# **Roles of Wnt Signaling Pathway and ROR2 Receptor in Embryonic Development: An Update Review Article**

# Rui Guo<sup>1</sup> and Quan Sheng Xing<sup>2</sup>

<sup>1</sup>Qingdao University, Qingdao, China. <sup>2</sup>Qingdao University-Affiliated Hospital of Women and Children, Qingdao, China.

**Epigenetics Insights** Volume 15: 1-9 © The Author(s) 2022 Article reuse guidelines: sagepub.com/journals-permissions DOI: 10.1177/25168657211064232



ABSTRACT: The Wht family is a large class of highly conserved cysteine-rich secretory glycoproteins that play a vital role in various cellular and physiological courses through different signaling pathways during embryogenesis and tissue homeostasis 3. Wnt5a is a secreted glycoprotein that belongs to the noncanonical Wht family and is involved in a wide range of developmental and tissue homeostasis. A growing body of evidence suggests that Wnt5a affects embryonic development, signaling through various receptors, starting with the activation of β-catenin by Wht5a. In addition to affecting planar cell polarity and Ca<sup>2+</sup> pathways, β-catenin also includes multiple signaling cascades that regulate various cell functions. Secondly, Wnt5a can bind to Ror receptors to mediate noncanonical Wnt signaling and a significant ligand for Ror2 in vertebrates. Consistent with the multiple functions of Wnt5A/Ror2 signaling, Wnt5A knockout mice exhibited various phenotypic defects, including an inability to extend the anterior and posterior axes of the embryo. Numerous essential roles of Wnt5a/Ror2 in development have been demonstrated. Therefore, Ror signaling pathway become a necessary target for diagnosing and treating human diseases. The Wnt5a- Ror2 signaling pathway as a critical factor has attracted extensive attention.

Email: xinggs0532@163.com

**KEYWORDS:** Wnt5a, Ror2, tetralogy of Fallot, β-catenin-mediated and β-catenin-independent

RECEIVED: May 21, 2021. ACCEPTED: November 15, 2021.

TYPE: Epigenetic changes and cardiometabolic risk factors - Review

FUNDING: The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: Guo Rui was supported by The National Natural Science Foundation of China (NSFC; Grant numbers 81770315 and 82071583) and Taishan Scholars Program (2019).

DECLARATION OF CONFLICTING INTERESTS: The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article CORRESPONDING AUTHOR: Quan Sheng Xing, Qingdao University-Affiliated Hospital of Women and Children, tongfu road 6, shibei district, Qingdao 266000, China.

# Overview

Every year, hundreds of thousands of children are born with congenital disabilities and leading infant mortality derivations. There are surgical interventions and medical treatment protocols for some congenital disabilities. However, many of these children live with congenital disabilities that leave them permanently disabled. The factors which might induce those congenital disabilities can be genetic, such as genetic mutations in specific genes or chromosomal abnormalities,<sup>1</sup> or those provoked by teratogens (environmental factors), such as by drugs, environmental chemicals, and abnormal concentrations of natural metabolites (such as folate deficiency).<sup>2-5</sup> Of all genetic birth anomalies, cardiac defects are the most predominant. Its incidence is up to 1% of live births is one of the leading causes of neonatal disability and death in China. Congenital cardiovascular disease includes various conditions, such as atrial septal defect, ventricular septal defect, patent ductus arteriosus, and tetralogy of Fallot.<sup>6</sup> These diseases seem to have little in common, but the underlying disease mechanisms are similar, and the onset is related to environmental and genetic factors.<sup>2</sup> It is essential to understand the heart's embryonic development to identify the underlying pathology of these defects'. The linear tube forms the mesoderm's anterior lateral plate during embryonic development before undergoing extensive remodeling. Additionally, the second heart field assigned a late differentiating lateral plate of mesoderm expands and initiates the right ventricle and atria's growth in the mammals. Formerly neural crest cell migration has occurred and contributes to heart formation, particularly the outflow tract. With the

continuous extending of the research on the pathogenesis of congenital heart disease (CHD), especially the rapid development of molecular biology and genetics, the pathogenic factors of CHD have been profoundly understood. Recently, the discovery of monoclonal genetic defects associated with an isolated illness or non-syndromic CHD has revealed critical molecular pathways in heart morphogenesis. Multiple genes are associated with CHD, such as NKX2.5,7 GATA4,8 MYH6,9 BMPR2,10 CRELD1 and ALK2, and NOTCH1.11 However, the genetic mutation of these genes causes severe cardiac defects.

Nonetheless, other associated factors influence cardiac mesoderm alteration. However, most of the patients with congenital heart disease, especially the genetic causes of sporadic patients, are still unclear. The study of Wnt signaling pathways has always been an area of concern. Therefore, this review article will outline Wnt signaling pathways, new cellular and molecular data to investigate the potential mechanisms in vertebrate models associated with congenital heart disease induction. In addition, we will specify brief details on environmental factors related to early heart development and new evidence on folic acid and inositol supplementation.

# Wnt Signaling Pathway

Since the first Wnt gene (Int-1) in 1982, Wnt proteins constitute an important family of signaling molecules that coordinate and influence many cellular biological and developmental processes.<sup>12</sup> A total of 19 Wnt proteins associated with this family and their function and related disease have been



Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (https://creativecommons.org/licenses/by-nc/4.0/) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access pages (https://us.sagepub.com/en-us/nam/open-access-at-sage).

mentioned in Table 1. The Wnt signaling pathway is a multichannel signal transduction pathway activated by binding ligands and proteins during embryonic development and tumorigenesis. Through this pathway, Wnt protein is secreted as cysteine-rich glycolipoproteins that's act as extracellular signals. This further transmits the signals into the cell by activating intracellular segments of receptors on the cell surface, playing an essential role in regulating cell proliferation, differentiation, survival, migration, polarity, and other physiological processes.<sup>3</sup> The Wnt pathway was designated as both the canonical ( $\beta$ -catenin-mediated) and non-canonical (β-catenin-independent) pathways. In addition to the listed in Table 1, there are some other ligand and receptor proteins associated with these pathways, such as Wnt receptor (Frizzled family proteins),13 LDL receptor-related protein (LRP), tyrosine kinase Ror,<sup>14</sup> Disheveled (DSH/DVL) protein, β-catenin, glycogen synthase kinase C3B (GSK-3B),<sup>15</sup> axin/conductin, and adenomatous polyposis coli (APC).<sup>16</sup> Furthermore, the non-canonical Wnt signals were additionally classified (based on downstream signaling) into the following categories: the non-canonical Planar cell polarity pathway (PCP pathway), the non-classical Wnt/Ca2+ pathway (Wnt/Ca2+ pathway), and intracellular pathways. The canonical Wnt signaling pathway is tangled in the cell cycle. The non-classical PCP pathway affects the cytoskeleton's formation, and the non-canonical Wnt signaling/calcium pathway involves DNA transcription. Besides, recent studies have pointed to the crucial role of Wnt signaling in maintaining adult homeostasis of stem cell pluripotency. Wnt binds to the Frizzled and LRP5/6 co-receptors to induce stable  $\beta$ -catenin proteins and enter the nucleus affecting the target transcription genes via the canonical Wnt signaling pathway.<sup>13</sup> In addition, it is highly genetically conserved in animals and is very similar in different animal species. Subsequently, overt Wnt signaling of uncontrolled embryonic development is a hallmark of congenital disabilities, cancer, and other degenerative diseases; understanding how this pathway is regulated is crucial,<sup>16,17</sup> as shown in Figure 1.

