#### **ORIGINAL COMMUNICATION**



# Quantitative <sup>1</sup>H and <sup>23</sup>Na muscle MRI in Facioscapulohumeral muscular dystrophy patients

Teresa Gerhalter<sup>1</sup> · Benjamin Marty<sup>2,3</sup> · Lena V. Gast<sup>1,6</sup> · Katharina Porzelt<sup>4</sup> · Rafael Heiss<sup>1</sup> · Michael Uder<sup>1</sup> · Stefan Schwab<sup>4</sup> · Pierre G. Carlier<sup>2,3</sup> · Armin M. Nagel<sup>1,5,6</sup> · Matthias Türk<sup>4</sup>

Received: 19 August 2020 / Revised: 29 September 2020 / Accepted: 1 October 2020 / Published online: 12 October 2020 © The Author(s) 2020

## Abstract

**Objective** Our aim was to assess the role of quantitative <sup>1</sup>H and <sup>23</sup>Na MRI methods in providing imaging biomarkers of disease activity and severity in patients with Facioscapulohumeral muscular dystrophy (FSHD).

**Methods** We imaged the lower leg muscles of 19 FSHD patients and 12 controls with a multimodal MRI protocol to obtain STIR- $T_2$ w images, fat fraction (FF), water  $T_2$  (w $T_2$ ), water  $T_1$  (w $T_1$ ), tissue sodium concentration (TSC), and intracellular-weighted sodium signal (inversion recovery (IR) and triple quantum filter (TQF) sequence). In addition, the FSHD patients underwent muscle strength testing.

**Results** Imaging biomarkers related with water mobility ( $wT_1$  and  $wT_2$ ) and ion homeostasis (TSC, IR, TQF) were increased in muscles of FSHD patients. Muscle groups with FF > 10% had higher  $wT_2$ ,  $wT_1$ , TSC, IR, and TQF values than muscles with FF < 10%. Muscles with FF < 10% resembled muscles of healthy controls for these MRI disease activity measures. However,  $wT_1$  was increased in few muscles without fat replacement. Furthermore, few STIR-negative muscles (n = 11/76) exhibited increased  $wT_1$ , TSC, IR or TQF. Increased  $wT_1$  as well as <sup>23</sup>Na signals were also present in muscles with normal  $wT_2$ . Muscle strength was related to the mean FF and all imaging biomarkers of tibialis anterior except  $wT_2$  were correlated with dorsal flexion.

**Conclusion** The newly evaluated imaging biomarkers related with water mobility  $(wT_1)$  and ion homeostasis (TSC, IR, TQF) showed different patterns compared to the established markers like FF in muscles of FSHD patients. These quantitative biomarkers could thus contain valuable complementary information for the early characterization of disease progression.

Keywords Facioscapulohumeral muscular dystrophy  $\cdot$  Quantitative MRI  $\cdot$  Sodium MRI  $\cdot$  Triple quantum filter  $\cdot$  Water T<sub>1</sub>

Ele arti sup	ctronic supplementary material The online version of this cle (https://doi.org/10.1007/s00415-020-10254-2) contains plementary material, which is available to authorized users.
	Teresa Gerhalter teresagerhalter@gmx.at
1	Institute of Radiology, University Hospital Erlangen, Friedrich-Alexander University Erlangen-Nuremberg (FAU), Maximiliansplatz 3, 91054 Erlangen, Germany
2	NMR Laboratory, Institute of Myology, Paris, France
3	NMR Laboratory, CEA/DRF, IBFJ/MIRCen, Paris, France
4	Department of Neurology, Friedrich-Alexander University Erlangen-Nuremberg (FAU), Erlangen, Germany
5	

- <sup>5</sup> Division of Medical Physics in Radiology, German Cancer Research Centre, Heidelberg, Germany
- <sup>6</sup> Institute of Medical Physics, Friedrich-Alexander University Erlangen-Nuremberg (FAU), Erlangen, Germany

# 

٩b	brev	<b>/iat</b> i	ions
----	------	---------------	------

FF	Fat fraction
FSHD	Facioscapulohumeral muscular dystrophy
GL	Gastrocnemius lateralis muscle
GM	Gastrocnemius medialis muscle
IQR	Interquartile range
IR	Inversion recovery
SOL	Soleus muscle
$STIR-T_2w$	Short $T_1$ inversion recovery $T_2$ -weighted
$wT_1$	Longitudinal relaxation time of water
wT <sub>2</sub>	Transverse relaxation time of water
TA	Tibialis anterior muscle
TP	Tibialis posterior muscle
TQF	Triple quantum filter
TSC	Tissue sodium concentration

#### Introduction

Facioscapulohumeral muscular dystrophy (FSHD) is characterized by a typically asymmetric progression of muscle weakness and wasting including muscular inflammation and fibrotic and fatty replacement [1]. Several clinical trials strive to develop treatment and therefore require reliable and accurate tools to monitor muscle response to interventions [2, 3].

Quantitative MRI allows the non-invasive assessment of the structure and function of individual muscles. The majority of MRI studies on muscle dystrophies have measured the fat fraction (FF) within the muscle [4]. However, clinical trials require biomarkers that detect subtle changes in muscle pathology prior to any irreversible fat replacement [5]. Some muscles without visible fatty replacement have shown  $T_2$ changes reflecting myocyte damage, edema, inflammation and/or hypervascularization [6, 7], and therefore reveal the initial stages of disease expression [8].

The longitudinal relaxation time of water (wT<sub>1</sub>) and <sup>23</sup>Na MRI have been used to study muscles of patients with other neuromuscular diseases [9–13]. Like transverse relaxation time (T<sub>2</sub>) quantification, water T<sub>1</sub> mapping enables the characterization of the muscle tissue at the microstructure level. Several studies have already endorsed T<sub>1</sub> mapping to quantify inflammation, necrosis or fibrosis in the myocardium [14].

In addition, the sodium distribution in the tissue as measured by <sup>23</sup>Na MRI is a sensitive marker of cell integrity and energy metabolism [15]. <sup>23</sup>Na MRI revealed an increase of tissue sodium concentration (TSC), which is a volume-weighted average of the intracellular and extracellular sodium concentrations, in the skeletal muscle tissue of Duchenne muscular dystrophy patients and in myotonic dystrophy patients [10–13]. Besides, a partial weighting towards the intracellular sodium can be achieved by relaxationweighted <sup>23</sup>Na MRI techniques such as inversion-recovery (IR) imaging [16-18] or triple-quantum filtered (TQF) imaging [19, 20]. Based on differences in  $T_1$  relaxation times, IR imaging can suppress signal from free sodium ions in fluid such as saline solution [17]. TQF imaging selectively detects signal from sodium ions in environments with restricted motion (e.g., caused by macromolecular interactions) [21]. However, also extracellular sodium ions can contribute to the TQF signal [22].

