with a range of 10-91% (Supplementary Table 1) with 8/17 cases having a del(17p) clone less than 25%. Patients in population based cohorts are still routinely screened for del(17p) by FISH, whilst testing for TP53 mutations can vary substantially by institution. This is despite very clear guidelines to the contrary (6, 14). Screening for only del(17p) in our study would have missed 50% of the alterations in the cohort (159/320). The relevance of FISH only based studies in the era of NGS is questionable as only a minority of p53 deficient cases are missed by NGS. In this series 0.7% of p53 deficient cases were missed by NGS of which 8 cases had a del(17p) clone size of less than 25%. Also recent data showing low-frequency del(17p) sub clones (<25% of CLL cells) in the absence of a TP53 mutation has been demonstrated to mirror that of cases with no del(17p) in the chemoimmunotherapy setting (12, 19). In the study by Do et al. 15/20 (75%) patients demonstrated a low frequency subclone of del(17p) (<25)). This is a well recognized phenomena in the literature with subset of patients with low frequency del(17p) clones having enhanced progression free survival (10). This subgroup of patients is enriched with a mutated IGHV gene and relatively few copy number alterations. The study by Do et al. represents a surprisingly high percentage of low level del (17p) not previously described and likely reflects the genomic composition of the elderly trial cohort in the study. In the current study, we demonstrated 28/144 (19%) cases where del(17p) <25% with 17 cases demonstrating a high burden (≥10%) mutations and 11 cases with low burden mutations (Figure 1B). Unfortunately clinical data was not available in this study to ascertain the IGHV status in the cohort of del(17p) subclones.

TP53 aberrations are still relevant in the era of novel therapies. Long term survival outcomes remain inferior in cohorts of patients with TP53 aberrations (20, 21). This is likely attributable to the role of p53 in the maintenance of genomic stability. It is well recognized that mutations in TP53 occur early in the disease progression proceeding the genomic instability generated by chromosomal abnormalities.

This has been further addresses in a recent study demonstrating that patients treated with single-agent ibrutinib carrying only a single TP53 hit have a superior long term response while multi-hit TP53 is associated with a shorter progression free and overall survival (22). In this scenario single hit CLL can be classified by the presence of either del (17p) or TP53 mut. Multi hit CLL arises when either del(17p) and TP53 mut occur together or when greater than one TP53 mutation is found. Whilst this is of interest it has yet to be verified in larger cohorts or indeed in separate treatment regimens. In our current study 55% (176/320) were single hit with 45% (144/320) of cases demonstrating a multi hit CLL. In this study ≥ 2 TP53 mutations were detected in 76 patients with the majority of cases in the TP53 mut/hol del(17p) wt cohort (46/76) with the remaining 30 cases in the TP53 mut/no del(17p) cohort.

This reinforces the need to redefine a VAF threshold to aid in the selection of TP53 mutated patients benefiting from targeted treatments.

In conclusion, in the largest series to date we have demonstrated the presence of low and high burden *TP53* mutations in a series of CLL cases. The use of NGS prevents cases being misclassified as normal TP53 due to its enhanced sensitivity. In the investigation of *TP53* aberrations, NGS is an important strategy for patient management in this setting.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**, further inquiries can be directed to the corresponding author/s.

ETHICS STATEMENT

Using the NHS Health Research Authority decision tools this study classes as clinical audit (i.e., no randomization of patients, no alteration to standard clinical care and informs practice in our setting) and therefore formal research ethics is not required.

AUTHOR CONTRIBUTIONS

MAC, AS and PT conceived the project. DW, LC, DC, PS, MM and DC performed data analysis. DD, SL, EE, AH, MRC, DO'S, JQ, PM, JS, KM, FF, SI, NC, AS and PT provided clinical input. All authors read and agreed with the manuscript.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fonc.2022.909615/full#supplementary-material

Supplementary Table 1 | TP53 mutations from mutated cohort.

Supplementary Table 2 | Sequencing metrics from Amplicon and Capture technologies.