# Wnt Signaling Pathway and Embryonic Cardiac Development

Genetics is dependent on the intensification of a small number of primitive germ cells (PGCs) in the early embryo. The derivative (stem cells) of the PGCs differentiates into multiple organ development. In mammals, stem cells or progenitor cells reside in a particular microenvironment termed a niche. Its microenvironment influences progenitor cell proliferation in different directions, including neuronal differentiation, liver, myogenic, chondrogenic, osteogenic, adipogenic, and cardiogenic cells.<sup>31</sup> The growth of progenitor cells is regulated by both circulating and local signaling pathways. These multiple regulatory pathways, including receptor tyrosine kinases (RTKs), Notch (N), Hedgehog (Hh), TGFβ, Jak/Stat, nuclear receptors, and Wnt, are involved in cell differentiation. During embryonic growth, the Wnt/b-catenin dependent pathway regulates the early differentiation of mouse embryonic stem cells.<sup>32</sup> Thus, Wnt and membrane protein receptors are essential regulators of heart development in vertebrates.<sup>4</sup> Furthermore, Wnt signaling activity is low in a healthy adult cardiovascular system under normal physiological conditions. Nevertheless, the reactivation of this pathway has been observed in cardiovascular disease.<sup>5</sup> Additionally, many fetal genomes are activated during the heart's pathological remodeling, including Wnt singling pathway. Experimental studies have shown that the application of Wnt signaling activators increases the myogenic differentiation of mesenchymal stem cells from cardiac patients. However, the Wnt pathway is active in developing the pericardial, the adjacent cardiac mesoderm, the endocardial cushions, and the early outflow tract. However, it retains almost no role in the developing ventricular myocardium.33 Surprisingly, several Wnt ligands have a high expression level in the heart's early development, including Wnt2, Wnt2b, and Wnt8a.34-36 Furthermore, cardiac genes' expression significantly increases via Wnt3A. On the other hand, inhibition of the Wnt signaling pathway resulted in the embryo's complete disappearance.17 These findings suggest that the canonical Wnt pathway not only drives the formation of embryonic stem cell cardiomyocytes but is also an essential pathway for development.

Similarly, the non-canonical Wnt pathways play an endorsement role in the fate of cardiomyocytes. The Wnt11-mediated mesenchymal stem cells have been shown to upregulate GATA-4, conceivably through signaling via the PKC or JNK pathways.8 Both Wnt5a and Wnt11 promote the fate of embryonic cardiac stem cells. However, there is little literature on the molecular and cellular mechanisms behind the influence of Wnt5a and Wnt11 on cardiac embryonic development. Furthermore, we used human embryonic stem cells (hESC) to systematically analyze the expression of endogenous Wnt signaling elements during the development of Human cardiomyocytes. So, our preliminary results showed the expression of Wnt3 and Wnt8a regulates the short axis and influences mesoderm development through FZD7.17,28,29 In addition, Wnt5a/5B regulates cardiovascular development through the Rror2 non-canonical signaling pathway. Besides, Wnt2, Wnt5a/5B, and Wnt11 regulate the differentiation of cardiomyocytes via FZD4, FZD6.24,25,28 The role of classical Wnt signaling in the ordered phase of vertebrate cardiogenesis has been confirmed. The more precise role of single Wnt signaling and Wnt receptor genes in atypical signaling pathways in human cardiomyocyte development has been identified.<sup>37</sup>

# Role of the Ror Receptor in the Cardiac Embryonic Development

Receptor tyrosine kinases (RTK) play a crucial role in cell development by inducing a cascade of homologous ligand binding signals that lead to cell proliferation and differentiation. The Ror-family receptor tyrosine kinases in mammals consist of Ror1 and Ror2, characterized by the extracellular

