RSC Advances



PAPER

Check for updates

Cite this: RSC Adv., 2022, 12, 22175

Oxa-Michael-based divergent synthesis of artificial glutamate analogs[†]

Shuntaro Tsukamoto, ^b^a Oriel Hlokoane, ^b^{ab} Kei Miyako, ^b^c Raku Irie, ^b^a Ryuichi Sakai ^c^c and Masato Oikawa ^b*^a

Herein we report stereoselective generation of two skeletons, 1,3-dioxane and tetrahydropyranol, by oxa-Michael reaction as the key reaction from δ -hydroxyenone. The construction of the 1,3-dioxane skeleton, achieved through hemiacetal formation followed by oxa-Michael reaction from δ -hydroxyenone, was exploited to access structurally diverse heterotricyclic artificial glutamate analogs. On the other hand, formation of a novel tetrahydro-2*H*-pyranol skeleton was accomplished by the inverse reaction order: oxa-Michael reaction followed by hemiacetal formation. Thus, this study succeeded in showing that structural diversity in a compound collection can be acquired by interchanging the order of just two reactions. Among the skeletally diverse, heterotricyclic artificial glutamate analogs synthesized in this study, a neuronally active compound named TKM-50 was discovered in the mice *in vivo* assay.

Received 17th June 2022 Accepted 2nd August 2022

DOI: 10.1039/d2ra03744k

rsc.li/rsc-advances

Ionotropic glutamate receptors (iGluRs) mediate the majority of the excitatory neurotransmissions such as learning, memory, and nociception in the mammalian central nervous system (CNS).¹ To study and control the function of iGluRs, specific glutamate analogs have been developed in natural product chemistry² and in medicinal chemistry.^{3,4} IKM-159 (Fig. 1A) is an artificial glutamate analog designed and developed based on dysiherbaine5,6 and kainic acid7 in our laboratories as an antagonist selective to (S)-2-amino-3-(3-hydroxy-5-methyl-4isoxazolvl)propionic acid (AMPA)-type iGluR.⁸⁻¹² The AMPA receptor consists of four subunits: GluA1, GluA2, GluA3, and GluA4.3 In vitro, IKM-159 selectively inhibits the GluA1/GluA2 heterodimer and GluA4 homodimer.10 In vivo, IKM-159 inhibits voluntary action of mice for 50 min to several hours upon intracerebroventricular injection. The potency and selectivity of IKM-159 are, however, not very satisfactory to selectively modulate the function of AMPA-type iGluR. As an attempt to improve the biological profiles of IKM-159, we have been studying its structural modification.

From the first-generation studies on structure–activity relationships (SARs) of IKM-159, it had been shown that the ring size and the heteroatom of the C-ring were important for neuroactivity of IKM-159.^{10,13} We then studied the secondgeneration SAR on the oxa analogs generated by a Prins-Ritter three-component coupling strategy, although all analogs were found to lose the original neuronal activity of IKM-159.¹⁴ Herein, we report our continuous effort along this line employing the homoallylic alcohol such as 5 and 7 (see Scheme 2) as the common intermediates.¹⁵

One of the strategies in this work is the thermodynamically controlled, stereoselective formation of 1,3-dioxane (1 in Fig. 1B) by hemiacetal formation followed by oxa-Michael reaction from δ -hydroxyenone derivative that we recently developed (Scheme 1).¹⁶ The other strategy is the novel stereoselective formation of tetrahydropyranol (2 in Fig. 1B) by the inverse reaction order; oxa-Michael reaction followed by hemiacetal formation (see Scheme 5). Thus, this study succeeded in showing that structural diversity in a compound collection can be acquired by interchanging the order of just two reactions; hemiacetal formation and oxa-Michael reaction. Among the skeletally diverse, heterotricyclic artificial glutamate analogs thus synthesized, a compound named TKM-50 (1ar) was discovered to be neuronally active in the mice *in vivo* assay.

The substrate used for the 1,3-dioxane formation was prepared from the known dimethyl ester 5 (Scheme 2).¹⁷

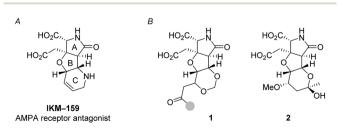
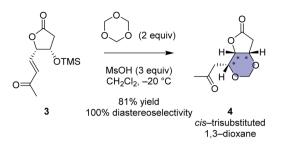


Fig. 1 Background (A) and summary (B) of this work. (A) AMPA-type iGluR antagonist IKM-159. (B) Artificial glutamate analogs 1 and 2, generated by oxa-Michael-based transformations (this work). The gray circle in 1 denotes the position for the structural diversity.

