

Supporting Information

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Nicotinamide Mononucleotide Ameliorates Sleep Deprivation-Induced Gut Microbiota Dysbiosis and Restores Colonization Resistance against Intestinal Infections

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Nicotinamide mononucleotide ameliorates sleep deprivation-induced gut microbiota dysbiosis and restores colonization resistance against intestinal infections

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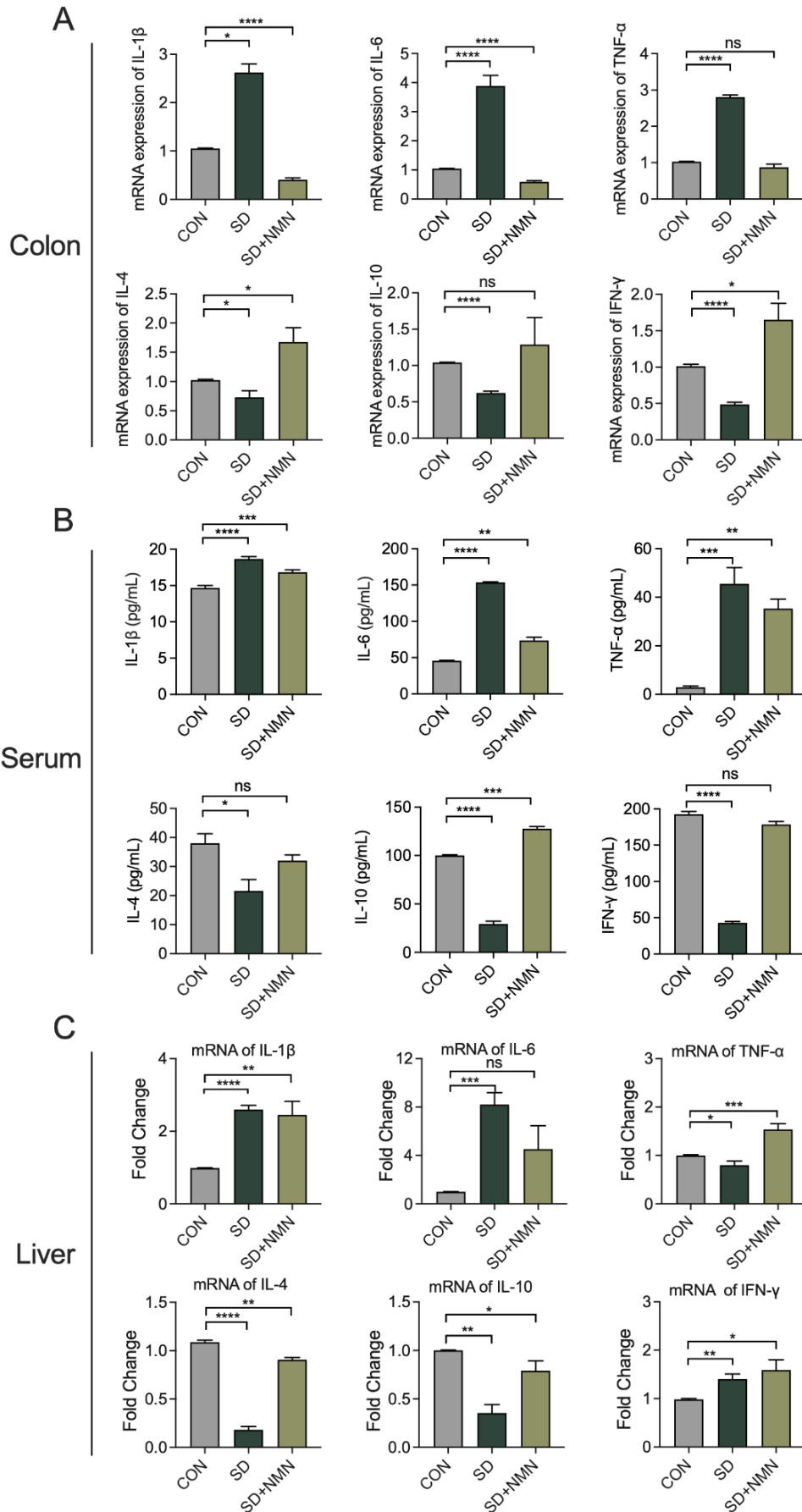


Figure S1. mRNA expression of inflammatory cytokines in the mice.

