

MINI REVIEW

Tumor-associated macrophages: An important player in breast cancer progression

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Abstract

Breast cancer is the most common form of malignant tumor in females, accounting for the second highest mortality among cancer patients. In the breast tumor microenvironment, tumor-associated macrophages (TAMs) are the most abundant immune cells, which regulate the progression of breast cancer. During breast cancer tumorigenesis and progression, TAMs support breast tumor growth by promoting angiogenesis and cancer cell metastasis, inducing cancer stemness, regulating energy metabolism, and supporting immune system suppression. TAMs exhibit a high degree of cellular plasticity. Repolarizing tumor-related macrophages into M1 macrophages can promote tumor regression. This study reviews the role and mechanism of action of TAMs in the development of breast cancer and establishes TAMs as effective targets for breast cancer treatment.

KEYWORDS

breast cancer, tumor immune microenvironment, tumor-associated macrophages (TAMs)

INTRODUCTION

The global cancer incidence and prevalence statistics of 2020 indicated that breast cancer has surpassed lung cancer to become the most common form of cancer prevalent in women today.¹ Histological classification of breast cancer based on the expression of estrogen receptor (ER), progesterone receptor (PR), and/or human epidermal growth factor receptor-2 (HER2) has been established as the gold standard of cancer diagnosis which is clinically used to classify breast cancer as luminal A, luminal B, HER2-positive and basal-like triple negative breast cancer (BL/TNBC).² Clinical efforts to characterize the features of the breast tumor microenvironment (TME) have confirmed the presence of tumor cells and the active recruitment of host immune cells such as tumor-associated macrophages (TAMs), T cells, natural killer (NK) cells, B cells, granulocytes, plasma cells, and basophils, respectively.³ TAMs are prominent components of the TME, comprising over 50% of the total infiltrating immune cells in some cases, and can affect the progression of breast cancer through diverse mechanisms.^{4,5} Macrophages as a heterogeneous cell population are differentiated into two functionally distinct subtypes which respond to different environmental factor-based

stimuli to form classical activated M1 macrophages or alternatively activated M2 macrophages, respectively.^{6,7}

Traditionally, M1 macrophages exert tumor-killing functions via cancer cell recognition and phagocytosis accompanied with production of proinflammatory cytokine molecules such as interferon γ (IFN- γ) and interleukin-12 (IL-12).⁸⁻¹⁰ During tumor progression, the number of M2 macrophages increase and they become the dominant type of TAM in the TME. M2 macrophages are generally regarded as “tumor promoters”, which support the progression of breast cancer by promoting tumor cell invasion and metastasis, angiogenesis, cancer stemness, regulating energy metabolism, and supporting immune system evasion.^{5,11-14} TAM-based infiltration in the primary tumor has been associated with inferior patient prognoses and treatment outcomes.^{5,11-14} In this review, we summarize the functional aspects of TAMs in the development of breast cancer as shown in Figure 1 which may be utilized in breast cancer diagnosis and prevention.

BREAST CANCER ANGIOGENESIS

Angiogenesis involves the formation of new blood vessels which support tumor growth and development. TAMs act



FIGURE 1 TAM-associated mechanisms which promote the development of breast cancer

as important players in angiogenesis by closely associating themselves with high-density vascular networks formed in breast cancer. In breast cancer TME, TAMs are an important source of vascular endothelial growth factor (VEGF).^{15,16} The interactions of VEGF with vascular endothelial growth factor receptors (VEGFRs) triggers angiogenesis in breast cancer. Therefore, inhibiting potential VEGF/VEGFR interactions can significantly block angiogenesis and tumor metastasis.^{17,18} The macrophage colony-stimulating factor or colony-stimulating factor 1 (CSF-1) drives the recruitment and the differentiation of macrophages towards a M2 phenotype. During neoplasm development in the mammary gland, the application of colony-stimulating factor 1 receptor (CSF1-R) inhibitors can

deplete TAMs to effectively inhibit metastasis, angiogenesis and reduce the invasiveness of the tumor.¹⁹

Hypoxia is a hallmark of the TME that promotes angiogenesis and leads to efficient recruitment of macrophages.²⁰ The hypoxic environment activates macrophages to transform into TAMs stimulating the upregulation of hypoxia-inducible factors (HIFs) in TAMs, which act as transcriptional activators of VEGF.^{21,22} VEGF facilitates hypoxic microenvironment-based angiogenesis which supports oxygen and nutrient delivery to the tumor, promoting its growth.^{21,22} Early evidence of the role of HIF signaling has been correlated with angiogenesis, inhibition of the HIF-1 α signaling which impedes angiogenesis and tumor growth. Interestingly, inhibition of the HIF-2 α signaling leads to the

formation of highly disordered blood vessels and aggravation of the hypoxic condition in the TME.²⁰ Additionally, breast cancer cells in the hypoxic TME upregulate the expression of activating transcription factor 4 (ATF4), a member of the ATF/cAMP response element-binding protein (CREB) family, which has been reported to be related to the recruitment of macrophages and promotion of angiogenesis which indirectly support tumor growth.²³ Thus, TAMs increase the malignancy of tumors by promoting angiogenesis.

