MicroRNA profile of hepatic epithelioid hemangioendothelioma: A case report

ASAHIRO MORISHITA¹, HISAKAZU IWAMA², HIROHITO YONEYAMA¹, TEPPEI SAKAMOTO¹, KOJI FUJITA¹, TAKAKO NOMURA¹, JOJI TANI¹, HISAAKI MIYOSHI¹, SHINTARO FUJIHARA¹, NORIKO NISHIYAMA¹, HIDEKI KOBARA¹, HIROHITO MORI¹, NAOKI YAMAMOTO³, KEIICHI OKANO³, YASUYUKI SUZUKI³, EMI IBUKI⁴, REIJI HABA⁴, TAKASHI HIMOTO⁵ and TSUTOMU MASAKI¹

¹Department of Gastroenterology and Neurology; ²Life Science Research Center; Departments of ³Gastroenterological Surgery and ⁴Diagnostic Pathology, Faculty of Medicine, Kagawa University, Takamatsu, Kagawa 761-0793;

⁵Department of Medical Technology, Kagawa Prefectural University of Health Sciences, Takamatsu, Kagawa 761-0123, Japan

Received July 28, 2015; Accepted September 1, 2016

DOI: 10.3892/ol.2017.5643

Abstract. A 72-year-old female was referred for further evaluation of epigastralgia. Abdominal contrast computed tomography revealed numerous tumors in the two lobes of the liver. Liver biopsy and immunohistochemical staining revealed that the tumor cells were positive for factor VIII-associated antigen, platelet endothelial cell adhesion molecule 1 and human hematopoietic progenitor cell antigen, concordant with a diagnosis of hepatic epithelioid hemangioendothelioma (HEH). To elucidate the etiology of HEH, particularly the microRNA (miRNA) profiles, tissue samples obtained from normal and tumor tissues were analyzed using a miRNA array system. A total of 14 miRNAs were significantly upregulated and 93 miRNAs were downregulated in the tumor tissues (P<0.01). Additionally, unsupervised hierarchical clustering analysis using Pearson's correlation revealed that the tumor tissues clustered separately from the normal tissues. The miRNA expression profile was analyzed in HEH and compared with angiosarcoma, which exhibits histology similar to HEH. Out of a total of 107 miRNAs, only miR-122-5p and miR-1290 demonstrated a differential expression pattern in angiosarcoma. Therefore, these miRNAs may be novel biological markers for the determination of a diagnosis of HEH in primary mesenchymal tumors of the liver. To the best of our knowledge, this study is the first report of a miRNA microarray analysis in HEH.

E-mail: asahiro@med.kagawa-u.ac.jp

Introduction

Epithelioid hemangioendothelioma (EHE) is a rare disease that was initially described by Weiss and Enzinger in 1982 (1). EHE develops from a malignant transformation of the vascular endothelium and is generally located in soft tissues and internal organs (1). Hepatic epithelioid hemangioendothelioma (HEH) is the most common type of EHE; forms of EHE in other organs, including the lungs, brain, spleen, bones, breast, heart and stomach have been reported only rarely (2-6). HEH is a rare sarcoma of the liver, which typically presents as a number of nodules and may be misdiagnosed as a metastatic carcinoma (7,8). Additionally, HEH may recur locally and metastasize to distant organs (8); HEH has previously been considered an intermediate malignancy (9). However, the World Health Organization has previously categorized EHE as having full malignant potential (10). Therefore, it is crucial to identify the grade of malignancy and to distinguish HEH from other types of epithelioid vascular tumors, including epithelioid hemangioma and epithelioid angiosarcoma. However, due to the morphological similarities, it is challenging to differentiate these tumors solely using histological features (7). The current study focused on microRNAs (miRNAs) and performed miRNA profiling, in order to identify novel specific biomarkers for the accurate diagnosis of HEH.