# Table 1. Wnt protein associated function, gene location, and related diseases.

ALIASES	GENE LOCATION (HUMAN) HTTPS://WWW. GENECARDS.ORG	FUNCTION	ASSOCIATED DISEASE
Proto-oncogene Wnt-1	12q13.12	Involve with oncogenesis, cells differentiation in the embryogenesis, formation of the embryonic dorsal neural tube, and initiation of mesencephalon and cerebellum	Joubert syndrome, autism, osteogenesis imperfecta, and type XV <sup>17</sup>
WNT2	7q31.2	Involve with oncogenesis, cell differentiation in the embryogenesis, cognitive/linguistic, and midbrain dopaminergic neuron development.	Autism <sup>18,19</sup>
Wnt-2b (formerly Wnt-13)	1p13.2	Controlling mesoderm specification and some aspects of brain, heart, and lung during gastrulation.	Diarrhea 9 (DIAR9) <sup>17</sup>
Wnt3	17q21.31-q21.32	Involve with oncogenesis, cell differentiation in the embryogenesis	Tetraamelia syndrome and tetra-amelia syndrome <sup>17</sup>
Wnt3a	1q42.13	Involve with oncogenesis, adipogenesis, and cell differentiation in the embryogenesis	Osteoporosis, juvenile, and hypotrichosis <sup>20</sup>
Wnt4	1p36.12	Aggravate the testis-determining factor, regulates endometrial stromal cell proliferation, survival, and differentiation	SERKAL syndrome, Mullerian aplasia, and hyperandrogenism <sup>21,22</sup>
Wnt5a	3p14.3	Cell differentiation in the embryogenesis, posterior development of the female reproductive tract, and cardiac outflow tract	Robinow syndrome, autosomal dominant, congenital heart disease <sup>23-26</sup>
Wnt5b	12p13.33	Cell differentiation in the embryogenesis, regulation of cardiac development	Type 2 diabetes, fallopian tube serous adenocarcinoma, and colorectal cancer <sup>17</sup>
Wnt6	2q35	Key roles in cell differentiation during embryogenesis, carcinogenesis, and inhibits the induction of cardiogenic mesoderm	Mullerian aplasia and hyperandrogenism, and gastric cancer <sup>23</sup>
Wnt7a	3p25.1	Roles in cell differentiation during embryogenesis, the development of the anterior-posterior axis in the female reproductive tract, and uterine smooth muscle	Fuhrmann syndrome, Schinzel phocomelia syndromes, fibular aplasia, or hypoplasia <sup>23</sup>
Wnt7b	22q13.31	Regulating of cell fate and patterning during embryogenesis, placental, lung, eye, dendrite, and bone formation along with kidney development	Beckwith-Wiedemann syndrome, microphthalmia, syndromic 9 and colobomatous microphthalmia <sup>27</sup>
Wnt8a	5q31.2	Development of early embryos and germ cell tumors	Exudative vitreoretinopathy 1 and Norrie disease.
Wnt8b	10q24.31	Roles in cell differentiation during embryogenesis	Exudative vitreoretinopathy 1, epilepsy, and Alzheimer's disease.
Wnt9a (formerly Wnt14)	1q42.13	Involves oncogenesis and the regulation of cell differentiation during embryogenesis	Gastric cancer and exudative vitreoretinopathy 1.
Wnt9b (formerly Wnt15)	17q21.32	Required for craniofacial development, the standard fusion of the palate, embryonic kidney, and urogenital tract development	Cleft lip and Mayer-Rokitansky- Kuster-Hauser syndrome.
Wnt10a	2q35	Involves oncogenesis and the regulation of cell differentiation during embryogenesis	Odontoonychodermal dysplasia, Schopf-Schulz-Passage syndrome, Tooth agenesis, selective, promyelocytic leukemia, and Burkitt's lymphoma.
Wnt10b (formerly Wnt12)	12q13.12	Involved in oncogenesis and the regulation of cell differentiation during embryogenesis, breast cancer, and governs adipogenesis	Breast cancer, split-hand/foot malformation, tooth agenesis
Wnt11	11q13.5	Development of skeleton, kidney, and regulates cardiac chamber	Fallopian tube serous adenocarcinoma and exudative vitreoretinopathy <sup>28,29</sup>
Wnt16	7q31.31	Embryonic regulation of cell growth and differentiation	Basal cell carcinoma and nodular basal cell carcinoma. <sup>30</sup>



**Figure 1.** Signaling pathways involved in CHD. In normal conditions, the Wnt/ $\beta$ -catenin pathway is kept inactive as the  $\beta$ -catenin is complex with APC/ Axin complex. When the Wnt or other specific ligands bind with membrane receptors, including frizzled and Ror, or by situations in which APC/Axin become unstable, cascades become activated and increase the free form of  $\beta$ -catenin and is not being phosphorylated, leading to accumulation in the cytoplasm. Eventually, this free  $\beta$ -catenin is associated with nuclease and promotes the expression of several Wnt target genes. APC helps Axin phosphorylate  $\beta$ -catenin and subsequent degradation of  $\beta$ -catenin through ubiquitination if the cell remains inactive. Wnt/ $\beta$ -catenin non-canonical signaling pathway is transduced independent from  $\beta$ -catenin activation. After ligation, DvI utilizing Daam 1 activates Rho kinase (RhoA). DvI also activates CaMK, calcineurin, and PKC, resulting in subsequent activation of JNK. The integrity of these pathways leading to cytoskeletal changes and stimulation of cell polarization and mutilations during gastrulation.

Frizzled-like cysteine-rich domain and membrane-proximal kringle domains. Ror (RTK) and Wnt/β-catenin signals can activate various intracellular signals, including the activation of the planar cell polarity pathway (PCP pathway).<sup>38,39</sup> Another mechanism of ROR2's action on the PCP signal is activating the classical PTK signaling pathway. By interacting with PTK7 and SFRPs, ROR2 promotes signal transduction to JNK. Therefore, the activation of the PCP pathway and typical RTK signaling pathway by ROR2 may be caused by different signals that are differentially regulated by SFRPs.<sup>13</sup>

The Ror-receptor has previously been multiple signaling functions that regulate several processes during embryonic development, such as developing the heart, nervous system, bone, and kidney. Furthermore, Ror2 knockout mice presented with dwarfism, facial anomalies, and cardiac septal defects. During embryonic development, the expression pattern of the Ror gene is significantly different between invertebrates and vertebrates.<sup>40</sup> In addition, Ror1 and Ror2 were initially being found to regulate the germinal bands of neurogenic ectoderm during Drosophila embryogenesis, suggesting their role in

neurodevelopment.<sup>41</sup> In vertebrates, Ror1 and Ror2 signals are widely expressed in chickens and mice during embryonic development, most notably in the central nervous system, heart, lung, kidney, early limb bud, and cartilage growth plate.<sup>26,42</sup> Furthermore, the PCP pathway controls the direction of the static cilia in the skin, hair, and inner ear and cell polarity and coordination in developing the stomach, nerves, intestine, and limbs.43 Besides, in humans, mutations in the Ror2 gene cause 2 distinct developmental syndromes, including recessive Robinow syndrome (RRS) and autosomal dominant type B1 brachydactyly.44,45 In addition, some patients have congenital heart defects, including atrial or ventricular septal defects and cardiac outflow tract defects.<sup>26,46</sup> Another study has shown that Ror2 gene mutations in mice with RRS exhibit multiple bone defects, mainly severe shortening of limbs and tails.44 Besides, mice with homozygous Ror2 mutations died shortly after birth and had cardiac septal defects, primarily ventricular septal defects. This indicates that Ror2 is essential in forming the heart, especially in developing the cardiac septum.47

## Role of Wnt in the Cardiovascular System

Vasculogenesis and angiogenesis control the formation of blood vessels. Vasculogenesis is the process of developing blood vessels from the de novo production of endothelial cells (ECs) in embryos. Later, angiogenesis takes place, forming new blood vessels from pre-existing.48 Various Wnt ligands appear to affect EC functions (WNT2, -2B, -3, -4, -5A, -7A, -8A, -9A, -9B, -10B, and -11) as mention in Table 1. Vessel maturation and stabilization of blood vessels are also controlled by parietal cells. Pericytes are important pericytes supporting capillaries.<sup>49</sup> Reduced pericyte coverage in the endodermis can lead to vascular loss. In pulmonary arterial hypertension (PAH), the expression of FZD genes required for WNT/PCP activation in the pericyte is reduced, which is an essential mechanism for EC motility and polarity.5,50 In vitro transfection of the mutant WNT5A reporter gene explains the functional significance of the corresponding mutation that influences amino acid substitution in the epidermal growth factor-like domain (Cys-ARG).51