BREAST CANCER CELL METASTASIS

Metastasis is the primary cause of death in breast cancer patients. TAMs play a key role in promoting metastasis and invasion in breast cancer. Targeting TAMs has previously been suggested as a potential therapeutic strategy for the treatment of metastatic breast cancer.^{5,24} TAMs facilitate tumor metastasis by chemokine (C-C motif) ligand2 (CCL2), CCL5, and CCL18, respectively. The functional mechanism of CCL2 involves the promotion of metastasis in breast cancer cells to bone and lung tissue. The CCL2-expressing breast tumor cells recruit C-C motif chemokine receptor 2+ (CCR2+) macrophages to accumulate in the lung and regulate osteoclast differentiation in the bone, playing a significant role in premetastatic niche formation by cancer cell colonization. Thus, inhibition of CCL2-CCR2 may effectively inhibit tumor metastasis.^{25,26} Breast cancer cells that secrete CCL5 act on mononuclear macrophages towards TAMs which can promote tumor migration and invasion.²⁷ CCL18 is abundantly released by TAMs, and its expression in TME is associated with tumor metastasis and decreased patient survival. The PYK2 N-terminal domain-interacting receptor 1 (PITPNM3), which is the functional receptor of CCL18, inhibits the metastatic and invasive effects exerted by CCL18.²⁸ Nie and colleagues reported the existence of positive feedback loops of CCL5-CCR5 and CCL18-PITPNM3 between malignant phyllodes tumors (PT) of the breast and TAMs, while assisting in maintaining TAM phenotype as well as PT aggressiveness. Their study reported the use of CCR5 inhibitor and CCL18 monoclonal antibody to double-block the CCL5-CCR5 and CCL18-PITPNM3 pathways, which led to significant suppression of tumor metastasis.²⁹ TAMs secrete cellular cytokines and surface receptors which are important factors promoting breast cancer metastasis. High epidermal growth factor (EGF) expression in TAMs activates epidermal growth factor receptors (EGFRs) in the cancer cells which in turn promotes metastasis and CSF-1 secretion. The CSF-1 recruits and activates TAMs to further secrete EGF, which suggests the existence of an EGF/CSF-1 positive feedback loop between TAMs and cancer cells. EGF induces the infiltration of breast cancer cells into the blood vessels, leading blood vessel metastasis.³⁰ A group of matrix-metalloproteinases (MMPs), such as MMP2, MMP7 and MMP9 are secreted by TAMs, which have been

demonstrated to be involved in the degradation matrix components of the TME, promoting the metastasis of tumor cells and the formation of the metastatic microenvironment.¹⁵ High expression of the scavenger receptor named macrophage receptor with collagenous structure (MARCO) by suppressive TAMs promotes tumor growth and metastasis. MARCO is closely associated with metastasis driving gene signatures for epithelial-mesenchymal-transition (EMT), and targeted blocking of MARCO expression can effectively inhibit tumor metastasis.³¹

TAMS PROMOTE BREAST CANCER CELL STEMNESS

TME consists of a large number of immunosuppressive cells (mainly TAMs). There is evidence to support that TAMs induce and maintain cancer stem cells (CSCs), thereby promoting tumorigenesis, proliferation, and self-renewal.^{5,32,33} There is a vast body of published literature that supports the involvement of the various TAM-based cytokines in the generation of breast CSCs. It was earlier thought that classical “M1” activation exerts antitumor effects via proinflammatory cytokines which prevent tumor progress. A recent study by Guo and colleagues showed that the proinflammatory effects of M1 can also trigger the expansion and self-renewal of CSCs. Coculture of breast cancer cells with M1 macrophages induced the formation of aldehyde dehydrogenase 1+ (ALDH1+) breast CSCs through inflammatory cytokine activation of the Lin-28B-let-7-HMGA2 pathway, and these breast CSCs were highly drug-resistant with elevated spheroid forming capability. Their study also suggested that the M1 phenotype repolarized into the M2 phenotype to maintain a high population of ALDH1+ breast CSCs.³⁴ The IL-6 from TAMs can promote the transformation of human and mouse nonstem cancer cells (NSCC) into CSCs by activating the JAK/STAT pathway to enhance the self-renewal and tumorigenic capacity of CSCs.^{35,36} Immune-suppressing M2-like macrophages in inflammatory breast cancer (IBC) have been found to secrete high levels of IL-8 and growth-regulated oncogene (GRO) chemokines which activate the STAT3 pathway, and are the main driving force for the formation of the CSCs.³⁷

Additionally, TAMs also promote breast cancer cell stemness by upregulating the expression of the SRY-related HMG-box (SOX) family of transcription factors (TFs) and surface receptors. The EGF secreted by the TAMs activates the EGFR/STAT3/SOX-2 paracrine pathway in the breast cancer, resulting in increased SOX-2 expression, which in turn enhances the CSC phenotype in the tumor cells.³⁸ The existing body of research on SOX-2, OCT-4 and NANOG suggests that early-stage breast tumors exhibit SOX-2 expression, with no expression of OCT-4 and NANOG. Overexpression of SOX-2 increased the spheroid-forming ability and self-renewal in CSCs,³⁹ suggesting that SOX-2 is a key molecule regulating the formation of CSCs in early breast cancer. Transforming growth factor- β (TGF- β)

upregulates SOX-4 expression during the EMT process, and SOX4 directly enhances the expression of histone methyltransferase EZH2. Overexpression of EZH2 is essential in stem cell self-renewal and the expansion of CSCs in breast cancer.^{40,41} The ephrin type-A receptor 4 (EPHA4) protein on the surface of TAMs is upregulated during EMT and binds directly to the receptor on cancer cells, which activates the NF- κ B pathway in cancer cells to facilitate the maintenance of homeostasis in the CSCs.¹⁰

TAMS MODULATE T CELL ACTIVITY TO INDUCE IMMUNOSUPPRESSIVE MICROENVIRONMENT IN BREAST CANCER

The immunomodulatory function of TAMs is a major mechanism of cancer disease progression and the main area of focus here is the regulation of the tumor-killing function of effector T cells.⁵ TAMs regulate arginine metabolism as an important way to suppress T cell function. The expression level of arginase-1 (ARG-1), a molecular marker of M2 macrophages, has been reported to be significantly higher in breast cancer patients compared with healthy controls. The level of L-arginine decreases to suppress the function of effector T cells in condition of ARG-1 hydrolyzes L-arginine.⁴² In addition to ARG-1, nitric oxide synthase (iNOS), a molecular marker of M1 macrophages, metabolizes L-arginine to form the product NO, which inhibits the function of effector T cells.¹⁰

