Original Article

Classic and non-classic 21-hydroxylase deficiency can be discriminated from P450 oxidoreductase deficiency in Japanese infants by urinary steroid metabolites

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Abstract. We previously reported a two-step biochemical diagnosis to discriminate classic 21-hydroxylase deficiency (C210HD) from P450 oxidoreductase deficiency (PORD) by using urinary steroid metabolites: the pregnanetriolone/tetrahydrocortisone ratio (Ptl/the cortisol metabolites 5aand 58-tetrahydrocortisone (sum of these metabolites termed THEs), and 118-hydroxyandrosterone (110HAn). The objective of this study was to investigate whether both C210HD and non-classic 21OHD (C+NC21OHD) could be biochemically differentiated from PORD. We recruited 55 infants with C210HD, 8 with NC210HD, 16 with PORD, 57 with transient hyper-17a-hydroxyprogesteronemia (TH17OHP), and 2,473 controls. All infants were Japanese with ages between 0-180 d. In addition to Ptl, THEs, and 110HAn, we measured urinary tetrahydroaldosterone (THAldo) and pregnenediol (PD5). The first step: by Ptl with the age-specific cutoffs 0.06 mg/g creatinine (0-10 d of age) and 0.3 mg/g creatinine (11–180 d of age), we were able to differentiate C+NC210HD and PORD from TH170HP and controls (0–10 d of age: 0.065-31 vs. < 0.001-0.052, 11-180 d of age: 0.40-42 vs. <0.001–0.086) with 100% sensitivity and specificity. The second step: by the 110HAn/THAldo or 110HAn/PD5 ratio with a cutoff of 0.80 or 1.0, we were able to discriminate between C+NC210HD and PORD (1.0-720 vs. 0.021-0.61 or 1.8-160 vs. 0.005-0.32, respectively) with 100% sensitivity and specificity. Ptl, 110HAn/THAldo, and 110HAn/PD5 could differentiate between C+NC210HD and PORD in Japanese infants.

Key words: urinary steroid metabolites, non-classical 21-hydroxylase deficiency, cytochrome P450 oxidoreductase deficiency

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Introduction

The clinical differential diagnoses of 21-hydroxylase deficiency (210HD) and cytochrome P450 oxidoreductase deficiency (PORD) are sometimes difficult since both deficiencies can have similar phenotypes and high levels of 17a-hydroxyprogesterone (170HP) in the blood. We previously reported specific cutoff(s) to discriminate between classic 210HD (C210HD) and PORD by using urinary steroid metabolites, i.e. the pregnanetriolone (Ptl)/ the cortisol metabolites 5α - and 5β -tetrahydrocortisone (sum of these metabolites termed THEs) ratio and 118-hydroxyandrosterone (110HAn), by using gas chromatography/mass spectrometry (GC/MS) (1). However, we did not investigate whether the cutoffs were able to discriminate between non-classic 210HD (NC210HD) and PORD. The prevalence of NC210HD is estimated at 1 case out of 2 million individuals in Japan (2), whereas it is reported to be 1 out of 1,000 in Caucasians (3, 4), and is considered to be the most common form of congenital adrenal hyperplasia. Patients with NC210HD have mildly impaired 21-hydroxylase activity leading to various symptoms from childhood to adulthood, such as precocious pubarche, acne, hirsutism, infertility, etc. (5, 6). Biochemical diagnosis of NC210HD is challenging because of the relatively mild glucocorticoid deficiency seen in patients. We previously reported that clinically diagnosed 210HD, including classic and non-classic 210HD (C+NC210HD), can be distinguished from transient hyper-17a-hydroxyprogesteronemia (TH170HP) and controls by Ptl measurements in GC/MS (7). Additionally, we reported in the same study that C+NC210HD could be differentiated from PORD by the ratio between 110HAn and pregnanediol, which is a metabolite of progesterone, in three infants between the ages of 1 and 3 months (7).

