



Using DREADDs to isolate internal clocks

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Although much is known about the psychophysics of timing and time perception, the ability to selectively modulate discrete neuronal populations subserving interval timing could advance the field in unprecedented ways (see Buhusi and Meck, 2005; Allman and Meck, 2011; Cheng et al., 2011). Current approaches used to delineate the neuronal correlates of timing are often restricted to pharmacological (Meck, 1996; Coull et al., 2011) or ablative approaches, both physical (Meck, 2006a,b; Wiener et al., 2008) and genetic (Sysoeva et al., 2010; Agostino et al., 2011; Meck et al., 2011; Wiener et al., 2011). Most, if not all, of the ablative approaches are limited to loss of function-type hypothesis testing and frequently have unintended off-target effects. The pharmacologic approach has inherent confounds in the non-specificity of small molecule ligands. One way to overcome these confounds is to use a system in which a designer receptor exclusively activated by designer drug (DREADD) is expressed in a cell-type specific manner via transgenic expression. This chemicogenetic approach is further described by the acronym of the first tool of its type: receptor activated solely by synthetic ligand (RASSL). In short, a receptor is engineered such that it is no longer activated by any endogenous ligands and instead activated by an otherwise inert exogenous ligand. The various RASSLs and DREADDs have been reviewed elsewhere (Pei et al., 2008). In short, there are three DREADDs available with proven functionality in neuronal environments – hM3Dq (Gq-coupled, promotes neuronal excitability), hM4Di (Gi-coupled, promotes neuronal inhibition), and the rM3Ds (Gs-coupled, promotes cAMP production). Here, we discuss this technology's potential in the field of interval timing.