Expression of immune checkpoints, such as programmed cell death protein 1 (PD-1), is an important way for TAMs to regulate the tumor-killing function of T cells.⁶ Several studies have investigated the ability of TAMs to modulate the expression of PD-1/programmed death-ligand 1 (PD-L1) via several cytokines in the breast TME. For example, IFN- γ secreted by TAMs activates the JAK/STAT3 and PI3K/AKT pathways to upregulate PD-L1 expression. TGF- β , a multifunctional cytokine, induces macrophage polarization to M2, thereby enhancing the suppressive activity of TAMs while inducing upregulation of PD-L1 promoting tumor escape. In the IL-6 deficient condition, PD-L1 expression was significantly upregulated, and treatment with an anti-PD-L1 antibody proved to be remarkably effective.¹⁴ Moreover, deficiency of macrophage common lymphatic endothelial and vascular endothelial receptor-1 (CLEVER-1) markedly impedes tumor development via activation of the tumor-killing ability in effector T cells.⁴³

The TAMs play an important role in cancer disease progression since they can exhaust CD8⁺ T cells, leading them to lose their ability to eliminate cancer cells.^{44,45} Thus, as a potential therapeutic rationale in the development of cancer immunotherapy, it is necessary to elucidate the mechanism by which TAMs cause T cell exhaustion. In the TNBC-based study conducted by Xu and colleagues, the interaction between TAMs and exhausted T cells were demonstrated using single-cell transcriptome analysis. The findings

indicated that lymphocyte activating 3 (LAG3) and T cell immunoglobulin and mucin domain-containing protein 3 (TIM3) were enriched during T cell exhaustion when compared to PD-1 and CTLA-4, providing targets for potential immune-based therapies.⁴⁶ Calcium/calmodulin-dependent protein kinase kinase (CaMKK2), highly expressed within TAMs in breast cancer, can suppress proliferation and T cell tumor killing function.⁴⁷ Additionally, high cyclooxygenase-2 (COX-2) expressing hepatocellular carcinoma cell lines can induce M2 TAMs polarization, which can contribute to the exhaustion of the antitumor abilities in activated CD8⁺ T cells.⁴⁸ When the number of TAMs in the stroma increases, the cells secrete STAT3 into the TME, causing CD8⁺ T cell exhaustion.⁴⁴ Similarly, Pu et al asserted that TAM-derived extracellular vesicles (EVs) promoted CD8⁺ T cell exhaustion in a hepatocellular carcinoma (HCC) mice model. The microRNA-21-5p (miR-21-5p) expression was upregulated in EVs that were carried into tumor tissues. Inhibition of miR-21-5p blocked the tumor-promoting effect of TAMs.⁴⁹ Another study demonstrated that exosomal microRNA-146a-5p (miR-146a-5p) from TAMs drives T cell exhaustion in HCC.⁵⁰

TAMs are involved in tumor immune regulation by numerous potential mechanisms. TAMs and myeloid-derived suppressor cells (MDSCs) exert their immunosuppressive effects in a cell contact-dependent manner. Skewed macrophages which transform into TAMs can be induced by MDSCs, and are characterized by downregulation of IL-12 expression. TAMs stimulate MDSCs to upregulate IL-10 expression, resulting in secretion of IL-12 in macrophages further downregulating, forming the self-perpetuating negative loop damage effector T cell function.⁵¹ TAMs blunt the function of effector T cells through secretion of IL-10 from TAMs inhibits IL-12 production by dendritic cells, leading to blunting of effector T cell function by TAMs.⁵² TAMs play a critical role in suppressing T cell recruitment; however, the potential mechanism of action is still unknown. Targeting the CSF1/CSF1R pathway can obstruct macrophage recruitment and enhance T cell infiltration during chemotherapy or high-dose irradiation. Similar results were observed when blocking the CCL2/CCR2 pathway which led to macrophage recruitment.⁵³ Additionally, classically activated macrophages can be induced by Th1 cytokines (IFN- γ and TNF- α), while alternatively activated macrophages can be induced by Th2 cytokines (IL-13 and IL-4).⁵⁴ As previously mentioned, TAMs are involved in the immunosuppression of breast cancer and can protect cancer cells.

TAMS REGULATE ENERGY METABOLISM IN BREAST CANCER CELLS

TAMs impact the overall metabolic profile of the TME through modulation of metabolic activities and metabolites which can influence tumor development.⁶ A large number of macrophages localize significantly in the hypoxic tumor regions and the lactic acid produced by glycolysis in the

cancer cells cause them to polarize into M2 phenotype.¹⁰ Lactate-activated macrophages promote the secretion of CCL5 by activating the Notch pathway. CCL5 play a key role in translating the metabolic communication between TAMs and breast cancers, increasing aerobic glycolysis, migration and invasiveness of the cancer cells. Blocking the CCL5-CCR5 axis with monoclonal antibodies disrupts the glycolytic metabolic cycle and inhibits cancer cell metastasis.⁵⁵ The G protein-coupled receptor 132 (Gpr132), expressed by TAMs in a high-lactate environment, is a key sensor of the rising lactate levels in TME, mediated the interaction between cancer cells and TAMs during metastasis. Lactate-activated Gpr132 is involved in M2-type macrophage polarization.⁵⁶ TAMs upregulate the expression of HIFs, which assists them in adapting to the hypoxic TME. The HIFs function as important regulatory players modulating the tumor energy metabolism.²¹ HIF-1 α drives glycolytic metabolism in metastatic breast cancer cells and promotes metastasis and colonization of cancer cells to the liver.⁵⁷ Chen and colleagues have demonstrated that RNA is also involved in aerobic glycolysis. TAMs promote aerobic glycolysis in breast cancer via HIF-1 α -stabilizing long noncoding RNA (HISLA). The lactate released from glycolytic tumor cells further induces expression of HISLA in macrophages, creating a positive feedback loop for glycolysis that enhances drug resistance in cancer cells.⁵⁸ Additionally, HIF-2 α expression activates mitochondrial oxidative phosphorylation in tumor cells and over-activation of mitochondrial oxidative phosphorylation is a marker for an aggressive form of breast cancer.⁵⁹ Therefore, HIFs are important energy metabolic targets for breast cancer treatment.

