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### Molecular Genetics and Metabolism Reports



journal homepage: www.elsevier.com/locate/ymgmr

## Case Report

# Novel homozygous *GLDC* variant causing late-onset glycine encephalopathy: A case report and updated review of the literature

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ARTICLE INFO	A B S T R A C T
Keywords:	Glycine encephalopathy (MIM #605899) is an autosomal recessive inborn error of metabolism caused by pathogenic variants in three genes <i>GLDC</i> , <i>AMT</i> , <i>GCSH</i> encoding glycine cleavage enzyme system. We report an 8-year-old boy with late-onset glycine encephalopathy who harbors a novel homozygous <i>GLDC</i> likely pathogenic variant c.707G > A p.(Arg236Gln). Polyhydramnios was noted at fetal ultrasound. He displayed global develo
glycine encephalopathy	
Polyhydramnios Chorioangioma	

trum of late-onset nonketotic hyperglycinemia.

#### 1. Introduction

Dandy-Walker malformation

Deleterious pathogenic variants in the glycine cleavage system P protein (GCSP) encoded by GLDC gene (MIM \*238300) have been reported to cause autosomal recessive glycine encephalopathy (MIM #605899) characterized by early-onset neonatal hypotonia, progressive lethargy, poor feeding, seizures, developmental delays, hyperactivity with or without choreatic movements. Of note, approximately 20% of patients with glycine encephalopathy harbor GLDC gene deletion. Brain MRI often shows thin and shortened corpus callosum, hydrocephalus, cerebral atrophy, ventricular dilatation, hypomyelination [1]. Most of patients with glycine encephalopathy have an uneventful pregnancy with normal delivery (Supplementary Table S1) [2-29]. Here, we document an 8-year-old-boy with late-onset nonketotic hyperglycinemia (NKH) in whom WES revealed a novel homozygous GLDC likely pathogenic variant c.707G > A p.(Arg236Gln). Second trimester fetal ultrasound showed polyhydramnios. To the best of our knowledge, this is the first report of late-onset glycine encephalopathy with abnormal prenatal findings.

#### 2. Clinical report and results

opmental delay, craniofacial dysmorphism, convulsions. Our report expands the phenotypic and genetic spec-

This 8-year-old boy presented for the genetic consultation because of global developmental delay and epilepsy. Fetal ultrasound at 26 weeks of gestation showed polyhydramnios and dilated umbilical veins (Fig. 1A, right image). He was born prematurely by caesarean section at 26 + 1 weeks of gestation to first-cousin consanguineous parents and presented with umbilical hernia, intraventricular hemorrhage at birth. His birth weight, length and occipitofrontal circumference were respectively 1060 g (> 50th percentile), 37.5 cm (> 50th percentile) and 25.5 cm (>50th percentile) with Apgar score of 8/7. Examination of the placenta demonstrated giant chorioangioma (Fig. 1A, left image). He displayed respiratory distress at 2-month-old, right hydrocele at 5month-old. Recurrent respiratory infections, peripheral hypertonia and axial hypotonia were also noted. Brain MRI showed Dandy-Walker malformation (hydrocephalus, cerebellar hypoplasia and retrocerebellar cyst), bilateral posterior periventricular leukomalacia (Fig. 1B). He experienced his first epilepsy at the age of 2-year-6-monthold which was controlled by treatment of valproate sodium (200 mg x2 per day) and levetiracetam (300 mg x2 per day). Electroencephalogram (EEG) showed multifocal spikes. He underwent endoscopic third

https://doi.org/10.1016/j.ymgmr.2023.100959

Received 30 December 2022; Received in revised form 31 January 2023; Accepted 31 January 2023

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ventriculocisternostomy for the treatment of hydrocephalus.