^eGraduate School of Nanobioscience, Yokohama City University, Seto 22-2, Kanazawa-ku, Yokohama 236-0027, Japan. E-mail: moikawa@yokohama-cu.ac.jp ^bDepartment of Pharmacy, National University of Lesotho, P.O. Roma 180, Maseru, Lesotho

^cFaculty of Fisheries Sciences, Hokkaido University, Hakodate 041-8611, Japan † Electronic supplementary information (ESI) available. See https://doi.org/10.1039/d2ra03744k

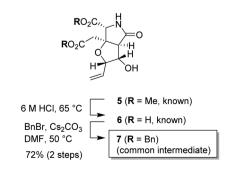


Scheme 1 Our recent work regarding stereoselective 1,3-dioxane formation.¹⁶ For clarity and comparison, enantiomers of the reported compounds are shown in this scheme.

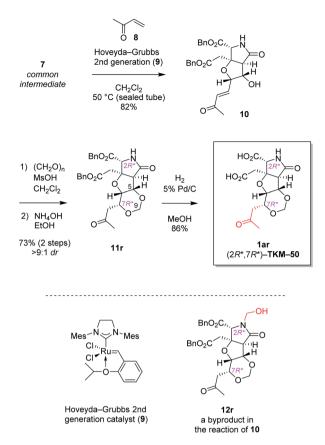
Exposure of dimethyl ester 5 to hydrochloric acid (6 M) at 65 °C provided dicarboxylic acid $6.^{17}$ Without purification, dicarboxylic acid 6 was treated with BnBr and Cs_2CO_3 to furnish the common intermediate 7 in 72% yield (2 steps).¹⁸

The alkene 7 was subjected to cross metathesis with methyl vinyl ketone (8) mediated by Hoveyda-Grubbs second generation catalyst (9)¹⁹ to provide enone 10 in 82% yield (Scheme 3). Upon exposure to paraformaldehyde as an equivalent of formaldehyde and 1,3,5-trioxane¹⁶ in the presence of MsOH, 1,3dioxane ring formed smoothly by oxa-Michael reaction to give rise to desired $(7R^*)$ -heterotricycle **11r** and the $(7S^*)$ epimer **11s** (structure not shown) in the ratio of >9:1, as well as the Nhydroxymethylated product 12r (see Scheme 3) and the $(7S^*)$ epimer 12s (structure not shown). Since we had found that alkaline hydrolysis is of use to remove the N-hydroxymethyl group, the mixture of hemiaminals (12r/12s) and free amides (11r/11s) was treated with ammonium hydroxide²⁰ to obtain free amide 11r in 73% isolated yield (2 steps), and free amide 11s in 10% yield (estimated by NMR, 2 steps). The formation of 1,3dioxane ring of 11r was determined by the HMBC correlations (Fig. 2A), and the stereochemical configuration was established by a ${}^{3}J_{H,H}$ value and NOESY correlations denoted in Fig. 2B. Both configuration and conformation of 11r are identical to those we observed recently in the simple case $(3 \rightarrow 4, \text{ see})$ Scheme 1),¹⁶ showing that the 1,3-dioxane formation in this study is also thermodynamically controlled (see below for the mechanism).

The proposed mechanism for the 1,3-dioxane formation is shown in Scheme 4A. Reaction of alcohol **10** and paraformaldehyde would form hemiacetal intermediate **A** under acidic conditions, which, then undergoes intramolecular oxa-



Scheme 2 Preparation of the common intermediate 7 (racemate).



Scheme 3 Stereoselective 1,3-dioxane formation leading to heterotricyclic artificial glutamate analog 1ar.^{*a*} ^{*a*}dr denotes the diastereoselectivity in the 1,3-dioxane formation.