(A) The mRNA expression of inflammatory cytokines in the colon of mice, including pro-inflammatory cytokines (IL-1 β , IL-6, TNF- α) and anti-inflammatory cytokines (IL-4, IL-10 and INF- γ). mRNA abundances were determined by RT-qPCR and relative gene expressions were normalized with β -actin.

(B) The protein abundance of pro-inflammatory cytokines (IL-1 β , IL-6, TNF- α) and anti-inflammatory cytokines (IL-4, IL-10 and INF- γ) in the serum from CON, SD and SD + NMN groups respectively. Data were representative of three independent experiments with three replicates per group.

(C) Fold changes of mRNA expression of inflammatory cytokines in the livers of mice, including pro-inflammatory cytokines (IL-1 β , IL-6, TNF- α) and anti-inflammatory cytokines (IL-4, IL-10 and INF- γ). mRNA abundances were determined by RT-qPCR and relative gene expressions were normalized with β -actin.

Data are presented as mean \pm SEM and statistical significance are identified by Student's *t*-test. Experiments were performed with biological replicates.

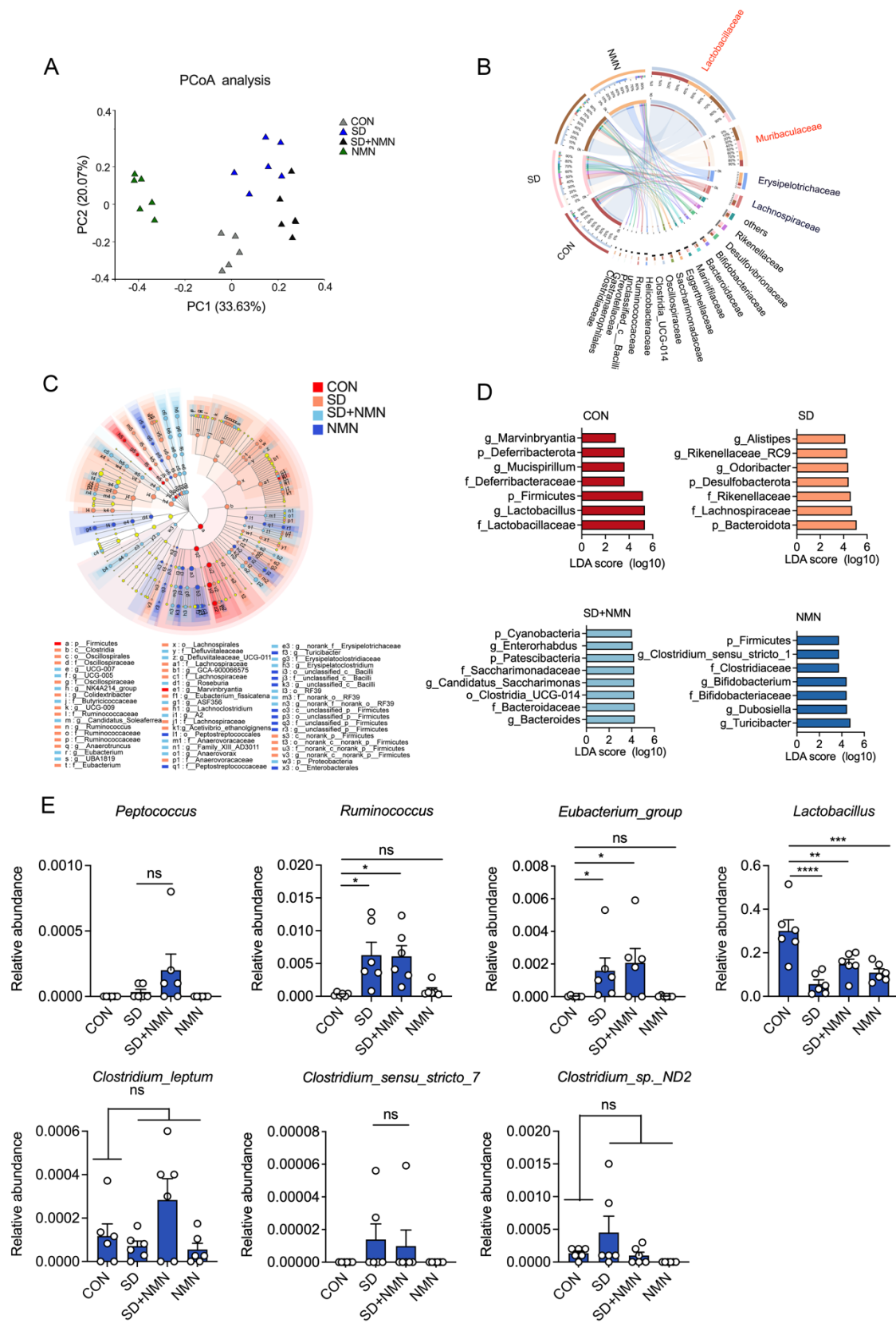


Figure S2. Microbial diversity analysis of altered gut microbiota among four groups.

(A) Principal component analysis (PCA) plot of CON, SD, SD + NMN and NMN samples along principal component (PC) 1 and 2, which explained 52.1% and 20.07% of the total variance, respectively.

(B) Shown are comparisons of the dominant bacterial population at the class level for each group. The width of the bars from each class indicates the relative abundance of that class in each group.

(C) Cladogram generated by LEfSe analysis showing enriched taxa in four groups, including CON (red), SD (orange), SD + NMN (light blue) and NMN (dark blue).

(D) LDA scores of enriched taxa from **C** ($LDA > 2$).

(E) Bile acids metabolism-related gut microbiota abundance in four groups.

Data in **(E)** were displayed as mean \pm SEM, and statistical significance were determined by unpaired *t*-test (* $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$; **** $P < 0.0001$; ns, not significant).