On last clinical assessment at the age of 8-year-4-month-old, his weight, height, and head circumference were 26 kg (50-75th percentile), 130 cm (50-75th percentile) and 53.5 cm (>50th percentile). He had global developmental delay, began to walk at 32 months and spoke his first word at 24 months. He also had severe learning disability, dyslexia, and dysgraphia. Deep tendon reflexes revealed hyperreflexia of the upper limbs as well as hypertonia of the lower extremities. Dysmorphic features included long face, high forehead, high arched eyebrows, bilateral ptosis, strabismus, low-set ears, long philtrum, thin upper lip, upturned corners of the mouth and bilateral camptodactyly (Fig. 1C). Ataxic gait and bilateral valgus were also noticed. 180 K array Comparative Genomic Hybridization (array-CGH) showed no chromosomal imbalances. Targeted next-generation sequencing of 95 genes involved in epilepsy revealed a normal result. Trio WES identified a novel homozygous missense likely pathogenic variant in GLDC gene c.707G > A p.(Arg236Gln) (Fig. 1D) (PM2, PM3, PM5, PP3, PP4 ACMG criteria, and 4 likely functional affect according to ABC functional "A" grading of ESHG). No GLDC intragenic deletion were detected (Supplementary Table S2). The parents were heterozygous. This variant allele was absent in 13,006 chromosomes from Exome Server Variant (https://evs.gs.washington.edu/EVS/) and found at a frequency of heterozygotes of  $1193 \times 10^{-5}$  in 251,476 control chromosomes (3 Molecular Genetics and Metabolism Reports 34 (2023) 100959

heterozygous individuals/251,476 subjects) in the gnomAD databases (https://gnomad.broadinstitute.org/). Moreover, the variant *GLDC* c.707G > A p.(Arg236Gln) is predicted to be deleterious by suite of bioinformatics prediction analysis programs. The substitution of a charged amino acid arginine with a polar amino acid glutamine at 236th position results in the decrease of the P protein stability (Supplementary Table S3). The Arg<sup>236</sup> is highly conserved during species and predicted to be intolerant to changes by MetaDome (https://stuart.radboudumc.nl/metadome/) and Phyre<sup>2</sup> (http://www.sbg.bio.ic.ac.uk/~phyre2/h tml/page.cgi?id=index) (Fig. 2). Molecular modelling of Arg236Gln based on the human glycine decarboxylase structure showing a complete reorganization of the non-covalent interactions of the region (Supplemental Fig. S1).

Metabolic investigation showed the plasma glycine level increased to 1029  $\mu$ mol/L (reference value 113–397  $\mu$ mol/L) and elevated urine glycine of 4280 mmol/mol creatinine (reference value <250 mmol/mol creatinine).

After 12 h and 30 min of fasting, blood redox status showed a normal result [lactic acid 1.2 mmol/L (reference value 0.5–1.8 mmol/L), py-ruvic acid 0.10 mmol/L (reference value 0.05–0.12 mmol/L), lactate/ pyruvate ratio 12 mmol/mmol (reference value 7–16), beta-hydroxybutyric acid (B) 0.02 mmol/L (reference value <0.22-), total ketone body 0.05 mmol/L (reference value 0.02–0.40 mmol/L),



**Fig. 1.** A. Placental examination showing moderate villous hydrops associated with large chorioangioma (left picture) and prenatal ultrasound showing polyhydramnios at 26 weeks of gestation (right picture). B. Brain MRI showing Dandy-Walker malformation with cerebellar hypoplasia (red asterisk). C. Front and lateral views of the patient showing facial dysmorphism including elongated face with high forehead, high arched eyebrows, ptosis and strabismus, low-set ears, long philtrum, thin upper lip, upturned corner of the mouth, and bilateral camptodactyly. D. Sanger sequencing identified a homozygous novel *GLDC* variant c.707G > A p.(Arg236Gln) in the propositus. The parents were heterozygous.

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A



**Fig. 2.** A. ClustalW showing the  $Arg^{236}$  is highly conserved among orthologs. B. Graphic representation of the mutation tolerance score of the affected  $Arg^{236}$  of the glycine cleavage system P protein obtained by MetaDome web server indicating that missense variants occurring at this position is intolerant. C. Phyre<sup>2</sup> showing 3D glycine cleavage system P protein structure, the replacement of the charged  $Arg^{236}$  by polar glutamine amino acid is moderately intolerant and represent with green color ( $Arg^{236}$  is surrounded by red color, mutational sensitivity score = 4).

acetoacetic acid (A) 0.03 mmol/L (reference value 0.00–0.15 mmol/L), (B/A) ratio 0.67 mmol/mmol (reference value >1), free fatty acid 0.66 mmol/L (reference value 0.20–1.20 mmol/L)]. Repeated plasma glycine showed hyperglycinemia of 1414  $\mu$ mol/L confirming the nonketotic hyperglycinemia without ketone bodies. Indeed, the analysis of amino acids in the plasma, along with the molecular analysis result and clinical manifestation, proved the diagnosis of nonketotic hyperglycinemia.

