

Evolving Understanding of T-cell Cosignaling Pathways

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e read with great interest the previously published article "Costimulation Blockade in Kidney Transplantation: An Update" written by Malvezzi et al.¹ In the setting of solid-organ transplantation, it represents a great progress to explore the blockade of some costimulation pathways as an efficient immunosuppressive tool instead of using calcineurin inhibitors. In the section of targeting costimulation pathways, Malvezzi et al¹ summarized main T-cell surface signaling molecules, including costimulation and coinhibition. Cosignaling molecules are controllers of T-cell responses to antigens. By selectively turning on and/or off costimulatory and coinhibitory pathways, the T-cell receptor (TCR) signal could be transiently regulated in positive and negative directions. cluster of differentiation (CD), CD28/CD154(CD40L)/inducible costimulator had a positive regulatory effect on T-cell activation, whereas cytotoxic T-lymphocyte-associated antigen-4/programmed death ligand-1 was negative. Based on this article, we will provide some progress in costimulation pathways over the past few years, as shown in Figure 1.

First, as we all know, programmed death 1 (PD-1) and its ligands, mainly PD-L1 and PD-L2, deliver inhibitory signals that regulate the balance among T-cell activation, tolerance, and exhaustion. PD-1 is a CD28 family member. PD-L1 (B7-H1) and PD-L2 (B7-DC) are B7 family members. Several previous findings pointed out a significant bidirectional inhibitory interaction between B7-1 (CD80) and PD-L1, which added an additional dimension to immunoregulatory functions of the B7:CD28 family.²⁻⁵ The competition assays in vitro showed that PD-1 and B7-1 competed for binding to PD-L1 and block each other's binding.^{6,7} Extremely low levels of PD-1 are sufficient for

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potent inhibition in the earliest stages of T-cell activation.⁸ However, this potent inhibition effect of PD-1 depends on the colocalization of TCR and CD28, which initiate the T-cell activation. Therefore, PD-1 plays an extremely important role in the negative regulation of T cells.^{9,10}

Second, the expression of PD-1 and its ligands is widespread. PD-1 can be expressed on T cells, B cells, natural killer T cells, activated monocytes, and dendritic cells (DCs). PD-1 is not expressed on resting T cells. It is induced to express after activation.^{3-5,11} The expression of PD-L1 and PD-L2 differ in patterns and vary across species. Research data on PD-L2 exhibited a smaller interspecies gap, whereas PD-L1 had a larger expression gap between mice and humans. PD-L1 is constitutively expressed on mouse T and B cells, DCs, macrophages, mesenchymal stem cells, and bone marrow-derived mast cells. However, it is minimally expressed on resting naive human CD4 and CD8 T cells.¹² It is upregulated on several cell types after activation.^{2,7} Compared with PD-L1, the expression of PD-L2 is increasingly restricted. It is induced to express on DCs, macrophages, and bone marrow-derived mast cells.^{2,5,9,11} Although PD-1 and PD-L1 are simultaneously expressed on T cells and PD-L1 has a high expression on activated T cells in mouse, and studies revealed that PD-L1 could attenuate TCR-mediated T-cell stimulation, PD-1 is the principal inhibitory molecule after T-cell activation, just like CTLA-4. Binding of PD-1 to its ligand PD-L1 or PD-L2 delivers a negative signal by the recruitment of Src homology 2-domain-containing tyrosine phosphatase 2 to the phosphorylated tyrosine residue in the cytoplasmic region.⁴ Moreover, the result that PD-L1 fails to inhibit PD-1 deficient T cells indicates that PD-L1 conveys suppression signal via its interaction with PD-1.³ Consequently, PD-1 plays a major role in activated T-cell inhibition.

