

Intestinal Permeability Is a Mechanical Rheostat in the Pathogenesis of Liver Cirrhosis

Norihisa Nishimura *[®], Kosuke Kaji, Koh Kitagawa, Yasuhiko Sawada, Masanori Furukawa, Takahiro Ozutsumi, Yukihisa Fujinaga [®], Yuki Tsuji, Hiroaki Takaya, Hideto Kawaratani [®], Kei Moriya, Tadashi Namisaki [®], Takemi Akahane [®], Hiroshi Fukui and Hitoshi Yoshiji

> Department of Gastroenterology, Nara Medical University, 840 Shijo-cho, Kashihara, Nara 634-8522, Japan; kajik@naramed-u.ac.jp (K.K.); kitagawa@naramed-u.ac.jp (K.K.); yasuhiko@naramed-u.ac.jp (Y.S.); furukawa@naramed-u.ac.jp (M.F.); ozutaka@naramed-u.ac.jp (T.O.); fujinaga@naramed-u.ac.jp (Y.F.); tsujih@naramed-u.ac.jp (Y.T.); htky@naramed-u.ac.jp (H.T.); kawara@naramed-u.ac.jp (H.K.); moriyak@naramed-u.ac.jp (K.M.); tadashin@naramed-u.ac.jp (T.N.); stakemi@naramed-u.ac.jp (T.A.); hfukui@naramed-u.ac.jp (H.F.); yoshijih@naramed-u.ac.jp (H.Y.)

* Correspondence: nishimuran@naramed-u.ac.jp; Tel.: +81-744-22-3051

Abstract: Recent studies have suggested that an alteration in the gut microbiota and their products, particularly endotoxins derived from Gram-negative bacteria, may play a major role in the pathogenesis of liver diseases. Gut dysbiosis caused by a high-fat diet and alcohol consumption induces increased intestinal permeability, which means higher translocation of bacteria and their products and components, including endotoxins, the so-called "leaky gut". Clinical studies have found that plasma endotoxin levels are elevated in patients with chronic liver diseases, including alcoholic liver disease and nonalcoholic liver disease. A decrease in commensal nonpathogenic bacteria including Ruminococaceae and Lactobacillus and an overgrowth of pathogenic bacteria such as Bacteroidaceae and Enterobacteriaceae are observed in cirrhotic patients. The decreased diversity of the gut microbiota in cirrhotic patients before liver transplantation is also related to a higher incidence of post-transplant infections and cognitive impairment. The exposure to endotoxins activates macrophages via Toll-like receptor 4 (TLR4), leading to a greater production of proinflammatory cytokines and chemokines including tumor necrosis factor-alpha, interleukin (IL)-6, and IL-8, which play key roles in the progression of liver diseases. TLR4 is a major receptor activated by the binding of endotoxins in macrophages, and its downstream signal induces proinflammatory cytokines. The expression of TLR4 is also observed in nonimmune cells in the liver, such as hepatic stellate cells, which play a crucial role in the progression of liver fibrosis that develops into hepatocarcinogenesis, suggesting the importance of the interaction between endotoxemia and TLR4 signaling as a target for preventing liver disease progression. In this review, we summarize the findings for the role of gut-derived endotoxemia underlying the progression of liver pathogenesis.

Keywords: leaky gut; endotoxins; alcoholic liver disease; nonalcoholic steatohepatitis; liver cirrhosis; hepatocarcinogenesis; Toll-like receptor 4 pathway

1. Introduction

The interaction between the intestine and liver via microbiologic features from the gut, referred to as the "gut–liver axis", has been recently demonstrated as contributing to the development of liver diseases that develop liver cirrhosis and hepatocellular carcinoma (HCC). Although there are many mechanisms underlying the pathogenesis of liver inflammation, fibrosis, and carcinogenesis, microbial metabolites and products derived from the intestinal tract are considered to be some of the major factors that accelerate the progression of liver diseases due to the close links between the liver and intestine.

In patients with liver diseases, the gut bacteria themselves, as well as their components, often translocate into the portal blood flow and directly reach the liver as a result of the dis-



Citation: Nishimura, N.; Kaji, K.; Kitagawa, K.; Sawada, Y.; Furukawa, M.; Ozutsumi, T.; Fujinaga, Y.; Tsuji, Y.; Takaya, H.; Kawaratani, H.; et al. Intestinal Permeability Is a Mechanical Rheostat in the Pathogenesis of Liver Cirrhosis. *Int. J. Mol. Sci.* **2021**, *22*, 6921. https://doi.org/10.3390/ijms22136921

Academic Editor: J. B. Bernd Helms

Received: 26 May 2021 Accepted: 24 June 2021 Published: 28 June 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). rupted intestinal barrier, or so-called bacterial translocation. In chronic liver disease (CLD) patients, dysbiosis is often observed and increases alongside the stages of liver fibrosis, leading to bacterial translocation including endotoxins. Endotoxins (lipopolysaccharides; LPS) are a well-known component of Gram-negative bacteria and work as pathogen-associated molecular patterns (PAMPs) for Toll-like receptor 4 (TLR4), which recognizes unique structural components of bacteria and drives innate immunity-producing inflammatory cytokines, such as tumor necrosis factor- α (TNF- α) and interleukin (IL)-6 [1]. In the liver, Kupffer cells (KCs) are regional macrophages that control the innate immune response and are activated by LPS binding, but LPS also stimulate hepatic nonimmune cells, including hepatic stellate cells (HSCs) and sinusoidal endothelial cells as they also have TLR4 [2–4]. In this review, we summarize the influence of gut-derived endotoxins on the progression of liver disease.

2. Mechanisms of Endotoxemia Derived from Gut Microbiota

2.1. Dysbiosis

Recent studies have demonstrated that the alteration in the gut microbiome referred to as "dysbiosis" is closely associated with the clinical stages of CLDs, including liver cirrhosis, the end stage of chronic hepatitis [5–7]. The role of dysbiosis in patients with CLDs has been reported by a study investigating the alteration in gut microbial diversity by 16S sequencing and a meta-analysis in some diseases such as liver cirrhosis, colorectal cancer, inflammatory bowel disease, obesity, and type 2 diabetes mellitus [5]. This study also suggested that liver fibrosis, the F4 stage of fibrosis, is the condition in which dysbiosis is linked to the pathogenesis, including in patients with nonalcoholic steatohepatitis (NASH) [8,9]. Likewise, an alteration in the diversity of the gut microbiota showed the most significant difference between healthy control and cirrhotic patients [5,6]. In patients with hepatitis B (HBV), the proportion of Bifidobacteria and Bacteroidetes decreased, whereas that of *Enterococcus*, *Enterobacteriaceae*, and *Proteobacteria* increased [10–12]. In patients with NASH and diabetes, obesity triggers dysbiosis, which leads to a decrease in the diversity of the gut microbiota and an increased ratio of *Firmicutes* to *Bacteroidetes* [13]. A decrease in Bacteroidetes has also been observed in the intestine of alcoholic patients [14]. Furthermore, patients with alcoholic liver disease demonstrated an increase in Veillonellaceae, which has been reported to induce highly systemic inflammation [15]. In Table 1, we list representative alterations in bacterial diversity according to types of CLDs. The evidence from these studies suggests that dysbiosis affects multiple cascades related to the progression of CLD, such as inflammation, fibrogenesis, regeneration, and immunity, leading to the development of HCC.

2.2. Small Intestinal Bacterial Overgrowth (SIBO)

Small intestinal bacterial overgrowth (SIBO) is a part of dysbiosis that occurs in the small intestine. It is characterized by an abnormal composition of microbiota in the small intestinal tract [16]. Recently, it has become known to be associated with various diseases and not just liver diseases. SIBO is considered as an important phenomenon that leads to an increased intestinal permeability via the impairment of tight junctions (TJs) by intestinal epithelial cell damage [17,18]. Since the intestinal overgrowth of *Escherichia coli* in patients with liver cirrhosis was reported in 2016, a recent study was the first to demonstrate the change in gut microbial diversity in cirrhotic and HCC patients and its possibility as a novel, noninvasive surrogate marker for the detection of HCC in patients with HBV infection [19,20].

An alteration of bile acid secretion is a major factor in inducing bacterial overgrowth. In patients with liver cirrhosis, as the secretion of bile acid from the liver is decreased, bacterial overgrowth and the change in the bacterial composition are promoted. It has further been demonstrated that the gastrointestinal transition time is longer in cirrhotic patients than in healthy controls because of the dysfunction of intestinal motility, which also leads to the promotion of bacterial overgrowth [21]. Another study reported an association

between proximal small intestinal motility and SIBO in patients with liver cirrhosis [22]. As gastric acid affects the regulation of microbiota and prevents the translocation of oral bacteria through the stomach, the inhibition of gastric acid production with the use of a proton pump inhibitor could also be implicated in the development of SIBO [23].