In addition, the thickening of the pulmonary arterial wall is observed in PAH due to the proliferation of vascular smooth muscle cells. This was associated with increased levels of β-catenin activity in pulmonary artery smooth muscle cells (PASMCs).<sup>52</sup> In addition, β-catenin is located at the adhesion junction of the Catenin /cadherin complex and plays a crucial role in cell adhesion and intercellular communication.53 mXina, a protein of the intercalated disk (a critical point for cardiac hypertrophy), contains a β-catenin-binding domain that plays a crucial role in bundling actin filaments.<sup>54</sup> Remarkably, accumulation of  $\beta$ -catenin was found in the intercalated disk of hypertrophic cardiomyopathy hamsters that possibly leads to stiffening of the myocardial wall and harmful structural.<sup>55</sup> However, in vitro, the exact roles of β-catenin are still not fully understood.<sup>56</sup> Further studies are needed to clarify the precise mechanisms by which  $\beta$ -catenin(and WNT signaling) reveal the effects in hypertrophic reactions.

#### The Wnt5a-Ror

Wnt5a is a member of the Wnt family and consists of 1172 adenines, 884 cytosines, 946 guanines, and 1172 thymine. In recent years, Wnt5a has attracted extensive attention due to its regulatory effects on both the canonical Wnt Wnt/βcatenin pathway and the non-canonical Wnt/Ca<sup>2+</sup> pathway. Previous studies have shown that Wnt5a is involved in intercellular signal transduction during embryogenesis. Again, Wnt5a produces a series of intracellular reactions by binding to Frizzled receptor (a 7-fold transmembrane receptor protein), which transmits signals into the cell.<sup>21</sup> Furthermore, Ror2 is a single transmembrane receptor protein containing a tyrosine kinase that acts by forming a complex with Wnt5a that transmits signals into cells. In addition, Ror2 and Vangl2 procedure a Wnt-induced receptor complex, and Wnt5a signaling controls cell polarity by regulating Vangl2 phosphorylation.<sup>57</sup>

As previously established, the Wnt5a-Ror signaling pathway is an important regulatory pathway for tissue regeneration, embryonic tissue formation, and reproduction. The Wnt5a facilitates tissue homeostasis by activating transforming growth factor-B (TGF-B). The core of the downstream TGF-B signaling pathway involves ROR1/2 and Smad proteins.58 Meanwhile, TGF- B enhanced the transcriptional activity of the Smad protein complex through ROR1/2 signaling. Besides, kinesin superfamily protein KIF26b acts as a downstream target of the Wnt5a-Ror pathway. Wnt5a-Ror induces degradation of KIF26B through a ubiquitin-proteasome system and modulates its cell stability through the canonical Wnt/βcatenin-dependent pathways. Wnt5a regulates epithelial-mesenchymal cell migration through this mechanism. Genetic distress of KiF26b function in vivo leads to axial malformation in embryos and loss of primitive germ cells, 2 phenotypic characteristics Wnt5a-RORs signal disruption. Wnt5a binds to Ror2 (as a ligand in PGC) to promote the polarization, elongation, and redirection of PGC. Furthermore, Wnt5a enhances the chemotaxis of PGC by redistributing intracellular Ror2.59 Besides, Wnt5a inhibits the Wnt signaling pathway through Ror2. It activates the expression of nuclear target genes, which is beneficial to cells' early and efficient migration and has advantages in cloning germ cells.60 However, mutations in Ror2 and Wnt5a signaling leads to the formation of developmental defects. In Ror2 mutant mice, the number of PGCs surviving is reduced due to dysregulated migration of PGC in the embryo. In conclusion, Wnt signals affect cell proliferation and differentiation, while Ror mainly controls the process of cell polarization migration.

The neural crest is a unique cell group of vertebrates originating from the neural plate. The craniofacial skeleton and cardiac outflow tract are all derivatives of the neural crest cells. Cranial neural crest cells migrate ventrally to the heart and form the cardiac outflow tract with the detached aorta and pulmonary trunk.<sup>61</sup> Ror2 expression is critical during mouse embryonic development in various tissues, including the skeletal system and internal organs. Furthermore, Ror2-deficient and Wnt5a-deficient mice were born with craniofacial abnormalities and significantly shortened limbs and tails. However, the craniofacial and cardiac defects of Wnt5a-deficient mice were more severe than those of Ror2-deficient mice. This is because the Wnt5A mutation induces the downregulation of cardiac neural rest through plexus protein 2 (PlxNA2), and the animals exhibit outflow tract defects similar to those of Ror2deficient mice.62

Furthermore, cardiac phenotypic defects in Wnt5a-deficient mice are attributed to defective Wnt/Ca<sup>2+</sup> signaling in CaMK2,<sup>25</sup> closely related to Wnt/PCP signaling.<sup>63</sup> Regarding the cardiac neural crest, Wnt/PCP signals are required to form other cardiac regions, contributing to the cardiac outflow tract formation.<sup>64</sup> Therefore, simultaneous craniofacial and cardiac outflow tract malformations in patients with RRS and corresponding mouse models suggest that the Wnt5a/Ror2 signaling pathway plays a crucial role in heart development. Mice deficient in both Wnt5a and Ror genes show abnormal bone and heart elevation.<sup>46</sup> This is because Wnt5a-Ror signals affect tissue pattern, cell polarity, and cell migration during organ development. Besides, Wnt5a-Ror signaling is active in adult uteri and crucial for early pregnancy. The Wnt5a-Ror signaling pathway promotes embryo implantation, and the disorder of this pathway leads to reduced fertility. An abnormal Wnt5a-Ror signal can lead to abnormal decidua growth. One of the main functions of the decidua is to regulate the entry of the trophoblast into the stroma and guide placenta formation. Implantation defects and decidual growth restriction were found in uteruses with varying Wnt5a levels.<sup>65</sup> However, the molecular mechanism of the Wnt5a-Ror pathway mediating these processes remains unclear.<sup>38</sup>