#### 3. Discussion

We report the new case of late-onset nonketotic hyperglycinemia with a confirmed novel homozygous missense variant in the GLDC gene. The variant c.707G > A p.(Arg236Gln) has never been reported in the public databases (homozygous GLDC variant c.707G > A p.(Arg236Gln) was absent in 264,482 control chromosomes). It is predicted to result from a substitution of a charged arginine amino acid from a polar glutamine amino acid at the 236th position of the glycine cleavage system P protein. The Arg<sup>236</sup> is highly conserved among orthologs, and its substitution is also predicted to be "intolerant" or "probably damaging" by MetaDome and Phyre<sup>2</sup> and decrease the stability of the P protein by a suite of bioinformatics analysis prediction (Fig. 2, Supplementary Table S2). We performed a PUBMED search of all studies including keywords "GLDC gene, glycine encephalopathy, nonketotic hyperglycinemia" from 1964 to 2022. Eligible publications had to include at least age of disease-onset, symptomatology, initial biochemical data [Cerebrospinal fluid (CSF) glycine, plasma glycine and CSF/ plasma ratio], EEG, brain imaging and molecular genetic result. Studies missing clinical phenotype and/or genetic information were all excluded.

Having reviewed the medical literature, only seventeen *GLDC*mutated patients with prenatal record have been documented so far and all of them have uneventful pregnancy with normal delivery [2–12]. Of interest, this is the first case-report of nonketotic hyperglycinemia with prenatal findings. The propositus was born at 26 + 1 weeks of gestation due to polyhydramnios and giant chorioangioma.

Of note, he was referred to the clinical genetics department because of his seizure and global developmental delay without metabolic investigation. He exhibited unusual, infrequent phenotype of nonketotic hyperglycinemia with the presence of antenatal polyhydramnios and chorioangioma, facial dysmorphism and Dandy-Walker malformation which were rarely observed in late-onset nonketotic hyperglycemic patient. Consequently, the diagnosis of nonketotic hyperglycinemia was not established in the absence of metabolic investigation. Trio whole-exome sequencing was performed to unravel the molecular pathogenesis of epilepsy associated with Dandy-Walker malformation in the propositus with the identification of a novel homozygous *GLDC* missense variant. In addition, he displayed particular prenatal phenotype which seems to be incompatible with attenuated nonketotic hyperglycinemia leading to suspect the presence of other variants in genes implicated in folic acid metabolism. However, neither pathogenic variants nor copynumber variations were detected in *MTHFR*, *MTR*, *MTRR* genes.

Indeed, the presence of non-specific phenotype and difficulty in obtaining CSF samples could hamper the clinical diagnosis of late-onset nonketotic hyperglycinemia and delay the therapeutic strategy [3]. The residual glycine cleavage system enzyme activity caused by different genetic variants can lead to distinct clinical presentation.

The majority of *GLDC*-mutated patients have no dysmorphic features or unavailable data of craniofacial dysmorphism. Hitherto, only two reported *GLDC*-mutated patients display minor dysmorphic features including low-set ears and micrognathia [13]. *GLDC* disease-causing pathogenic variants account for approximately 80% NKH cases. 85% patients had severe neonatal NKH and up to 30% patients died during the neonatal period making difficult to assess, continue regular followup and evaluate the craniofacial dysmorphism, particularly in neonates with minor craniofacial changes [1,11,13,22,23,29]. So, the detailed data of facial dysmorphic features are commonly not available in most cases (Supplementary Table S1).