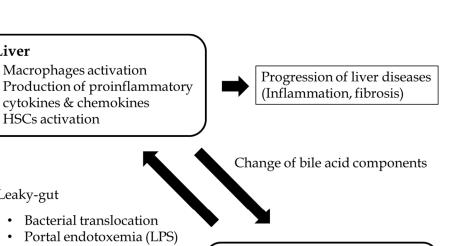
Type of Diseases	Bacteria				
	Phylum	Class	Order	Family	Genus (Species)
ALD	Firmicutes Bacteroidetes↓ Proteobacteria↑	Bacilli Clostridia Negativicutes Bacteroidia proteobacteria ↑	Lactobacilales Clostridiales Selenomonadales Bacteroidales Enterobacteriales	Lactobacillaceae Clostridiaceae Ruminococcaceae↓ Lachnospiraceae Veillonellaceae↑ Bacteroidaceae↓ Porphyromonadaceae↓ Enterobacteriaceae↑	Lactobacillus ↓ Clostridium ↓ Faecalibacterium ↓ Dorea ↑ Blautia ↑ Klebsiella ↑
NAFLD	Firmicutes Bacteroidetes ↑↓ Proteobacteria ↑	Bacilli Clostridia↓ Bacteroidia proteobacteria↑	Lactobacilales Clostridiales J Bacteroidales Enterobacteriales	Lactobacillaceae ↑ Clostridiaceae Ruminococcaceae↓ Lachnospiraceae Bacteroidaceae↓ Porphyromonadaceae ↑↓ Enterobacteriaceae ↑↓	Lactobacillus ↑ Clostridium Faecalibacterium ↓ Dorea ↑ Bacteroides ↓ Escherichia ↑
HBV	Firmicutes Bacteroidetes↓ Proteobacteria↑ Actinobacteria	Bacilli Clostridia Bacteroidia Proteobacteria Betaproteobacteria Actinobacteriia	Lactobacilales Clostridiales Bacteroidales Enterobacteriales Neisseriales Bifidobacteriales	Lactobacillaceae↓ Enterococcaceae↑ Ruminococcaceae Bacteroidaceae Enterobacteriaceae↑ Neiseriaceae↑ Bifidobacteriaceae↓	Lactobacillus↓ Enterococcus↑ Faccalibacterium Bacteroides Bifidobacterium↓
нсу	Firmicutes↓ Bacteroidetes↑ Proteobacteria↑ Actinobacteria	Bacilli Bacteroidia Proteobacteria Actinobacteriia	Lactobacilales Bacillales Enterobacteriales Bifidobacteriales	Lactobacillaceae Enterococcaceae ↑ Stphylococcaceae Enterobacteriaceae ↑ Bifidobacteriaceae	Lactobacillus Enterococcus ↑ Staphylococcus ↑ Bifidobacterium
РВС	Firmicutes Bacteroidetes †↓ Proteobacteria ↑ Fusobacteria Acidobacteria ↓	Bacilli Clostridia Negativicutes Proteobacteria Gammaproteobacteria Fusobacteriia	Lactobacilales Clostridiales Selenomonadales Enterobacteriales Pasteurellales Fusobacteriales	Lactobacillaceae Clostridiaceae Ruminococcaceae Lachnospiraceae Veillonellaceae Enterobacteriaceae ↑ Pasteurellaceae Fusobacteriaceae	Lactobacillus ↑ Eubacterium ↑ Clostridium ↓ Ruminococcus ↓ Lachnobacterium ↓ Veillonella ↑ Actinobacillus ↑ Haemophilus ↑ Fusobacterium ↑↓
AIH	Firmicutes Actinobacteria Bacteroidetes Proteobacteria	Bacilli Actinobacteriia Negativicutes Proteobacteria	Lactobacilales Bifidobacteriales Selenomonadales Enterobacteriales	Lactobacillaceae Bifidobacteriaceae Veillonellaceae Streptococcaceae Enterobacteriaceae	Lactobacillus †↓ Bifidobacterium↓ Veillonella † Streptococcus † Klebsiella †

Table 1. Representative changes in microbial diversity according to types of CLDs.

2.3. Disruptions in Intestinal Barrier Function

The intestinal barrier is composed of three different components, which are physical, immunologic, and microbial barriers. The physical barrier contains a mucus layer and epithelial elements. The epithelium of the intestine has occlusive intracellular junctions, which are the so-called tight junctions (TJs) [24]. TJs are composed of three types of transmembrane proteins associated with scaffolding proteins that link to the actin cytoskeleton such as occludin, claudins, and junctional adhesion molecules [25]. In particular, claudins are the main molecules within the TJ structure and produce charge-selective pores that regulate the transportation of ions and small particles across the epithelial layers. These proteins bind scaffolding proteins such as zonula occludins 1 (ZO-1), ZO-2, and ZO-3, which connect transmembrane proteins protect the translocation of bacterial components and their metabolites.

The causes of intestinal permeability vary and remain to be elucidated. Physical damage to the intestinal epithelium, disruption to the TJ, and alterations in the thickness of the mucus layer can result in increased intestinal leakiness [29]. Dysfunctions in either the innate or adaptive immune response can affect the increase in microbe translocation [30]. Dysbiosis can also promote inflammation in the intestinal epithelium and barrier dysfunction [31]. The increased PAMPs such as LPS, bacterial RNAs, and viral RNAs translocate from the intestine through the intestinal barrier and enter the liver, and they stimulate immune cells such as KCs via binding to TLR4 on their membrane, resulting in the induction of hepatic inflammation that progresses to liver injury and diseases (Figure 1) [31,32].





(Bacterial overgrowth (SIBO))

Impaired gut barrier function (Increased intestinal permeability)

Figure 1. Schema of the interaction between the gut and liver in CLDs. Lifestyle factors such as drinking alcohol and consuming a high-fat diet alter the diversity of the gut microbiota and result in bacterial overgrowth, increasing bacterial translocation including endotoxins (lipopolysaccharides; LPS). LPS can translocate into the portal vein via the disruption of the gut barrier, eventually reaching into the liver as a result of increased intestinal permeability, strongly activating regional liver immune cells, mainly Kupffer cells, which produce proinflammatory cytokines and chemokines. LPS also stimulate the activation of hepatic stellate cells, which play a main role in the progression of liver fibrosis via the production of extracellular matrix proteins such as collagens.

3. The Role of Endotoxemia in the Progression of Liver Pathogenesis

3.1. Alcoholic Liver Disease

Liver

Leaky-gut

High-fat diet

Excessive consumption of alcohol is an important cause of chronic hepatitis that develops into cirrhosis and the occurrence of HCC based on cirrhosis [33,34]. Continuous alcohol intake promotes bacterial overgrowth in both the small and large intestines [35-37]. Moreover, drinking alcohol induces bacterial translocation into the portal vein, which plays a crucial role in alcoholic liver injury [38]. A previous study reported that alcohol-induced cirrhotic patients have SIBO [34]. In addition, in these patients, the serum endotoxin levels are elevated even at the early stage of alcoholic liver disease [39]. Leclercq et al. indicated that almost half of the patients with alcohol dependence have increased intestinal permeability, even at the early stage of liver fibrosis (F0–1) [40]. Those authors also demonstrated that the total amount of gut bacteria, especially Faecalibacterium prausnitzii in the Ruminococcaceae family, was negatively correlated with intestinal permeability, whereas the genera Dorea and Blautia were positively correlated with intestinal permeability. Kakiyama et al. also reported that Bacteroidaceae and Porphyromonadaceae were decreased and Veillonellaceae was increased in patients with alcoholic liver cirrhosis as compared with those with cirrhosis due to other etiologies [15]. In particular, Veillonellaceae could lead to highly systemic inflammation and endotoxemia [15]. The evidence from these studies suggests that dysbiosis is closely associated with increased intestinal permeability and portal endotoxemia.

Endotoxemia derived from the Gram-negative bacteria of the Proteobacteria phylum is considered to be a driver of increased hepatic inflammation. Dysbiosis, namely, a decrease in commensal bacteria, induces a disruption in the intestinal TJ barrier [41]. Experimentally, supplementation of the probiotic *Lactobacillus rhamnosus GG* improved downregulated TJ protein expression and attenuated endotoxemia in alcohol-fed mice [42]. This finding could

corroborate that improvement of dysbiosis contributes to preventing the disruption of the gut barrier function.

Gut-derived endotoxins from gut bacteria play a crucial role in the pathogenesis of alcoholic liver injury [43,44]. When endotoxins enter into the liver, they initially activate KCs, which are regional macrophages in the liver, via binding of TLR4 on their membrane, leading to an induction of inflammatory cytokines and chemokines including TNF- α , IL-1, and IL-6, mediated by the activation of the nuclear factor– κ B pathway (Figure 2) [45]. In addition, alcohol induces the LPS binding protein, which is closely related to the effect of endotoxins. LPS–TLR4 binding on the surface of KCs also induces reactive oxygen species (ROS) production, which in turn induces migration of T lymphocytes and neutrophils and HSC activation [1,2]. Likewise, because nonimmune cells such as HSCs and liver sinusoidal endothelial cells have TLR4, the exposure of endotoxins to their cells further promotes fibrotic activity via the activation of these cells [4].

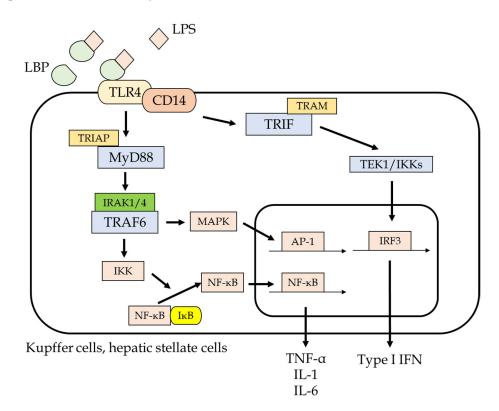


Figure 2. Intracellular signaling of the TLR4 pathway via LPS binding. Liver cells such as Kupffer cells and hepatic stellate cells contain TLR4, which mediates the innate immunity response. LPS binding to LPS binding protein, which is a translocator for LPS, attaches to TLR4 and stimulates its downstream signal, including NF- κ B and mitogen-activated protein kinase, which induces the transcription of proinflammatory cytokines and chemokines such as TNF- α , IL-1, and IL-6. These inflammatory cytokines cause liver inflammation as well as activating hepatic stellate cells, leading to excessive production of the extracellular matrix, which suggests that chronic exposure of LPS can result in the progression of liver fibrosis, culminating in liver cirrhosis.

3.2. NASH

Nonalcoholic fatty liver disease (NAFLD) is considered to be the manifestation of metabolic syndrome in the liver. Recently, it has been suggested that the definition and term NAFLD itself should be changed to "metabolic-associated fatty liver disease" [46]. Various studies have indicated that the gut microbiota are closely associated with the pathogenesis of the development of NASH [47].