# Myoinositol and Folic Acid Role in Embryogenesis

Many studies have confirmed that folic acid (FA) has been generally accepted as a preventive effect on neurodevelopmental abnormalities. Folic acid deficiency and elevated folic acid levels impair the Wnt pathway function in developmental signaling pathways. Depending on the individual genotype, FA supplementation may be detrimental to neurodevelopment and embryogenesis. A recent review article suggested that multivitamins may also reduce the risk for certain heart defects but at higher concentrations than those used for neural tube defects.66 Furthermore, a cohort study in Denmark and Norway has shown that FA is not associated with the risk of heart defects (including severe defects and septal defects) in offspring.67 However, another study has confirmed that FA is related to decreased risk of congenital heart defects.68 It is suggested that women of reproductive age take FA supplements before pregnancy to prevent neural tube defects, including spina bifida and anencephaly. Although the link between the Wnt pathway and FA is not well established, further studies are needed to elucidate it. Inositol is essential in the inositol-lipid cycle and provides metabolic substrates for the signal transduction process, including the Wnt pathway.<sup>69</sup> Furthermore, the depletion of inositol in the Wnt pathway resulted in the cell's inhibition of GSK3B.70 Wnt3A triggers the phosphatidylinositol signal attached to the G protein to generate inositol polyphosphates instantaneously, including inositol pentaphosphate (IP5). Further, IP5 inhibits GSK-3β activity. By blocking IP5 formation inhibits the buildup of βcatenin.71 The association between the inositol phosphatidyl pathway and its intermediates with the canonical Wnt signaling pathway may explain the severity of heart defects and different phenotypes. Besides, several studies have found that once mothers increase their folic acid intake, their risk of other congenital disabilities, such as cleft palate, appears to be decline.72 The addition of myoinositol combined with folate resulted in even better protection of the chick embryo from the adverse effects of the environmental factors.62,63

There are some natural compounds other than folic acid and myoinositol, phytochemicals, especially polyphenols, are among the most studied groups of the flavonoids and nonflavonoids subgroups. These drugs overcome the need to introduce foreign compounds with multiple complications into individuals.<sup>73</sup> They are generally non-toxic, more readily available, and less costly than synthetic drugs, preventing several diseases in healthy individuals. Recent studies have also identified several potent inhibitors of the Wnt/ $\beta$ -catenin and hedgehog pathways, including curcumin,<sup>74</sup> resveratrol, epigallocatechin-3-gallate (EGCG), lycopene, and retinoids.<sup>74-76</sup> Analysis of these plant components revealed several unique inhibitors of the Wnt/ $\beta$ -catenin and hedgehog signaling pathways.<sup>77</sup>

## **Epigenetic Regulation of Ror Receptor Pathway**

Epigenetic regulations such as DNA methylation and histone modification closely coordinate the transcriptional activity of genes, and specific epigenetic structures are necessary to maintain normal cell function.78 DNA methylation occurs in the covalent modification of cytosine 5-C, mainly in the context of cytosine guanine dinucleotide.79 The mechanism for establishing and maintaining DNA methylation is well established and involves 3 mammalian methyltransferases (Dnmts): Dnmt1, 3A, and 3B.80 Germ-line deletion studies in mice have identified an essential role for Dnmts in embryonic heart development.<sup>81</sup> Recent genome-wide DNA methylation analysis studies have identified extensive changes in DNA methylation patterns during early embryonic heart development and revealed potentially essential associations with changes in cardiogenic gene transcription.82 Furthermore, dynamic changes in DNA methylation patterns are associated with changes in gene expression in human heart failure and dilated cardiomyopathy, and inhibition of DNA methylation may have a cardioprotective effect in norepinephrine-induced hypertrophy ischemic heart disease.83 Inadequate epigenetic modification control activates oncogenes and inhibits gene inactivation, leading to the development of various cancers.<sup>84</sup> It has recently been reported that colon cancer occurs through the accumulation of abnormal DNA methylation and the destruction of histone coding. Abnormal DNA methylation has been detected in normal mucosa in early colon cancer.85 Overall hypomethylation leads to chromosomal instability and oncogene activation. The promoter -CpG island is a CpG dinucleotide-rich region, and changes in its methylation intensity alter essential tumor suppressor genes.86

In addition to DNA methylation, histone modification is considered to be a key regulator of gene activity. Acetylation, methylation, and phosphorylation of histone-specific residues, such as Lys 4, Lys 9, and Ser 10, provide chemical driving forces for chromatin configuration that prevent or allow the initiation of gene transcription.<sup>87</sup> Increased histone deacetylase activity such as HDAC3 in the nucleus is associated with decreased histone acetylation.<sup>88</sup> In addition, the cross-talk between DNA methylation and histone modification is a hot topic in recent years. On the other hand, DNA methylation also affects histone methylation. It has been shown that nucleosomes composed of acetylated histones assemble unmethylated DNA.<sup>89</sup>

## **Summary and Prospect**

In the past decade, few studies have been published on the characteristics of Wnt signaling in general and its role in cardiovascular disease in particular. In this review, we focused on the part of Wnt signaling in congenital heart diseases, cardiovascular diseases, including atherosclerosis and cardiac hypertrophy. However, underlying standard features in the pathological mechanisms of these diseases seem to process such as cell proliferation and differentiation, which the role of Wnt signaling has been demonstrated. This explains the regulatory role of Wnt signaling pathways in these diverse cardiovascular conditions.

The Wnt family is a large class of highly conserved cysteinerich secretory glycoproteins that play a vital role in various cellular and physiological courses through different signaling pathways during embryogenesis and tissue homeostasis.<sup>3</sup> Wnt5a is a secreted glycoprotein that belongs to the non-canonical Wnt family and is involved in a wide range of developmental and tissue homeostasis. A growing body of evidence suggests that Wnt5a affects embryonic development, signaling through various receptors, starting with the activation of  $\beta$ -catenin by Wnt5a. In addition to affecting planar cell polarity and Ca<sup>2+</sup> pathways, β-catenin also includes multiple signaling cascades that regulate various cell functions. Secondly, Wnt5a can bind to Ror receptors to mediate non-canonical Wnt signaling and is a significant ligand for Ror2 in vertebrates. Consistent with the multiple functions of Wnt5A/Ror2 signaling, Wnt5A knockout mice exhibited various phenotypic defects, including an inability to extend the anterior and posterior axes of the embryo. Numerous essential roles of Wnt5a/Ror2 in development have been demonstrated. The Wnt5a-Ror2 signaling pathway as a critical factor has attracted extensive attention. Although the pathogenicity of the Ror2 receptor in congenital heart disease has not been thoroughly studied, the study of Wnt5a signaling and the Ror2 receptor still has a long way to go.