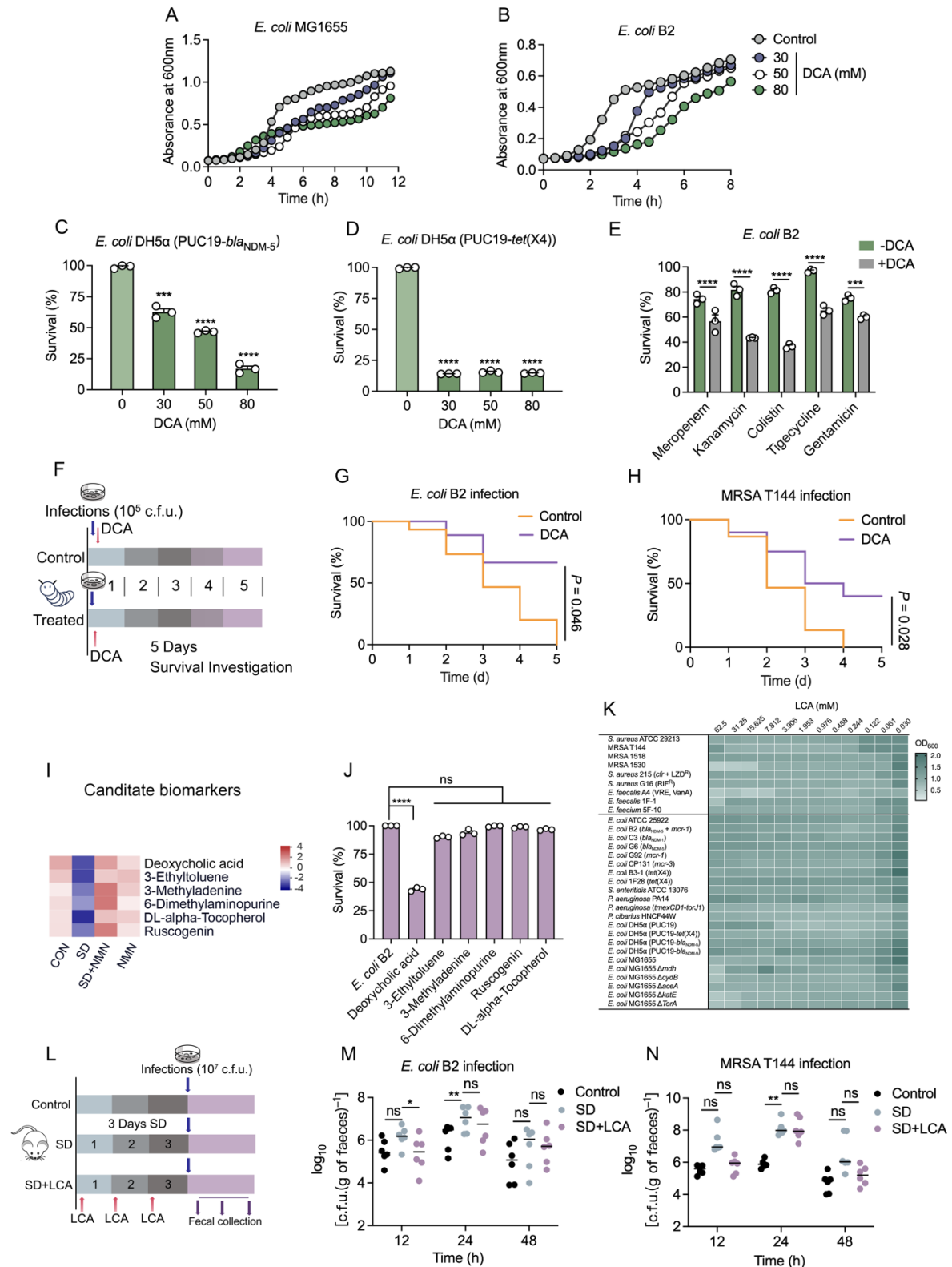


Figure S3. Antimicrobial activity analysis of DCA and LCA.

(A and B) Representative growth curves of *E. coli* MG1655 and *E. coli* B2 in the presence of varying concentrations of DCA. The curves represent averaged values with a <5% margin of error.

(C and D) Percent survival of engineered *E. coli* DH5 α (PUC19-*bla*_{NDM-5}) and *E. coli* DH5 α (PUC19-*tet*(X4)) treated by increasing concentrations of DCA.

(E) DCA exhibits the synergistic effect with different classes of antibiotics, including meropenem, kanamycin, colistin, tigecycline and gentamicin, against *E. coli* B2.

(F) Protocol of the therapeutic potential assessment of single DCA administration in larvae.

(G and H) Monotherapy of DCA (20 μ M/kg) improves the survival rate of *Galleria mellonella* larvae after 5 days post-infection by *E. coli* B2 or MRSA T144 (10⁵ CFUs per larvae).

(I) The potential biomarkers that decreased in the SD group but elevated in the SD+NMN group were presented in the form of a heatmap. Blue to red colors correspond to low to high abundance.