As compared with lean NAFLD individuals, NAFLD patients with obesity have a tendency to have SIBO and leaky gut. An initial preliminary study demonstrated that a

greater population of *Firmicutes* and a smaller population of *Bacteroidetes* were observed in patients with obesity as compared with healthy controls [48]. It was then established that the *Firmicutes/Bacteroidetes* ratio was increased in the obese population in both rodents and humans [49,50]. A greater Firmicutes/Bacteroidetes ratio and increased Proteobacteria were associated with negative health effects, such as induction of systemic inflammatory activity by the increased gut permeability [51]. Miele et al. first reported that the expression of the TJ protein ZO-1 was decreased in patients with NAFLD as compared with healthy subjects [52]. A higher serum level of endotoxins was observed not only in adult NASH patients but also in children with NASH as compared with those without NASH [53-55]. Patients with NASH also displayed a higher expression level of TLR4 in the liver than in those without NASH [56]. The involvement of TLR4 signaling has also been reported in a study demonstrating the relationship between TLR4 mutation and NAFLD [55]. An experimental study using obese mice demonstrated that increased intestinal permeability via the downregulation of occludins and ZO-1 on the intestinal epithelium is closely associated with portal endotoxemia as well as the elevation of serum inflammatory cytokine levels in obese mice [57]. In obese rats, the liver has an elevated sensitivity against LPS exposure and a reduction in the phagocytic function of KCs [58]. Moreover, a high-fat diet also promotes the translocation of bacterial products including living bacteria through the intestinal mucosa, suggesting the importance of the gut-liver axis in liver injury in NAFLD patients [59].

To date, various studies have already demonstrated the importance of the gut–liver axis and the role of TLR4 signaling on the progression of NAFLD. It was previously reported that mice fed a methionine choline-deficient diet demonstrated steatohepatitis, portal endotoxemia, and elevation of TLR4 expression in the liver, whereas TLR4 mutant mice showed less tissue damage and lipid accumulation in the liver [60]. Our study demonstrated that rats fed a choline-deficient L-amino acid-defined diet, which mimics the NASH liver, showed greater α -smooth muscle actin expression and enhanced LPS binding protein mRNA levels in the liver tissue, increased intestinal permeability, and decreased TJ protein expression in the intestine [61]. In contrast, oral medication with poorly absorbable antibiotics inhibited LPS–TLR4 signaling and suppressed the progression of liver fibrosis [61]. This evidence supports the idea that the prevention of LPS stimulation could be a useful therapeutic strategy for patients with CLD.

3.3. Viral Hepatitis

Hepatitis A and E viral infections cause acute hepatitis, which can be self-clearing. Both A and E hepatitis viruses are transmitted through the mouth into the gastrointestinal tract and may affect the diversity of the gut microbiota. Although supplementation of beneficial bacteria such as *Enterococcus faecium* was shown to contribute to the removal of the hepatitis E virus from the intestine of pigs, this effect remains unclear in humans [62].

In contrast, hepatitis B and C viruses are major viruses that cause chronic hepatitis. It is also observed that patients with chronic hepatitis have higher bacterial translocation from the gut [63]. It was recently reported that commensal microbiota play an important role in both the viral host cell interaction and viral replication. Some bacterial species including *Neisseria, E. coli, Enterobacteriaceae, E. faecalis, F. prausnitzii,* and *Gemella* have been identified as having a responsible role in the progression of hepatitis B and C infection [64,65].

3.3.1. Hepatitis B Viral Infection

Dysbiosis can influence the progression of disease pathogenesis, resulting in liver failure. In patients with HBV, bacteria producing LPS are enriched. It has been demonstrated that the proportions of *F. prausnitzii*, *E. faecalis*, *Enterobacteriaceae*, *Bifidobacteria*, and *Lactobacillus* were markedly changed in HBV cirrhotic patients [66]. Oral dysbiosis has also been reported during HBV infection, as a reduction in *Bacteroidetes* and an increase in *Proteobacteria*. Another study suggested the positive correlation of *Neisseriaceae* with the level of serum HBV DNA [67].

In contrast, Lu et al. reported that cirrhotic HBV patients exhibited a marked decrease in the ratio of *Bifidobacteriaceae* to *Enterobacteriaceae*. Another study of cirrhotic patients with HBV also found a decrease in *Bifidobacteria* and *Lactobacillus*, whereas the levels of *Enterococcus* and *Enterobacteriaceae* were significantly increased as compared with the healthy population.

3.3.2. Hepatitis C Viral Infection

In most patients with hepatitis C virus (HCV), the amount of *Enterobacteriaceae* and *Bacterioidetes* increased, whereas that of *Firmicutes* decreased. Previous studies found that plasma levels of LPS in HCV patients are elevated due to the promotion of bacterial translocation and intestinal inflammation [68,69]. As the production of bile acids is important for the gut microbial composition [70], several pathogenic bacteria including *Enterobacteriaceae*, *Enterococcus*, and *Staphylococcus* prevent the production of bile acid in patients with HCV, and the use of oral direct-acting antivirals reversed the alteration in the gut microbiota. That study further revealed that oral direct-acting antivirals could be helpful in improving the gut microflora through a reduction in intestinal inflammation via mediating the TNF- α level [71].

Several beneficial bacteria have been reported to have a protective effect during HCV infection. These bacteria including *Bifidobacterium* spp. and *L. acidophilus* play a supportive role in the antiviral effect, even in antibacterial activities [72]. The immune response by natural killer cells can be extended by protective bacteria such as probiotics in HCV-infected patients, and they also promote the cytotoxic effect of natural killer cells to inhibit HCV replication [73,74].

3.4. Autoimmune Liver Diseases

Primary biliary cholangitis (PBC) is one of the common autoimmune liver diseases characterized by hepatobiliary injury, which shows progressive nonsuppurative destruction of small intrahepatic bile ducts, resulting in cholestasis, inflammation, and fibrosis. An increased diversity of *Eubacterium* and *Veillonella* and a decreased diversity of *Fusobacterium* in the oral microbiota were demonstrated in Japanese patients with PBC as compared with healthy individuals [75]. This report also demonstrated that an increase in *Veillonella* is positively correlated with IL-1 β and IL-8 levels and the relative abundance of *Lactobacillales* in feces [75].

In a Chinese study of patients with PBC, *Acidobacteria*, *Lachnobacterium* sp., *Bacteroides eggerthii*, and *Ruminococcus bromii*, which are considered to be potentially beneficial bacteria, were reported to be decreased as compared with healthy controls, whereas pathogenic bacteria including *Enterobacteriaceae*, *Neisseriaceae*, *Veillonella*, *Actinobacillus pleuropneumoniae*, and *Haemophilus parainfluenzae* were increased in the feces of these patients [76]. Another study of PBC patients using 16S rRNA sequence analyses derived from fecal microbiota indicated a decrease in *Bacteroidetes* and an increase in *Fusobacteria* and *Proteobacteria* spp. [77]. Our recent study using the 16S rDNA sequence also revealed that bacterial diversity was lower in PBC patients with a decreased abundance of *Clostridium* and an increase in *Lactobacillus* [78]. As ursodeoxycholic acid (UDCA) is a common medication for PBC, we also investigated the difference in the microbial alteration between UDCA responders and nonresponders. We found a smaller population of the genus *Faecalibacterium* in the UDCA nonresponder group as compared with the UDCA responder group [78].

Autoimmune hepatitis (AIH) is another major autoimmune liver disease, but the pathogenesis of AIH still remains unclear. The first clinical study from Lin et al. revealed that the decreased abundance of *Bifidobacteria* and *Lactobacillus* is involved in the development of AIH, along with increasing plasma LPS levels in the later stage of AIH [79]. A recent study reported that as compared with healthy controls, AIH patients had a reduced diversity of intestinal microbiota and a change in bacteria species including *Streptococcus*, *Veillonella*, *Klebsiella*, and *Lactobacillus* [80]. Another study also reported that AIH patients

demonstrated augmented intestinal permeability and bacterial translocation, which were associated with the severity of AIH activity, but not the stage of fibrosis [81].

In the experimental AIH model, germ-free mice demonstrated a much smaller amount of inflammatory cytokines and chemokines, including TNF- α , IL-4, interferon- γ , monocyte chemotactic protein-1, C-X-C motif chemokine 1, granulocyte colony-stimulating factor, and eotaxin. Concanavalin A (ConA)-induced apoptosis of liver cells was also significantly prevented in germ-free BALB/c mice as compared with positive controls [82]. Moreover, in conventional mice with ConA injection, plasma LPS levels were significantly higher than those in germ-free mice with ConA [82]. These data might explain the crucial role of LPS in ConA-induced liver injury in the mimicry of the pathogenesis of AIH.

3.5. Liver Cirrhosis and Its Complications

Liver cirrhosis is the end stage (F4) of liver fibrosis, which is characterized by the conversion of the normal liver architecture and regenerative nodules surrounded by tissue fibrosis and liver dysfunction [83]. Its complications, including ascites, varices, spontaneous bacterial peritonitis (SBP), and HCC, often require management during the clinical course, as they can result in death. As patients with liver cirrhosis are basically in an immunosuppressive state, infections can increase the rate of morbidity and mortality in cirrhotic patients [84]. An increase in the incidence of systemic endotoxemia in patients with liver cirrhosis has already been demonstrated using the Limulus amebocyte lysate test [85]. Likewise, subsequent quantitative endotoxin assays have revealed an elevation in plasma endotoxin levels along with liver fibrosis development to liver cirrhosis [86,87]. Another study reported the endotoxin levels in the portal blood flow of cirrhotic patients, suggesting that systemic endotoxemia is caused by the increased translocation of endotoxins from the gut via gut barrier dysfunction [88]. Taking into consideration the accumulating evidence, endotoxemia via leaky gut appears to be closely associated not only with SBP and liver fibrosis but also with the hyperdynamic state as well as renal, cardiac, pulmonary, and coagulation disturbances [1]. The disruption of the gut barrier is considered to be a pathological factor that can exaggerate liver cirrhosis and its complications [89]. Increased intestinal permeability is reported to be associated with the Child-Pugh classification, which indicates the stage of liver cirrhosis [90–92]. Thus, it is important to investigate the change in gut microbial diversity because of the key roles of bacterial translocation and endotoxemia in the pathogenesis of complications resulting from liver cirrhosis [93].