Quantitative MRI measures such as <sup>1</sup>H relaxometry and sodium content might therefore be promising early indicators of pathological changes in FSHD. To test this hypothesis, we sought to evaluate the sensitivity of commonly used biomarkers, such as FF, T<sub>2</sub>-weighting, and wT<sub>2</sub>, and novel MRI techniques, such as wT<sub>1</sub> and <sup>23</sup>Na MRI, to analyze pathological changes in patients with FSHD.

#### Methods

#### Study design and subject cohort

The study was approved by the local ethics committee (Number: 10 18 B), and written informed consent was obtained from all participants prior to examination. Between May 2018 and October 2019, we included 19 patients (age  $48.1 \pm 12.7$  years) with genetically confirmed FSHD. 12 ageand sex-matched volunteers (age  $46.6 \pm 14.2$  years) were recruited as controls. Exclusion criteria were contraindications for MRI (e.g., claustrophobia or metallic implants) and an age younger than 18. A neurologist scored manually the strength using the Medical Research Council grading system ranging from 0 (no muscle contraction) to 5 (maximal muscle strength) for the foot dorsal and plantar flexors [23]. MR data were then acquired from the less affected leg or leg without any confounding factor such as former vein thrombosis or lymphatic fistula. In patients with no difference in leg muscle strength, the right leg was scanned. In addition, patients provided a genetic test with the D4Z4 fragment length analyzed by EcoRI-digestion. The neurologist also collected clinically relevant data (self-reported age at disease onset, total disease duration, duration of muscle weakness of lower legs, and medication) by a personal interview and from available medical records.

#### **MRI** acquisitions

MRI was performed on a 3 T whole-body MR system (Magnetom Skyra, Siemens Healthineers, Erlangen, Germany). Total scan time was between 60 and 75 min, including  $B_0$  shimming and repositioning between the <sup>1</sup>H and <sup>23</sup>Na parts of the protocol.

A 15-channel quadrature knee coil (Siemens Healthcare, Erlangen, Germany) was used for <sup>1</sup>H MRI. The center of the coil was positioned at the biggest circumference of the lower leg. This position was marked to ensure the same positioning for <sup>23</sup>Na MRI, which was peformed during the second part of the protocol. FF was measured using a 3D 3-point Dixon method (three echo times, 2.75/3.95/5.15 ms; repetition time, 10 ms; flip angle, 3°; 64 slices of 5 mm thickness and with a  $1.3 \times 1.3$  mm<sup>2</sup> resolution; total acquisition time, 3 min 12 s) [24]. From the same position, short  $T_1$  inversion recovery  $T_2$ -weighted (STIR- $T_2$ w) images were acquired with an inversion time of 220 ms to null the fat signal (echo time, 69 ms; repetition time, 6.9 s; flip angle,  $145^{\circ}$ ; 23 slices with 5 mm thickness and  $1 \times 1$  mm<sup>2</sup> resolution; acquisition time, 3 min 29 s) and to depict possible edematous/inflammatory/ nectrotic processes within muscle tissue. A multi-spin-echo sequence was acquired with 32 echoes (echo times ranging from 9.5 ms to 304 ms) to determine the  $wT_2$  (repetition

time, 3 s; 5 slices with a  $1.4 \times 1.4 \text{ mm}^2$  resolution and 10 mm thickness; acquisition time, 3 min 41 s). We measured wT<sub>1</sub> with a MR fingerprinting sequence allowing a drastic reduction of acquisition time (train of 1400 spokes; 3 slices with a  $1 \times 1 \text{ mm}^2$  resolution and 10 mm thickness; acquisition time, 30 s) [9].

Sodium imaging was performed using a single-resonant (<sup>23</sup>Na) birdcage knee radiofrequency coil (Stark Contrast, Erlangen, Germany) and NaCl reference tubes for <sup>23</sup>Na signal quantification (20 mM and 40 mM NaCl without and with 5% agarose). Axial <sup>1</sup>H images were acquired with the body radiofrequency coil using a gradient-echo sequence to provide anatomical reference for the sodium images and a global flip angle calibration and a manual B<sub>0</sub> shim using the <sup>1</sup>H MRI-based B<sub>0</sub>-shimming routine of the manufacturer were performed [13]. The <sup>23</sup>Na imaging part measured the TSC and intracellular-weighted sodium signal using two relaxation-weighted <sup>23</sup>Na MRI techniques (IR and TQF imaging). All <sup>23</sup>Na images were acquired using a density-adapted 3D-radial readout scheme with anisotropic resolution [25]. For TSC assessment, we obtained spin density-weighted <sup>23</sup>Na images using an echo time of 0.3 ms (repetition time, 120 ms; 5384 projections with 384 radial samples; flip angle,  $80^{\circ}$ ; nominal resolution,  $3 \times 3 \times 15$  mm<sup>3</sup>; acquisition time, 10 min 46 s). IR images were acquired with an inversion time of 34 ms to reduce <sup>23</sup>Na signal originating from free sodium ions (nominal resolution,  $4 \times 4 \times 20$ mm<sup>3</sup>; 4760 projections with 384 radial samples; inversion flip angle, 180°; acquisition time, 9 min 50 s) [18]. For 15 FSHD patients and 7 healthy subjects, the <sup>23</sup>Na protocol was extended with a TQF sequence [26], with the preparation time set to 4 ms as published in [27] (1056 projections with 384 radial samples; zero-filled matrix size,  $60 \times 60 \times 20$ ; nominal spatial resolution,  $4 \times 4 \times 12$  mm<sup>3</sup>; acquisition time, 12 min 40 s).

#### **Data analysis**

For the semi-qualitative analysis, a radiologist with 6 years of experience in reading musculoskeletal MRIs evaluated eight different lower leg muscles: tibialis anterior, extensor halluces/digitorum, peroneus, tibialis posterior, gastrocnemius medialis, gastrocnemius lateralis, soleus, and flexor halluces/digitorum longus. The fat replacement of these muscles (fat score) was scored with an established five-point scale ranging from 0 (not-fat replaced) to 4 (completely fat replaced) by Goutallier et al. using the first echo of the multi-spin-echo sequence [28]. Similarly, STIR-T<sub>2</sub>w intensity was rated on a five-point scale from 0, normal appearance, to 4, severe involvement of entire muscle, based on the scale described by Wang et al. [29]. All imaging slices of each muscle were considered for the grading.