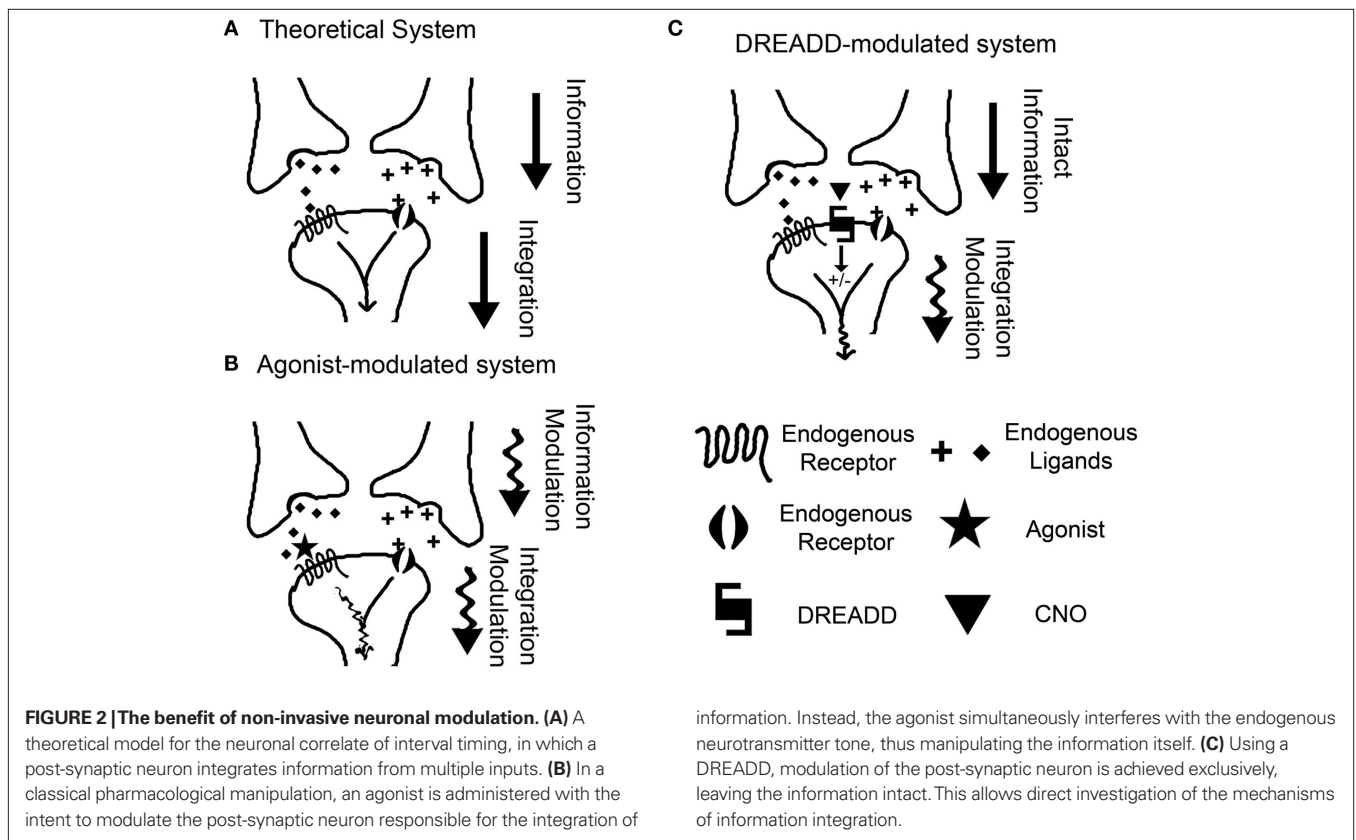
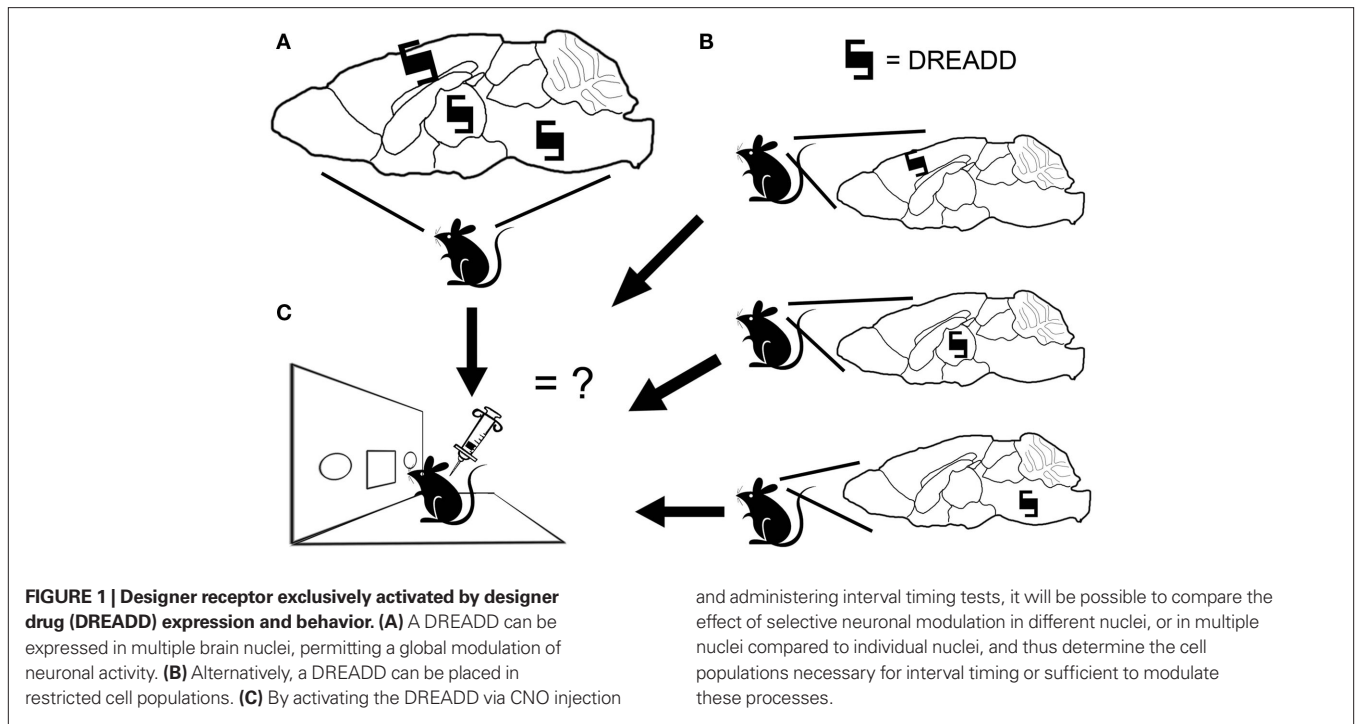
Cell-type specificity of signaling has been a hurdle for delineating the neuroanatomical substrates of timing (Cheng et al., 2011). Using the chemicogenetic approach, it is possible to express a receptor in a select population of neurons and then turn that receptor on via peripheral administration