3.5.1. Ascites and SBP

SBP is one of the most important infections in cirrhotic patients with ascitic fluid, which might be caused by bacterial translocation from the intestine. It has been reported that enteric Gram-negative bacteria, mainly the *Enterobacteriaceae* family, comprise the vast majority of causative microorganisms of SBP [94]. The impairment of small intestinal motility and SIBO are frequently observed in patients with liver cirrhosis with a history of SBP [95]. Bacterial DNA has also been identified as a factor associated with proinflammatory cytokine levels in ascites [96]. The presence of bacterial DNA in ascites and blood was observed in one third of cirrhotic patients with culture-negative ascites, which could worsen intrahepatic endothelial function and peripheral vasodilation [97,98]. Proton pump inhibitors (PPIs) are potent inducers of increased bacterial translocation, which might be associated with the promotion of intestinal overgrowth. Cirrhotic patients who use PPIs have demonstrated an increase in the prevalence of SBP [99]. The evidence from these studies shows that bacterial overgrowth could increase the risk of SBP via bacterial translocation resulting from intestinal hyperpermeability [100,101].

3.5.2. Portal Hypertension

Along with the development of liver fibrosis, the portal blood pressure is elevated in patients with CLD as a result of the elevation in liver stiffness. As mentioned above, cirrhotic patients have an impaired gut barrier, resulting in bacterial translocation. An experimental study demonstrated that intraperitoneal injection of LPS can elevate the portal pressure and variceal bleeding, and increased portal hypertension can increase intestinal permeability [102–104]. Bacterial translocation induces the production of not only proinflammatory cytokines but also nitric oxide, endothelin, and cyclooxygenases [105–107]. These products also impair the contractility of the mesenteric vessels, which further exaggerates the elevation in the portal blood pressure [108,109].

Clinically, several studies, including our current study, have demonstrated that cirrhotic patients experience an elevation in plasma endotoxin levels and increased intestinal permeability after variceal hemorrhage [110,111]. Moreover, the results of a multivariate statistical analysis revealed that excessive intestinal permeability was an independent risk factor for infections such as SBP [110,112]. In these patients, poorly unabsorbable antibiotics including rifaximin could be useful for preventing SBP and the re-rupture of gastrointestinal varices [113,114]. The accumulating evidence suggests that there is a malignant circulation that induces portal hypertension via LPS stimulation through the gut–liver axis.

3.5.3. Hepatic Encephalopathy

Hepatic encephalopathy (HE) is a major complication of end-stage liver cirrhosis, which induces symptoms due to an abnormal elevation in plasma ammonia levels, leading to confusion, lethargy, sleep disturbances, and coma [115]. An experimental animal model using germ-free mice in the mimicry of liver cirrhosis displayed a lower concentration of serum ammonia and reduced the level of neuroinflammation as compared with conventional mice [116]. This study also showed that the enrichment of *Lactobacillae* is positively associated with increased neuroinflammation in conventional mice with liver cirrhosis. In addition, LPS permeabilizes the blood–brain barrier in the same manner as the gut barrier and affects the brain microglia through the production of nitric oxide, which induces the swelling of astrocytes in patients with HE [117,118]. Likewise, an association was reported between endotoxemia and minimal HE and an increase in the incidence of overt HE [117].

There is no significant difference in the profiles of fecal microbiota between cirrhotic patients with HE and those without HE [12,119]. However, the diversity of *Alcaligeneceae* in HE patients was shown to be increased as compared with normal controls. *Alcaligeneceae* is reported to contribute to the production of ammonia, and this change was observed only among most HE patients taking PPIs. In contrast, none of the healthy controls taking PPIs demonstrated an increase in this alteration [12]. It has also been demonstrated that SIBO is closely correlated with minimal HE [120]. Bajaj et al. reported a higher abundance of fecal *Enterobacteriaceae*, which is a major bacterium that produces a large amount of endotoxins [121].

3.6. HCC

Changes in the gut microbiota affect the hepatic tumor environment. Hepatic inflammation caused by leaky gut can drive tumorigenesis in the liver. The results of several animal experiments have suggested that LPS–TLR4 signaling can induce the promotion of HCC development [122–124]. It has also been reported that HCC model mice treated with antibiotics and germ-free mice demonstrated less carcinogenesis. In this study, TLR4 signaling in the liver cells was partially mediated by the inhibition of hepatocyte apoptosis and growth signals, including epiregulin derived from HSCs [124]. Dysbiosis-induced bacterial metabolites, such as deoxycholic acid, are additional factors that promote hepatocarcinogenesis related to gut-derived endotoxemia, which could enhance liver inflammation via TLR pathways [125].

In clinical studies, a higher abundance of intestinal *E. coli* was found to be associated with the presence of HCC and reported to produce endotoxins [20]. Animal studies have also indicated that endotoxins play an important role in the progression of HCC [126]. These studies further suggested that an increase in fecal *Bacteroides* is probably accompanied by an elevation in proinflammatory cytokine levels such as IL-8 and IL-13, activated

monocytes, and monocytic myeloid-derived suppressor cells, which play a role in the hepatocarcinogenesis of NAFLD patients [7]. *Klebsiella-* and *Haemophilus*-producing LPS are also reported to be increased in patients with HCC in the early stage as compared with non-HCC controls [19,127,128]. This growing evidence may suggest the possibility of a noninvasive predictive marker for the detection of HCC incidence in the future.

4. Conclusions

Various studies have revealed that gut-derived endotoxins from the gut microbiota could be closely associated with CLDs. Lifestyle factors, such as dietary factors and alcohol drinking, are fundamental therapeutic targets for preventing the pathogenesis of liver diseases in terms of the gut–liver axis. Dysbiosis and leaky gut are induced by alcohol and a high-calorie/high-fat diet, which results in liver inflammation and liver fibrosis progression, eventually progressing to liver cirrhosis, especially in patients with alcoholic liver disease and NAFLD. As mentioned above, it is already known that endotoxemia is associated with the occurrence of cirrhotic complications in patients with liver cirrhosis via activation of TLR4 signaling in KCs and HSCs. This gut–liver axis resulting from gut dysbiosis might also induce the development of HCC. Considering these findings, the gut microbiota and gut-derived endotoxemia could be useful therapeutic targets in the management of the progression of liver pathogenesis, and further investigation in this field is expected in the future.

Funding: This research received no external funding.

Conflicts of Interest: The authors declare no conflict of interest for this article.

References

- 1. Fukui, H. Gut-liver axis in liver cirrhosis: How to manage leaky gut and endotoxemia. *World J. Hepatol.* **2015**, *7*, 425–442. [CrossRef] [PubMed]
- 2. Guo, J.; Friedman, S.L. Toll-like receptor 4 signaling in liver injury and hepatic fibrogenesis. *Fibrogenes Tissue Repair* **2010**, *3*, 21. [CrossRef] [PubMed]
- 3. Seki, E.; Schnabl, B. Role of innate immunity and the microbiota in liver fibrosis: Crosstalk between the liver and gut. *J. Physiol.* **2012**, *590*, 447–458. [CrossRef]
- Seo, Y.S.; Shah, V.H. The role of gut-liver axis in the pathogenesis of liver cirrhosis and portal hypertension. *Clin. Mol. Hepatol.* 2012, *18*, 337–346. [CrossRef]
- 5. Pasolli, E.; Truong, D.T.; Malik, F.; Waldron, L.; Segata, N. Machine Learning Meta-analysis of Large Metagenomic Datasets: Tools and Biological Insights. *PLoS Comput. Biol.* 2016, 12, e1004977. [CrossRef] [PubMed]
- 6. Qin, N.; Yang, F.; Li, A.; Prifti, E.; Chen, Y.; Shao, L.; Guo, J.; Le Chatelier, E.; Yao, J.; Wu, L.; et al. Alterations of the human gut microbiome in liver cirrhosis. *Nature* **2014**, *513*, 59–64. [CrossRef] [PubMed]
- Ponziani, F.R.; Bhoori, S.; Castelli, C.; Putignani, L.; Rivoltini, L.; Del Chierico, F.; Sanguinetti, M.; Morelli, D.; Sterbini, F.P.; Petito, V.; et al. Hepatocellular Carcinoma Is Associated with Gut Microbiota Profile and Inflammation in Nonalcoholic Fatty Liver Disease. *Hepatology* 2019, 69, 107–120. [CrossRef]
- 8. Loomba, R.; Seguritan, V.; Li, W.; Long, T.; Klitgord, N.; Bhatt, A.; Dulai, P.S.; Caussy, C.; Bettencourt, R.; Highlander, S.K.; et al. Gut microbiome-based metagenomic signature for non-invasive detection of advanced fibrosis in human nonalcoholic fatty liver disease. *Cell Metab.* **2017**, *25*, e1055. [CrossRef]
- 9. Caussy, C.; Tripathi, A.; Humphrey, G.; Bassirian, S.; Singh, S.; Faulkner, C.; Bettencourt, R.; Rizo, E.; Richards, L.; Xu, Z.Z.; et al. A gut microbiome signature for cirrhosis due to nonalcoholic fatty liver disease. *Nat. Commun.* **2019**, *10*, 1–9. [CrossRef]
- 10. Xu, M.; Wang, B.; Fu, Y.; Chen, Y.; Yang, F.; Lu, H.; Chen, Y.; Xu, J.; Li, L. Changes of fecal Bifidobacterium species in adult patients with hepatitis B virus-induced chronic liverdisease. *Microb. Ecol.* **2012**, *63*, 304–313. [CrossRef]
- Wu, Z.-W.; Lu, H.-F.; Wu, J.; Zuo, J.; Chen, P.; Sheng, J.-F.; Zheng, S.-S.; Li, L.-J. Assessment of the Fecal Lactobacilli Population in Patients with Hepatitis B Virus-Related Decompensated Cirrhosis and Hepatitis B Cirrhosis Treated with Liver Transplant. *Microb. Ecol.* 2012, 63, 929–937. [CrossRef] [PubMed]
- 12. Bajaj, J.S.; Ridlon, J.M.; Hylemon, P.B.; Thacker, L.R.; Heuman, D.M.; Smith, S.; Sikaroodi, M.; Gillevet, P.M. Linkage of gut microbiome with cognition in hepatic encephalopathy. *Am. J. Physiol. Liver Physiol.* 2012, 302, G168–G175. [CrossRef] [PubMed]
- 13. Chakraborti, C.K. New-found link between microbiota and obesity. *World J. Gastrointest. Pathophysiol.* **2015**, *6*, 110–119. [CrossRef] [PubMed]
- Dubinkina, V.B.; Tyakht, A.V.; Odintsova, V.; Yarygin, K.S.; Kovarsky, B.A.; Pavlenko, A.V.; Ischenko, D.S.; Popenko, A.S.; Alexeev, D.G.; Taraskina, A.Y.; et al. Links of gut microbiota composition with alcohol dependence syndrome and alcoholic liver disease. *Microbiome* 2017, 5, 1–14. [CrossRef] [PubMed]