(J) Evaluation of the antimicrobial effect of the candidate metabolites against *E. coli* B2 and only DCA (50 mM) exhibits significant antibacterial properties.

(K) LCA sensitivity analysis in response to individual microbiota strains, including a panel of clinically drug-resistant strains. The value of OD₆₀₀ can be compared relative to a light-green and dark-green gradient. Light green has the lowest value and dark green has the highest value.

(L) Experimental protocols of LCA-supplied intestinal infection model. Three-days LCA administration: 100 mg/kg/day (*i.p.*).

(M and N) Bacterial loads in the feces of mice at 12, 24, 48 h post infection by *E. coli* B2 (**M**) or MRSA T144 (**N**).

Values are presented as mean \pm SEM. Statistical significance was determined by unpaired *t*-test and denoted as follows: **P* < 0.05; ***P* < 0.01; ****P* < 0.001; ns, not significant.

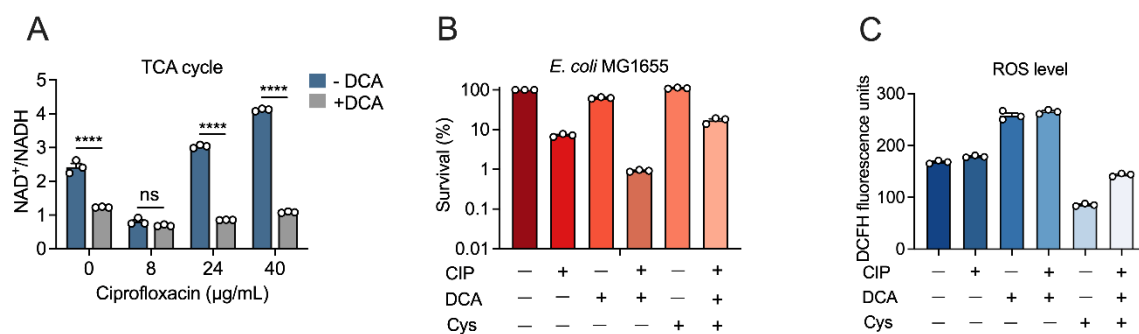


Figure S4. Biochemical factors determination of *E. coli* MG1655 in the presence of DCA and CIP.