Case report

A 72-year-old female was referred to Kagawa University Hospital in 2014 (Kagawa, Japan) for further evaluation of epigastralgia. The patient had no significant medical history and no familial history of genetic disorders or cancers. Abdominal contrast computed tomography (CT) revealed a number of liver tumors (Fig. 1A); multiple nodules with capsules located peripherally in the liver were apparent, with coarse calcification and peripheral enhancement following the administration of contrast medium. A liver biopsy and immunohistochemical staining revealed that the tumor cells were positive for factor VIII-associated antigen (FVIII-rAg),

Correspondence to: Dr Asahiro Morishita, Department of Gastroenterology and Neurology, Faculty of Medicine, Kagawa University, 1750-1 Ikenobe, Miki-cho, Kida-gun, Takamatsu, Kagawa 761-0793, Japan

Key words: microRNA profile, hepatic epithelioid hemangioendothelioma, angiosarcoma, hemangioma, multiple liver tumors

platelet endothelial cell adhesion molecule 1 (CD31) and human hematopoietic progenitor cell antigen (CD34), consistent with a diagnosis of HEH (Fig. 1B). Primary antibodies against human FVIII-rAg (Dako, Tokyo, Japan), CD31 (Dako, JC70A, Tokyo, Japan), and CD34 (Leica Microsystems, Tokyo, Japan) were added to paraffin-embedded liver sections. Secondary antibodies, including ImmPRESS[™] REAGENT Anti-Rat Ig and ImmPRESS[™] REAGENT Anti-Rabbit Ig, were also used (Vector Laboratories, Inc., Burlingame, CA, USA) and incubated with a DAB Substrate kit (Vector Laboratories, Inc.).

The patient was treated with a partial ultrasonography-guided hepatectomy for the multiple HEHs. Currently the patient is still alive and has been relapse-free for 2 years post-surgery.

To elucidate the etiology of HEH, particularly the miRNA profiles, tissue samples obtained from three separate sections of normal and tumor tissues were analyzed. Total RNA was extracted from the human liver samples using the miRNeasy Mini kit (Qiagen, Inc., Venlo, Netherlands), according to the manufacturer's instructions. Following RNA measurement with an RNA 6000 Nano kit (Agilent Technologies, Santa Clara, CA, USA), the samples were labeled using a miRCURY Hy3 Power Labeling kit (Exiqon, Vedbaek, Denmark) and then hybridized onto a mouse miRNA Oligo chip (version 19; Toray Industries, Inc., Tokyo, Japan). Scanning was conducted with the 3D-Gene Scanner 3000 (Toray Industries, Inc.). The 3D-Gene extraction version 1.2 software (Toray Industries, Inc.) was used to read the raw intensities of the image. To determine changes in miRNA expression between normal tissue and tumor tissue samples, the raw data were analyzed via GeneSpringGX version 10.0 (Agilent Technologies, Inc., Tokyo, Japan).

A total of 14 miRNAs were significantly upregulated and 93 miRNAs were significantly downregulated in the tumor tissues, as compared with the normal tissues (Table I). Additionally, these data were organized using unsupervised hierarchical clustering and analysis of variance (ANOVA) functions in the GeneSpring software. Unsupervised hierarchical clustering was performed by using the clustering function (condition tree) with Pearson's correlation. Two-way ANOVA and asymptotic P-value computation without any error correction on the samples were performed to identify the miRNAs that varied most prominently across the different groups. The P-value cutoff was set as 0.05. In the present study, the miRNAs in tumor tissues clustered separately from those in normal tissues (Fig. 2). The microarray data used in the present study were submitted as a complete data set to the NCBI Gene Expression Omnibus (no. GSE66794). Each expression of 107 miRNAs, which were statistically up-and down-regulated in HEH tissues, was analyzed as compared to that profile in angiosarcoma which exhibits a similar histology to HEH by using 'Sarcoma-microRNA Database'. The sarcoma miRNA expression database was accessed through http://www. oncomir.umn.edu/.

Of a total of 107 significantly upregulated and downregulated miRNAs identified in HEH, only miR-122-5p and miR-1290 demonstrated a differential expression pattern in angiosarcoma (Table I).



Figure 1. Clinicopathological features of the present case of hepatic epithelioid hemangioendothelioma. (A) Computed tomography revealed hypodense tumors in the two lobes (left panel). In the portal phase, the center of the tumor was enhanced in the right lobe (right panel; red arrow). (B) In the H&E-stained section, tumor cells were observed to form intracytoplasmic lumina, and solitary, alveolar and single-file patterns were detected. Immunohistochemical staining demonstrated that the tumor cells were positive for FVIII-rAg, CD31 and CD34. FVIII-rAg, factor VIII-associated antigen; CD31, platelet endothelial cell adhesion molecule 1; CD34, human hematopoietic progenitor cell antigen; H&E, hematoxylin and eosin.