- Kakiyama, G.; Hylemon, P.B.; Zhou, H.; Pandak, W.M.; Heuman, D.M.; Kang, D.J.; Takei, H.; Nittono, H.; Ridlon, J.M.; Fuchs, M.; et al. Colonic inflammation and secondary bile acids in alcoholic cirrhosis. *Am. J. Physiol. Liver Physiol.* 2014, 306, G929–G937. [CrossRef]
- 16. Drasar, B.S.; Shiner, M. Studies on the intestinal flora: Part II Bacterial flora of the small intestine in patients with gastrointestinal disorders. *Gut* **1969**, *10*, 812–819. [CrossRef]
- 17. Unno, N.; Wang, H.; Menconi, M.J.; Tytgat, S.H.; Larkin, V.; Smith, M.; Morin, M.J.; Chavez, A.; Hodin, R.A.; Fink, M.P. Inhibition of inducible nitric oxide synthase ameliorates endotoxin-induced gut mucosal barrier dysfunction in rats. *Gastroenterology* **1997**, *113*, 1246–1257. [CrossRef]
- Zolotarevsky, Y.; Hecht, G.; Koutsouris, A.; Gonzalez, D.E.; Quan, C.; Tom, J.; Mrsny, R.J.; Turner, J.R. A membrane-permeant peptide that inhibits MLC kinase restores barrier function in in vitro models of intestinal disease. *Gastroenterology* 2002, 123, 163–172. [CrossRef]
- 19. Ren, Z.; Li, A.; Jiang, J.; Zhou, L.; Yu, Z.; Lu, H.; Xie, H.; Chen, X.; Shao, L.; Zhang, R.; et al. Gut microbiome analysis as a tool towards targeted non-invasive biomarkers for early hepatocellular carcinoma. *Gut* **2019**, *68*, 1014–1023. [CrossRef]
- Grąt, M.; Wronka, K.M.; Krasnodębski, M.; Masior, Ł.; Lewandowski, Z.; Kosińska, I.; Grąt, K.; Stypułkowski, J.; Rejowski, S.; Wasilewicz, M.; et al. Profile of gut microbiota associated with the presence of hepatocellular cancer in patients with liver cirrhosis. *Transplant. Proc.* 2016, 48, 1687–1691. [CrossRef]
- Van Thiel, D.H.; Fagiuoli, S.; Wright, H.I.; Chien, M.C.; Gavaler, J.S. Gastrointestinal transit in cirrhotic patients: Effect of hepatic encephalopathy and its treatment. *Hepatology* 1994, 19, 67–71. [CrossRef] [PubMed]
- 22. Chesta, J.; Defilippi, C. Abnormalities in proximal small bowel motility in patients with cirrhosis. *Hepatology* **1993**, *17*, 828–832. [PubMed]
- 23. Lombardo, L.; Foti, M.; Ruggia, O.; Chiecchio, A. Increased Incidence of Small Intestinal Bacterial Overgrowth during Proton Pump Inhibitor Therapy. *Clin. Gastroenterol. Hepatol.* **2010**, *8*, 504–508. [CrossRef]
- 24. Farquhar, M.G.; Palade, G.E. JUNCTIONAL COMPLEXES IN VARIOUS EPITHELIA. J. Cell Biol. 1963, 17, 375–412. [CrossRef]
- Van Itallie, C.M.; Anderson, J.M. Architecture of tight junctions and principles of molecular composition. *Semin. Cell Dev. Biol.* 2014, 36, 157–165. [CrossRef]
- Simon, D.B.; Lu, Y.; Choate, K.A.; Vekazquez, H.; Al-Sabban, E.; Praga, M.; Casari, G.; Bettinelli, A.; Colussi, G.; Rodriguez-Soriano, J.; et al. Paracellin-1, a renal tight junction protein required for paracellular Mg2b resorption. *Science* 1999, 285, 103–106. [CrossRef]
- 27. Shen, L.; Weber, C.R.; Raleigh, D.R.; Yu, D.; Turner, J.R. Tight Junction Pore and Leak Pathways: A Dynamic Duo. *Annu. Rev. Physiol.* **2011**, *73*, 283–309. [CrossRef]
- Luissint, A.-C.; Parkos, C.A.; Nusrat, A. Inflammation and the Intestinal Barrier: Leukocyte–Epithelial Cell Interactions, Cell Junction Remodeling, and Mucosal Repair. *Gastroenterology* 2016, 151, 616–632. [CrossRef] [PubMed]
- Hiippala, K.; Jouhten, H.; Ronkainen, A.; Hartikainen, A.; Kainulainen, V.; Jalanka, J.; Satokari, R. The Potential of Gut Commensals in Reinforcing Intestinal Barrier Function and Alleviating Inflammation. *Nutrients* 2018, 10, 988. [CrossRef]
- 30. Hendrikx, T.; Schnabl, B. Antimicrobial proteins: Intestinal guards to protect against liver disease. *J. Gastroenterol.* **2018**, *54*, 209–217. [CrossRef] [PubMed]
- Tripathi, A.; Debelius, J.; Brenner, D.A.; Karin, M.; Loomba, R.; Schnabl, B.; Knight, R. The gut–liver axis and the intersection with the microbiome. *Nat. Rev. Gastroenterol. Hepatol.* 2018, *15*, 397–411. [CrossRef] [PubMed]
- 32. Wang, H.J.; Gao, B.; Zakhari, S.; Nagy, L.E. Inflammation in Alcoholic Liver Disease. *Annu. Rev. Nutr.* 2012, 32, 343–368. [CrossRef]
- Gao, B.; Bataller, R. Alcoholic Liver Disease: Pathogenesis and New Therapeutic Targets. *Gastroenterology* 2011, 141, 1572–1585. [CrossRef] [PubMed]
- 34. Hartmann, P.; Seebauer, C.T.; Schnabl, B. Alcoholic Liver Disease: The Gut Microbiome and Liver Cross Talk. *Alcohol. Clin. Exp. Res.* **2015**, *39*, 763–775. [CrossRef] [PubMed]
- 35. Bode, J.C.; Bode, C.; Heidelbach, R.; Dürr, H.K.; Martini, G.A. Jejunal microflora in patients with chronic alcohol abuse. *Hepatogastroenterology* **1984**, *31*, 30–34.
- 36. Morencos, F.C.; de las Heras Castano, G.; Martin Ramos, L.; López Arias, M.J.; Ledesma, F.; Romero, F.P. Small bowel bacterial over-growth in patients with alcoholic cirrhosis. *Dig. Dis. Sci.* **1995**, *40*, 1252–1256. [CrossRef]
- Hartmann, P.; Chen, P.; Wang, H.J.; Wang, L.; McCole, D.F.; Brandl, K.; Stärkel, P.; Belzer, C.; Hellerbrand, C.; Tsukamoto, H.; et al. De-ficiency of intestinal mucin-2 ameliorates experimental alcoholic liver disease in mice. *Hepatology* 2013, 58, 108–119. [CrossRef]
- Vassallo, G.; Mirijello, A.; Ferrulli, A.; Antonelli, M.; Landolfi, R.; Gasbarrini, A.; Addolorato, G. Review article: Alcohol and gut microbiota–the possible role of gut microbiota modulation in the treatment of alcoholic liver disease. *Aliment. Pharmacol. Ther.* 2015, 41, 917–927. [CrossRef]
- 39. Fukui, H.; Brauner, B.; Bode, J.; Bode, C. Plasma endotoxin concentrations in patients with alcoholic and non-alcoholic liver disease: Reevaluation with an improved chromogenic assay. *J. Hepatol.* **1991**, *12*, 162–169. [CrossRef]
- Leclercq, S.; Matamoros, S.; Cani, P.D.; Neyrinck, A.; Jamar, F.; Stärkel, P.; Windey, K.; Tremaroli, V.; Bäckhed, F.; Verbeke, K.; et al. Intestinal permeability, gut-bacterial dysbiosis, and behavioral markers of alcohol-dependence severity. *Proc. Natl. Acad. Sci. USA* 2014, 111, E4485–E4493. [CrossRef] [PubMed]