Moreover, brain imaging usually showed cerebral volume decrease or brain atrophy, ventricular dilatation, white matter abnormalities, hypogenesis of the corpus callosum, short and thin corpus callosum in severe NKH, diffusion restriction in the posterior limb of the internal capsule, focal cerebral hemorrhage, and diffuse hypomyelination. Cerebellar atrophy was reported only in one glycine encephalopathy patient (Supplementary Table S1) [1,30]. Our patient is the second case presenting with Dandy-Walker malformation [31]. Our study shows that apneic episode/respiratory distress is one of the most common clinical features which was observed in 37% patients with glycine encephalopathy (Supplementary Table S1).

In conclusion, this is the first report of late-onset nonketotic hyperglycinemia presenting abnormal prenatal ultrasound findings with polyhydramnios, giant chorioangioma leading to the preterm labor. We also highlight the importance of WES in the diagnosis of inherited metabolic disorders, especially in the case of unspecific phenotype and difficulty in obtaining CSF samples for metabolic investigation. Our observations confirm the clinical heterogeneity of nonketotic hyperglycinemia and expand the phenotypic and molecular spectrum of *GLDC*-related glycine encephalopathy.

Supplementary data to this article can be found online at https://doi.org/10.1016/j.ymgmr.2023.100959.

#### **Funding sources**

The authors received no financial support for this study.

#### Code availability statement

Not applicable.

#### Authors' contribution

M.T.H designed the study and wrote the manuscript. J-M.D.S.G and E.L. performed the molecular analysis. A.P. performed the prenatal ultrasound. D·D-L.C and H·B performed patient's follow-up and critically revised the manuscript.

#### Ethical approval statement

This work is not clinical research and considered as routine clinical care.

#### **Consent for publication**

Written informed consent of this case report was obtained from the patient's parents who have given their permission to publish the patients' photographs.

#### **Declaration of Competing Interest**

The authors declared no conflicts of interest.

#### Data availability

The data was submitted in LOVD databases (Leiden Open Databases): Individual #00424557: https://databases.lovd.nl/shared/individuals/ 00424557.

#### Acknowledgement

We would like to thank the patients and his family members for participation in the study.

#### References

- [1] N.A.A. Azize, W.Z.W. Ngah, Z. Othman, N.M. Desa, C.B. Chin, Z.M. Yunus, A. Mohan, T.S. Hean, S.Z.S. Zakaria, N. Lock-Hock, Mutation analysis of glycine decarboxylase, aminomethyltransferase and glycine cleavage system protein-H genes in 13 unrelated families with glycine encephalopathy, J. Hum. Genet. 59 (2014) 593–597.
- [2] C.F. Chiu, J.L. Lin, J.J. Lin, M.H. Tseng, F.S. Lo, M.C. Chiang, Nonketotic hyperglycinemia of infants in Taiwan, Pediatr. Neonatol. 57 (2016) 420–426.