REPOLARIZATION OF TAMs INTO M1 TYPE MACROPHAGES EXERT TUMOR-KILLING EFFECTS

The altered TME causes the TAMs to polarize into M1 macrophages and mediate an antitumor immune response.¹¹ M1 macrophages highly express proinflammatory factors, such as IL-6, IL-12, iNOS, reactive oxygen species (ROS), TNF- α , which can exert effects to kill tumor cells.⁸ M1 macrophages have a stronger antigen-presenting ability because they express major histocompatibility complex (MHC) class II.⁵⁴ Repolarization of TAMs into M1 macrophages can inhibit tumor progression by exploiting the plasticity of TAMs.¹⁹ The maintenance of the immunosuppressive phenotype of TAMs is closely related to the NF- κ B signaling pathway. When NF- κ B signaling is inhibited specifically in TAMs, they repolarize to M1 macrophages and abundantly secrete IL-12. The IL-12 can activate and recruit NK cells to perform tumor-killing functions in advanced tumors.⁶⁰ Upregulation of miR-155 expression levels drives repolarization of TAMs to M1 macrophages to regain tumor-killing functions.⁶¹ Paclitaxel converts TAMs into M1 macrophages via the Toll-like receptor 4 (TLR4) pathway.⁶² Exosomes of M1 macrophages have been reported to enhance the

therapeutic effect of paclitaxel in breast cancer through macrophage-mediated inflammation.⁶³ The combination of anti-CD40 with anti-CSF-1R immunotherapy has been reported to prompt the TAMs to polarize towards a proinflammatory phenotype with antitumor functionality, significantly enhancing the antitumor response and prolonging the survival in patients.⁶⁴ A recent study showed that anti-Her2 antibody alone was able to upregulate PD-L1 in macrophages which led to immunosuppression and poor prognosis. Interestingly, a combination of therapeutic antibodies and anti-PD-L was shown to be beneficial.⁶⁵ Traditional Chinese medicine can promote repolarization of TAMs, and may serve as a novel treatment modality for breast cancer treatment in the future.⁶⁶ An important example of an effective Chinese herbal medicine-based anticancer agent is emodin, which exerts antitumor effects in breast cancer by inhibiting the TGF- β 1 production in the macrophages which in turn suppresses TAM polarization.^{32,67} Additionally, XIAOPI formula (XPS) is being extensively used as a promising traditional Chinese medicine-based therapy in breast cancer treatment. Baohuoside I (BHS) is the key bioactive compound of XPS. Functional studies have revealed that BHS can suppress the M2 phenotype polarization of TAMs to significantly inhibit the migration and invasion of breast cancer cells.⁶⁸

FUTURE DEVELOPMENT

One of the main limitations of targeting TAMs for cancer therapy is the lack of reliable and specific markers. Cassetta et al. used multicolor flow cytometric analysis to determine the sialic acid-binding Ig-like lectin 1 (SIGLEC1) protein expression in breast cancer patients and found that SIGLEC1 was exclusively expressed by TAMs. Furthermore, in the circulation, both classical and nonclassical monocytes exhibited low expression of SIGLEC1, with no difference between cancer and noncancer patients, indicating the specificity of SIGLEC1 to macrophages/TAMs.⁶⁹ Additionally, breast cancer cells overexpress CD24, while TAMs express high levels of Siglec-10. Genetic ablation of Siglec-10 robustly resulted in a macrophage-dependent reduction of tumor growth.⁷⁰ This study emphasizes the existing knowledge concerning the role of TAMs in breast cancer and attempts to identify unique genes expressed by human TAMs to uncover novel therapeutic targets.

CONCLUSIONS

This article summarizes the roles played by TAMs in breast cancer development by promoting TME angiogenesis and cancer cell metastasis, inducing cancer cell stemness, regulating energy metabolism, and supporting immune system suppression. Macrophage function and polarization are regulated by multiple TME-based factors. TAMs are important players in tumor progression which should be explored with

the aim of developing improved therapies for breast cancer treatment. Macrophages can adopt different states of activation. Repolarization of TAMs into antitumorigenic M1 macrophages is a very promising therapeutic option. A recent study conducted by Xiao et al. showed the presence of a high proportion of M2-like TAMs reaching 43.1% in control groups. In comparison, M2-like TAMs decreased to 10.7% in the treatment group by M2 repolarizing to M1. In addition, the proportion of M1 macrophages increased from 10.2% to 58%,⁷¹ which apparently contributed to the effective inhibition of tumor growth and metastasis with low immune side-effects. In future, combination treatment modalities involving traditional chemotherapeutic drugs and traditional Chinese medicine targeted at promoting repolarization of TAMs can serve as a novel treatment modality for breast cancer treatment. Therefore, exploring the role and mechanism of action of TAMs in the development of breast cancer can provide a foundation for better treatment of breast cancer.

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CONFLICT OF INTEREST

The authors confirm that there are no conflicts of interest.