The objective of this study was to investigate whether C+NC21OHD could be biochemically differentiated from PORD in Japanese infants. In addition to Ptl, THEs, and 110HAn, we focused on the pregnenolone (P5) metabolite pregnenediol (PD5), and the aldosterone metabolite tetrahydroaldosterone (THAldo). We focused on these metabolites because in PORD, (i) blood P5 was shown to be higher (8, 9), and (ii) blood aldosterone and urinary THAldo were shown to be normal or slightly higher, respectively, compared to that in normal subjects (7, 9, 10).

Materials and Methods

All legal guardian(s) gave written informed consent and the study was approved by the Institutional Review Boards at Keio University Hospital and Keio University School of Medicine. We recruited 55 infants with C210HD (gestational age, 35–41 wk; birth weight, 1,658– 4,174 g), 8 infants with NC210HD (37–40 wk; 2,704–3,408 g), 16 infants with PORD (34–41 wk; 1,018-3,418 g), 57 infants with TH170HP (37-41 wk, 2,062–4,980 g), and 2,473 controls (34–41 wk, 770-4,610 g). All infants were Japanese with ages between 0-180 d, the period during which most patients with C210HD or PORD are diagnosed (7, 11). The diagnosis of 210HD and PORD was confirmed by CYP21A2 and POR gene analyses, respectively. Notably, all patients with NC210HD were positive in newborn massscreening in Japan. Patients with 210HD having normal genitalia and elevated dried blood spot 170HP (positive results in newborn mass-screening), but without any evidence of salt wasting (low serum sodium, high serum potassium, high plasma renin activity, etc.) were classified as NC210HD. Any subjects with abnormal physical findings except for external genitalia were excluded. None of the subjects received antenatal or perinatal glucocorticoid before urine sampling.

We measured urinary steroid metabolites by GC/MS (12). The 21-deoxycortisol metabolite Ptl, and the cortisol metabolites 5α -tetrahydrocortisone and 5β -tetrahydrocortisone (hereafter referred



Fig. 1. A steroid metabolic map. Solid arrow, steroid synthesis; open arrow, steroid metabolism; solid line, impaired 21-hydroxylase activity; open line, impaired 17-hydroxylase/17,20lvase activity. First step, differentiation of C+NC210HD and from PORD TH170HP and the control. Second step, discrimination between C+NC210HD and PORD. Both 21-hydroxylase and 17-hydroxylase/17,20-lyase activity are reduced in PORD, whereas only 21-hydroxylase is reduced in C+NC210HD. Preg, pregnenolone; Prog, progesterone; DOC, deoxycorticosterone; Aldo, aldosterone; 170HPreg, 17α-hydroxypregnenolone; 11DOF. 11-deoxycortisol; DHEA, dehydroepiandrosterone; AD4, androstendione.

to collectively as THEs) were measured and the ratio of Ptl to the cortisol metabolites (Ptl/ THEs) (1, 13, 14) was calculated to differentiate C210HD, NC210HD, and PORD from TH170HP and control (Fig. 1). Ptl was considered equal to 0.001 mg/g creatinine for calculations in infants whose Ptl was under the detection limit (< 5pg/injection). In addition to the metabolite of 118-hydroxyandrostenedione, 110HAn, (1), we measured THAldo and PD5. These steroids and their metabolites were previously shown to be increased in PORD (8-10). We then calculated the 110HAn/THAldo and 110HAn/PD5 ratios to discriminate between C+NC210HD and PORD (Fig. 1). Quantification and quality ions of each metabolite were as follows (m/z): Ptl 449, 359;

5αTHE 488, 578; 56THE 488, 578; 11OHAn 448, 358; THAldo 506 (quantified ion only); PD5 372, 462. Urinary creatinine was measured by IATRO-LQ CRE (A)II (LSI Medience Co., Tokyo, Japan). Urinary steroid concentration was expressed relative to urinary creatinine (mg/g creatinine).

Statistical analysis was performed using the Mann-Whitney U test. A p value of < 0.05 was considered statistically significant.