Michael reaction to give **11r** and **11s**. Since the second conjugate addition is generally a thermodynamically controlled, reversible process, production of more stable ($7R^*$) isomer **11r** predominated over the ($7S^*$) epimer **11s**, as discussed also in our preliminary study.¹⁶ It should be also noted here that, in that preliminary study employing a simple substrate, the (7S) epimer had not been obtained.¹⁶ Generation of the less stable ($7S^*$) epimer **11s** in this study would be due to unfavorable steric interactions between the acetyl group and the benzyl ester on the near side in **11r** (Scheme 4B), that make the energy difference between the two diastereomers (**11r** and **11s**) smaller.

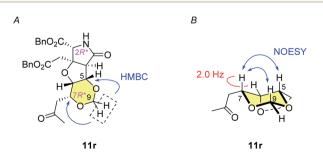
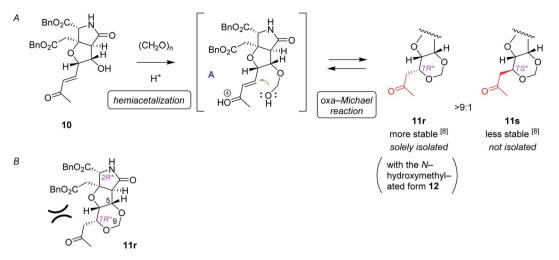


Fig. 2 Structure analysis of 1,3-dioxane **11r**. (A) HMBC correlations in **11r** indicates formation of the 1,3-dioxane ring. (B) Small ${}^{3}J_{H,H}$ value and NOESY correlations show the configuration and the conformation of **11r**.



Scheme 4 The plausible mechanism of 1,3-dioxane ring formation. (A) Stepwise mechanism that consists of hemiacetal formation followed by intramolecular oxa-Michael reaction. (B) The steric repulsion included in the stable isomer **11**r.

Then two benzyl groups of **11r** were removed by hydrogenolysis²¹ to cleanly provide glutamate analog **1ar** ($(2R^*, 7R^*)$ -TKM-50) in 86% yield (Scheme 3).

With the same reaction sequences for **1ar** (Scheme 3), two more analogs **1br** and **1cr** were furthermore synthesized (Fig. 3). The marked decrease in diastereoselectivity in these oxa-Michael reactions (see Fig. 3) suggests that the steric repulsion between the pentyl/methoxyphenyl group and the benzyl ester on the near side is extremely large. The minor ($7S^*$) diastereomers obtained in these oxa-Michael reactions were also isolated and deprotected to give **1bs** and **1cs** (see the ESI†), which were subjected to *in vivo* assay (see below).

We also found that another skeleton can be constructed from δ -hydroxyenone being used for 1,3-dioxane formation, under alkaline hydrolytic conditions. Thus, as shown in Scheme 5, the δ -hydroxyenone 13 derived from homoallylic alcohol 5 by cross metathesis was selectively transformed into cyclic hemiacetal 2 in 53% yield (1 M LiOH in water, MeOH, rt). In this transformation, dimethyl ester and δ -hydroxyenone moiety independently suffer hydrolysis and cyclization, respectively, to generate glutamate analog 2 efficiently. The configuration of 2 was determined by combined analysis of NMR and DFT calculation (see the ESI†). $^{\rm 22}$

The plausible mechanism for the hemiacetal formation is shown in Scheme 6. In view of the fact that the hydroxy and carbonyl groups are located apart in 13, the six-membered-ring formation should take place after saturation of the *trans*-alkene. It is, therefore, supposed that oxa-Michael reaction of MeOH to enone 13 first generates saturated ketone C *via* enolate B.²³ Under alkaline conditions, the alkoxide C intramolecularly attacks carbonyl group to give rise to hemiacetal 2. Considering the fact that oxa-Michael reaction and the acetalization are thermodynamically controlled, reversible processes, energetically favorable diastereomer 2 would have been obtained predominantly (see the ESI† for discussions on thermodynamic stability of 2). A related example had been reported in 1992 by Shing *et al.*²⁴

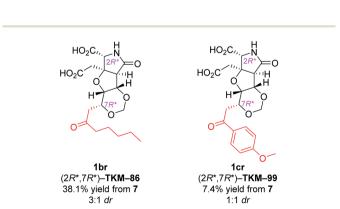
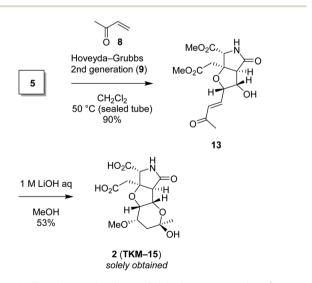
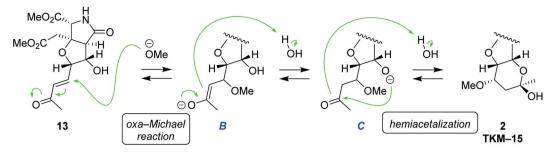