- Wang, Y.; Kirpich, I.; Liu, Y.; Ma, Z.; Barve, S.; McClain, C.J.; Feng, W. Lactobacillus rhamnosus GG Treatment Potentiates Intestinal Hypoxia-Inducible Factor, Promotes Intestinal Integrity and Ameliorates Alcohol-Induced Liver Injury. *Am. J. Pathol.* 2011, 179, 2866–2875. [CrossRef] [PubMed]
- 42. Bull-Otterson, L.; Feng, W.; Kirpich, I.; Wang, Y.; Qin, X.; Liu, Y.; Gobejishvili, L.; Joshi-Barve, S.; Ayvaz, T.; Petrosino, J.; et al. Meta-genomic analyses of alcohol induced pathogenic alterations in the intestinal microbiome and the effect of Lactobacillus rhamnosus GG treatment. *PLoS ONE* **2013**, *8*, e53028. [CrossRef] [PubMed]
- 43. Mutlu, E.A.; Gillevet, P.M.; Rangwala, H.; Sikaroodi, M.; Naqvi, A.; Engen, P.A.; Kwasny, M.; Lau, C.K.; Keshavarzian, A. Colonic mi-crobiome is altered in alcoholism. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2012**, 302, G966–G978. [CrossRef] [PubMed]
- 44. Woodhouse, C.A.; Patel, V.C.; Singanayagam, A.; Shawcross, D.L. Review article: The gut microbiome as a therapeutic target in the pathogenesis and treatment of chronic liver disease. *Aliment. Pharmacol. Ther.* **2018**, *47*, 192–202. [CrossRef] [PubMed]
- 45. Szabo, G. Gut–Liver Axis in Alcoholic Liver Disease. Gastroenterology 2015, 148, 30–36. [CrossRef]
- Eslam, M.; Newsome, P.N.; Sarin, S.K.; Anstee, Q.M.; Targher, G.; Romero-Gomez, M.; Zelber-Sagi, S.; Wong, V.W.-S.; Dufour, J.-F.; Schattenberg, J.M.; et al. A new definition for metabolic dysfunction-associated fatty liver disease: An international expert consensus statement. *J. Hepatol.* 2020, 73, 202–209. [CrossRef]
- 47. Ilan, Y. Leaky gut and the liver: A role for bacterial translocation in nonalcoholic steatohepatitis. *World J. Gastroenterol.* **2012**, *18*, 2609–2618. [CrossRef]
- Ley, R.E.; Turnbaugh, P.J.; Klein, S.; Gordon, J.I. Microbial ecology: Human gut microbes associated with obesity. *Nature* 2006, 444, 1022–1023. [CrossRef]
- 49. Clarke, S.F.; Murphy, E.F.; Nilaweera, K.; Ross, P.R.; Shanahan, F.; O'Toole, P.W.; Cotter, P.D. The gut microbiota and its relationship to diet and obesity: New insights. *Gut Microbes* **2012**, *3*, 186–202. [CrossRef]
- 50. Turnbaugh, P.J.; Ley, R.E.; Mahowald, M.A.; Magrini, V.; Mardis, E.R.; Gordon, J.I. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nat. Cell Biol.* **2006**, 444, 1027–1031. [CrossRef]
- 51. Panasevich, M.R.; Peppler, W.T.; Oerther, D.; Wright, D.C.; Rector, R.S. Microbiome and NAFLD: Potential influence of aerobic fitness and lifestyle modification. *Physiol. Genom.* **2017**, *49*, 385–399. [CrossRef]
- 52. Miele, L.; Valenza, V.; La Torre, G.; Montalto, M.; Cammarota, G.; Ricci, R.; Mascianà, R.; Forgione, A.; Gabrieli, M.L.; Perotti, G.; et al. Increased intestinal permeability and tight junction alterations in nonalcoholic fatty liver disease. *Hepatology* **2009**, *49*, 1877–1887. [CrossRef]
- 53. Alisi, A.; Manco, M.; De Vito, R.; Piemonte, F.; Nobili, V. Endotoxin and Plasminogen Activator Inhibitor-1 Serum Levels Associated With Nonalcoholic Steatohepatitis in Children. *J. Pediatr. Gastroenterol. Nutr.* **2010**, *50*, 645–649. [CrossRef]
- 54. Farhadi, A.; Gundlapalli, S.; Shaikh, M.; Frantzides, C.; Harrell, L.; Kwasny, M.M.; Keshavarzian, A. Susceptibility to gut leakiness: A possible mechanism for endotoxaemia in non-alcoholic steatohepatitis. *Liver Int.* **2008**, *28*, 1026–1033. [CrossRef] [PubMed]
- 55. Sharifnia, T.; Antoun, J.; Verriere, T.G.C.; Suarez, G.; Wattacheril, J.; Wilson, K.T.; Peek, R.M.; Abumrad, N.N.; Flynn, C.R. Hepatic TLR4 signaling in obese NAFLD. *Am. J. Physiol. Liver Physiol.* 2015, 309, G270–G278. [CrossRef] [PubMed]
- 56. Kapil, S.; Duseja, A.; Sharma, B.K.; Singla, B.; Chakraborti, A.; Das, A.; Ray, P.; Dhiman, R.K.; Chawla, Y. Small intestinal bacterial overgrowth and toll-like receptor signaling in patients with non-alcoholic fatty liver disease. *J. Gastroenterol. Hepatol.* 2015, 31, 213–221. [CrossRef] [PubMed]
- 57. Brun, P.; Castagliuolo, I.; Di Leo, V.; Buda, A.; Pinzani, M.; Palù, G.; Martines, D. Increased intestinal permeability in obese mice: New evidence in the pathogenesis of nonalcoholic steatohepatitis. *Am. J. Physiol. Liver Physiol.* 2007, 292, G518–G525. [CrossRef] [PubMed]
- 58. Yang, S.Q.; Lin, H.Z.; Lane, M.D.; Clemens, M.; Diehl, A.M. Obesity increases sensitivity to endotoxin liver injury: Implications for the pathogenesis of steatohepatitis. *Proc. Natl. Acad. Sci. USA* **1997**, *94*, 2557–2562. [CrossRef]
- 59. Aron-Wisnewsky, J.; Gaborit, B.; Dutour, A.; Clement, K. Gut microbiota and non-alcoholic fatty liver disease: New insights. *Clin. Microbiol. Infect.* **2013**, *19*, 338–348. [CrossRef]
- 60. Rivera, C.A.; Adegboyega, P.; van Rooijen, N.; Tagalicud, A.; Allman, M.; Wallace, M. Toll-like receptor-4 signaling and Kupffer cells play pivotal roles in the pathogenesis of non-alcoholic steatohepatitis. *J. Hepatol.* **2007**, *47*, 571–579. [CrossRef]
- 61. Douhara, A.; Moriya, K.; Yoshiji, H.; Noguchi, R.; Namisaki, T.; Kitade, M.; Kaji, K.; Aihara, Y.; Nishimura, N.; Takeda, K.; et al. Reduction of en-dotoxin attenuates liver fibrosis through suppression of hepatic stellate cell activation and remission of intestinal permeability in a rat non-alcoholic steatohepatitis model. *Mol. Med. Rep.* **2015**, *11*, 1693–1700. [CrossRef] [PubMed]
- Kreuzer, S.; Machnowska, P.; Aßmus, J.; Sieber, M.; Pieper, R.; Schmidt, M.F.; Brockmann, G.A.; Scharek-Tedin, L.; Johne, R. Feeding of the probiotic bacterium Enterococcus faecium NCIMB 10415 differentially affects shedding of enteric viruses in pigs. *Veter Res.* 2012, 43, 58. [CrossRef] [PubMed]
- 63. Li, D.K.; Yan, P.; Chung, R.T.; Butt, A.A.; Abou-Samra, A.-B. Proton pump inhibitors are associated with accelerated development of cirrhosis, hepatic decompensation and hepatocellular carcinoma in noncirrhotic patients with chronic hepatitis C infection: Results from ERCHIVES. *Aliment. Pharmacol. Ther.* **2017**, *47*, 246–258. [CrossRef] [PubMed]
- 64. Chen, Y.; Ji, F.; Guo, J.; Shi, D.; Fang, D.; Li, L. Dysbiosis of small intestinal microbiota in liver cirrhosis and its association with etiology. *Sci. Rep.* **2016**, *6*, 34055. [CrossRef]
- 65. Mohamadkhani, A. On the potential role of intestinal microbial community in hepatocarcinogenesis in chronic hepatitis B. *Cancer Med.* **2018**, *7*, 3095–3100. [CrossRef]