Images for fat fraction and  $wT_2$  measurement were processed with custom written Python scripts.  $wT_2$  values were calculated based on a tri-exponential fitting procedure to account for fat replacment [30]. The sodium and  $wT_1$  data sets were reconstructed offline with custom written MAT-LAB scripts (MathWorks, Natick, MA, USA). The signal evolution from the MR fingerprinting were matched to a two-component model for water-fat separation and to obtain  $wT_1$  values [9]. Sodium signals were calibrated using the standard deviation of the background noise and the mean signal of two agarose phantoms (20 and 40 mM NaCl in 5% agarose). ROIs were delineated on the middle slice of the multi-spin-echo images and on the middle slice of the <sup>1</sup>H anatomical reference image for the <sup>23</sup>Na part. All ROIs were outlined conservatively within the muscle.

As fat tissue has lower sodium concentrations than muscle tissue [13, 31], we performed a fat correction for the <sup>23</sup>Na measures to consider partial volume effects of fat replaced muscles as previously described in [13]. Briefly, the estimates of TSC, IR, and TQF in muscle tissue were corrected for the non-negligible sodium signal of fat using the sodium signal of fat and FF of the muscle obtained by the Dixon acquisitions. In the following, the fat-corrected values are presented as TSC, IR, and TQF, respectively. The unit of TSC will be given in mM and refers to the sodium content in mmol per volume of tissue. In addition, the fat-corrected ratios were calculated and will be presented as IR/TSC and TQF/TSC, respectively.

For further analysis, the individual muscles of the FSHD patients were divided into three groups according to their individual intramuscular FF (low: FF < 10%, medium: 10% < FF < 50%, and high: FF > 50%). To analyse the presence of damage, the positive STIR-T<sub>2</sub>w muscles were grouped according to their signal intensity (low: STIR 1, medium: STIR 2, and high: STIR 3 and 4) [29].

#### **Statistical analysis**

Statistical analysis was performed using MATLAB (Math-Works, Natick, MA, USA) and descriptive statistics including boxplots of the average MRI metrics for every ROI were used to evaluate each outcome. Thresholds for aberrant values of any outcome measure were defined as the mean of the control cohort plus two standard deviations. Nonparametric statistics were used as not all the measures from the patient group were normally distributed as assessed by the Lilliefors test. Wilcoxon rank-sum tests compared all MRI parameters for each muscle between dystrophic and control groups. Relationships of MRI parameters with clinical measures of strength were determined using Spearman correlation coefficients for ordinal or non-normally distributed data. False discovery rate (FDR) correction was applied for multiple comparisons [32]. A multiple linear regression model investigated the association between age, duration of leg muscle weakness, and D4Z4 fragment length (independent variables) and the mean FF (dependent variable). The square root transformation was used for variance-stabilization of the mean FF. A p value < 0.05 was considered significant for all statistical tests.

## Results

#### Clinical characteristics of the study cohort

Nineteen FSHD patients (all FSHD1, mean age,  $48.1 \pm 12.7$  years; age range, 25–67 years; 4 females and 15 males; median disease duration, 19.5 years; range, 2 to > 50 years) and 12 healthy volunteers (mean age,  $46.6 \pm 14.2$  years; age range 25–70 years; 4 females and 8 males) were enrolled for this prospective MRI study. Detailed patient demographic and clinical characteristics are compiled in Table 1. All patients were ambulant at least for short distances (more than 10 m). According to the Medical Research Council score, strength of the ankle dorsal flexors and ankle plantar flexors ranged from 1 to 5 and 3 to 5, respectively. In seven patients, both ankle dorsal flexors and ankle plantar flexors showed normal strength. The mean D4Z4 fragment length was  $28 \pm 7$  (range 17–38) for 18 patients. In patient 15, D4Z4-repeat contraction was not proven in the patient but in his child. However, limited genetic testing confirmed moderate hypomethylation of the distal 4qA-D4Z4-repeat in the patient himself matching the diagnosis of FSHD. Information on their medication are provided in the Supplemental data (Table e-1).

#### **Qualitative MRI changes**

The semi-quantitative scoring revealed that 12 FSHD patients had at least one fat-replaced muscle (Table e-2 in supplemental data). The median fat score of all muscles and patients was 0 [Interquartile range (IQR) 0–3] with a median of 3 affected muscles (IQR 0–6). The gastrocnemius medialis (GM, n = 11/19, median fat score 3 [IQR 0–4]) and tibialis anterior (TA, n = 10/19, median fat score 2 [IQR 0–4]) were the most affected muscles, whereas the soleus [SOL, n = 8/19, median fat score 0 (IQR 0–3)] and gastrocnemius lateralis [GL n = 6/19, median fat score 0 (IQR 0–1)] were less often affected by fat replacement and the tibialis posterior (TP) was only mildly fat-replaced in two patients.

STIR-T<sub>2</sub>w imaging showed that 13 out of 18 FSHD patients had at least 1 muscle with hyperintensities. For one patient, no STIR images were acquired due to time constraints. The median STIR score of all muscles and patients was 0 (IQR 0–1) with 2 (IQR 0–4.3) number of STIR-positive muscles. Overall, the GL (STIR 0: n = 6/18,

STIR 1: n = 8/18, STIR 2: n = 4/18, STIR 3: n = 0/18, STIR 4: n = 0/18) was the most frequent STIR-positive muscle, followed by GM (STIR 0: n = 8/18, STIR 1: n = 5/18, STIR 2: n = 4/18, STIR 3: n = 1/18, STIR 4: n = 0/18) and TA (STIR 0: n = 11/18, STIR 1: n = 1/18, STIR 2: n = 2/18, STIR 3: n = 4/18, STIR 4: n = 0/18). Five patients had a STIR-positive SOL (STIR 0: n = 13/18, STIR 1: n = 2/18, STIR 2: n = 2/18, STIR 3: n = 0/18). Five patients had a STIR-positive SOL (STIR 0: n = 13/18, STIR 1: n = 2/18, STIR 2: n = 2/18, STIR 3: n = 0/18, STIR 4: n = 1/18), while no patient had a STIR-positive TP. 12% of these STIR-positive muscles (n = 19/162) had no fat replacement (fat score 0). While all muscles of the healthy controls had a fat score of 0, some muscles had a STIR score of 1 (Peroneus n = 2/12, GM = 11/12, GL = 11/12, see Table e-3 in supplemental data).

Following the qualitative analysis, we chose to investigate quantitatively muscles that were more affected (GM, TA), moderately affected (SOL), and almost spared (TP) and were big enough for the ROI analysis of <sup>23</sup>Na MRI.

# Impact of fat on <sup>23</sup>Na signals

In completely fat-replaced muscles, the sodium signal intensities were close to the signal intensities of subcutaneous fat tissue (Fig. e-1 in supplemental data). Quantification of the sodium signals in the fat-replaced GM muscle of the 59-year-old patient (#6) resulted in 11.2 mM TSC, 0.26 a.u. IR signal, and 0.24 a.u. TQF signal, while the subcutaneous fat yielded 10.6 mM TSC, 0.21 a.u. IR signal, and 0.15 a.u. TQF signal. The sodium values were then corrected for the fat replacement as previously published (TSC<sub>fat</sub>= $7.0 \pm 1.0$  mM, IR<sub>fat</sub>= $0.10 \pm 0.04$  a.u., and TQF<sub>fat</sub>= $0.17 \pm 0.01$  a.u.) [13].