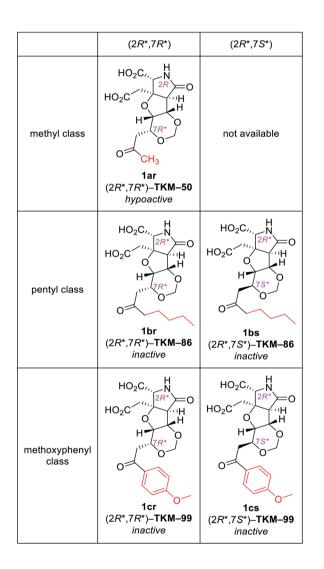
Fig. 3 Other 1,3-dioxane analogs synthesized by the intramolecular oxa-Michael reaction.^a ^adr denotes the diastereoselectivity in the 1,3-dioxane formation.



Scheme 5 The heterotricyclic artificial glutamate analog 2, constructed by intermolecular oxa-Michael reaction of MeOH followed by acetalization.



Scheme 6 The plausible mechanism for hemiacetal formation under alkaline conditions.



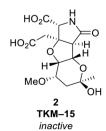


Fig. 4 The *in vivo* activities on mice.

Behavioral activities of all six compounds upon intracerebroventricular (i.c.v.) injection were evaluated in mice (Fig. 4).²⁵ Injection of **1ar** (TKM-50, 50 µg per mouse) resulted in loss of voluntary motor activity for 10 min after injection and then ataxia-like motions were recorded, thus annotated as hypoactive. The hypoactivity observed for **1ar** (TKM-50) is thus somewhat weaker than IKM-159 which causes loss of mice spontaneous activity for up to 4 h.¹² Other congeners, however, did not cause any noticeable behavioral changes at the same dose tested.

Conclusions

In this paper, we reported synthesis of skeletally diverse artificial glutamate analogs from a common precursor. Since we employed thermodynamically controlled, reversible process for the key cyclizations, most of the reactions proceeded stereoselectively. The cases that were less selective (**1br** and **1cr** in Fig. 3) could even be reasonably explained, supporting the origin of the stereoselectivity we proposed in Scheme 4.¹⁶

It is of interest to note that the formed skeleton changes significantly, just by interchanging the order of the oxa-Michael reaction and the hemiacetalization (see Schemes 4 and 6). Therefore, it is expected that our methodology is generally of use for discovery of biologically active small molecules.²⁶ In fact, we succeeded in identifying neuroactive compound (**1ar**, TKM-50) in this study.

We are currently working on the construction of a larger compound library using this methodology and the development of alternative methodology for generation of other skeletons. The results will be reported in due course.

Author contributions

ST: investigation, writing the first draft and editing; OH: investigation and editing; KM: investigation; RI: formal analysis and editing; RS: funding acquisition, investigation and writing the first draft; MO: conceptualization, formal analysis, funding acquisition, project administration, supervision, writing the first draft and final editing.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was supported by the grant for Academic Research Promotion (No. SG2803) of Yokohama City University, Japan. The JSPS grant-in-aid for scientific research (15H04546, 18H0460009, 19H03040, 20H0475800) is also gratefully acknowledged. The financial support from the government of Japan through Japan International Cooperation Agency (JICA) is gratefully acknowledged by O.H.