Third, the interaction on T-cell responses is comprehensive complex. It is also related to binding affinity, the expression position, spatial conformation, etc. As far as PD-1 and its ligands are concerned, although PD-1 seems to be an exclusive coinhibitor, PD-L1 and PD-L2 have dual functions of coinhibitors and costimulators. A series of in vivo studies indicate that endogenous PD-L1 could act as coinhibitor^{2,3,13} or costimulatory,^{6,14-17} whereas PD-L2 mainly acts as costimulatory.^{18,19} Some observations strongly suggested that PD-L1 and PD-L2 costimulated T-cell growth through a non–PD-1 receptor.^{20,21} From an affinity point of view, molecular pairs were ranked according to their binding affinity as B7-1: CTLA-4 > PD-1: PD-L1/PD-L2 > PD-L1: B7-1 > B7-1: CD28,^{2,7} which implies that interactions between cosignaling pathway participants on T cells are sequential. Between T cells and antigen-presenting



FIGURE 1. Major T-cell cosignaling pathways. Black lines indicate stimulating signals on T cells, whereas red lines indicate inhibitory signals. CD, cluster of differentiation; CTLA-4, cytotoxic t-lymphocyte-associated antigen-4; ICOS, inducible costimulator; ICOS-L, ICOS ligand; MHC, major histocompatibility complex; PD-1, programmed death 1; PD-L1, programmed death ligand-1; TCR, T-cell receptor.

cells (APCs), it is possible that PD-L1 mainly on APC cell preferentially binds PD-1 on T cell, whereas B7-1 (mainly on APC cell) preferentially binds CTLA-4 on T cell. When PD-L1 and B7-1 are simultaneously expressed on the same cell or expressed on 2 different cells, their interactions are different in the aspect of spatial conformation. The interaction between PD-L1 and B7-1 occurs in cis when they are located in the same cell.^{7,22,23} However, binding between PD-L1 and B7-1 on different cells still remains controversial.²⁴ Some reports suggested that PD-L1 and B7-1 on different cells bound in trans,^{2,23,25} whereas others indicated that the interaction occurred not in trans.^{7,23}

Apart from cosignaling pathway participants (CD28, CD80/86, CTLA-4, PD-1, PD-1L, and inducible costimulator ligand) mentioned above, several novel negative immune regulation molecules have been introduced over the past few years, such as lymphocyte activation gene-3 (LAG-3 or CD223), T-cell immunoglobulin and mucin-domain–containing-3, T-cell immunoglobulin and immunoreceptor tyrosinebased inhibitory motif domain, V-type immunoglobulin domain-containing suppressor of T-cell activation, and so on.²⁶ These negative regulatory molecules provide new targets for inhibiting T-cell activation and are worth our concern.

In summary, there is a dynamic and complex interaction network among PD-1, PD-L1/PD-L2/, B7-1 and CTLA-4. PD-L1 has dual functions as coinhibitor and costimulator.

REFERENCES

- Malvezzi P, Jouve T, Rostaing L. Costimulation blockade in kidney transplantation: an update. *Transplantation*. 2016;100:2315–2323.
- Butte MJ, Keir ME, Phamduy TB, et al. Programmed death-1 ligand 1 interacts specifically with the B7-1 costimulatory molecule to inhibit T cell responses. *Immunity*. 2007;27:111–122.