Overall, this review provides new insights into the role of DNA methylation in the regulation of neonatal cardiac maturation. However, further research is needed to clarify the mechanisms that induce specific methylation changes in embryonic heart development. Further analysis of DNA methylation during embryonic heart development may better understand the mechanisms that drive cardiac cell cycle arrest and binucleate formation.

## Acknowledgements

The authors thank Xing Quan Sheng for the helpful discussion and for desinging the manuscript. We are thank full to Xing Yuhan, Li Guoju, and Du Zhanhui for their helpful suggestion and reviewing the article. All the authors have no conflict of interest.

#### **ORCID** iD

Rui Guo 问 https://orcid.org/0000-0003-0671-9975

#### REFERENCES

- Ewer AK, Middleton LJ, Furmston AT, et al. Pulse oximetry screening for congenital heart defects in newborn infants (PulseOx): a test accuracy study. *Lancet*. 2011;378:785-794.
- Oyen N, Poulsen G, Wohlfahrt J, Boyd HA, Jensen PK, Melbye M. Recurrence of discordant congenital heart defects in families. *Circ Cardiovasc Genet*. 2010;3:122-128.
- Angers S, Moon RT. Proximal events in Wnt signal transduction. Nat Rev Mol Cell Biol. 2009;10:468-477.
- Moon RT, Brown JD, Torres M. WNTs modulate cell fate and behavior during vertebrate development. *Trends Genet*. 1997;13:157-162.
- Foulquier S, Daskalopoulos EP, Lluri G, Hermans KCM, Deb A, Blankesteijn WM. WNT signaling in cardiac and vascular disease. *Pharmacol Rev.* 2018; 70:68-141.
- Zhong X, Zhao X, Liu Z, Guo Y, Ma L. Childhood disability and its associated perinatal characteristics in Bao'an district of Shenzhen, China. *BMC Public Health.* 2020;20:1540.
- Bouveret R, Waardenberg AJ, Schonrock N, et al. NKX2-5 mutations causative for congenital heart disease retain functionality and are directed to hundreds of targets. *eLife*. 2015;4:e06942.
- Peng L, Qian M, Liu Z, et al. Deacetylase-independent function of SIRT6 couples GATA4 transcription factor and epigenetic activation against cardiomyocyte apoptosis. *Nucleic Acids Res.* 2020;48:4992-5005.
- Kim MS, Fleres B, Lovett J, et al. Contractility of induced pluripotent stem cellcardiomyocytes with an MYH6 head domain variant associated with hypoplastic left heart syndrome. *Front Cell Dev Biol.* 2020;8:440.
- Theilmann AL, Hawke LG, Hilton LR, et al. Endothelial BMPR2 loss drives a proliferative response to BMP (bone morphogenetic protein) 9 via prolonged canonical signaling. *Arterioscler Thromb Vasc Biol*. 2020;40:2605-2618.
- Richards AA, Garg V. Genetics of congenital heart disease. *Curr Cardiol Rev.* 2010;6:91-97.
- Nusse R, Varmus H. Three decades of Whts: a personal perspective on how a scientific field developed. *EMBO J.* 2012;31:2670-2684.
- Brinkmann EM, Mattes B, Kumar R, et al. Secreted frizzled-related protein 2 (sFRP2) redirects non-canonical Wnt signaling from Fz7 to Ror2 during vertebrate gastrulation. J Biol Chem. 2016;291:13730-13742.
- Nye DMR, Albertson RM, Weiner AT, et al. The receptor tyrosine kinase Ror is required for dendrite regeneration in Drosophila neurons. *PLoS Biol.* 2020;18: e3000657.
- González-Sancho JM, Brennan KR, Castelo-Soccio LA, Brown AM. Wnt proteins induce dishevelled phosphorylation via an LRP5/6- independent mechanism, irrespective of their ability to stabilize beta-catenin. *Mol Cell Biol.* 2004; 24:4757-4768.
- Baranov PV, Wills NM, Barriscale KA, et al. Programmed ribosomal frameshifting in the expression of the regulator of intestinal stem cell proliferation, adenomatous polyposis coli (APC). *RNA Biol.* 2011;8:637-647.
- Kemp C, Willems E, Abdo S, Lambiv L, Leyns L. Expression of all Wnt genes and their secreted antagonists during mouse blastocyst and postimplantation development. *Dev Dyn*. 2005;233:1064-1075.
- Lin PI, Chien YL, Wu YY, et al. The WNT2 gene polymorphism associated with speech delay inherent to autism. *Res Dev Disabil*. 2012;33:1533-1540.
- Sonderegger S, Pollheimer J, Knöfler M. Wnt signalling in implantation, decidualisation and placental differentiation—review. *Placenta*. 2010;31: 839-847.
- Sebastian A, Hum NR, Murugesh DK, Hatsell S, Economides AN, Loots GG. Wnt co-receptors Lrp5 and Lrp6 differentially mediate Wnt3a signaling in osteoblasts. *PLoS One*. 2017;12:e0188264.
- 21. Vainio S, Heikkilä M, Kispert A, Chin N, McMahon AP. Female development in mammals is regulated by Wnt-4 signalling. *Nature*. 1999;397:405-409.
- 22. Atli MO, Guzeloglu A, Dinc DA. Expression of wingless type (WNT) genes and their antagonists at mRNA levels in equine endometrium during the estrous cycle and early pregnancy. *Anim Reprod Sci.* 2011;125:94-102.
- Cooke PS, Spencer TE, Bartol FF, Hayashi K. Uterine glands: development, function and experimental model systems. *Mol Hum Reprod.* 2013;19:547-558.
- Gray CA, Taylor KM, Ramsey WS, et al. Endometrial glands are required for preimplantation conceptus elongation and survival. *Biol Reprod.* 2001;64: 1608-1613.
- 25. Schleiffarth JR, Person AD, Martinsen BJ, et al. Wnt5a is required for cardiac outflow tract septation in mice. *Pediatr Res.* 2007;61:386-391.
- Stricker S, Verhey van Wijk N, Witte F, Brieske N, Seidel K, Mundlos S. Cloning and expression pattern of chicken Ror2 and functional characterization of

8

truncating mutations in Brachydactyly type B and Robinow syndrome. *Dev Dyn.* 2006;235:3456-3465.