Discussion

HEH is a rare hepatic, vascular, soft-tissue tumor with malignant potential; the incidence of HEH is <1/100,000 (11). The majority of patients present with nonspecific symptoms. HEH was first identified incidentally by imaging, including ultrasonography and computed tomography (12). The etiological factors of HEH require further study; however, various risk factors have been identified, including oral contraceptives, alcohol, trauma, viral infections and chronic liver diseases (11,12). Thorotrast (thorium dioxide) (13) and vinyl chloride (14) have also been reported as risk factors.

Laboratory findings, including biochemical examination of blood, are typically non-diagnostic in cases of EHE. Tumor markers, including α -fetoprotein, carcinoembryonic antigen and carbohydrate antigen 19-9 have a potential role in excluding the diagnosis of other hepatic neoplasms (15,16). Examination of blood samples has revealed that an increase in alkaline phosphatase is the most frequently observed abnormality of HEH (12). Radiological studies, including CT, magnetic resonance imaging and angiography indicate that there are two subtypes of HEH: The nodular subtype manifests as multifocal nodules in the early stages of HEH, and these nodules grow and eventually coalesce, whereas the

Table I. Cor	nparison	of	miRNAs	between	normal	and	HEF
tumor tissues	s (P<0.01).					

H Table I. Continued.

miRNA	P-value	Fold change (T/N) mean ± SD
hsa-miR-145-5p	0.0059	26.4833±12.9514
hsa-miR-155-5p	0.0099	20.5192±8.659
hsa-miR-181a-5p	0.0071	13.1044±4.978
hsa-miR-23b-3p	0.0096	12.362±4.993
hsa-miR-140-3p	0.0076	12.7157±5.4437
hsa-miR-125b-5p	0.0056	11.0117±4.9514
hsa-miR-4324	0.0096	10.1675±4.1997
hsa-miR-214-3p	0.0056	12.0701±7.5642
hsa-miR-26a-5p	0.0058	10.9722±5.6222
hsa-miR-126-3p	0.0064	8.2668±3.5601
hsa-miR-130b-3p	0.0074	8.3378±4.1174
hsa-miR-6822-5p	0.0055	7.8268±3.3700
hsa-let-7a-5p	0.0058	5.2697±1.8241
hsa-miR-34c-3p	0.0043	1.6188±0.1349

B, Downregulated miRNAs

		Fold change (T/N),
miRNA	P-value	mean ± SD
hsa-miR-4463	0.0076	0.5185±0.1658
hsa-miR-4713-5p	0.0049	0.4745±0.1267
hsa-miR-3191-5p	0.0090	0.4616±0.1886
hsa-miR-296-3p	0.0059	0.441±0.1595
hsa-miR-1290ª	0.0089	0.4589±0.1978
hsa-miR-6716-5p	0.0066	0.4225±0.1967
hsa-miR-4442	0.0016	0.4199±0.1121
hsa-miR-4534	0.0056	0.4224±0.1531
hsa-miR-4484	0.0019	0.4176±0.1387
hsa-miR-615-3p	0.0065	0.3907±0.114
hsa-miR-6791-5p	0.0067	0.4044 ± 0.1801
hsa-miR-636	0.0006	0.3943±0.0982
hsa-miR-6808-5p	0.0076	0.4024±0.143
hsa-miR-4800-3p	0.0039	0.3808±0.1669
hsa-miR-6855-3p	0.0022	0.3805±0.1372
hsa-miR-4525	0.0014	0.3655±0.0679
hsa-miR-6165	0.0053	0.3614±0.1487
hsa-miR-7114-5p	0.0065	0.3395±0.128
hsa-miR-6879-5p	0.0012	0.3438±0.1193
hsa-miR-3622a-5p	0.0078	0.3342±0.1694
hsa-miR-4497	0.0081	0.3358±0.1514
hsa-miR-8089	0.0024	0.3367±0.1433
hsa-miR-6880-5p	0.0025	0.3261±0.1262
hsa-miR-7854-3p	0.0068	0.3168±0.1401
hsa-miR-6895-5p	0.0027	0.316±0.1378
hsa-miR-885-5p	0.0080	0.3281±0.1762
hsa-miR-7110-5p	0.0078	0.3198±0.1633
hsa-miR-6877-5p	0.0062	0.3072±0.1433