- Freeman GJ, Long AJ, Iwai Y, et al. Engagement of the PD-1 immunoinhibitory receptor by a novel B7 family member leads to negative regulation of lymphocyte activation. *J Exp Med*. 2000;192:1027–1034.
- Keir ME, Butte MJ, Freeman GJ, et al. PD-1 and its ligands in tolerance and immunity. Annu Rev Immunol. 2008;26:677–704.
- Freeman GJ. Structures of PD-1 with its ligands: sideways and dancing cheek to cheek. Proc Natl Acad Sci USA. 2008;105:10275–10276.
- Haile ST, Bosch JJ, Agu NI, et al. Tumor cell programmed death ligand 1-mediated T cell suppression is overcome by coexpression of CD80. *J Immunol.* 2011;186:6822–6829.
- Chaudhri A, Xiao Y, Klee AN, et al. PD-L1 Binds to B7-1 Only In Cis on the Same Cell Surface. *Cancer Immunol Res.* 2018;6:921–929.
- Chemnitz JM, Parry RV, Nichols KE, et al. SHP-1 and SHP-2 associate with immunoreceptor tyrosine-based switch motif of programmed death 1 upon primary human T cell stimulation, but only receptor ligation prevents T cell activation. *J Immunol.* 2004;173:945–954.
- Boussiotis VA. Molecular and biochemical aspects of the PD-1 checkpoint pathway. N Engl J Med. 2016;375:1767–1778.
- Okazaki T, Honjo T. PD-1 and PD-1 ligands: from discovery to clinical application. *Int Immunol.* 2007;19:813–824.
- Chen L. Co-inhibitory molecules of the B7-CD28 family in the control of T-cell immunity. *Nat Rev Immunol.* 2004;4:336–347.
- Brown JA, Dorfman DM, Ma FR, et al. Blockade of programmed death-1 ligands on dendritic cells enhances T cell activation and cytokine production. *J Immunol*. 2003;170:1257–1266.
- Latchman Y, Wood CR, Chernova T, et al. PD-L2 is a second ligand for PD-1 and inhibits T cell activation. *Nat Immunol.* 2001;2:261–268.
- Kanai T, Totsuka T, Uraushihara K, et al. Blockade of B7-H1 suppresses the development of chronic intestinal inflammation. J Immunol. 2003;171:4156–4163.
- Subudhi SK, Zhou P, Yerian LM, et al. Local expression of B7-H1 promotes organ-specific autoimmunity and transplant rejection. *J Clin Invest*. 2004;113:694–700.
- Dong H, Strome SE, Salomao DR, et al. Tumor-associated B7-H1 promotes T-cell apoptosis: a potential mechanism of immune evasion. *Nat Med*. 2002;8:793–800.
- Haile ST, Horn LA, Ostrand-Rosenberg S. A soluble form of CD80 enhances antitumor immunity by neutralizing programmed death ligand-1 and simultaneously providing costimulation. *Cancer Immunol Res.* 2014;2:610–615.

- Shin T, Kennedy G, Gorski K, et al. Cooperative B7-1/2 (CD80/CD86) and B7-DC costimulation of CD4+ T cells independent of the PD-1 receptor. J Exp Med. 2003;198:31–38.
- Liu X, Gao JX, Wen J, et al. B7DC/PDL2 promotes tumor immunity by a PD-1-independent mechanism. J Exp Med. 2003;197:1721–1730.
- Wang S, Bajorath J, Flies DB, et al. Molecular modeling and functional mapping of B7-H1 and B7-DC uncouple costimulatory function from PD-1 interaction. *J Exp Med*. 2003;197:1083–1091.
- 21. Greenwald RJ, Freeman GJ, Sharpe AH. The B7 family revisited. Annu Rev Immunol. 2005;23:515–548.
- Nishimura CD, Pulanco MC, Cui W, et al. PD-L1 and B7-1 Cisinteraction: new mechanisms in immune checkpoints and immunotherapies. *Trends Mol Med.* 2021;27:207–219.
- Zhao Y, Lee CK, Lin CH, et al. PD-L1:CD80 Cis-heterodimer triggers the co-stimulatory receptor CD28 while repressing the inhibitory PD-1 and CTLA-4 pathways. *Immunity*. 2019;51:1059– 1073.e9.
- Sugiura D, Maruhashi T, Okazaki IM, et al. Restriction of PD-1 function by cis-PD-L1/CD80 interactions is required for optimal T cell responses. *Science*. 2019;364:558–566.
- Park JJ, Omiya R, Matsumura Y, et al. B7-H1/CD80 interaction is required for the induction and maintenance of peripheral T-cell tolerance. *Blood*. 2010;116:1291–1298.
- Morad G, Helmink BA, Sharma P, et al. Hallmarks of response, resistance, and toxicity to immune checkpoint blockade. *Cell*. 2021;184:5309–5337.