- 66. Lu, H.; Wu, Z.; Xu, W.; Yang, J.; Chen, Y.; Li, L. Intestinal Microbiota Was Assessed in Cirrhotic Patients with Hepatitis B Virus Infection. *Microb. Ecol.* 2011, 61, 693–703. [CrossRef]
- 67. Zhao, Y.; Mao, Y.-F.; Tang, Y.-S.; Ni, M.-Z.; Liu, Q.-H.; Wang, Y.; Feng, Q.; Peng, J.-H.; Hu, Y.-Y. Altered oral microbiota in chronic hepatitis B patients with different tongue coatings. *World J. Gastroenterol.* **2018**, *24*, 3448–3461. [CrossRef]
- Dolganiuc, A.; Norkina, O.; Kodys, K.; Catalano, D.; Bakis, G.; Marshall, C.; Mandrekar, P.; Szabo, G. Viral and Host Factors Induce Macrophage Activation and Loss of Toll-Like Receptor Tolerance in Chronic HCV Infection. *Gastroenterology* 2007, 133, 1627–1636. [CrossRef]
- 69. Inoue, T.; Nakayama, J.; Moriya, K.; Kawaratani, H.; Momoda, R.; Ito, K.; Lio, E.; Nojiri, S.; Fujiwara, K.; Yoneda, M.; et al. Gut dysbiosis associated with hepatitis C virus in-fection. *Clin. Infect. Dis.* **2018**, *67*, 869–877. [CrossRef]
- 70. Ponziani, F.R.; Putignani, L.; Sterbini, F.P.; Petito, V.; Picca, A.; Del Chierico, F.; Reddel, S.; Calvani, R.; Marzetti, E.; Sanguinetti, M.; et al. Influence of hepatitis C virus eradication with direct-acting antivirals on the gut microbiota in patients with cirrhosis. *Aliment. Pharmacol. Ther.* **2018**, *48*, 1301–1311. [CrossRef]
- Pérez-Matute, P.; Íñiguez, M.; Villanueva-Millán, M.J.; Recio-Fernández, E.; Vázquez, A.M.; Sánchez, S.C.; Morano, L.E.; Oteo, J.A. Short-term effects of direct-acting antiviral agents on inflammation and gut microbiota in hepatitis C-infected patients. *Eur. J. Intern. Med.* 2019, 67, 47–58. [CrossRef]
- 72. Dore, G.J.; Ward, J.; Thursz, M. Hepatitis C disease burden and strategies to manage the burden (Guest Editors Mark Thursz, Gregory Dore and John Ward). *J. Viral Hepat.* **2014**, *21* (Suppl. 1), 1–4. [CrossRef]
- Doskali, M.; Tanaka, Y.; Ohira, M.; Ishiyama, K.; Tashiro, H.; Chayama, K.; Ohdan, H. Possibility of adoptive immunotherapy with pe-ripheral blood derived CD3–CD56+ and CD3+CD56+ cells for inducing anti-hepatocellular carcinoma and anti-hepatitis C virus activity. *J. Immunother.* 2011, 34, 129–138. [CrossRef] [PubMed]
- 74. Preveden, T.; Scarpellini, E.; Milić, N.; Luzza, F.; Abenavoli, L. Gut microbiota changes and chronic hepatitis C virus infection. *Expert Rev. Gastroenterol. Hepatol.* **2017**, *11*, 813–819. [CrossRef] [PubMed]
- 75. Abe, K.; Takahashi, A.; Fujita, M.; Imaizumi, H.; Hayashi, M.; Okai, K.; Ohira, H. Dysbiosis of oral microbiota and its association with salivary immunological biomarkers in autoimmune liver disease. *PLoS ONE* **2018**, *13*, e0198757. [CrossRef]
- Lv, L.-X.; Fang, D.-Q.; Shi, D.; Chen, D.-Y.; Yan, R.; Zhu, Y.-X.; Chen, Y.-F.; Shao, L.; Guo, F.-F.; Wu, W.-R.; et al. Alterations and correlations of the gut microbiome, metabolism and immunity in patients with primary biliary cirrhosis. *Environ. Microbiol.* 2016, 18, 2272–2286. [CrossRef]
- 77. Tang, R.; Wei, Y.; Li, Y.; Chen, W.; Chen, H.; Wang, Q.; Yang, F.; Miao, Q.; Xiao, X.; Zhang, H.; et al. Gut microbial profile is altered in primary biliary cholangitis and partially restored after UDCA therapy. *Gut* **2018**, *67*, 534–541. [CrossRef]
- 78. Furukawa, M.; Moriya, K.; Nakayama, J.; Inoue, T.; Momoda, R.; Kawaratani, H.; Namisaki, T.; Sato, S.; Douhara, A.; Kaji, K.; et al. Gut dysbiosis associated with clinical prognosis of patients with primary biliary cholangitis. *Hepatol. Res.* 2020, 50, 840–852. [CrossRef]
- 79. Lin, R.; Zhou, L.; Zhang, J.; Wang, B. Abnormal intestinal permeability and microbiota in patients with autoimmune hepatitis. *Int. J. Clin. Exp. Pathol.* **2015**, *8*, 5153–5160.
- 80. Wei, Y.; Li, Y.; Yan, L.; Sun, C.; Miao, Q.; Wang, Q.; Xiao, X.; Lian, M.; Li, B.; Chen, Y.; et al. Alterations of gut microbiome in autoimmune hepatitis. *Gut* 2020, *69*, 569–577. [CrossRef] [PubMed]
- Goeser, F.; Münch, P.; Lesker, T.R.; Lutz, P.L.; Krämer, B.; Kaczmarek, D.J.; Finnemann, C.; Nischalke, H.D.; Geffers, R.; Parcina, M.; et al. Neither black nor white: Do altered intestinal microbiota reflect chronic liver disease severity? *Gut* 2020. [CrossRef]
- 82. Wei, Y.; Zeng, B.; Chen, J.; Cui, G.; Lu, C.; Wu, W.; Yang, J.; Wei, H.; Xue, R.; Bai, L.; et al. Enterogenous bacterial glycolipids are required for the generation of natural killer T cells mediated liver injury. *Sci. Rep.* **2016**, *6*, 36365. [CrossRef]
- 83. Pinzani, M.; Rosselli, M.; Zuckermann, M. Liver cirrhosis. Best Pract. Res. Clin. Gastroenterol. 2011, 25, 281–290. [CrossRef]
- 84. Tandon, P.; Garcia-Tsao, G. Bacterial Infections, Sepsis, and Multiorgan Failure in Cirrhosis. *Semin. Liver Dis.* **2008**, *28*, 026–042. [CrossRef]
- 85. Prytz, H.; Holst-Christensen, J.; Korner, B.; Liehr, H. Portal venous and systemic endotoxemia in patients without liver disease and systemic endotoxemia in patients with cirrhosis. *Scand. J. Gastroenterol.* **1976**, *11*, 857–863. [CrossRef]
- 86. Fukui, H.; Tsujita, S.; Matsumoto, M.; Kitano, H.; Hoppou, K.; Morimura, M.; Takaya, A.; Okamoto, S.; Tsujii, T.; Bode, C. Endotoxemia in chronic hepatitis and cirrhosis: Epiphenomenon or of pathological relevence? In *Gut and the Liver*; Blum, H., Bode, C., Bode, J.C., Sartor, R.B., Eds.; Kluwer Academic Publishers: Dordrecht, The Netherlands, 1998; pp. 251–262.
- Lin, R.S.; Lee, F.Y.; Lee, S.D.; Tsai, Y.T.; Lin, H.C.; Lu, R.H.; Hsu, W.C.; Huang, C.C.; Wang, S.S.; Lo, K.J. Endotoxemia in patients with chronic liver diseases: Relationship to severity of liver diseases, presence of esophageal varices, and hyperdynamic circulation. *J. Hepatol.* 1995, 22, 165–172. [CrossRef]
- 88. Fukui, H. Endotoxin and Other Microbial Translocation Markers in the Blood: A Clue to Understand Leaky Gut Syndrome. *Cell. Mol. Med. Open Access* **2016**, *2*, 3. [CrossRef]
- 89. Kalaitzakis, E. Gastrointestinal dysfunction in liver cirrhosis. World J. Gastroenterol. 2014, 20, 14686–14695. [CrossRef]
- 90. Campillo, B.; Pernet, P.; Bories, P.; Richardet, J.; Devanlay, M.; Aussel, C. Intestinal permeability in liver cirrhosis: Relationship with severe septic complications. *Eur. J. Gastroenterol. Hepatol.* **1999**, *11*, 755–759. [CrossRef]
- 91. Pascual, S.; Such, J.; Esteban, A.; Zapater, P.; Casellas, J.A.; Aparicio, J.R.; Girona, E.; Gutierrez, A.; Carnices, F.; Palazon, J.M. Intestinal permeability is increased in patients with advanced cirrhosis. *Hepatogastroenterology* **2003**, *50*, 1482–1486.