Notes and references

- 1 G. Riedel, B. Platt and J. Micheau, *Behav. Brain Res.*, 2003, **140**, 1–47.
- 2 G. T. Swanson and R. Sakai, in *Marine Toxins as Research Tools*, ed. N. Fusetani and W. Kem, Springer Berlin Heidelberg, Berlin, Heidelberg, 2009, pp. 123–157.
- 3 P. Stawski, H. Janovjak and D. Trauner, *Bioorg. Med. Chem.*, 2010, **18**, 7759–7772.
- 4 S. K. Bagal, A. D. Brown, P. J. Cox, K. Omoto, R. M. Owen,
 D. C. Pryde, B. Sidders, S. E. Skerratt, E. B. Stevens,
 R. I. Storer and N. A. Swain, *J. Med. Chem.*, 2013, 56, 593–624.
- 5 R. Sakai, H. Kamiya, M. Murata and K. Shimamoto, J. Am. Chem. Soc., 1997, 119, 4112–4116.
- 6 R. Sakai, T. Koike, M. Sasaki, K. Shimamoto, C. Oiwa, A. Yano, K. Suzuki, K. Tachibana and H. Kamiya, *Org. Lett.*, 2001, 3, 1479–1482.
- 7 A. F. Parsons, Tetrahedron, 1996, 52, 4149-4174.
- 8 M. Ikoma, M. Oikawa, M. B. Gill, G. T. Swanson, R. Sakai,
 K. Shimamoto and M. Sasaki, *Eur. J. Org. Chem.*, 2008, 2008, 5215–5220.
- 9 M. Oikawa, M. Ikoma, M. Sasaki, M. B. Gill, G. T. Swanson, K. Shimamoto and R. Sakai, *Eur. J. Org. Chem.*, 2009, **2009**, 5531–5548.
- 10 M. B. Gill, S. Frausto, M. Ikoma, M. Sasaki, M. Oikawa, R. Sakai and G. T. Swanson, *Br. J. Pharmacol.*, 2010, **160**, 1417–1429.
- 11 M. Oikawa, M. Ikoma, M. Sasaki, M. B. Gill, G. T. Swanson, K. Shimamoto and R. Sakai, *Bioorg. Med. Chem.*, 2010, 18, 3795–3804.
- 12 L. Juknaite, Y. Sugamata, K. Tokiwa, Y. Ishikawa, S. Takamizawa, A. Eng, R. Sakai, D. S. Pickering,

K. Frydenvang, G. T. Swanson, J. S. Kastrup and M. Oikawa, *J. Med. Chem.*, 2013, **56**, 2283–2293.

- 13 M. Oikawa, Y. Kasori, L. Katayama, E. Murakami, Y. Oikawa and Y. Ishikawa, *Synthesis*, 2013, **45**, 3106–3117.
- 14 M. Chiba, Y. Ishikawa, R. Sakai and M. Oikawa, *ACS Comb. Sci.*, 2016, **18**, 399–404.
- 15 Racemate synthesis was studied in this study, however (2*R*)enantiomer has been shown to be neuronally active in IKM-159. See ref. 12.
- 16 O. Hlokoane, S. Tsukamoto, R. Irie and M. Oikawa, *Chem. Lett.*, 2021, **50**, 1464–1466.
- 17 M. Chiba, C. Fujimoto, R. Sakai and M. Oikawa, *Bioorg. Med. Chem. Lett.*, 2015, 25, 1869–1871.
- 18 J. C. Lee, Y. S. Oh, S. H. Cho and J. D. Lee, Org. Prep. Proced. Int., 1996, 28, 480–483.
- 19 S. B. Garber, J. S. Kingsbury, B. L. Gray and A. H. Hoveyda, *J. Am. Chem. Soc.*, 2000, **122**, 8168–8179.
- 20 Z. Galla, E. Forró and F. Fülöp, *Tetrahedron: Asymmetry*, 2016, 27, 729–731.
- 21 W. H. Hartung and R. Simonoff, Org. React., 1953, 7, 263-326.
- 22 In our preliminary experiments, this transformation can be realized also in the presence of other alcohols such as EtOH or BnOH instead of MeOH.
- 23 When monitoring the progress of the reaction $13 \rightarrow 2$ by LCMS and TLC, oxa-Michael product which is a protonated form for intermediated C (see Scheme 6) was detected as the reaction intermediate. Finally, the reaction product was converged to 2 *via* the oxa-Michael product, supporting the reaction is based on a stepwise mechanism we proposed in Scheme 6. This observation will be summarized and discussed in detail in a separate paper in the future.
- 24 T. K. M. Shing, Z.-H. Zhou and T. C. W. Mak, J. Chem. Soc., Perkin Trans. 1, 1992, 1907–1910.
- 25 H. Uchimasu, K. Matsumura, M. Tsuda, K. Kumagai, M. Akakabe, M. J. Fujita and R. Sakai, *Tetrahedron*, 2016, 72, 7185–7193.
- 26 C. J. Gerry and S. L. Schreiber, *Nat. Rev. Drug Discovery*, 2018, 17, 333–352.