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REFERENCES

- Sung H, Ferlay J, Siegel RL, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *Cancer J Clin*. 2021;2021(0):1–41. Epub 2021/02/05. PubMed PMID: 33538338.
- Jiang G, Zhang S, Yazdanparast A, et al. Monocyte-derived macrophages promote breast cancer bone metastasis outgrowth. *J Exp Med*. 2020;217(11): Epub 2020/08/12. PubMed PMID: 32780802; PubMed Central PMCID: PMCPCMC7596825.
- Wagner J, Rapsomaniki MA, Chevrier S, et al. A single-cell atlas of the tumor and immune ecosystem of human breast cancer. *Cell*. 2019;177(5):1330–45.e18. Epub 2019/04/16. PubMed PMID: 30982598; PubMed Central PMCID: PMCPCMC6526772.
- Qiu SQ, Waaijer SJH, Zwager MC, et al. Tumor-associated macrophages in breast cancer: innocent bystander or important player? *Cancer Treat Rev*. 2018;70:178–89. Epub 2018/09/19. PubMed PMID: 30227299.
- Chen Y, Song Y, Du W, et al. Tumor-associated macrophages: an accomplice in solid tumor progression. *J Biomed Sci*. 2019;26(1):78 Epub 2019/10/21. PubMed PMID: 31629410; PubMed Central PMCID: PMCPCMC6800990.
- Vitale I, Manic G, Coussens LM, et al. Macrophages and metabolism in the tumor microenvironment. *Cell Metab*. 2019;30(1):36–50. Epub 2019/07/04. PubMed PMID: 31269428.
- Pan Y, Yu Y, Wang X, et al. Tumor-associated macrophages in tumor immunity. *Front Immunol*. 2020;11:583084 Epub 2020/12/29. PubMed PMID: 33365025; PubMed Central PMCID: PMCPCMC7751482.
- Sousa S, Brion R, Lintunen M, et al. Human breast cancer cells educate macrophages toward the M2 activation status. *Breast Cancer Res*. 2015;17(1):101 Epub 2015/08/06. PubMed PMID: 26243145; PubMed Central PMCID: PMCPCMC4531540.
- Yang Y, Guo J, Huang L. Tackling TAMs for cancer immunotherapy: it's nano time. *Trends Pharmacol Sci*. 2020;41(10):701–14. Epub 2020/09/19. PubMed PMID: 32946772; PubMed Central PMCID: PMCPCMC7652091.
- Choi J, Gyamfi J, Jang H, et al. The role of tumor-associated macrophage in breast cancer biology. *Histol Histopathol*. 2018;33(2):133–45. PubMed PMID: 28681373.
- Yu T, Di G. Role of tumor microenvironment in triple-negative breast cancer and its prognostic significance. *Chin J Cancer Res = Chung-kuo yen cheng yen chiu*. 2017;29(3):237–52. Epub 2017/07/22. PubMed PMID: 28729775; PubMed Central PMCID: PMCPCMC5497211.
- Ma RY, Zhang H, Li XF, et al. Monocyte-derived macrophages promote breast cancer bone metastasis outgrowth. *J Exp Med*. 2020;217(11). Epub 2020/08/12. PubMed PMID: 32780802; PubMed Central PMCID: PMCPCMC7596825.
- Linde N, Casanova-Acebes M, Sosa MS, et al. Macrophages orchestrate breast cancer early dissemination and metastasis. *Nat Commun*. 2018;9(1):21 Epub 2018/01/04. PubMed PMID: 29295986; PubMed Central PMCID: PMCPCMC5750231.
- Santoni M, Romagnoli E, Saladino T, et al. Triple negative breast cancer: key role of tumor-associated macrophages in regulating the activity of anti-PD-1/PD-L1 agents. *Biochim Biophys Acta Rev Cancer*. 2018;1869(1):78–84. Epub 2017/11/12. PubMed PMID: 29126881.
- Salmaninejad A, Valilou SF, Soltani A, et al. Tumor-associated macrophages: role in cancer development and therapeutic implications. *Cell Oncol (Dordr)*. 2019;42(5):591–608. Epub 2019/05/31. PubMed PMID: 31144271.
- Dallavalasa S, Beeraka NM, Basavaraju CG, et al. The role of tumor associated macrophages (TAMs) in cancer progression, Chemoresistance, angiogenesis and metastasis - current status. *Curr Med Chem*. 2021;xxxx(xx):1–34. Epub 2021/07/26. PubMed PMID: 34303328.
- Farzaneh Behelgard M, Zahri S, Gholami Shahvir Z, et al. Targeting signaling pathways of VEGFR1 and VEGFR2 as a potential target in the treatment of breast cancer. *Mol Biol Rep*. 2020;47(3):2061–71. Epub 2020/02/20. PubMed PMID: 32072404.
- Song Y, Tang C, Yin C. Combination antitumor immunotherapy with VEGF and PIGF siRNA via systemic delivery of multi-functionalized nanoparticles to tumor-associated macrophages and breast cancer cells. *Biomaterials*. 2018;2018(185):117–32. Epub 2018/09/22. PubMed PMID: 30241030.
- Genard G, Lucas S, Michiels C. Reprogramming of tumor-associated macrophages with anticancer therapies: radiotherapy versus chemo- and immunotherapies. *Front Immunol*. 2017;8:828 Epub 2017/08/05. PubMed PMID: 28769933; PubMed Central PMCID: PMCPCMC5509958.
- LaGory EL, Giaccia AJ. The ever-expanding role of HIF in tumour and stromal biology. *Nat Cell Biol*. 2016;18(4):356–65. Epub 2016/03/31. PubMed PMID: 27027486; PubMed Central PMCID: PMCPCMC4898054.
- Fang HY, Hughes R, Murdoch C, et al. Hypoxia-inducible factors 1 and 2 are important transcriptional effectors in primary macrophages experiencing hypoxia. *Blood*. 2009;114(4):844–59. Epub 2009/05/21. PubMed PMID: 19454749; PubMed Central PMCID: PMCPCMC2882173.
- Tamura R, Tanaka T, Akasaki Y, et al. The role of vascular endothelial growth factor in the hypoxic and immunosuppressive tumor microenvironment: perspectives for therapeutic implications. *Med Oncol (Northwood, London, England)*. 2019;37(1):2 Epub 2019/11/13. doi: 10.1007/s12032-019-1329-2. PubMed PMID: 31713115.