REFERENCES

- Gonzalez D, Martinez P, Wade R, Hockley S, Oscier D, Matutes E, et al. Mutational Status of the TP53 Gene as a Predictor of Response and Survival in Patients With Chronic Lymphocytic Leukemia: Results From the LRF CLL4 Trial. J Clin Oncol (2011) 29(16):2223–9. doi: 10.1200/JCO.2010.32.0838
- Stilgenbauer S, Schnaiter A, Paschka P, Zenz T, Rossi M, Döhner K, et al. Gene Mutations and Treatment Outcome in Chronic Lymphocytic Leukemia: Results From the CLL8 Trial. *Blood* (2014) 123:3247–54. doi: 10.1182/blood-2014-01-546150
- Farooqui MZ, Valdez J, Martyr S, Aue G, Saba N, Niemann CU, et al. Ibrutinib for Previously Untreated and Relapsed or Refractory Chronic Lymphocytic Leukaemia With TP53 Aberrations: A Phase 2, Single-Arm Trial. Lancet Oncol (2015) 16(2):169–76. doi: 10.1016/S14702045(14)71182-9
- Furman RR, Sharman JP, Coutre SE, Cheson BD, Pagel JM, Hillmen P, et al. Idelalisib and Rituximab in Relapsed Chronic Lymphocytic Leukemia. N Engl J Med (2014) 370(11):997–1007. doi: 10.1056/NEJMoa1315226
- Roberts AW, Davids MS, Pagel JM, Kahl BS, Puvvada SD, Gerecitano JF, et al. Targeting BCL2 With Venetoclax in Relapsed Chronic Lymphocytic Leukemia. N Engl J Med (2016) 374(4):311–22. doi: 10.1056/NEJMoa1513257
- Malcikova J, Tausch E, Rossi D, Sutton LA, Soussi T, Zenz T, et al. European Research Initiative on Chronic Lymphocytic Leukemia (ERIC) — TP53 Network. ERIC Recommendations for TP53 Mutation Analysis in Chronic Lymphocytic Leukemia-Update on Methodological Approaches and Results Interpretation. Leukemia (2018) 32(5):1070–80. doi: 10.1038/s41375-017-0007-7
- Campo E, Cymbalista F, Ghia P, Jäger U, Pospisilova S, Rosenquist R, et al. TP53 Aberrations in Chronic Lymphocytic Leukemia: An Overview of the Clinical Implications of Improved Diagnostics. Haematologica (2018) 103 (12):1956–68. doi: 10.3324/haematol.2018.187583
- Malcikova J, Smardova J, Rocnova L, Tichy B, Kuglik P, Vranova V, et al. Monoallelic and Biallelic Inactivation of TP53 Gene in Chronic Lymphocytic Leukemia: Selection, Impact on Survival, and Response to DNA Damage. Blood (2009) 114(26):5307–14. doi: 10.1182/blood-2009-07-234708
- Catovsky D, Richards S, Matutes E, Oscier D, Dyer M, Bezares RF, et al. Assessment of Fludarabine Plus Cyclophosphamide for Patients With Chronic Lymphocytic Leukaemia (the LRF CLL4 Trial): A Randomised Controlled Trial. *Lancet* (2007) 370:230–9. doi: 10.1016/S0140-6736(07) 61125-8
- Tam CS, Shanafelt TD, Wierda WG, Abruzzo LV, Van Dyke DL, O'Brien S, et al. De Novo Deletion 17p13.1 Chronic Lymphocytic Leukemia Shows Significant Clinical Heterogeneity: The M. D. Anderson and Mayo Clinic Experience. Blood (2009) 114(5):957–64. doi: 10.1182/blood-2009-03-210591
- Rossi D, Khiabanian H, Spina V, Ciardullo C, Bruscaggin A, Famà R, et al. Clinical Impact of Small TP53 Mutated Subclones in Chronic Lymphocytic Leukemia. *Blood* (2014) 123(14):2139–47. doi: 10.1182/blood-2013-11-539776
- Malcikova J, Pavlova S, Kunt Vonkova B, Radova L, Plevova K, Kotaskova J, et al. Low-Burden TP53 Mutations in CLL: Clinical Impact and Clonal Evolution Within the Context of Different Treatment Options. (Blood) (2021), 2020009530. 138(25):2670–85 doi: 10.1182/blood.2020009530
- Bomben R, Rossi FM, Vit F, Bittolo T, D'Agaro T, Zucchetto A, et al. TP53
 Mutations With Low Variant Allele Frequency Predict Short Survival in
 Chronic Lymphocytic Leukemia. Clin Cancer Res (2021) 27(20):5566–75.
 doi: 10.1158/1078-0432.CCR-21-0701
- Hallek M, Cheson BD, Catovsky D, Caligaris-Cappio F, Dighiero G, Döhner H, et al. iwCLL Guidelines for Diagnosis, Indications for Treatment, Response Assessment, and Supportive Management of CLL. *Blood* (2018) 131 (25):2745–60. doi: 10.1182/blood-2017-09-806398
- 15. Stewart JP, Gazdova J, Darzentas N, Wren D, Proszek P, Fazio G, et al. Validation of the EuroClonality-NGS DNA Capture Panel as an Integrated