- [3] Y. Cao, L. Meng, Y. Zhang, J. Jiao, W. Pu, L. Ma, Novel GLDC compound heterozygous variant leading to Nonketotic hyperglycinemia: case report and literature review, Front. Pediatr. 9 (2021), 725930.
- [4] S. Meyer, C. Acquaviva, M.G. Shamdeen, D. Haas, C. Vianey-Saban, A novel mutation in a neonate with Nonketotic hyperglycinemia, Pediatr. Neurol. 43 (2010) 363–367.
- [5] E. Kose, U. Yis, S. Hiz, Arslan N (2017) A novel mutation in the glycine decarboxylase gene in patient with non-ketotic hyperglycinemia, Neurosciences 22 (2017) 131–133.
- [6] A. Dinopoulos, S. Kure, G. Chuck, K. Sato, D.L. Gilbert, Y. Matsubara, T. Degrauw, Glycine decarboxylase mutations: A distinctive phenotype of nonketotic hyperglycinemia in adults, Neurology. 64 (2005) 1255–1257.
- [7] W. Khraim, B. Abu-Libdeh, S. Ayesh, I. Dweikat, Clinical heterogeneity of glycine encephalopathy in three Palestinian siblings: A novel mutation in the glycine decarboxylase (*GLDC*) gene, Brain and Development 39 (2017) 601–605.
- [8] S. Kure, S.H. Korman, J. Kanno, A. Narisawa, M. Kubota, T. Takayanagi, M. Takayanagi, T. Saito, A. Matsui, F. Kamada, Y. Aoki, T. Ohura, Y. Matsubara, Rapid diagnosis of glycine encephalopathy by <sup>13</sup>C-Glycine breath test, Ann. Neurol. 59 (2006) 862–867.
- [9] P.S. Kruszka, B. Kirmse, D.J. Zand, K. Cusmano-Ozog, E. Spector, J.L.V. Hove, K. A. Chapman, Concurrent non-ketotic hyperglycinemia and propionic acidemia in an eight-year-old boy, Mol. Genet. Metab. Rep. 1 (2014) 237–240.
- [10] L. Sellner, E. Edkins, L. Greed, B. Lewis, Detection of mutations in the glycine decarboxylase gene in patients with nonketotic hyperglycinemia, Mol. Genet. Metab. 84 (2005) 167–171.
- [11] C. Brunel-Guitton, B. Casey, M. Coulter-Mackie, H. Vallance, D. Hewes, S. Stockler-Ipsiroglu, S. Mercimek-Mahmutoglu, Late-onset nonketotic hyperglycinemia caused by a novel homozygous missense mutation in the GLDC gene, Mol. Genet. Metab. 103 (2011) 193–196.
- [12] H. Flusser, S.H. Korman, K. Sato, Y. Matsubara, A. Galil, S. Kure, Mild glycine encephalopathy in a large kindred due to a silent exonic *GLDC* splice mutation, Neurology. 64 (2005) 1426–1430.
- [13] H. Bayrak, Y. Yildiz, A. Olgaç, C.S. Kasapkara, A. Küçükcongar, A. Zenciroglu, D. Yüksel, S. Ceylaner, M. Kiliç, Genotypic and phenotypic features in Turkish patients with classic nonketotic hyperglycinemia, Met. Brain. Dis. 36 (2021) 1213–1222.
- [14] S. Kanekar, D. Byler, Characteristic MRI findings in neonatal nonketotic hyperglycinemia due to sequence changes in GLDC gene encoding the enzyme glycine decarboxylase, Metab. Brain Dis. 28 (2013) 717–720.
- [15] J.N. Brenton, R.S. Rust, Late-onset hyperglycinemia with a heterozygous novel point mutation of the GLDC gene, Pediatr. Neurol. 50 (2014) 536–538.
- [16] S. Kure, A. Ichinohe, K. Kojima, K. Sato, Z. Kizaki, F. Inoue, C. Yamanaka, Y. Matsubara, Mild variant of nonketotic hyperglycinemia with typical neonatal presentations: mutational and in vitro expression analyses in two patients, J. Pediatr. 144 (2004) 827–829.
- [17] I. Bravo-Alonso, R. Navarrete, L. Arribas-Carreira, A. Perona, D. Abia, M.L. Couce, A. Garcia-Cazorla, A. Morais, R. Domingo, M.A. Ramos, M.A. Swanson, L.J.K. V. Hove, M. Ugarte, B. Perez, C. Peréz-Cerda, P. Rodriguez-Pombo, Nonketotic hyperglycinemia: functional assessment of missense variants in GLDC to understand phenotypes of the disease, Hum. Mut. 38 (2017) 678–691.