- 27. Zhang M, Sun C, Liu R, et al. Phenotypes and epigenetic errors in patients with Beckwith-Wiedemann syndrome in China. *Transl Pediatr.* 2020;9:653-661.
- 28. Touma M, Kang X, Gao F, et al. Wht11 regulates cardiac chamber development and disease during perinatal maturation. *JCI Insight*. 2017;2:e94904.
- 29. Eisenberg CA, Eisenberg LM. WNT11 promotes cardiac tissue formation of early mesoderm. *Dev Dyn.* 1999;216:45-58.
- Stewart CL, Kaspar P, Brunet LJ, et al. Blastocyst implantation depends on maternal expression of leukaemia inhibitory factor. *Nature*. 1992;359:76-79.
- West-Livingston LN, Park J, Lee SJ, Atala A, Yoo JJ. The role of the microenvironment in controlling the fate of bioprinted stem cells. *Chem Rev.* 2020; 120:11056-11092.
- McCubrey JA, Rakus D, Gizak A, et al. Effects of mutations in Wnt/β-catenin, hedgehog, Notch and PI3K pathways on GSK-3 activity-diverse effects on cell growth, metabolism and cancer. *Biochim Biophys Acta*. 2016;1863:2942-2976.
- Cohen ED, Wang Z, Lepore JJ, et al. Wnt/beta-catenin signaling promotes expansion of isl-1-positive cardiac progenitor cells through regulation of FGF signaling. J Clin Investig. 2007;117:1794-1804.
- Monkley SJ, Delaney SJ, Pennisi DJ, Christiansen JH, Wainwright BJ. Targeted disruption of the Wnt2 gene results in placentation defects. *Development*. 1996;122:3343-3353.
- Jaspard B, Couffinhal T, Dufourcq P, Moreau C, Duplàa C. Expression pattern of mouse sFRP-1 and mWnt-8 gene during heart morphogenesis. *Mech Dev.* 2000;90:263-267.
- Zakin LD, Mazan S, Maury M, Martin N, Guénet JL, Brûlet P. Structure and expression of Wnt13, a novel mouse wnt2 related gene. *Mech Dev.* 1998; 73:107-116.
- Mazzotta S, Neves C, Bonner RJ, Bernardo AS, Docherty K, Hoppler S. Distinctive roles of canonical and noncanonical wnt signaling in human embryonic cardiomyocyte development. *Stem Cell Reports*. 2016;7:764-776.
- Liu Y, Ross JF, Bodine PV, Billiard J. Homodimerization of Ror2 tyrosine kinase receptor induces 14-3-3(beta) phosphorylation and promotes osteoblast differentiation and bone formation. *Mol Endocrinol.* 2007;21:3050-3061.
- Sakane H, Yamamoto H, Matsumoto S, Sato A, Kikuchi A. Localization of glypican-4 in different membrane microdomains is involved in the regulation of Wnt signaling. *J Cell Sci.* 2012;125:449-460.
- Laird DJ, Altshuler-Keylin S, Kissner MD, Zhou X, Anderson KV. Ror2 enhances polarity and directional migration of primordial germ cells. *PLoS Genet*. 2011;7:e1002428.
- Oishi I, Sugiyama S, Liu ZJ, Yamamura H, Nishida Y, Minami Y. A novel Drosophila receptor tyrosine kinase expressed specifically in the nervous system. Unique structural features and implication in developmental signaling. *J Biol Chem.* 1997;272:11916-11923.
- DeChiara TM, Kimble RB, Poueymirou WT, et al. Ror2, encoding a receptorlike tyrosine kinase, is required for cartilage and growth plate development. *Nat Genet.* 2000;24:271-274.
- Feike AC, Rachor K, Gentzel M, Schambony A. Wnt5a/Ror2-induced upregulation of xPAPC requires xShcA. *Biochem Biophys Res Commun.* 2010; 400:500-506.
- Oldridge M, Fortuna AM, Maringa M, et al. Dominant mutations in ROR2, encoding an orphan receptor tyrosine kinase, cause brachydactyly type B. *Nat Genet.* 2000;24:275-278.
- Gao B, Song H, Bishop K, et al. Wnt signaling gradients establish planar cell polarity by inducing Vangl2 phosphorylation through Ror2. *Dev Cell*. 2011; 20:163-176.
- Takeuchi S, Takeda K, Oishi I, et al. Mouse Ror2 receptor tyrosine kinase is required for the heart development and limb formation. *Genes Cells*. 2000; 5:71-78.
- Nomi M, Oishi I, Kani S, et al. Loss of mRor1 enhances the heart and skeletal abnormalities in mRor2-deficient mice: redundant and pleiotropic functions of mRor1 and mRor2 receptor tyrosine kinases. *Mol Cell Biol.* 2001;21:8329-8335.
- Patan S. Vasculogenesis and angiogenesis as mechanisms of vascular network formation, growth and remodeling. J Neurooncol. 2000;50:1-15.
- Rüger BM, Buchacher T, Dauber EM, et al. De novo vessel formation through cross-talk of blood-derived cells and mesenchymal stromal cells in the absence of pre-existing vascular structures. *Front Bioeng Biotechnol.* 2020;8:602210.
- 50. Yuan K, Shamskhou EA, Orcholski ME, et al. Loss of endothelium-derived Wnt5a Is associated with reduced pericyte recruitment and small vessel loss in pulmonary arterial hypertension. *Circulation*. 2019;139:1710-1724.
- Karuna EP, Choi SS, Scales MK, et al. Identification of a WNT5A-responsive degradation domain in the Kinesin superfamily protein KIF26B. *Genes*. 2018;9:196.
- 52. Takahashi J, Orcholski M, Yuan K, de Jesus Perez V. PDGF-dependent  $\beta$ -catenin activation is associated with abnormal pulmonary artery smooth muscle cell proliferation in pulmonary arterial hypertension. *FEBS Lett.* 2016;590:101-109.