of clozapine-N-oxide (CNO). The cell-type specificity is dependent upon the transgenic approach used, though the current genetic toolkit appears to be more than adequate for the study of interval timing. To date, there has been success expressing DREADDs using virally mediated approaches (Ferguson et al., 2011; Krashes et al., 2011; Sasaki et al., 2011), single-transgenic approaches (Guettier et al., 2009), and intersectional transgenic approaches (Alexander et al., 2009; Ray et al., 2011). With these approaches, it should be possible to place a DREADD into a subpopulation of neurons and then determine how this population modulates interval timing. Furthermore, the identical signaling-type initiated via the DREADDs in various subpopulations would allow us to piece together the circuitry by expressing the DREADD in multiple subpopulations either simultaneously or separately (Figure 1).

In addition to the cell-type specificity possible with DREADDs, this approach also affords us the ability to silently and independently modulate neuronal signaling. An inherent confound in the pharmacologic approach to neuronal modulation is the intrinsic interference with the endogenous tone of the system being investigated. By definition, any agonist interacting with a receptor competes with the endogenous ligand for that receptor. Thus, the experimental modulation is no longer limited to the modulation of the neuronal population induced by the ligand–receptor pair, but instead also includes interference in the homeostatic, basal signaling state of the system. As it is hypothesized that information is encoded in patterns of neuronal firing and the subsequent phasic neurotransmitter presence (Johnson, 2000), it can be seen that interfering with the temporal signaling nature of the basal state can be detrimental in our quest to determine the neuronal correlates of time (Figure 2). The DREADD overcomes this confound by the use of a non-native receptor which has no basal tone. Take, for example, the neurons of the striatum, a well renowned candidate for the

epicenter of interval timing circuitry (Matell et al., 2003; Meck et al., 2008). In addition to the technical difficulty of selectively manipulating these neurons through pharmacological means (local microinjections), the key confound is that any pharmacological manipulation will interfere with the afferent tone on the striatal synapses (Figure 2B). Thus, the experimental manipulation is no longer limited to striatal medium spiny neuron (MSN) modulation, but instead now includes modulation of the afferent tone associated with the neurotransmitter of the targeted receptor. Thus, the experimental manipulation is affecting both the hypothetical integrator of information (the MSN) and the information itself (the phasic, temporally encoded presynaptic release of neurotransmitter). With the DREADD approach, it is possible to modulate the activity of the MSNs independent of the endogenous tone, leaving the theoretical temporally encoded information in place (Figure 2C). This provides us the unique opportunity to reductively piece together each component of the neuronal substrates of timing.

Finally, the usability of the chemicogenetic approach is another salient aspect when considering its application in the field of interval timing. Indeed, other approaches for cell-type specific control of neuronal signaling exist, most notably that of optogenetics (Yizhar et al., 2011). The optogenetic approach has many features that lend well to interval timing research – namely, the exclusive control of neuronal firing in the subsecond range. Both optogenetics and chemicogenetics achieve cell-type specific control of signaling through transgenic expression of a non-native receptor, so both approaches leave the endogenous tone intact. The chemicogenetic approach, in its simplest form (a single-transgenic mouse), requires a simple peritoneal injection of CNO to modulate cell-type specific neuronal signaling. This simplicity is an experimental aspect that cannot be ignored considering the exclusive use of behavior paradigms in the study of interval timing.



In conclusion, it can be seen that by combining DREADDs and transgenic technology in the chemogenetic approach it will

be possible to further delineate the neuronal substrates of interval timing. The cell-type specific control in combination with the

independent modulation of neuronal signaling afford a level of precision that is almost requisite in the investigation of such

an intertwined neuronal process as that of interval timing. We propose an initial experiment here: investigating the functional role of the MSNs in temporal integration (see Matell and Meck, 2004; Oprisan and Buhusi, 2011). It can be seen that this technology is well suited for use in this field, and will hopefully advance our understanding of the neural basis of timing and time perception (Gibbon et al., 1997; Gu et al., 2011).

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