Results

Differentiation of C+NC21OHD and PORD from TH17OHP and controls

Results of Ptl and Ptl/THEs are shown in Table 1 and Fig. 2. Both Ptl and Ptl/THEs showed similar overlap between C+NC210HD, PORD, TH170HP, and control within 10 days of age by uniform cutoff through 0-180 d of age (Ptl 0.1 and Ptl/THEs 0.020). We then separately set the cutoff for 0–10 d of age and 11–180 d of age. Ptl differentiated C+NC210HD and PORD from TH17OHP and control with 100% (95% confidence interval (CI): 97.6–100%) sensitivity and 100% (95% CI: 99.9-100%) specificity using the 0.06 mg/g creatinine (0-10 d of age) and 0.3 mg/g creatinine (11–180 d of age) cutoffs. Ptl/THEs differentiated with 100% (95% CI: 96.5-100%) sensitivity and 99.9% (95% CI: 99.8–99.9%) specificity using the 0.01 (0–10 d of age) and 0.02 (11-180 d of age) cutoffs.

Discrimination between C+NC21OHD and PORD

Table 2 and Fig. 3 show the results of urinary 110HAn in C+NC210HD and PORD. 110HAn discriminated between C210HD and PORD with 96.8% (95% CI: 93.3–96.8%) sensitivity and 100% (95% CI: 86.1–100%) specificity using the 0.35 mg/g creatinine cutoff. We then focused on the aldosterone and P5 metabolites, THAldo and PD5. Although both metabolites showed significantly higher distribution in PORD than in C+NC210HD (THAldo p < 0.001, PD5p < 0.001),

	Ptl (mg/g creatinine)	Ptl/THEs	
C+NC21OHD PORD TH17OHPnemia Controls	$\begin{array}{rrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrr$	$\begin{array}{cccc} 2.8 & (0.13-20) \\ 0.13 & (0.010-0.23) \\ < 0.001 & (< 0.001-0.006) \\ < 0.001 & (< 0.001-0.027) \end{array}$	

Table 1Results of Ptl and Ptl/THEs in C+NC210HD, PORD, TH170HP,
and controls

mean (range).

Table 2 Results of 110HAn, THAldo, and PD5 in C+NC210HD and PORD

	110HAn	THAldo	PD5
C+NC21OHD PORD	2.3 (0.16–22) 0.037 (0.007–0.22)	$\begin{array}{c} 0.093 \ (0.005 - 0.80) \\ 0.34 \ \ (0.074 - 0.94) \end{array}$	$\begin{array}{c} 0.123 \ (0.012 - 1.3) \\ 0.63 \ \ (0.32 - 2.7) \end{array}$

mean (range), mg/g creatinine.



Fig. 2. Urinary Ptl and Ptl/THEs in infants with C+NC210HD, PORD, TH170HP, and controls. The upper graph is for Ptl and the lower one is for Ptl/THEs. Lines indicate cutoffs: Ptl 0.05 mg/g creatinine (0–10 d of age) and 0.1 mg/g creatinine (11–180 d of age), Ptl/THEs 0.01 (0–10 d of age) and 0.02 (11–180 d of age). Closed circle, C210HD; gray circle, NC210HD; triangle, PORD; open square, TH170HP; dot, control.



Fig. 3. Urinary 110HAn in infants with C210HD, NC210HD, and PORD. Line indicates cutoff of 0.35 mg/g creatinine. Closed circle, C210HD; gray circle, NC210HD; triangle, PORD.

there was clear overlap between the two groups (Table 2). Table 3 and Fig. 4 show the results of urinary 110HAn/THAldo and 110HAn/PD5 ratio calculations. 110HAn/THAldo discriminated with 100% (95% CI: 97.2–100%) sensitivity and 100% (95% CI: 88.9–100%) specificity using the 0.80 cutoff. 110HAn/PD5 discriminated with 100% (95% CI: 97.2–100%) sensitivity and 100% (95% CI: 88.9–100%) sensitivity and 100% (95% CI: 88.9–100%) specificity using the 1.0 cutoff.