- 92. Scarpellini, E.; Valenza, V.; Gabrielli, M.; Lauritano, E.C.; Perotti, G.; Merra, G.; Lago, A.D.; Ojetti, V.; Ainora, M.E.; Santoro, M.; et al. Intestinal Permeability in Cirrhotic Patients with and Without Spontaneous Bacterial Peritonitis: Is the Ring Closed? *Am. J. Gastroenterol.* **2010**, *105*, 323–327. [CrossRef]
- Bajaj, S.; Heuman, D.M.; Hylemon, P.B.; Sanyal, A.J.; White, M.B.; Monteith, P.; Noble, N.A.; Unser, A.B.; Daita, K.; Fisher, A.R.; et al. The cirrhosis dysbiosis ratio defines changes in the gut microbiome associated with cirrhosis and its complications. *J. Hepatol.* 2014, 60, 940–947. [CrossRef]
- 94. Campillo, B.; Richardet, J.-P.; Kheo, T.; Dupeyron, C. Nosocomial Spontaneous Bacterial Peritonitis and Bacteremia in Cirrhotic Patients: Impact of Isolate Type on Prognosis and Characteristics of Infection. *Clin. Infect. Dis.* **2002**, *35*, 1–10. [CrossRef]
- 95. Chang, C.-S.; Chen, G.-H.; Lien, H.-C.; Yeh, H.-Z. Small intestine dysmotility and bacterial overgrowth in cirrhotic patients with spontaneous bacterial peritonitis. *Hepatology* **1998**, *28*, 1187–1190. [CrossRef]
- 96. Francés, R.; González-Navajas, J.M.; Zapater, P.; Muñoz, C.; Cano, R.; Pascual, S.; Santana, F.; Márquez, D.; Pérez-Mateo, M.; Such, J. Translocation of bacterial DNA from Gram-positive microorganisms is associated with a species-specific inflammatory response in serum and ascitic fluid of patients with cirrhosis. *Clin. Exp. Immunol.* 2007, 150, 230–237. [CrossRef]
- Such, J.; Francés, R.; Muñoz, C.; Zapater, P.; Casellas, J.A.; Cifuentes, A.; Rodriguez-Valera, F.; Pascual, S.; Sola-Vera, J.; Carnicer, F.; et al. Detection and identification of bacterial DNA in patients with cirrhosis and culture-negative, nonneutrocytic ascites. *Hepatology* 2002, *36*, 135–141. [CrossRef]
- Bellot, P.; García-Pagán, J.C.; Francés, R.; Gonzalez-Abraldes, J.; Navasa, M.; Pérez-Mateo, M.; Such, J.; Bosch, J. Bacterial DNA translocation is associated with systemic circulatory abnormalities and intrahepatic endothelial dysfunction in patients with cirrhosis. *Hepatology* 2010, 52, 2044–2052. [CrossRef] [PubMed]
- Goel, G.A.; Deshpande, A.; Lopez, R.; Hall, G.S.; van Duin, D.; Carey, W.D. Increased Rate of Spontaneous Bacterial Peritonitis Among Cirrhotic Patients Receiving Pharmacologic Acid Suppression. *Clin. Gastroenterol. Hepatol.* 2012, 10, 422–427. [CrossRef] [PubMed]
- Aldersley, M.A.; Howdle, P.D. Intestinal permeability and liver disease. *Eur. J. Gastroenterol. Hepatol.* 1999, 11, 401–404. [CrossRef]
 [PubMed]
- 101. Thalheimer, U.; Triantos, C.K.; Samonakis, D.N.; Patch, D.; Burroughs, A.K. Infection, coagulation, and variceal bleeding in cirrhosis. *Gut* 2005, *54*, 556–563. [CrossRef] [PubMed]
- 102. Lumsden, A.B.; Henderson, J.M.; Kutner, M.H. Endotoxin levels measured by a chromogenic assay in portal, hepatic and peripheral venous blood in patients with cirrhosis. *Hepatology* **1988**, *8*, 232–236. [CrossRef] [PubMed]
- 103. Chiva, M.; Guarner, C.; Peralta, C.; Llovet, T.; Gómez, G.; Soriano, G.; Balanzó, J. Intestinal mucosal oxidative damage and bacterial translocation in cirrhotic rats. *Eur. J. Gastroenterol. Hepatol.* **2003**, *15*, 145–150. [CrossRef] [PubMed]
- Clements, W.D.B.; Erwin, P.; McCaigue, M.D.; Halliday, I.; Barclay, G.R.; Rowlands, B.J. Conclusive evidence of endotoxaemia in biliary obstruction. *Gut* 1998, 42, 293–299. [CrossRef]
- 105. Pannen, B.H.; Bauer, M.; Zhang, J.X.; Robotham, J.L.; Clemens, M.G. Endotoxin pretreatment enhances portal venous contractile response to endothelin-1. *Am. J. Physiol. Circ. Physiol.* **1996**, 270, H7–H15. [CrossRef] [PubMed]
- Xu, J.; Cao, H.; Liu, H.; Wu, Z.-Y. Role of nitric oxide synthase and cyclooxygenase in hyperdynamic splanchnic circulation of portal hypertension. *Hepatobiliary Pancreat. Dis. Int.* 2008, 7, 503–508.
- 107. Dong, T.; Aronsohn, A.; Reddy, K.G.; Te, H.S. Rifaximin Decreases the Incidence and Severity of Acute Kidney Injury and Hepatorenal Syndrome in Cirrhosis. *Dig. Dis. Sci.* **2016**, *61*, 3621–3626. [CrossRef]
- 108. Wiest, R.; Das, S.; Cadelina, G.; Garcia-Tsao, G.; Milstien, S.; Groszmann, R.J. Bacterial translocation in cirrhotic rats stimulates eNOS-derived NO production and impairs mesenteric vascular contractility. *J. Clin. Investig.* **1999**, *104*, 1223–1233. [CrossRef]
- Tazi, K.A.; Moreau, R.; Hervé, P.; Dauvergne, A.; Cazals-Hatem, D.; Bert, F.; Poirel, O.; Rabiller, A.; Lebrec, D. Norfloxacin Reduces Aortic NO Synthases and Proinflammatory Cytokine Up-Regulation in Cirrhotic Rats: Role of Akt Signaling. *Gastroenterology* 2005, 129, 303–314. [CrossRef]
- Kim, B.I.; Kim, H.J.; Park, J.H.; Park, D.I.; Cho, Y.K.; Sohn, C.I.; Jeon, W.K.; Kim, D.J.; Kim, H.S. Increased intestinal permeability as a predictor of bacterial infections in patients with decompensated liver cirrhosis and hemorrhage. *J. Gastroenterol. Hepatol.* 2010, 26, 550–557. [CrossRef]
- 111. Fukui, H.; Matsumoto, M.; Tsujita, S.; Takaya, A.; Kojima, H.; Matsumura, M.; Tsujii, T. Plasma endotoxin concentration and endo-toxin binding capacity of plasma acute phase proteins in cirrhotics with variceal bleeding: An analysis by new methods. *J. Gastroenterol. Hepatol.* **1994**, *9*, 582–586. [CrossRef]
- 112. Aguirre Valadez, J.M.; Rivera-Espinosa, L.; Méndez-Guerrero, O.; Chávez-Pacheco, J.L.; García Juárez, I.; Torre, A. Intestinal per-meability in a patient with liver cirrhosis. *Ther. Clin. Risk Manag.* **2016**, *12*, 1729–1748. [CrossRef]
- 113. Hou, M.-C.; Lin, H.-C.; Liu, T.-T.; Kuo, B.I.-T.; Lee, F.-Y.; Chang, F.-Y.; Lee, S.-D. Antibiotic prophylaxis after endoscopic therapy prevents rebleeding in acute variceal hemorrhage: A randomized trial. *Hepatology* **2004**, *39*, 746–753. [CrossRef]
- 114. Bernard, B.; Grangé, J.D.; Khac, E.N.; Amiot, X.; Opolon, P.; Poynard, T. Antibiotic prophylaxis for the prevention of bacterial infec-tions in cirrhotic patients with gastrointestinal bleeding: A meta-analysis. *Hepatology* 1999, 29, 1655–1661. [CrossRef] [PubMed]
- 115. Fasullo, M.; Rau, P.; Liu, D.Q.; Holzwanger, E.; Mathew, J.P.; Guilarte-Walker, Y.; Szabo, G. Proton pump inhibitors increase the se-verity of hepatic encephalopathy in cirrhotic patients. *World J. Hepatol.* **2019**, *11*, 522–530. [CrossRef] [PubMed]

- 116. Kang, D.J.; Betrapally, N.; Ghosh, S.A.; Sartor, R.B.; Hylemon, P.B.P.B.; Gillevet, P.M.P.M.; Sanyal, A.J.A.J.; Heuman, D.M.D.M.; Carl, D.; Zhou, H.; et al. Gut microbiota drive the development of neuroinflammatory response in cirrhosis in mice. *Hepatology* 2016, 64, 1232–1248. [CrossRef] [PubMed]
- Luo, M.; Guo, J.-Y.; Cao, W.-K. Inflammation: A novel target of current therapies for hepatic encephalopathy in liver cirrhosis. World J. Gastroenterol. 2015, 21, 11815–11824. [CrossRef] [PubMed]
- 118. Jayakumar, A.R.; Rao, K.V.R.; Norenberg, M.D. Neuroinflammation in Hepatic Encephalopathy: Mechanistic Aspects. *J. Clin. Exp. Hepatol.* **2015**, *5*, S21–S28. [CrossRef]
- 119. Bajaj, J.S.; Hylemon, P.B.; Ridlon, J.M.; Heuman, D.M.; Daita, K.; White, M.B.; Monteith, P.; Noble, N.A.; Sikaroodi, M.; Gillevet, P.M. Colonic mucosal microbiome differs from stool microbiome in cirrhosis and hepatic encephalopathy and is linked to cognition and inflammation. *Am. J. Physiol. Liver Physiol.* **2012**, *303*, G675–G685. [CrossRef]
- Gupta, A.; Dhiman, R.K.; Kumari, S.; Rana, S.; Agarwal, R.; Duseja, A.; Chawla, Y. Role of small intestinal bacterial overgrowth and delayed gastrointestinal transit time in cirrhotic patients with minimal hepatic encephalopathy. *J. Hepatol.* 2010, 53, 849–855.
 [CrossRef]
- 121. Bajaj, J.S. Review article: Potential mechanisms of action of rifaximin in the management of hepatic encephalopathy and other complications of cirrhosis. *Aliment. Pharmacol. Ther.* **2015**, *43*, 11–26. [CrossRef]
- 122. Zamparelli, M.S.; Rocco, A.; Compare, D.; Nardone, G. Faculty Opinions recommendation of the gut microbiota: A new potential driving force in liver cirrhosis and hepatocellular carcinoma. *United Eur. Gastroenterol. J.* 2017, *5*, 944–953. [CrossRef] [PubMed]
- 123. Yu, L.-X.; Yan, H.-X.; Liu, Q.; Yang, W.; Wu, H.-P.; Dong, W.; Tang, L.; Lin, Y.; He, Y.-Q.; Zou, S.-S.; et al. Endotoxin accumulation prevents carcinogen-induced apoptosis and promotes liver tumorigenesis in rodents. *Hepatology* 2010, 52, 1322–1333. [CrossRef] [PubMed]
- 124. Dapito, D.H.; Mencin, A.; Gwak, G.Y.; Pradere, J.P.; Jang, M.K.; Mederacke, I.; Caviglia, J.M.; Khiabanian, H.; Adeyemi, A.; Bataller, R.; et al. Promotion of hepatocellular carcinoma by the intestinal microbiota and TLR4. *Cancer Cell* 2012, 21, 504–516. [CrossRef]
- Yu, L.X.; Schwabe, R.F. The gut microbiome and liver cancer: Mechanisms and clinical translation. *Nat. Rev. Gastroenterol. Hepatol.* 2017, 14, 527–539. [CrossRef]
- 126. Fukui, H. Improve gut microbiome: A new horizon of cancer therapy. *Hepatobiliary Surg. Nutr.* 2017, *6*, 424–428. [CrossRef] [PubMed]
- 127. Gupta, H.; Youn, G.S.; Shin, M.J.; Suk, K.T. Role of Gut Microbiota in Hepatocarcinogenesis. *Microorganisms* 2019, 7, 121. [CrossRef] [PubMed]
- 128. Adolph, T.E.; Grander, C.; Moschen, A.R.; Tilg, H. Liver–Microbiome Axis in Health and Disease. *Trends Immunol.* **2018**, *39*, 712–723. [CrossRef] [PubMed]