# Quantitative <sup>1</sup>H and <sup>23</sup>Na analysis

Examples of STIR-T<sub>2</sub>w images with the corresponding FF, wT<sub>2</sub>, wT<sub>1</sub>, and sodium maps of one young healthy volunteer in comparison with three FSHD patients of 25, 46, and 58 years of age are shown in Fig. 1. Imaging of the lower leg of the 25-year-old patient (disease onset before school, but no weakness of the legs) showed only two mildly STIR-positive muscles and no increased quantitative MRI measure. In contrast, fat replacement as well as edema-like changes, increased  $wT_2$ ,  $wT_1$ , and sodium signals were present in the 46- and 58-year-old FSHD patients. Both patients exhibit several STIR-positive muscles and increased quantitative indices despite the difference in reported disease duration (32 years and 5 years, respectively) and duration of leg weakness (10 years and 5 years, respectively). Furthermore, the TQF maps showed a different contrast in some muscle compartments compared to IR. Nevertheless, the <sup>23</sup>Na signal originating from the vessels and saline phantoms was

FSHD patient	Age (years)	Sex	Disease dura- tion (years)	Fragment length by EcoR1-digestion (kb)	Side of examined leg	Duration of lower leg weakness (years)	Dorsal flexion	Plantar flexion	Fat-replaced muscles	Positive STIR- T <sub>2</sub> w muscles
_	66	М	40	38	R	40	4	5	8	0
2	46	Μ	32	19	L	10	2	3	5	5
3	31	М	9–13	31	L	0	5	5	0	2
4	56	М	16	35 and 32	R	10	2–3	5	4	4
5	65	Μ	10	27	R	10	2	3	8	6
6	59	ц	> 50	24	R	20	1	4	6	1
7	49	Μ	13	24	L	8	4	5	2	1
8	35	Μ	21	33	R	0	5	5	0	2
6	58	ц	18	36	L	0	5	5	0	0
10	49	Μ	39	37	L	11	2	5	ĸ	4
11	58	Μ	5	38	R	5	2	4	4	3
12	67	Μ	> 50	33	R	40	2	3-4	6	0
13	31	ц	21–25	17	R	0	5	5	0	0
14	25	ц	15-19	20	L	0	5	5	0	2
15	61	Μ	2	n/a; *	L	0	5	5	6	5
16	33	М	8	22	R	0	5	5	0	0
17	46	М	30	21	R	30	4	4	n	4
18	45	М	5-30	33	L	2	3	4	0	2
19	55	Μ	25	20	L	6	2	5	3	5
MRI and clinical	l data were acqu	ired fron	1 the less affected	leg (side of examined leg	). The duration	of muscle weakness for t	he examined leg is	also listed		

Table 1 Patients demographics, clinical data, and qualitative MRI findings of Facioscapulohumeral muscular dystrophy (FSHD) cohort

 $\underline{\textcircled{O}}$  Springer

\*Moderate hypomethylation of distal 4qA-D4Z4-repeat

R right leg, L left leg, n/a not available



**Fig. 1** Examples of acquired MR images and maps.  $T_2$ -weighted STIR images are plotted with the corresponding fat fraction, water  $T_2$ , water  $T_1$ , and tissue sodium concentration (TSC), inversion-recovery (IR), and triple-quantum filter (TQF) signal maps in a 30-year-old healthy volunteer (top row) and three Facioscapulohumeral muscular dystrophy (FSHD) patients (2nd–4th row). Note that the 25-year-old patient (#14) presented normal appearing quantitative MRI maps, and only the GM and GL on the STIR image were rated as a one (indicated by orange arrow). On the other hand, fat replacement, moderate to severe edema-like changes, and increased sodium signals were

successfully suppressed with both intracellular-weighted sequences.

For FSHD patients, the fat fraction as assessed in all analyzed 76 muscles by the Dixon method was significantly increased in the GM, SOL, and TA muscles (total n = 36/76 muscles) in contrast to the wT<sub>2</sub> values that remained in the normal range in most of the investigated muscles (Fig. 2). However, several individual muscles of patients showed elevated wT<sub>1</sub> (n = 20/63), wT2 (n = 14/74), TSC (n = 21/72), IR (n = 21/72), and TQF (n = 26/60) values compared to controls. Significant increases were observed for the wT<sub>1</sub> in the GM and TA and the TQF signal in the TA and a significant drop in the IR of the TP and the IR/TSC ratio of the TA. The descriptive statistics of all quantitative MRI parameters are compiled in the Supplemental Material (Table e-4).

To further investigate the relation of fat replacement and STIR scaling with the quantitative indices, we grouped the four muscles (TA, TP, GM, and SOL) in normal appearing,

present in selective muscles of the older FSHD patients (disease duration 32 years and 5 years, respectively). While in the 46-year-old patient (#2), the sodium alterations coincided with muscular edema, TQF signal increases without visual edema occurred in the GM of the 58-year-old patient (#11) (see white arrow). Some areas with increased higher TQF signal appeared normal or showed only slightly increased signal intensities in the IR maps. With both methods, the IR and TQF, the <sup>23</sup>Na signal from vessels and the saline solution phantoms was suppressed sufficiently

mildly, moderately, and severely affected. The alterations of wT<sub>2</sub>, wT<sub>1</sub>, and the different sodium indices according to the intramuscular fat fraction and STIR scores are plotted in Fig. 3. Muscles with FF < 10% resembled muscles of healthy controls for almost all disease activity indices. Only few muscles with FF < 10% had increased  $wT_1$  and wT<sub>2</sub> as compared to controls. The TSC and IR did drop in muscles with low FF (both p < 0.05). In contrary, muscles with FF > 10% had significant higher wT<sub>1</sub>, TSC, IR, and TQF values than muscles of the control cohort (all p < 0.05). While the IR/TSC ratio dropped for moderately fat-replaced muscles (p < 0.05), the TQF/TSC ratio increased for those muscles (p < 0.05). STIR-positive muscles showed changes compared to controls for all quantitative disease activity indices, especially in STIR > 1 muscle (STIR 2 and STIR 3 and 4: wT<sub>1</sub>, wT<sub>2</sub>, TSC, TQF, IR/TSC: *p* < 0.05; STIR 3&4: IR: *p* < 0.05; STIR 2: TQF/TSC: *p* < 0.05). Furthermore, some STIR-negative muscles exhibited increases for  $wT_1$ ,