(A) Intracellular NAD⁺ /NADH ratio in the presence of CIP with or without DCA.

(B) Comparison of *E. coli* MG1655 survival in the presence of combination treatment with or without L-cys.

(C) Comparison of ROS levels in *E. coli* MG1655 in the presence of combination treatment with or without L-cys.

Values are presented as mean \pm SEM. Statistical significance was determined by unpaired *t*-test and denoted as follows: **P* < 0.05; ***P* < 0.01; ****P* < 0.001; ns, not significant.

Supplementary Tables

Supplementary Table 1 Bacterial strains used in this study.

Organism and phenotypes	Sources/references
Gram-positive bacteria	
<i>S. aureus</i> ATCC 29213	ATCC
MRSA T144	[1]
MRSA 1518	[1]
MRSA 1530	In this study
<i>S. aureus</i> 215 (<i>cfr</i> + LZD ^R)	[1]
<i>S. aureus</i> G16 (RIF ^R)	[1]
<i>E. faecalis</i> A4 (<i>tet</i> (A), <i>vanA</i> , Van ^R)	[2]
<i>E. faecalis</i> 1F-1	[2]
<i>E. faecium</i> 5F-10	[2]
Gram-negative bacteria	
<i>E. coli</i> ATCC 25922	ATCC
<i>E. coli</i> B2 (<i>bla</i> _{NDM-5} + <i>mcr</i> -1)	[3]
<i>E. coli</i> C3 (<i>bla</i> _{NDM-1})	[4]
<i>E. coli</i> G6 (<i>bla</i> _{NDM-5})	[4]
<i>E. coli</i> G92 (<i>mcr</i> -1)	In this study
<i>E. coli</i> CP131 (<i>mcr</i> -3)	In this study
<i>E. coli</i> B3-1 (<i>tet</i> (X4))	[4]
<i>E. coli</i> 1F28 (<i>tet</i> (X4))	In this study
<i>S. enteritidis</i> ATCC 13076	ATCC
<i>P. aeruginosa</i> PA14	In this study
<i>P. aeruginosa</i> (VIM + <i>tmexCD1-torJ1</i>)	[5]
<i>P. cibarius</i> HNC44W (<i>bla</i> _{NDM-1} + <i>tet</i> (X6))	[6]
<i>E. coli</i> DH5α (PUC19)	In this study
<i>E. coli</i> DH5α (PUC19- <i>tet</i> (X4))	In this study
<i>E. coli</i> DH5α (PUC19- <i>bla</i> _{NDM-5})	In this study
<i>E. coli</i> DH5α (PUC19- <i>bla</i> _{NDM-9})	In this study
<i>E. coli</i> MG1655	[7]
<i>E. coli</i> MG1655Δ <i>mdh</i>	[7]
<i>E. coli</i> MG1655Δ <i>cydB</i>	[7]
<i>E. coli</i> MG1655Δ <i>aceA</i>	[7]
<i>E. coli</i> MG1655Δ <i>katE</i>	[7]
<i>E. coli</i> MG1655Δ <i>torA</i>	[7]

ATCC, American type culture collection; MRSA, Methicillin-resistant *staphylococcus aureus*; LZD^R, linezolid resistance; Van^R, vancomycin resistance;

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Supplementary Table 2 Antimicrobial spectrum of DCA against bacterial strains.

Strains	DCA (MIC, mM)
Gram-positive bacteria	
<i>S. aureus</i> ATCC 29213	0.97
MRSA T144	0.97
Gram-negative bacteria	
<i>E. coli</i> ATCC 25922	125
<i>E. coli</i> DH5 α (PUC19- <i>tet</i> (X4))	125
<i>E. coli</i> DH5 α (PUC19- <i>bla</i> _{NDM-5})	125
<i>E. coli</i> DH5 α (PUC19- <i>bla</i> _{NDM-9})	125

Supplementary Table 3 RT-qPCR primers used in this study.