23. Liu C, Li Z, Wang L, et al. Activating transcription factor 4 promotes angiogenesis of breast cancer through enhanced macrophage recruitment. *BioMed Res Int.* 2015;2015:974615 Epub 2015/04/18. PubMed PMID: 25883982; PubMed Central PMCID: PMCPCMC4391610.
24. Sánchez-González I, Bobien A, Molnar C, et al. miR-149 suppresses breast cancer metastasis by blocking paracrine interactions with macrophages. *Cancer Res.* 2020;80(6):1330–41. Epub 2020/01/09. PubMed PMID: 31911555.
25. Lu X, Kang Y. Chemokine (C-C motif) ligand 2 engages CCR2+ stromal cells of monocytic origin to promote breast cancer metastasis to lung and bone. *J Biol Chem.* 2009;284(42):29087–96. Epub 2009/09/02. PubMed PMID: 19720836; PubMed Central PMCID: PMCPCMC2781454.
26. Li D, Ji H, Niu X, et al. Tumor-associated macrophages secrete CC-chemokine ligand 2 and induce tamoxifen resistance by activating PI3K/Akt/mTOR in breast cancer. *Cancer Sci.* 2020;111(1):47–58. Epub 2019/11/12. PubMed PMID: 31710162; PubMed Central PMCID: PMCPCMC6942430.
27. An G, Wu F, Huang S, et al. Effects of CCL5 on the biological behavior of breast cancer and the mechanisms of its interaction with tumor-associated macrophages. *Oncol Rep.* 2019;42(6):2499–511. Epub 2019/10/04. PubMed PMID: 31578575; PubMed Central PMCID: PMCPCMC6826325.
28. Chen J, Yao Y, Gong C, et al. CCL18 from tumor-associated macrophages promotes breast cancer metastasis via PITPNM3. *Cancer Cell.* 2011;19(4):541–55. Epub 2011/04/13. doi: 10.1016/j.ccr.2011.02.006. PubMed PMID: 21481794; PubMed Central PMCID: PMCPCMC3107500.PMCPCMC3107500.
29. Nie Y, Huang H, Guo M, et al. Breast phyllodes tumors recruit and repolarize tumor-associated macrophages via secreting CCL5 to promote malignant progression, which can be inhibited by CCR5 inhibition therapy. *Clin Cancer Res: Off J Am Assoc Cancer Res.* 2019; 25(13):3873–86. Epub 2019/03/21. PubMed PMID: 30890553.
30. Condeelis J, Pollard JW. Macrophages: obligate partners for tumor cell migration, invasion, and metastasis. *Cell.* 2006;124(2):263–6. Epub 2006/01/28. PubMed PMID: 16439202.
31. Georgoudaki AM, Prokopec KE, Boura VF, et al. Reprogramming tumor-associated macrophages by antibody targeting inhibits cancer progression and metastasis. *Cell Rep.* 2016;15(9):2000–11. Epub 2016/05/24. PubMed PMID: 27210762.
32. Liu Q, Hodge J, Wang J, et al. Emodin reduces breast cancer lung metastasis by suppressing macrophage-induced breast cancer cell epithelial-mesenchymal transition and cancer stem cell formation. *Theranostics.* 2020;10(18):8365–81. Epub 2020/07/30. PubMed PMID: 32724475; PubMed Central PMCID: PMCPCMC7381725.
33. Liu D, Lu Q, Wang X, et al. LSECtin on tumor-associated macrophages enhances breast cancer stemness via interaction with its receptor BTN3A3. *Cell Res.* 2019;29(5):365–78. Epub 2019/03/13. PubMed PMID: 30858559; PubMed Central PMCID: PMCPCMC6796923.
34. Guo L, Cheng X, Chen H, et al. Induction of breast cancer stem cells by M1 macrophages through Lin-28B-let-7-HMGA2 axis. *Cancer Lett.* 2019;452:213–25. Epub 2019/03/29. PubMed PMID: 30917918.
35. Iliopoulos D, Hirsch HA, Wang G, et al. Inducible formation of breast cancer stem cells and their dynamic equilibrium with non-stem cancer cells via IL6 secretion. *Proc Natl Acad Sci US A.* 2011;108(4):1397–402. Epub 2011/01/12. PubMed PMID: 21220315; PubMed Central PMCID: PMCPCMC3029760.
36. Iliopoulos D, Hirsch HA, Struhl K. An epigenetic switch involving NF-kappaB, Lin28, Let-7 MicroRNA, and IL6 links inflammation to cell transformation. *Cell.* 2009;139(4):693–706. Epub 2009/11/03. PubMed PMID: 19878981; PubMed Central PMCID: PMCPCMC2783826.
37. Valeta-Magara A, Gadi A, Volta V, et al. Inflammatory breast cancer promotes development of M2 tumor-associated macrophages and cancer mesenchymal cells through a complex chemokine network. *Cancer Res.* 2019;79(13):3360–71. Epub 2019/05/03. PubMed PMID: 31043378; PubMed Central PMCID: PMCPCMC7331114.
38. Yang J, Liao D, Chen C, et al. Tumor-associated macrophages regulate murine breast cancer stem cells through a novel paracrine EGFR/Stat3/Sox-2 signaling pathway. *Stem Cells (Dayton, Ohio).* 2013;31(2):248–58. Epub 2012/11/22. PubMed PMID: 23169551.