**Fig. 2** Boxplots showing quantitative <sup>1</sup>H and <sup>23</sup>Na MRI measures of analyzed muscles. Fat fraction (FF), water  $T_1$  and  $T_2$ , and <sup>23</sup>Na indices of individual muscles in patients with FSHD (orange) and healthy volunteers (blue). The dystrophic muscles were characterized by fatty replacement as well as increased water  $T_2$ , water  $T_1$  and sodium signals compared to the controls. The decrease in the IR/TSC ratio was significant in the tibialis anterior (TA). In the patient group, the gastrocnemius medialis (GM), soleus (SOL), and TA muscles were more

affected than the tibialis posterior (TP) muscle. The middle line in the box depicts the median value and the top and bottom edges of the box the 25th and 75th percentiles of the data, respectively. The whiskers extend to the most extreme data points not considering outliers. *TSC* tissue sodium concentration, *IR* inversion-recovery signal, *TQF* triple-quantum filter signal, \*p < 0.05 difference between control and FSHD

wT<sub>2</sub>, and all <sup>23</sup>Na indices compared to controls. Abnormal wT<sub>1</sub>, wT<sub>2</sub>, and <sup>23</sup>Na indices had an increased STIR score only when FF was low or moderate, since only three muscles had a STIR score > 0 while FF > 50%.

## Correlation of sodium homeostasis and water relaxometry

The different disease activity indices were correlated to analyze their relationship with each other (Fig. 4). wT<sub>1</sub> showed stronger correlation with TSC (R=0.55, p < 0.01) than with wT<sub>2</sub> (R=0.43, p < 0.01). wT<sub>2</sub> correlated the most with TSC (R=0.54, p < 0.01), before TQF (R=0.31, p < 0.01) and IR (R=0.23, p=0.02). Also, elevated wT<sub>1</sub>, TSC, IR, and TQF signals could occur at normal wT<sub>2</sub>. These alterations at normal wT<sub>2</sub> occurred in all four muscles, while the GM was the most prominent muscle to have increased wT<sub>1</sub> or <sup>23</sup>Na indices. These observations were not identical with a positive STIR-T<sub>2</sub>w muscle; eight muscles had at least one increased index at normal wT<sub>2</sub>. Also, none of these muscles belonged

to a clinical positive patient with a weakness of the lower leg. Furthermore, the TSC signal showed the strongest correlation with the IR (R=0.82, p<0.01) followed by the correlation with the TQF signal (R=0.54, p<0.01), whereas the TQF correlated less with the IR (R=0.50, p<0.01). Multiple comparison correction did not change the significance level of the correlations. Correlations and corrected p values are compiled in the Supplemental Material (Table e-5).

#### Association of MRI data and clinical measures

In addition, we aimed to describe the concordance between MRI measures and assessment of muscle function and clinical data. Our multiple linear regression model, which included duration of leg muscle weakness, age, and fragment length as independent variables (n = 18 FSHD patients), showed that age and duration of leg weakness had a significant influence (coefficient 0.010 with p < 0.001 and 0.008 with p = 0.002, respectively) on the square root transformed average FF. An influence of the fragment length on the fat fraction was also suggested (coefficient – 0.006 with p = 0.088). Our model explained



**Fig. 3** Analysis of muscles grouped according of their FF and STIR score. <sup>1</sup>H and <sup>23</sup>Na MRI indices in the leg muscle of FSHD patients (orange-brown) and controls (CTR, blue). The middle line in the box depicts the median value and the top and bottom edges of the box the 25th and 75th percentiles of the data, respectively. The whiskers extend to the most extreme data points not considering outliers. In the top row, the muscles of FSHD patients were gathered in three groups according to their intramuscular fat fraction (low FF: FF<10%, med. FF: 10% <FF<50%, and high FF: FF>50%). Muscles with FF<10% resembled muscles of healthy controls for all disease activ-

ity indices except the TSC and IR. In the bottom row, the muscles of FSHD patients were grouped according to the STIR- $T_2w$  signal intensity (STIR 0: normal appearance, STIR 1: very mild hyperintensities, STIR 2: mild diffuse/some moderate elevation, and STIR 3 and 4: moderate/severe involvement). Few negative STIR- $T_2w$  muscles (STIR 0) had increased disease activity indices. Both groups with FF > 10% had higher water  $T_2$ , water  $T_1$ , and sodium values than muscles with FF < 10%. *TSC* tissue sodium concentration, *IR* inversion-recovery signal, *TQF* triple-quantum filter signal, \*p < 0.05

84% of the variability of the FF (adjusted  $R^2 = 0.835$ ). The muscle strength was related to the mean FF of the GM and SOL for the plantar flexion (R = -0.52, p = 0.02) and the FF of TA for the dorsal flexion (R = -0.79, p < 0.01) (Fig. e-2 in supplemental data). Furthermore, all disease indices of TA except wT<sub>2</sub> were correlated with dorsal flexor strength (wT<sub>1</sub> R = -0.67, p < 0.01; wT<sub>2</sub> R = -0.25, p = 0.32; TSC R = -0.75, p < 0.01; IR R = -0.69, p = 0.02; TQF R = -0.59, p = 0.03).

Finally, seven patients showed no decline in strength (dorsal flexion 5 and plantar flexion 5). However, imaging of one of these patients (61 years, #15) revealed that the GM and SOL are completely or partly replaced by fat. Moreover, the part of the SOL that was spared from fat replacement was severely affected by edema with alterations in all quantitative disease activity indices (Fig. e-3 in supplemental data). One patient (25 years, #14) without strength decline exhibited increased TQF/TSC ratio in the TP and TA. The rest of this subgroup did not show any abnormal quantitative measures. Conversely, another patient (45 years, #18) had decreased muscle strength (dorsal flexion 3 and plantar flexion 4) while the muscles were not replaced by fat and only the TA showed an increased TQF.

# Discussion

In this study, we systematically characterized the status of calf muscle tissue in FSHD patients using different quantitative MRI techniques. To do so, we combined different <sup>1</sup>H and <sup>23</sup>Na MRI methods to assess fat replacement, STIRpositive, and normal appearing muscles in patients with FSHD. For the first time, <sup>23</sup>Na MRI as well as wT<sub>1</sub> measurements were reported for FSHD revealing the presence of sodium and wT<sub>1</sub> anomalies in skeletal muscle tissue of FSHD patients.