- Genomic Tool for Lymphoproliferative Disorders. . *Blood Adv* (2021) 5 (16):3188–98. doi: 10.1182/bloodadvances.2020004056
- Zenz T, Vollmer D, Trbusek M, Smardova J, Benner A, Soussi T, et al. TP53
 Mutation Profile in Chronic Lymphocytic Leukemia: Evidence for a Disease
 Specific Profile From a Comprehensive Analysis of 268 Mutations. *Leukemia* (2010) 24(12):2072–9. doi: 10.1038/leu.2010.208
- Malcikova J, Pavlova S, Kozubik KS, Pospisilova S. TP53 Mutation Analysis in Clinical Practice: Lessons From Chronic Lymphocytic Leukemia. *Hum Mutat* (2014) 35(6):663–71. doi: 10.1002/humu.22508
- Nadeu F, Delgado J, Royo C, Baumann T, Stankovic T, Pinyol M, et al. Clinical Impact of Clonal and Subclonal TP53, SF3B1, BIRC3, NOTCH1, and ATM Mutations in Chronic Lymphocytic Leukemia. *Blood* (2016) 127:2122–30. doi: 10.1182/blood-2015-07-659144
- Do C, Best OG, Thurgood L, Hotinski A, Apostolou S, Mulligan SP, et al. Insight Into Del17p Low-Frequency Subclones in Chronic Lymphocytic Leukaemia (CLL): Data From the Australasian Leukaemia and Lymphoma Group (ALLG)/CLL Australian Research Consortium (CLLARC) CLL5 Trial. Br J Haematol (2021) 193(3):556–60. doi: 10.1111/bjh.17394
- O'Brien S, Furman RR, Coutre S, Flinn IW, Burger JA, Blum K, et al. Single-Agent Ibrutinib in Treatment-Naïve and Relapsed/Refractory Chronic Lymphocytic Leukemia: A 5-Year Experience. *Blood* (2018) 131(17):1910–9. doi: 10.1182/blood-2017-10-810044
- Kater AP, Wu JQ, Kipps T, Eichhorst B, Hillmen P, D'Rozario J, et al. Venetoclax Plus Rituximab in Relapsed Chronic Lymphocytic Leukemia: 4-Year Results and Evaluation of Impact of Genomic Complexity and Gene Mutations From the MURANO Phase III Study. J Clin Oncol (2020) 38 (34):4042–54. doi: 10.1200/JCO.20.00948
- Brieghel C, Aarup K, Torp MH, Andersen MA, Yde CW , Tian X, et al. Clinical Outcomes in Patients With Multi-Hit TP53 Chronic Lymphocytic Leukemia Treated With Ibrutinib. Clin Cancer Res (2021) 27(16):4531–8. doi: 10.1158/1078-0432.CCR-20-4890

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