		Fold change (T/N),
miRNA	P-value	mean \pm SD
hsa-miR-3194-5p	0.0050	0.3039±0.1679
hsa-miR-1469	0.0008	0.2858±0.1107
hsa-miR-5010-5p	0.0048	0.2964±0.1442
hsa-miR-370-3p	0.0043	0.2966±0.1479
hsa-miR-4749-5p	0.0099	0.3068±0.1709
hsa-miR-6076	0.0084	0.2912±0.1304
hsa-miR-664b-5p	0.0081	0.2968±0.1903
hsa-miR-4745-5p	0.0043	0.276±0.0822
hsa-miR-3714	0.0009	0.2668±0.0619
hsa-miR-6824-5p	0.0053	0.2791±0.0895
hsa-miR-6825-5p	0.0070	0.2823±0.1436
hsa-miR-6889-5p	0.0007	0.2673±0.1206
hsa-miR-5585-3p	0.0071	0.2779±0.1291
hsa-miR-4428	0.0023	0.2574±0.1187
hsa-miR-184	0.0069	0.2667 ± 0.0888
hsa-miR-642b-3p	0.0068	0.2536±0.1318
hsa-miR-3621	0.0014	0.2451±0.0755
hsa-miR-6769a-5p	0.0045	0.2436±0.1112
hsa-miR-1247-3p	0.0002	0.2335 ± 0.0803
hsa-miR-6757-5p	0.0074	0.2381±0.1004
hsa-miR-1233-5p	0.0009	0.2357+0.0937
hsa-miR-8060	0.0031	0.2276±0.0573
hsa-miR-6825-3p	0.0041	0.2303±0.0685
hsa-miR-1254	0.0050	0.2409±0.1457
hsa-miR-3925-5p	0.0096	0.248±0.0242
hsa-miR-6851-5p	0.0047	0.2196+0.1064
hsa-miR-6748-5p	0.0081	0.2358±0.1399
hsa-miR-4257	0.0071	0.2184±0.090
hsa-miR-6743-5p	0.0017	0.2173±0.0827
hsa-miR-564	0.0020	0.2139±0.0959
hsa-miR-8063	0.0060	0.2046±0.1137
hsa-miR-1229-5p	0.0044	0.2149±0.0950
hsa-miR-6124	0.0072	0.2171±0.1188
hsa-miR-4792	0.0006	0.1942±0.072
hsa-miR-4476	0.0019	0.2045±0.0877
hsa-miR-4481	0.0094	0.1983±0.0423
hsa-miR-4435	0.0025	0.1843±0.0804
hsa-miR-3150a-3p	0.0043	0.1788±0.0622
hsa-miR-122-5p ^a	0.0094	0.1912±0.0978
hsa-miR-6750-5p	0.0097	0.1705±0.0376
hsa-miR-4498	0.0011	0.1771±0.0938
hsa-miR-6775-3p	0.0038	0.167±0.0633
hsa-miR-6870-5p	0.0032	0.1774±0.0767
hsa-miR-3682-3p	0.0044	0.1664±0.0194
hsa-miR-765	0.0033	0.1515±0.081
hsa-miR-8071	0.0045	0.1602±0.0773
hsa-miR-6782-5p	0.0042	0.1646±0.0752
hsa-miR-1236-5p	0.0006	0.157±0.0564
hsa-miR-7150	0.0022	0.161±0.0669
hsa-miR-4756-5p	0.0024	0.1585±0.081
hsa-miR-6891-5p	0.0073	0.1647±0.0828
1		

Ί	ab	le I.	Continued	Ι.
---	----	-------	-----------	----

miRNA	P-value	Fold change (T/N), mean ± SD
hsa-miR-4638-3p	0.0009	0.1561±0.0716
hsa-miR-6769b-5p	0.0031	0.1561±0.0639
hsa-miR-6871-5p	0.0022	0.1439±0.0416
hsa-miR-4443	0.0071	0.1427±0.0639
hsa-miR-513a-5p	0.0038	0.1287±0.0552
hsa-miR-8078	0.0061	0.1079±0.0482
hsa-miR-6842-5p	0.0012	0.0943±0.0427
hsa-miR-194-5p	0.0015	0.0948±0.0434
hsa-miR-6893-5p	0.0045	0.0858±0.0367
hsa-miR-192-5p	0.0016	0.0816±0.0414
hsa-miR-3162-5p	0.0063	0.0799±0.0308
hsa-miR-4450	0.0052	0.0558±0.0073
hsa-miR-3678-3p	0.0026	0.0546±0.0283
hsa-miR-3649	0.0045	0.0417±0.0192