Involvement of lower leg muscles in patients with FSHD, especially of the GM and TA, is a typical clinical and MRI finding and has also been observed in our study [33–36]. We showed as previously reported that fatty replacement of the muscle increases with age and with disease duration and correlates inversely with muscle flexor strength [37]. While



**Fig. 4** Relationship of water  $T_1$ , water  $T_2$ , and <sup>23</sup>Na signals. The dashed lines indicate the mean plus two standard deviations of the control values. Aberrant water  $T_1$ , tissue sodium concentration (TSC), inversion-recovery (IR), and triple-quantum filter (TQF) signal could occur also in "quiescent" muscles characterized by normal water  $T_2$  values. GM was the muscle with most frequent aberrant water  $T_1$ 

(n=4), TSC (n=5), IR (n=6), and TQF (n=6) at normal water T<sub>2</sub>, followed by TA (T<sub>1</sub>/TSC/IR/TQF n=3/2/2/3), Sol (T<sub>1</sub>/TSC/IR/TQF n=0/1/3/2), and TP (T<sub>1</sub>/TSC/IR/TQF n=1/1/1/1). TSC correlated better with IR than with TQF, whereas IR did not correlate that well with TQF. *GM* gastrocnemius medialis, *Sol* soleus, *TA* tibialis anterior, *TP* tibialis posterior

the TA is the sole muscle involved in the dorsal flexion, the plantar flexor comprises the GM, GL, and SOL. The involvement of three muscles could allow a compensation of an affected muscle by the others and thus explains the observed weaker relation with the measured plantar flexor strength than with the dorsal flexor in our study. Furthermore, for two patients, muscle strength was not in accordance with fat replacement.

The muscles of FSHD patients were mostly either highor low-fat replaced; few were moderately fat replaced (10% < FF < 50%). In a longitudinal study, FF quantification detected muscle changes in FSHD patients without any loss of muscle function [7]. Since fat measurement (qualitative or quantitative) are capturing the muscle wasting too late, scientists are seeking for other earlier markers before clinical involvement of the leg muscles. In concordance with other studies, STIR-T<sub>2</sub>w lesions were present in some muscles before significant fat replacement [6, 38]. Previously, the presence of these hyperintense lesions was linked to increased fat deposition at follow-up [38]. In line, our FSHD cohort showed several muscles with STIR-T<sub>2</sub>w hyperintensities, however also almost all controls had at least one STIR-positive calf muscle providing false-positive findings and thus suggesting a hypersensitivity of STIR-T<sub>2</sub>w imaging. Thus, we wanted to provide quantitative markers of active disease injury by assessing the relaxation times of the muscles. Isolated muscles in our cohort showed increased wT<sub>1</sub> and wT<sub>2</sub> values without significant fat replacement. Furthermore, these quantitative variables were also increased in STIR-negative muscles. The sensitivity of the relaxation times to the underlying disease activity seems to differ from each other, as some muscles had increased  $wT_1$  values at normal  $wT_2$  values.

<sup>23</sup>Na MRI was able to detect sodium anomalies in skeletal muscle tissue of FSHD patients, but the cause of changes in sodium signals is not fully clear. Water relaxometry was also able to detect changes related to the water mobility in the present FSHD cohort. For the TSC as for  $wT_1$  and  $wT_2$ , the extracellular and intracellular compartments contribute both to the signal, and derangements such as edema, fibrosis, inflammation, or membrane leakiness could contribute to altered signal. Edema as observed on the STIR-T<sub>2</sub>w images concurrently occurred with higher TSC, wT<sub>1</sub>, and wT<sub>2</sub> values. Although all three indices are non-specific and reflect inflammation, edema, necrosis, or fibrotic changes, they could show different behavior in the same muscle. While wT<sub>2</sub> signal as extracted from the T<sub>2</sub> signal decay represents essentially the intracellular wT2, TSC and wT1 likely capture more the vascular compartment. Moreover, corticosteroid treatment has been shown to decrease the wT<sub>2</sub> values at least in Duchenne muscular dystrophy patients [39]. Similar effects from the patients' medication could have influenced water relaxation times and <sup>23</sup>Na homeostasis in our study; nevertheless, medical histories did not reveal any medication obviously affecting the sodium or wT<sub>1</sub> imaging.

To better understand the origin of sodium signal changes, we measured the intracellular-weighted sodium signal by two different MRI methods. First, the IR sequence, which is the more established method, reduced the <sup>23</sup>Na signal from extracellular edema and blood vessels exploiting the differences in  $T_1$  of sodium [18, 40]. Secondly, TQF imaging suffers from a poorer signal-to-noise ratio and detects only signal from sodium ions experiencing slow isotropic or anisotropic motion [41]. Nevertheless, both methods can only achieve a weighting towards the intracellular compartment [13, 42–44]. Irrespective of the technique, our FSHD patients showed high intracellular-weighted signals in some muscle compartments, thus pointing to a role of increased intracellular sodium in FSHD especially amid muscle wasting (10% < FF < 50%). Since IR and TQF are more weighted to the intracellular compartment while TSC is more related to the vascular compartment, we expected the IR and TQF to be closely correlated with each other and less correlated with TSC. However, the correlation between IR and TQF was the weakest of all and IR was very strongly correlated to TSC. This observation could be related to the mechanism of the different techniques. While IR is optimized to suppress signal originating from sodium ions in free solution such as saline solution, the TQF sequence filters the signal of nuclei (ions) involved in slow molecular motion. The latter pulse sequence probably shows a stronger dependency on the strength of the motion restriction (i.e., change of correlation time) [45]. Changes in the environment such as myocyte damage or fibrosis could change the interactions of sodium with the environment. However, the signal behavior of TQF imaging is still subject of current research.

Muscles of FSHD patients without fat replacement (FF < 10%) had normal sodium levels, in contrast what has been reported for Duchenne patients [13]. In addition, wT<sub>1</sub> and wT<sub>2</sub> values were in the normal range for most FF < 10%muscles. This might be explained by the muscle pathology and the disease course of FSHD. A normal-appearing muscle of a Duchenne muscular dystrophy patient is undergoing waves of necrosis and regeneration while a non-fatty replaced FSHD muscle might be even close to normal. In Duchenne muscular dystrophy, the affected dystrophin complex interacts also with the sodium channel, which could also contribute to the difference in sodium imaging between normal-appearing muscles of a Duchenne patient and FSHD muscles. Since most participating FSHD patients were already in a more advanced stage of the disease, the affected muscles were already replaced at least partly by fatty tissue. A 4-month follow-up MRI study has suggested that fat replacement occurs first distally in the muscle with altered metabolic profile, but then rapidly progresses over the whole muscle [37]. This might create a challenge to detect the starting point of the dystrophic changes.

This report presents the initial results of our FSHD study; we showed that sodium anomalies and <sup>1</sup>H relaxation time differences are present in FSHD patients and can be measured in vivo on a clinical 3 T scanner. Though other muscles are also involved, we only examined the lower leg due to the geometry of our <sup>23</sup>Na coil. Time constraints prevented us from acquiring TQF and STIR-T<sub>2</sub>w images from all subjects. Thus, the number of examined muscles differ between MRI measures to due missing data points (TQF and wT<sub>1</sub> not acquired for all patients, sodium acquisition failed once due to problems with coil) or because the fitting procedure failed due to high fat infiltration (wT<sub>2</sub>). Furthermore, results regarding disease duration should be interpreted with caution since the retrospective determination of the onset presented a challenge despite medical records and self-reports.