39. Leis O, Eguiara A, Lopez-Arribillaga E, et al. Sox2 expression in breast tumours and activation in breast cancer stem cells. *Oncogene.* 2012;31(11):1354–65. Epub 2011/08/09. PubMed PMID: 21822303.
40. Tiwari N, Tiwari VK, Waldmeier L, et al. Sox4 is a master regulator of epithelial-mesenchymal transition by controlling Ezh2 expression and epigenetic reprogramming. *Cancer Cell.* 2013;23(6):768–83. Epub 2013/06/15. PubMed PMID: 23764001.
41. Chang CJ, Yang JY, Xia W, et al. EZH2 promotes expansion of breast tumor initiating cells through activation of RAF1- β -catenin signaling. *Cancer Cell.* 2011;19(1):86–100. Epub 2011/01/11. PubMed PMID: 21215703; PubMed Central PMCID: PMCPCMC3041516.
42. de Boniface J, Mao Y, Schmidt-Mende J, et al. Expression patterns of the immunomodulatory enzyme arginase 1 in blood, lymph nodes and tumor tissue of early-stage breast cancer patients. *Oncoimmunology.* 2012;1(8):1305–12. Epub 2012/12/18. PubMed PMID: 23243594; PubMed Central PMCID: PMCPCMC3518503.
43. Viitala M, Virtakoivu R, Tadayon S, et al. Immunotherapeutic blockade of macrophage cleaver-1 reactivates the CD8(+) T-cell response against immunosuppressive tumors. *Clin Cancer Res Off J Am Assoc Cancer Res.* 2019;25(11):3289–303. Epub 2019/02/14. PubMed PMID: 30755440.
44. Farhood B, Najafi M, Mortezaee K. CD8(+) cytotoxic T lymphocytes in cancer immunotherapy: a review. *J Cell Physiol.* 2019;234(6):8509–21. Epub 2018/12/07. PubMed PMID: 30520029.
45. Hossain MA, Liu G, Dai B, et al. Reinvigorating exhausted CD8(+) cytotoxic T lymphocytes in the tumor microenvironment and current strategies in cancer immunotherapy. *Med Res Rev.* 2021;41(1):156–201. Epub 2020/08/28. PubMed PMID: 32844499.
46. Li C, Xu L. Single-cell transcriptome analysis reveals the M2 macrophages and exhausted T cells and intratumoral heterogeneity in triple-negative breast cancer. *Anticancer Agents Med Chem.* 2021;xxx(xx): 1–20. Epub 2021/06/20. PubMed PMID: 34145996.
47. Racioppi L, Nelson ER, Huang W, et al. CaMKK2 in myeloid cells is a key regulator of the immune-suppressive microenvironment in breast cancer. *Nat Commun.* 2019;10(1):2450 Epub 2019/06/06. PubMed PMID: 31164648; PubMed Central PMCID: PMCPCMC6547743 D.P.M. have applied for a patent covering the use of CaMKK2 inhibitors alone or in combination with immunotherapy for the treatment of cancer. Title: “CaMKK2 inhibitor compositions and methods of using the same. Racioppi, L., Nelson, E.R., Huang, W., Chao, N. and McDonnell, D.P. Provisional Patent Application No.: 62/371,309; August 5, 2016. The remaining authors declare no competing financial interests.
48. Xun X, Zhang C, Wang S, et al. Cyclooxygenase-2 expressed hepatocellular carcinoma induces cytotoxic T lymphocytes exhaustion through M2 macrophage polarization. *Am J Trans Res.* 2021;13(5): 4360–75. Epub 2021/06/22. PubMed PMID: 34150019; PubMed Central PMCID: PMCPCMC8205841.
49. Pu J, Xu Z, Nian J, et al. M2 macrophage-derived extracellular vesicles facilitate CD8+T cell exhaustion in hepatocellular carcinoma via the miR-21-5p/YOD1/YAP/ β -catenin pathway. *Cell Death Discovery.* 2021;7(1):182 Epub 2021/07/21. PubMed PMID: 34282135; PubMed Central PMCID: PMCPCMC8289864.
50. Yin C, Han Q, Xu D, et al. SALL4-mediated upregulation of exosomal miR-146a-5p drives T-cell exhaustion by M2 tumor-associated macrophages in HCC. *Oncoimmunology.* 2019;8(7):1601479 Epub 2019/05/31. PubMed PMID: 31143524; PubMed Central PMCID: PMCPCMC6527304.
51. Ugel S, de Sanctis F, Mandrizzato S, et al. Tumor-induced myeloid deviation: when myeloid-derived suppressor cells meet tumor-associated macrophages. *J Clin Invest.* 2015;125(9):3365–76. Epub 2015/09/02. PubMed PMID: 26325033; PubMed Central PMCID: PMCPCMC4588310.
52. Ruffell B, Chang-Strachan D, Chan V, et al. Macrophage IL-10 blocks CD8+ T cell-dependent responses to chemotherapy by suppressing IL-12 expression in intratumoral dendritic cells. *Cancer Cell.* 2014;