^amiRNAs were differentially expressed in angiosarcoma compared with HEH. miRNA, microRNA; HEH, hepatic epithelioid hemangioendothelioma; T/N, tumor/normal tissue; SD, standard deviation.

diffuse subtype manifests as large confluent masses preferentially involving the peripheral liver (17). In the present case, multiple nodules with capsules were observed, located peripherally in the liver, appearing with coarse calcification and peripheral enhancement when imaged using a contrast medium (Fig. 1A).

A definitive diagnosis of HEH requires histopathological examination; the predominant histological features include nests and cords of epithelioid endothelial cells and the presence of intracytoplasmic lumina. CD31, CD34 and FVIII-rAg are established endothelial cell markers that are commonly used for the diagnosis of EHE (18). As shown in Fig. 1B, the immunohistochemistry of HEH tissue samples revealed the presence of these endothelial markers.

EHE has previously been considered as an intermediate malignancy; however, the World Health Organization has now classified EHE as having full malignant potential (9,10). The metastatic rate of EHE is ~25% and mortality occurs in ~15% of all cases (1,9). By contrast, angiosarcoma of deep soft tissues metastasizes in ~50% of epithelioid angiosarcomas, and mortality occurs in over half of patients within a year of the diagnosis (19). Prognostic factors to distinguish the two biological subsets of tumors require further study to be identified.

In the present study, miRNA profiles were examined using tumor tissue samples and adjacent normal tissue samples to identify the distinguishing features of EHE and angiosarcoma. Unsupervised hierarchical clustering analysis using Pearson's correlation demonstrated that the tumor tissues clustered separately from the normal tissues (Fig. 2). Of the 107 significantly upregulated and downregulated miRNAs identified in HEH, only miR-122-5p and miR-1290 demonstrated a differential expression pattern



Figure 2. Hierarchical clustering of miRNAs between normal and tumor tissues. Liver tissue miRNA clustering was performed according to the expression profiles of 107 differentially expressed miRNAs between normal and tumor tissues. The analyzed tissue samples are presented in the columns and the miRNAs are presented in the rows. The miRNA-clustering tree is presented on the left and the sample-clustering tree is presented at the top. The color scale depicts the relative expression levels of the miRNAs in the patient tissue samples, with red representing a high expression level and blue representing a low expression level. miRNA.

between HEH and angiosarcoma (Table I). miR-122-5p has previously been identified to be downregulated in patients with hepatocellular carcinoma, as compared with the healthy controls (20). The expression of miR-1290 has also previously been demonstrated to be associated with breast cancer prognosis (21). Interestingly, those previous reports indicate that miR-122-5p and miR-1290 were down-regulated in epithelial cancer cells. Therefore, the results of the present study suggest that EHE has similar miRNA expression patterns to epithelial cancer cells and that these expression patterns in EHE may form a different phenotype from that in angiosarcoma. In conclusion, miRNA profiling has potential as a novel biological diagnostic tool for HEH. Furthermore, miR-122-5p and miR-1290 are potential biomarkers for the accurate diagnosis of HEH in primary mesenchymal tumors of the liver.