## Conclusion

These results demonstrated that  $wT_1$  and sodium alterations are present in the muscles of FSHD patients compared with healthy controls. Specifically, fat replaced muscles showed changes in  $wT_1$ , TSC, and intracellular-weighted <sup>23</sup>Na signals. Further work is needed to investigate the time scale of these disturbances in FSHD patients and the potential of quantitative <sup>1</sup>H and <sup>23</sup>Na MRI as sensitive biomarkers for intervention. Nevertheless, our study shows that quantitative MRI could be more accurate than visual scores for the status of FSHD, which might be a promising tool for further interventional studies. **Acknowledgement** This project was supported by the Johannes and Frieda Marohn Foundation (to A.N., 2016). We thank the Imaging Science Institute (Erlangen, Germany) for providing us with measurement time at the 3T MRI system. We are also grateful to the volunteers, as well as the patients and their families, for their participation.

Author contributions Design and conceptualized study: TG, PC, AN, MT; acquisition of data: TG, LG, KP, MT; formal analysis and investigation: TG, BM, RH, AN, MT; writing—original draft preparation: TG; writing—review and editing: all authors; funding acquisition: AN; resources: MU, SS. All authors read and approved the final manuscript.

**Funding** Open Access funding enabled and organized by Projekt DEAL. This project was supported by the Johannes and Frieda Marohn Foundation (to A.N., 2016).

## **Compliance with ethical standards**

**Conflicts of interest** The authors declare that they have no conflict of interest.

Ethics approval This human study has been approved by the local ethics committee (Number: 10\_18 B), and have therefore been performed in accordance with the ethical standards laid down in the 1964 Declaration of Helsinki and its later amendments.

**Consent to participate** Written informed consent was obtained from all participants prior to examination.

Consent for publication Not applicable.

Availability of data and material Anonymized data can be made available to qualified investigators upon request to the corresponding author.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

# References

- Sacconi S, Salviati L, Desnuelle C (2015) Facioscapulohumeral muscular dystrophy. Biochim Biophys Acta Mol Basis Dis 1852:607–614. https://doi.org/10.1016/j.bbadis.2014.05.021
- Hamel J, Tawil R (2018) Facioscapulohumeral muscular dystrophy: update on pathogenesis and future treatments. Neurotherapeutics 15:863–871. https://doi.org/10.1007/s13311-018-00675 -3
- 3. Tawil R, Padberg GW, Shaw DW et al (2016) Clinical trial preparedness in facioscapulohumeral muscular dystrophy: Clinical, tissue, and imaging outcome measures 29–30 May 2015,

Rochester, New York. Neuromuscul Disord 26:181–186. https:// doi.org/10.1016/j.nmd.2015.10.005

- Carlier PG, Marty B, Scheidegger O et al (2016) Skeletal muscle quantitative nuclear magnetic resonance imaging and spectroscopy as an outcome measure for clinical trials. J Neuromuscul Dis 3:1–28. https://doi.org/10.3233/JND-160145
- Strijkers GJ, Araujo ECA, Azzabou N et al (2019) Exploration of new contrasts, targets, and mr imaging and spectroscopy techniques for neuromuscular disease – a workshop report of working group 3 of the biomedicine and molecular biosciences COST action BM1304 MYO-MRI. J Neuromuscul Dis 6:1–30. https:// doi.org/10.3233/JND-180333
- Friedman SD, Poliachik SL, Carter GT et al (2012) The magnetic resonance imaging spectrum of facioscapulohumeral muscular dystrophy. Muscle Nerve 45:500–506. https://doi.org/10.1002/ mus.22342
- Mul K, Vincenten SCC, Voermans NC et al (2017) Adding quantitative muscle MRI to the FSHD clinical trial toolbox. Neurology 89:2057–2065. https://doi.org/10.1212/WNL.000000000004647
- Dahlqvist JR, Andersen G, Khawajazada T et al (2019) Relationship between muscle inflammation and fat replacement assessed by MRI in facioscapulohumeral muscular dystrophy. J Neurol 266:1127–1135. https://doi.org/10.1007/s00415-019-09242-y
- Marty B, Carlier PG (2020) MR fingerprinting for water T1 and fat fraction quantification in fat infiltrated skeletal muscles. Magn Reson Med 83:621–634. https://doi.org/10.1002/mrm.27960
- Weber M-A, Nagel AM, Wolf MB et al (2012) Permanent muscular sodium overload and persistent muscle edema in Duchenne muscular dystrophy: a possible contributor of progressive muscle degeneration. J Neurol 259:2385–2392. https://doi.org/10.1007/ s00415-012-6512-8
- Weber M-A, Nagel AM, Jurkat-Rott K, Lehmann-Horn F (2011) Sodium (23Na) MRI detects elevated muscular sodium concentration in Duchenne muscular dystrophy. Neurology 77:2017–2024. https://doi.org/10.1212/WNL.0b013e31823b9c78
- Constantinides CD, Gillen JS, Boada FE et al (2000) Human skeletal muscle: sodium MR imaging and quantification-potential applications in exercise and disease. Radiology 216:559–568. https://doi.org/10.1148/radiology.216.2.r00j146559
- Gerhalter T, Gast LV, Marty B et al (2019) 23 Na MRI depicts early changes in ion homeostasis in skeletal muscle tissue of patients with duchenne muscular dystrophy. J Magn Reson Imaging 50:1103–1113. https://doi.org/10.1002/jmri.26681
- Germain P, El GS, Jeung M-Y et al (2014) Native T1 mapping of the heart – a pictorial review. Clin Med Insights Cardiol 84:S19005. https://doi.org/10.4137/CMC.S19005
- Madelin G, Regatte RR (2013) Biomedical applications of sodium MRI in vivo. J Magn Reson Imaging 38:511–529. https://doi. org/10.1002/jmri.24168
- Kline RP, Wu EX, Petrylak DP et al (2000) Rapid in vivo monitoring of chemotherapeutic response using weighted sodium magnetic resonance imaging. Clin Cancer Res 6:2146–2156
- Stobbe R, Beaulieu C (2005) In vivo sodium magnetic resonance imaging of the human brain using soft inversion recovery fluid attenuation. Magn Reson Med 54:1305–1310. https://doi.org/10.1002/mrm.20696
- Nagel AM, Amarteifio E, Lehmann-horn F, Jurkat-rott K (2011) 3 tesla sodium inversion recovery magnetic resonance imaging allows for improved visualization of intracellular sodium content changes in muscular channelopathies. Invest Radiol 46:759–766. https://doi.org/10.1097/RLI.0b013e31822836f6
- Pekar J, Leigh JS (1986) Detection of biexponential relaxation in sodium-23 facilitated by double-quantum filtering. J Magn Reson 69:582–584. https://doi.org/10.1016/0022-2364(86)90180-0
- 20. Jaccard G, Wimperis S, Bodenhausen G (1986) Multiple-quantum NMR spectroscopy of S=3/2 spins in isotropic phase: a new probe

for multiexponential relaxation. J Chem Phys 85:6282. https://doi. org/10.1063/1.451458