- 26(5):623–37. Epub 2014/12/03. PubMed PMID: 25446896; PubMed Central PMCID: PMC4254570.
53. DeNardo DG, Ruffell B. Macrophages as regulators of tumour immunity and immunotherapy. *Nat Rev Immunol.* 2019;19(6):369–82. Epub 2019/02/06. PubMed PMID: 30718830; PubMed Central PMCID: PMC67339861.
54. Laoui D, Movahedi K, van Overmeire E, et al. Tumor-associated macrophages in breast cancer: distinct subsets, distinct functions. *Int J Dev Biol.* 2011;55(7–9):861–7. Epub 2011/12/14. PubMed PMID: 22161841
55. Lin S, Sun L, Lyu X, et al. Lactate-activated macrophages induced aerobic glycolysis and epithelial-mesenchymal transition in breast cancer by regulation of CCL5-CCR5 axis: a positive metabolic feedback loop. *Oncotarget.* 2017;8(66):110426–43. Epub 2018/01/05. PubMed PMID: 29299159; PubMed Central PMCID: PMC5746394.
56. Chen P, Zuo H, Xiong H, et al. Gpr132 sensing of lactate mediates tumor-macrophage interplay to promote breast cancer metastasis. *Proc Natl Acad Sci U S A.* 2017;114(3):580–5. Epub 2017/01/05. PubMed PMID: 28049847; PubMed Central PMCID: PMC5255630.
57. Dupuy F, Tabariès S, Andrzejewski S, et al. PDK1-dependent metabolic reprogramming dictates metastatic potential in breast cancer. *Cell Metab.* 2015;22(4):577–89. Epub 2015/09/15. PubMed PMID: 26365179.
58. Chen F, Chen J, Yang L, et al. Extracellular vesicle-packaged HIF-1 α -stabilizing lncRNA from tumour-associated macrophages regulates aerobic glycolysis of breast cancer cells. *Nat Cell Biol.* 2019;21(4):498–510. Epub 2019/04/03. PubMed PMID: 30936474.
59. Chiavarina B, Martinez-Outschoorn UE, Whitaker-Menezes D, et al. Metabolic reprogramming and two-compartment tumor metabolism: opposing role(s) of HIF1 α and HIF2 α in tumor-associated fibroblasts and human breast cancer cells. *Cell Cycle (Georgetown, Tex).* 2012; 11(17):3280–9. Epub 2012/08/17. PubMed PMID: 22894905; PubMed Central PMCID: PMC3466527.
60. Hagemann T, Lawrence T, McNeish I, et al. "Re-educating" tumor-associated macrophages by targeting NF-kappaB. *J Exp Med.* 2008; 205(6):1261–8. Epub 2008/05/21 PubMed PMID: 18490490; PubMed Central PMCID: PMC2413024.
61. Cai X, Yin Y, Li N, et al. Re-polarization of tumor-associated macrophages to pro-inflammatory M1 macrophages by microRNA-155. *J Mol Cell Biol.* 2012;4(5):341–3. Epub 2012/07/27. doi: 10.1093/jmcb/mjs044. PubMed PMID: 22831835
62. Wanderley CW, Colón DF, Luiz JPM, et al. Paclitaxel reduces tumor growth by reprogramming tumor-associated macrophages to an M1 profile in a TLR4-dependent manner. *Cancer Res.* 2018;78(20):5891–900. Epub 2018/08/15. PubMed PMID: 30104241.
63. Wang P, Wang H, Huang Q, et al. Exosomes from M1-polarized macrophages enhance paclitaxel antitumor activity by activating macrophages-mediated inflammation. *Theranostics.* 2019;9(6):1714–27. Epub 2019/05/01. PubMed PMID: 31037133; PubMed Central PMCID: PMC6485189.
64. Wiehagen KR, Girgis NM, Yamada DH, et al. Combination of CD40 agonism and CSF-1R blockade reconditions tumor-associated macrophages and drives potent antitumor immunity. *Cancer Immunol Res.* 2017;5(12):1109–21. Epub 2017/11/04. PubMed PMID: 29097420.
65. Su S, Zhao J, Xing Y, et al. Immune checkpoint inhibition overcomes ADCP-induced immunosuppression by macrophages. *Cell.* 2018; 175(2):442–57.e23. Epub 2018/10/06. PubMed PMID: 30290143
66. Wang Y, Smith W, Hao D, et al. M1 and M2 macrophage polarization and potentially therapeutic naturally occurring compounds. *Int. Immunopharmacol.* 2019;70:459–66. Epub 2019/03/13. doi: 10.1016/j.intimp.2019.02.050. PubMed PMID: 30861466.
67. Jia X, Yu F, Wang J, et al. Emodin suppresses pulmonary metastasis of breast cancer accompanied with decreased macrophage recruitment and M2 polarization in the lungs. *Breast Cancer Res Treat.* 2014; 148(2):291–302. Epub 2014/10/15. PubMed PMID: 25311112; PubMed Central PMCID: PMC4224983.
68. Wang S, Wang N, Huang X, et al. Baohuoside i suppresses breast cancer metastasis by downregulating the tumor-associated macrophages/C-X-C motif chemokine ligand 1 pathway. *Phytomedicine: Int J Phytother and phytopharm.* 2020;78:153331 Epub 2020/09/11. PubMed PMID: 32911383.
69. Cassetta L, Fragkogianni S, Sims AH, et al. Human tumor-associated macrophage and monocyte transcriptional landscapes reveal cancer-specific reprogramming, biomarkers, and therapeutic targets. *Cancer Cell.* 2019;35(4):588–602.e10. Epub 2019/04/02. PubMed PMID: 30930117; PubMed Central PMCID: PMC6472943.
70. Barkal AA, Brewer RE, Markovic M, et al. CD24 signalling through macrophage Siglec-10 is a target for cancer immunotherapy. *Nature.* 2019;572(7769):392–6. Epub 2019/08/02. PubMed PMID: 31367043; PubMed Central PMCID: PMC6697206.
71. Xiao H, Guo Y, Li B, et al. M2-like tumor-associated macrophage-targeted Codelivery of STAT6 inhibitor and IKK β siRNA induces M2-to-M1 repolarization for cancer immunotherapy with low immune side effects. *ACS Cent Sci.* 2020;6(7):1208–22. Epub 2020/07/30. PubMed PMID: 32724855; PubMed Central PMCID: PMC7379385.

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