References

- 1. Weiss SW and Enzinger FM: Epithelioid hemangioendothelioma: A vascular tumor often mistaken for a carcinoma. Cancer 50: 970-981, 1982.
- 2. Tiu CM, Chou YH, Wang HT and Chang T: Epithelioid hemangioendothelioma of spleen with intrasplenic metastasis: Ultrasound and computed-tomography appearance. Comput Med Imaging Graph 16: 287-290, 1992.
- Lee KC, Ng WF and Chan JK: Epithelioid haemangioendothelioma presenting as a gastric polyp. Histopathology 12: 335-337, 1988.
- Ellis GL and Kratochvil FJ III: Epithelioid hemangioendothelioma of the head and neck: A clinicopathologic report of twelve cases. Oral Surg Oral Med Oral Pathol 61: 61-68, 1986.
 Marchiano D, Fisher F and Hofstetter S: Epithelioid hemangio-
- Marchiano D, Fisher F and Hofstetter S: Epithelioid hemangioendothelioma of the heart with distant metastases. A case report and literature review. J Cardiovasc Surg (Torino) 34: 529-533, 1993.
- Kamath SM, Nagaraj HK and Mysorekar VV: Hepatic and adrenal hemangioendothelioma-a case report. J Clin Diagn Res 7: 2583-2584, 2013.
- Lee SE, Park HY, Kim S, Bang H, Min JH and Choi YL: Epithelioid hemangioendothelioma with extensive cystic change and CAMTA1 rearrangement. Pathol Int 63: 502-505, 2013.
- Pokharna RK, Garg PK, Gupta SD, Dutta U and Tandon RK: Primary epithelioid haemangioendothelioma of the liver: Case report and review of the literature. J Clin Pathol 50: 1029-1031, 1997.
- Mentzel T, Beham A, Calonje E, Katenkamp D and Fletcher CD: Epithelioid hemangioendothelioma of skin and soft tissues: Clinicopathologic and immunohistochemical study of 30 cases. Am J Surg Pathol 21: 363-374, 1997.
- Moulai N, Chavanon O, Guillou L, Noirclerc M, Blin D, Brambilla E and Lantuejoul S: Atypical primary epithelioid hemangioendothelioma of the heart. J Thorac Oncol 1: 188-189, 2006.
- Neofytou K, Chrysochos A, Charalambous N, Dietis M, Petridis C, Andreou C and Petrou A: Hepatic epithelioid hemangioendothelioma and the danger of misdiagnosis: Report of a case. Case Rep Oncol Med 2013: 243939, 2013.

- 12. Mehrabi A, Kashfi A, Fonouni H, Schemmer P, Schmied BM, Hallscheidt P, Schirmacher P, Weitz J, Friess H, Buchler MW and Schmidt J: Primary malignant hepatic epithelioid hemangioendothelioma: A comprehensive review of the literature with emphasis on the surgical therapy. Cancer 107: 2108-2121, 2006.
- Soslow RA, Yin P, Steinberg CR and Yang GC: Cytopathologic features of hepatic epithelioid hemangioendothelioma. Diagn Cytopathol 17: 50-53, 1997.
- Darras T, Moisse R and Colette JM: Epithelioid hemangioendothelioma of the liver. J Belge Radiol 71: 722-723, 1988.
- 15. Okuda K, Kotoda K, Obata H, Hayashi N and Hisamitsu T: Clinical observations during a relatively early stage of hepatocellular carcinoma, with special reference to serum alpha-fetoprotein levels. Gastroenterology 69: 226-234, 1975.
- Haglund C, Lindgren J, Roberts PJ and Nordling S: Difference in tissue expression of tumour markers CA 19-9 and CA 50 in hepatocellular carcinoma and cholangiocarcinoma. Br J Cancer 63: 386-389, 1991.
- Miller WJ, Dodd GD III, Federle MP and Baron RL: Epithelioid hemangioendothelioma of the liver: Imaging findings with pathologic correlation. AJR Am J Roentgenol 159: 53-57, 1992.
- Makhlouf HR, Ishak KG and Goodman ZD: Epithelioid hemangioendothelioma of the liver: A clinicopathologic study of 137 cases. Cancer 85: 562-582, 1999.
- Meis-Kindblom JM and Kindblom LG: Angiosarcoma of soft tissue: A study of 80 cases. Am J Surg Pathol 22: 683-697, 1998.
- Tan Y, Ge G, Pan T, Wen D and Gan J: A pilot study of serum microRNAs panel as potential biomarkers for diagnosis of nonalcoholic fatty liver disease. PLoS One 9: e105192, 2014.
 Endo Y, Yamashita H, Takahashi S, Sato S, Yoshimoto N,
- 21. Endo Y, Yamashita H, Takahashi S, Sato S, Yoshimoto N, Asano T, Hato Y, Dong Y, Fujii Y and Toyama T: Immunohistochemical determination of the miR-1290 target arylamine N-acetyltransferase 1 (NAT1) as a prognostic biomarker in breast cancer. BMC Cancer 14: 990, 2014.