- Bhatt T, Rizvi A, Batta SP, Kataria S, Jamora C. Signaling and mechanical roles of E-cadherin. *Cell Commun Adhes*. 2013;20:189-199.
- Choi S, Gustafson-Wagner EA, Wang Q, et al. The intercalated disk protein, mXinalpha, is capable of interacting with beta-catenin and bundling actin filaments [corrected]. *J Biol Chem.* 2007;282:36024–36036.
- Masuelli L, Bei R, Sacchetti P, et al. Beta-catenin accumulates in intercalated disks of hypertrophic cardiomyopathic hearts. *Cardiovasc Res.* 2003;60: 376-387.
- Pai SG, Carneiro BA, Mota JM, et al. Wnt/beta-catenin pathway: modulating anticancer immune response. *J Hematol Oncol.* 2017;10:101.
- Butler MT, Wallingford JB. Planar cell polarity in development and disease. Nat Rev Mol Cell Biol. 2017;18:375-388.
- Miyoshi H, Ajima R, Luo CT, Yamaguchi TP, Stappenbeck TS. Wnt5a potentiates TGF-β signaling to promote colonic crypt regeneration after tissue injury. *Science*. 2012;338:108-113.
- Green J, Nusse R, van Amerongen R. The role of Ryk and Ror receptor tyrosine kinases in wnt signal transduction. *Cold Spring Harb Perspect Biol.* 2014; 6:014.
- Cantú AV, Altshuler-Keylin S, Laird DJ. Discrete somatic niches coordinate proliferation and migration of primordial germ cells via Wnt signaling. J Cell Biol. 2016;214:215-229.
- 61. Keyte AL, Alonzo-Johnsen M, Hutson MR. Evolutionary and developmental origins of the cardiac neural crest: building a divided outflow tract. *Birth Defects Res C Embryo Today.* 2014;102:309-323.
- Hamblet NS, Lijam N, Ruiz-Lozano P, et al. Dishevelled 2 is essential for cardiac outflow tract development, somite segmentation and neural tube closure. *Development*. 2002;129:5827-5838.
- Gentzel M, Schille C, Rauschenberger V, Schambony A. Distinct functionality of dishevelled isoforms on Ca<sup>2+</sup>/calmodulin-dependent protein kinase 2 (Cam-KII) in Xenopus gastrulation. *Mol Biol Cell*. 2015;26:966-977.
- 64. Sinha T, Wang B, Evans S, Wynshaw-Boris A, Wang J. Disheveled mediated planar cell polarity signaling is required in the second heart field lineage for outflow tract morphogenesis. *Dev Biol.* 2012;370:135-144.
- Cha J, Bartos A, Park C, et al. Appropriate crypt formation in the uterus for embryo homing and implantation requires Wnt5a-ROR signaling. *Cell Rep.* 2014;8:382-392.
- Obeid R, Holzgreve W, Pietrzik K. Folate supplementation for prevention of congenital heart defects and low birth weight: an update. *Cardiovasc Diagn Ther.* 2019;9:S424-S433.
- Øyen N, Olsen SF, Basit S, et al. Association between maternal folic acid supplementation and congenital heart defects in offspring in birth cohorts from Denmark and Norway. J Am Heart Assoc. 2019;8:e011615.
- Li X, Li S, Mu D, et al. The association between periconceptional folic acid supplementation and congenital heart defects: a case-control study in China. Prev Med. 2013;56:385-389.
- Greene ND, Leung KY, Copp AJ. Inositol, neural tube closure and the prevention of neural tube defects. *Birth Defects Res.* 2017;109:68-80.
- Yu W, Greenberg ML. Inositol depletion, GSK3 inhibition and bipolar disorder. *Future Neurol.* 2016;11:135-148.
- Gao Y, Wang HY. Inositol pentakisphosphate mediates Wnt/beta-catenin signaling. J Biol Chem. 2007;282:26490-26502.
- Linask KK, Huhta J. Folate protection from congenital heart defects linked with canonical wnt signaling and epigenetics. *Curr Opin Pediatr.* 2010;22: 561-566.
- 73. Ullah A, Munir S, Badshah SL, et al. Important flavonoids and their role as a therapeutic agent. *Molecules*. 2020;25:5243.
- Kumar V, Vashishta M, Kong L, et al. The role of Notch, Hedgehog, and Wnt signaling pathways in the resistance of tumors to anticancer therapies. *Front Cell* Dev Biol. 2021;9:650772.
- Yuan L, Zhou M, Huang D, et al. Resveratrol inhibits the invasion and metastasis of colon cancer through reversal of epithelial-mesenchymal transition via the AKT/GSK-3β/snail signaling pathway. *Mol Med Rep.* 2019;20:2783-2795.
- Zhang H, Nan W, Wang S, et al. Epigallocatechin-3-Gallate promotes the growth of mink hair follicles through sonic Hedgehog and protein kinase B signaling pathways. *Front Pharmacol.* 2018;9:674.
- Farahmand L, Darvishi B, Majidzadeh A, Madjid Ansari K, A, Naturally occurring compounds acting as potent anti-metastatic agents and their suppressing effects on Hedgehog and WNT/β-catenin signalling pathways. *Cell Prolif.* 2017;50:e12299.
- Patel DJ. A structural perspective on readout of epigenetic histone and DNA methylation marks. *Cold Spring Harb Perspect Biol.* 2016;8:a018754.
- 79. Liu Y, Zhang Y, Yin J, et al. Distinct H3K9me3 and DNA methylation modifications during mouse spermatogenesis. *J Biol Chem.* 2019;294:18714-18725.
- Ren W, Gao L, Song J. Structural basis of DNMT1 and DNMT3A-mediated DNA methylation. *Genes.* 2018;9:620.
- 81. Cui D, Xu X. DNA methyltransferases, DNA methylation, and age-associated cognitive function. *Int J Mol Sci.* 2018;19:1315.

- Zhou J, Xiong Y, Dong X, et al. Genome-wide methylation analysis reveals differentially methylated CpG sites and altered expression of heart developmentassociated genes in fetuses with cardiac defects. *Exp Ther Med.* 2021;22:1032.
- Tabish AM, Arif M, Song T, et al. Association of intronic DNA methylation and hydroxymethylation alterations in the epigenetic etiology of dilated cardiomyopathy. *Am J Physiol Heart Circ Physiol.* 2019;317:H168-H180.
- Llinàs-Arias P, Esteller M. Epigenetic inactivation of tumour suppressor coding and non-coding genes in human cancer: an update. *Open Biol.* 2017;7:170152.
- Hanley MP, Hahn MA, Li AX, et al. Genome-wide DNA methylation profiling reveals cancer-associated changes within early colonic neoplasia. *Oncogene*. 2017;36:5035-5044.
- Li Z, Ren T, Li W, et al. Association between the methylation statuses at CpG sites in the promoter region of the SLCO1B3, RNA expression and color change in blue eggshells in lushi chickens. *Front Genet.* 2019;10:161.
- Nowak SJ, Corces VG. Phosphorylation of histone H3 correlates with transcriptionally active loci. *Genes Dev.* 2000;14:3003-3013.
- Patil H, Wilks C, Gonzalez RW, Dhanireddy S, Conrad-Webb H, Bergel M. Mitotic activation of a novel Histone deacetylase 3-linker Histone H1.3 protein complex by protein kinase CK2. *J Biol Chem.* 2016;291:3158-3172.
- Du J, Johnson LM, Jacobsen SE, Patel DJ. DNA methylation pathways and their crosstalk with histone methylation. *Nat Rev Mol Cell Biol.* 2015;16: 519-532.