- Hoesl MAU, Kleimaier D, Hu R et al (2019) 23 Na Triple-quantum signal of in vitro human liver cells, liposomes, and nanoparticles: cell viability assessment vs. separation of intra- and extracellular signal. J Magn Reson Imaging 50:435–444. https:// doi.org/10.1002/jmri.26666
- Jelicks LA, Gupta RK (1993) On the extracellular contribution to multiple quantum filtered 23Na NMR of perfused rat heart. Magn Reson Med 29:130–133
- Dyck PJ, Boes CJ, Mulder D et al (2005) History of standard scoring, notation, and summation of neuromuscular signs. A current survey and recommendation. J Peripher Nerv Syst 10:158–173. https://doi.org/10.1111/j.1085-9489.2005.0010206.x
- Glover GH, Schneider E (1991) Three-point Dixon technique for true water/fat decomposition with B0 inhomogeneity correction. Magn Reson Med 18:371–383. https://doi.org/10.1002/ mrm.1910180211
- Nagel AM, Laun FB, Weber MA et al (2009) Sodium MRI using a density-adapted 3D radial acquisition technique. Magn Reson Med 62:1565–1573. https://doi.org/10.1002/mrm.22157
- Hancu I, Boada F, Shen G (1999) Three-dimensional triple-quantum-filtered 23Na imaging of in vivo human brain. Magn Reson Med 42:1146–1154
- 27. Gast LV, Gerhalter T, Hensel B et al (2018) Double quantum filtered 23Na MRI with magic angle excitation of human skeletal muscle in the presence of B0 and B1 inhomogeneities. NMR Biomed 31:e4010. https://doi.org/10.1002/nbm.4010
- Goutallier D, Postel JM, Bernageau J et al (1994) Fatty muscle degeneration in cuff ruptures. Pre- and postoperative evaluation by CT scan. Clin Orthop Relat Res 1:78–83
- Wang LH, Friedman SD, Shaw D et al (2019) MRI-informed muscle biopsies correlate MRI with pathology and DUX4 target gene expression in FSHD. Hum Mol Genet 28:476–486. https://doi. org/10.1093/hmg/ddy364
- Azzabou N, Loureiro de Sousa P, Caldas E, Carlier PG (2015) Validation of a generic approach to muscle water T2 determination at 3T in fat-infiltrated skeletal muscle. J Magn Reson Imaging 41:645–653. https://doi.org/10.1002/jmri.24613
- Umathum R, Rösler MB, Nagel AM (2013) In Vivo 39K MR imaging of human muscle and brain. Radiology 269:569–576. https://doi.org/10.1148/radiol.13130757
- Benjamini Y, Hochberg Y (1995) Controlling the false discovery rate: a practical and powerful approach to multiple testing. J R Stat Soc Ser B 57:289–300. https://doi.org/10.1111/j.2517-6161.1995. tb02031.x
- 33. Mair D, Huegens-Penzel M, Kress W et al (2017) Leg Muscle involvement in facioscapulohumeral muscular dystrophy:

comparison between Facioscapulohumeral Muscular Dystrophy types 1 and 2. Eur Neurol 77:32–39. https://doi.org/10.1159/00045 2763

- Kan HE, Klomp DWJ, Wohlgemuth M et al (2010) Only fat infiltrated muscles in resting lower leg of FSHD patients show disturbed energy metabolism. NMR Biomed 23:563–568. https://doi. org/10.1002/nbm.1494
- Kan HE, Scheenen TWJ, Wohlgemuth M et al (2009) Quantitative MR imaging of individual muscle involvement in facioscapulohumeral muscular dystrophy. Neuromuscul Disord 19:357–362. https://doi.org/10.1016/j.nmd.2009.02.009
- Olsen DB, Gideon P, Jeppesen TD, Vissing J (2006) Leg muscle involvement in facioscapulohumeral muscular dystrophy assessed by MRI. J Neurol 253:1437–1441. https://doi.org/10.1007/s0041 5-006-0230-z
- Janssen BH, Voet NBM, Nabuurs CI et al (2014) Distinct disease phases in muscles of facioscapulohumeral dystrophy patients identified by MR detected fat infiltration. PLoS ONE 9:e85416. https://doi.org/10.1371/journal.pone.0085416
- Monforte M, Laschena F, Ottaviani P et al (2019) Tracking muscle wasting and disease activity in facioscapulohumeral muscular dystrophy by qualitative longitudinal imaging. J Cachexia Sarcopenia Muscle 10:1258–1265. https://doi.org/10.1002/jcsm.12473
- Arpan I, Willcocks RJ, Forbes SC et al (2014) Examination of effects of corticosteroids on skeletal muscles of boys with DMD using MRI and MRS. Neurology 83:974–980. https://doi. org/10.1212/WNL.00000000000775
- Amarteifio E, Nagel AM, Weber M, Lehmann-horn F (2012) Hyperkalemic periodic paralysis and permanent weakness : 3-T overload — initial results. Radiology 264:154–163
- Navon G, Shinar H, Eliav U, Seo Y (2001) Multiquantum filters and order in tissues. NMR Biomed 14:112–132
- Schepkin VD, Choy IO, Budinger TF et al (1998) Sodium TQF NMR and intracellular sodium in isolated crystalloid perfused rat heart. Magn Reson Med 39:557–563
- 43. Navon G (1993) Complete elimination of the extracellular 23Na NMR signal in triple quantum filtered spectra of rat hearts in the presence of shift reagents. Magn Reson Med 30:503–506. https ://doi.org/10.1002/mrm.1910300415
- 44. Eykyn TR, Aksentijević D, Aughton KL et al (2015) Multiple quantum filtered 23Na NMR in the Langendorff perfused mouse heart: Ratio of triple/double quantum filtered signals correlates with [Na]i. J Mol Cell Cardiol 86:95–101. https://doi. org/10.1016/j.yjmcc.2015.07.009
- Benkhedah N, Bachert P, Nagel AM (2014) Two-pulse biexponential-weighted 23Na imaging. J Magn Reson 240:67–76. https ://doi.org/10.1016/j.jmr.2014.01.007