	110HAn/THAldo	110HAn/PD5
C+NC21OHD PORD	$\begin{array}{ccc} 20 & (1.0-720) \\ 0.15 & (0.021-0.61) \end{array}$	18 (1.8–160) 0.059 (0.005–0.32)

Table 3Results of 110HAn/THAldo and 110HAn/PD5 in
C+NC210HD and PORD



mean (range).

Fig. 4. Urinary 110HAn/THAldo and 110HAn/ PD5 in infants with C210HD, NC210HD, and PORD. The upper graph is for 110HAn/ THAldo and the lower one is for 110HAn/ PD5. Lines indicate cutoffs: 110HAn/ THAldo 0.80, 110HAn/PD5 1.0. Closed circle, C210HD; gray circle, NC210HD; triangle, PORD.

Discussion

We demonstrated that a two-step biochemical diagnosis using urinary steroid metabolites is useful for diagnosis of 210HD and PORD when patients with NC210HD were included among the subjects. We propose a two-step biochemical diagnosis using Ptl for the first step, and 11OHAn/THAldo or 11OHAn/PD5 for the second step, because these two markers showed no overlap in each step.

In the first step, we set the age-specific cutoff of Ptl to differentiate C+NC210HD and PORD from TH170HP and the control. We separately set the cutoff at 0–10 and 11–180 d of age because patients with 210HD and PORD who were 0–10 d old showed lower Ptl values. Assuming that steroid metabolic enzyme activity in the liver was immature at early ages after birth, we could explain the imperfectness of the uniform cutoff through 0–180 d of age.

In the second step, a downstream/upstream metabolites ratio, 110HAn/THAldo or 110HAn/ PD5, was more useful than the single metabolite, 110HAn. Steroid downstream/upstream metabolite ratios have previously been used as markers of enzyme defects (15, 16). Indeed, as mentioned in the Introduction, we had used the ratio between 110HAn and pregnanediol (PD), a metabolite of progesterone, for distinguishing PORD from 210HD (7). As PD measurement is sometimes problematic in our GC/MS methods in newborns (our unpublished data), we used a single metabolite, 110HAn, to discriminate between C210HD and PORD (1). In this study, we chose metabolites of aldosterone and pregnenolone, THAldo, and PD5, for the following two reasons. First, aldosterone and pregnenolone are upstream of 17a-hydroxylase (Fig.1). Second, THAldo and PD5 can be measured in all newborn infants (our unpublished data).

In this study, we recruited 0–180 d old infants because most patients with C210HD or PORD

are diagnosed in this period (7, 11). Although it was reported that patients with PORD who were above 180 d of age showed similar trends in urinary steroid metabolites (i.e., high Ptl and normal-range 110HAn) (11), further analysis is required to determine whether our cutoffs can be applicable to infants over 180 d of age.

This method has two advantages compared to repeated 17OHP measurement: it is a single assay and offers the noninvasiveness of urine sampling. Thus, this method is a potential option for scrutiny of newborn mass-screened positive patients together with liquid chromatography/ tandem mass spectrometry (17, 18) and genetic analysis (19, 20).

Some limitations of this study should be discussed. First, we do not know whether our two-step method is applicable to all cases of NC210HD. In this study, NC210HD patients had positive results in a newborn mass-screening program in Japan. A few NC210HD cases have been reported to be positive in newborn massscreening programs (21), but most were negative because of the relatively low baseline levels of 170HP (5). Positive NC210HD may possess less 21-hydroxylase activity than negatives ones; i.e., their Ptl, 110HAn, and PD5 may be higher or THEs and THAldo may be lower. Second, as we described in a previous study (1), our data in Japanese infants may not apply to other ethnicities because of differences in common POR mutations and their residual activities in PORD (9, 22, 23). Additional studies are required for non-Japanese individuals. Third, preterm infants were not included in this study whose gestational age was less than 34 wk. Those infants might have more immature steroid metabolism in the liver, theoretically leading to lower Ptl and 110HAn compared with the subjects of this study.

In conclusion, we demonstrated a two-step biochemical diagnosis of C+NC210HD and PORD by urinary steroid profiling using Ptl, THAldo, PD5, and 110HAn.

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