- [18] J.R. Toone, D.A. Applegarth, S. Kure, M.B. Coulter-Mackie, P. Sazegar, K. Kojima, A. Ichinohe, Novel mutations in the P-protein (glycine decarboxylase) gene in patients with glycine encephalopathy (non-ketotic hyperglycinemia), Mol. Genet. Metab. 76 (2002) 243–249.
- [19] S.H. Korman, A. Boneh, A. Ichinohe, K. Kojima, K. Sato, Z. Ergaz, J.M. Gomori, A. Gutman, S. Kure, Persistent NKH with transient or absent symptoms and a homozygous GLDC mutation, Ann. Neurol. 56 (2004) 139–143.
- [20] S.L. Nickerson, S. Balasubramaniam, P.A. Dryland, J.M. Love, M.P. Kava, D. R. Love, D.O. Prosser, Two novel GLDC mutations in a neonate with nonketotic hyperglycinemia, J. Pediatr. Genet. 5 (2016) 174–180.
- [21] J.M. Love, D. Prosser, D.R. Love, K.P. Chintakindi, A.B. Dalal, S. Aggarwal, A novel Glycine decarboxylase gene mutation in an Indian family with nonketotic hyperglycinemia, J. Child Neurol. 29 (2014) 122–127.
- [22] C. Conter, M.O. Rolland, D. Cheillan, V. Bonnet, I. Maire, R. Froissart, Genetic heterogeneity of the *GLDC* gene in 28 unrelated patients with glycine encephalopathy, J. Inherit. Metab. Dis. 29 (2006) 135–142.
- [23] S. Kure, K. Kato, A. Dinopoulos, C. Gail, T.J. deGrauw, J. Christodoulou, V. Bzduch, R. Kalmanchey, G. Fekete, A. Trojovsky, B. Plecko, G. Breningstal, J. Tohyama, Y. Aoki, Y. Matsubara, Comprehensive mutation analysis of GLDC, AMT, and GCSH in nonketotic hyperglycinemia, Hum. Mut. 27 (2006) 343–352.
- [24] T.F. Tramontana, T.E. Wilson, B.E. Hainline, Consideration of a metabolic disorder in the differential of mild developmental delay: A case of nonketotic hyperglycinemia revisited 36 years later, JIMD. Rep. 59 (2021) 16–19.
- [25] F. Ezgu, B. Ciftci, B. Topçu, G. Adiyaman, H. Gökmnoglu, A. Küçükçongar, C. Kasapkara, biberoglu G, Tümer L, Hasanoglu A., Diagnosis of glycine encephalopathy in a pediatric patient by detection of a *GLDC* mutation during initial next generation DNA sequencing, Metab. Brain Dis. 29 (2014) 211–213.
- [26] N. Mitta, R.N. Menon, A. McTaque, A. Radhakrishnan, S. Sundaram, A. Cherian, G. K. Madhavilatha, A.U. Mannan, S. Nampoothiri, S.V. Thomas, Genotype-phenotype correlates of infantile-onset developmental and epileptic encephalopathy syndromes in South India: A single centre experience, Epilepsy Res. (2020) 166. http://doi. https://doi.org/10.1016/j.eplepsyres.2020.106398.
- [27] S. Kure, K. Kojima, A. Ichinohe, T. Maeda, R. Kalmanchey, G. Gekete, S.Z. Berg, J. Filiano, Y. Aoki Y, Suzuki Y, Izumi T, Matsubara Y., Heterozygous GLDC and GCSH gene mutations in transient neonatal hyperglycinemia, Ann. Neurol. 52 (2002) 643–646.

#### M.-T. Huynh et al.

- [28] A. Boneh, S.H. Korman, K. Sato, J. Kanno, Y. Matsubara, I. Lerer, Z. Ben-Neriah, S. Kure, A single nucleotide substitution that abolishes the initiator methionine codon of the *GLDC* gene is prevalent among patients with glycine encephalopathy in Jerusalem, J. Hum. Genet. 50 (2005) 230–234.
- [29] J. Kanno, T. Hutchin, F. Kamada, A. Narisawa, Y. Aoki, Y. Matsubara, S. Kure, Genomic deletion within *GLDC* is a major cause of non-ketotic hyperglycinemia, J. Med. Genet. 44 (2007), e69.
- [30] V.N. Stence, Z.L. Fenton, C. Levek, S. Tong, R.C. Coughlin, B.J. Hennermann, B. S. Wortmann, Van Hove LKJ, Brain imaging in classic nonketotic hyperglycinemia: quantitative analysis and relation to phenotype, J. Inherit. Metab. Dis. 42 (2019) 438–450.
- [31] L. Manel, Z.B. Houneida, A. Habib, B. Dejla, K. Chekib, A rare inborn error of metabolism associated with a Dandy-Walker malformation, Acta Neurol. Belg. 112 (2012) 425–426.