



Complete Genome Sequence of the Livestock-Associated Methicillin-Resistant Strain *Staphylococcus aureus* subsp. *aureus* 08S00974 (Sequence Type 398)

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ABSTRACT We report here the complete genome sequence of the livestock-associated methicillin-resistant *Staphylococcus aureus* strain 08S00974 from sequence type 398 (ST398 LA-MRSA) isolated from a fattening pig at a farm in Germany.

Staphylococcus aureus subsp. *aureus* strain 08S00974 is a methicillin-resistant member of sequence type (ST) 398. This livestock-associated clonal lineage, which first became known for its widespread colonization of industrially raised pigs (1), exhibits an extended host spectrum genotype (2). A broad host range in combination with a continuous acquisition of virulence and antibiotic-resistance genes from hospital-associated clones of *Staphylococcus aureus* make the members of this sequence type a serious threat to public health (3). To aid the studies of host adaptation and antimicrobial resistance, we sequenced the complete genome of *S. aureus* subsp. *aureus* strain 08S00974. This strain was originally isolated from a fattening pig raised at a farm in Germany and has been used for host colonization kinetics and susceptibility studies (4).

The reads were obtained using a PacBio RS II sequencer using P6/C4 chemistry (10-kb insert library; average read length, 93.93 bp; 289,303 reads) and an Illumina MiSeq (TruSeq DNA PCR-free library; 300-bp paired-end reads; 518,176 read pairs). A single contig was produced during the *de novo* assembly using Canu (5) and circularized using Circlator (6). Alignments produced by mapping Illumina reads to the assembly using the Burrows–Wheeler alignment method (7) were used to correct errors with Pilon (8). The resulting complete genome was annotated using Prokka (9).

The ST398 08S00974 chromosomal genome is 2,849,409 bp (G+C content, 32.9%) and possesses 2,551 open reading frames, 19 rRNAs, and 62 tRNAs. Prophage sequence prediction server PHAST (10) identified one intact 73.6-kb prophage sequence in the region between 1,830,776 bp and 1,904,406 bp, which bears similarity to the staphylococcal phage StauST398-2 (NC_021323). The strain belongs to ST398, *spa*-type t011, and SCC*mec* type V, according to sequence typing analysis using Ridom SeqSphere+ (Ridom GmbH, Germany) (11). The strain harbors a complete beta-lactamase operon (*blaZ*, *blaI*, *blaR*), genes associated with resistance phenotypes toward tetracycline (*tetK*, *tetM*), and licomycin/streptogramin (*vgaA*). Genes associated with the immune evasion cluster and exfoliative toxins described so far were not detected.

Accession number(s). The complete genome sequence of *S. aureus* subsp. *aureus* strain 08S00974 has been deposited in GenBank under the accession number [CP020019](https://www.ncbi.nlm.nih.gov/nuccore/CP020019).

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REFERENCES

1. Cuny C, Friedrich A, Kozytska S, Layer F, Nübel U, Ohlsen K, Strommenger B, Walther B, Wieler L, Witte W. 2010. Emergence of methicillin-resistant *Staphylococcus aureus* (MRSA) in different animal species. *Int J Med Microbiol* 300:109–117. <https://doi.org/10.1016/j.ijmm.2009.11.002>.
2. Walther B, Monecke S, Ruscher C, Friedrich AW, Ehrlich R, Slickers P, Soba A, Wleklinski CG, Wieler LH, Lübke-Becker A. 2009. Comparative molecular analysis substantiates zoonotic potential of equine methicillin-resistant *Staphylococcus aureus*. *J Clin Microbiol* 47:704–710. <https://doi.org/10.1128/JCM.01626-08>.
3. Diene SM, Corvaglia AR, François P, van der Mee-Marquet N; Regional Infection Control Group of the Centre Region. 2017. Prophages and adaptation of *Staphylococcus aureus* ST398 to the human clinic. *BMC Genomics* 18:133. <https://doi.org/10.1186/s12864-017-3516-x>.
4. Szabó I, Beck B, Friese A, Fetsch A, Tenhagen BA, Roesler U. 2012. Colonization kinetics of different methicillin-resistant *Staphylococcus aureus* sequence types in pigs and host susceptibilities. *Appl Environ Microbiol* 78:541–548. <https://doi.org/10.1128/AEM.05327-11>.
5. Koren S, Walenz BP, Berlin K, Miller JR, Bergman NH, Phillippy AM. 2017. Canu: scalable and accurate long-read assembly via adaptive k-mer weighting and repeat separation. *Genome Res* [Epub ahead of print]. <https://doi.org/10.1101/gr.215087.116>.
6. Hunt M, Silva ND, Otto TD, Parkhill J, Keane JA, Harris SR. 2015. Circlator: automated circularization of genome assemblies using long sequencing reads. *Genome Biol* 16:294. <https://doi.org/10.1186/s13059-015-0849-0>.
7. Li H. 2013. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. *Genomics arXiv:13033997*. <http://arxiv.org/abs/1303.3997>.
8. Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. *PLoS One* 9:e112963. <https://doi.org/10.1371/journal.pone.0112963>.
9. Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <https://doi.org/10.1093/bioinformatics/btu153>.
10. Zhou Y, Liang Y, Lynch KH, Dennis JJ, Wishart DS. 2011. PHAST: a fast phage search tool. *Nucleic Acids Res* 39:W347–W352. <https://doi.org/10.1093/nar/gkr485>.
11. Jünemann S, Sedlazeck FJ, Prior K, Albersmeier A, John U, Kalinowski J, Mellmann A, Goesmann A, von Haeseler A, Stoye J, Harmsen D. 2013. Updating benchtop sequencing performance comparison. *Nat Biotechnol* 31:294–296. <https://doi.org/10.1038/nbt.2522>.