Genes	Sequence (5'→3')
<i>acrB</i> -F	AAGAAGCTACCCGTAAGTCG
<i>acrB</i> -R	AGTAGAACCGCCAAAGAAGG
<i>acrD</i> -F	TGGAATCGTTAGTGAAGCAG
<i>acrD</i> -R	CAGCCAGACACAGGAATAC
<i>emrB</i> -F	TCTCATTGGCGGAAATAATCAG
<i>emrB</i> -R	TAAACCCGTTCAACCCGAAT
<i>mdfA</i> -F	TTGATTGGGTTCTACTTCG
<i>mdfA</i> -R	CCAGACAGGTGACGATAAAC
<i>macB</i> -F	GGCGTCTTGAAAACGTGTTGA
<i>macB</i> -R	GTCACTGACACCAGCATAATA
<i>mdtK</i> -F	TAATGTTCGTGCTTCCAATG
<i>mdtK</i> -R	CATACAGACACCCACCATAA
<i>gltA</i> -F	CACCCTCAACGGGGATACAG
<i>gltA</i> -R	GATGCGGTTGAAGTGAAGCC
<i>acnA</i> -F	CGTATCTCGACGAACTGGGG
<i>acnA</i> -R	CGTTTCGATAGGATCGGGCA
<i>icd</i> -F	CCCGAACACTGGCAAAGAGA
<i>icd</i> -R	GCCAGGGCGTCAGAAATGTA
<i>sdhC</i> -F	GCAGTTTGCTCACGTCAAGG
<i>sdhC</i> -R	TCACGGAAATGATGCTGGCT
<i>fumA</i> -F	CCTTACCGCTCTGGCGTATC
<i>fumA</i> -R	TGCGAGAAGTGAAAGCACGA
<i>mdh</i> -F	TACTTCGCTCACGACATCCG
<i>mdh</i> -R	CCTGACGGTTGATCTTCGCT
<i>aceA</i> -F	CGGTTATTGGCGGTCACTCT
<i>aceA</i> -R	CGTTCTGGATGCGTTTGGTC
<i>aceB</i> -F	CGCGAAATATCCGGGCAAAC
<i>aceB</i> -R	GCCAGGGTGATGAACTGGAA
<i>nuoA</i> -F	AGTCATCGCTCATCACTGGG
<i>nuoA</i> -R	CGACCGCCTAAAAACCAACC
<i>cyoA</i> -F	CCGAACTGGTCACACTCCAA
<i>cyoA</i> -R	TGGTAATGGGCTTCTCGTCG
<i>appC</i> -F	TCGAATCCACCTTTGTCGGG
<i>appC</i> -R	GATTTGAACCGAACGCCACC
<i>metB</i> -F	GTCGTTGCCGGAAAATCAGG
<i>metB</i> -R	GCCAGCGTAAACAACGACAG
<i>malY</i> -F	AAGCAGGCTGATGGTTGGTT
<i>malY</i> -R	CATCGCACGTCCACACTTTC

<i>cysK</i> -F	TTATTGCTGGCGTTGGGACT
<i>cysK</i> -R	GCTCAACGGCGACAGAGATA
<i>IL-1β</i> -F	ATGAAAGACGGCACACCCAC
<i>IL-1β</i> -R	GCTTGTGCTCTGCTTGTGAG
<i>TNF-α</i> -F	AGGCACTCCCCCAAAGAT
<i>TNF-α</i> -R	TGAGGGTCTGGGCCATAGAA
<i>IFN-γ</i> -F	GCTTTGCAGCTCTTCCTCA
<i>IFN-γ</i> -R	CTTTTGCCAGTTCCTCCAG
<i>IL-6</i> -F	TGCAAGAGACTTCCATCCAGT
<i>IL-6</i> -R	GTGAAGTAGGGAAGGCCG
<i>IL-4</i> -F	CCATATCCACGGATGCGACA
<i>IL-4</i> -R	AAGCACCTTGGAAGCCCTAC
<i>IL-10</i> -F	GGCGCTGTCATCGATTTCTC
<i>IL-10</i> -R	ATGGCCTTGTAGACACCTTGG
<i>β-actin</i> -F	GTCCACCTTCCAGCAGATGT
<i>β-actin</i> -R	GAAAGGGTGTAACGCAGC
