



Draft Genome Sequence of *Pseudomonas mosselii* Gil3, Isolated from Catfish and Antagonistic against Hypervirulent *Aeromonas hydrophila*

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Pseudomonas mosselii Gil3 was isolated from a catfish that survived from lethal challenge with hypervirulent *Aeromonas hydrophila* (vAh). When assayed *in vitro*, the bacterium showed antagonism against vAh. Sequence analysis revealed that the genome of *P. mosselii* Gil3 encodes numerous aromatic metabolism pathways and proteins for biosynthesis of antimicrobial compounds.

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nvironmental bacteria within the *Pseudomonas putida* group (1) colonize diverse terrestrial and aquatic habitats and are of interest for their biodegradation properties (2), their antagonistic secondary metabolites (3), and their potential in biotechnological applications (4, 5). A P. putida-like strain, Gil3, was isolated from the gill tissue of a channel catfish (Ictalurus punctatus) that survived from a lethal challenge of hypervirulent Aeromonas hydrophila (vAh) (6). This strain showed an antagonistic effect against vAh in in-vitro assays and was initially identified as P. putida Gil3 based on 16S rRNA homology to P. putida BW11M1 (7) and Pseudomonas sp. 250J (8), both of the latter being recognized as antibiotic-producing soil/rhizosphere isolates. The identity of this strain was reclassified as P. mosselii Gil3 (also P. putida BW11M1 as P. mossselii BW11M1) when the calculated value of average nucleotide identity was compared with that of the type species within the P. putida group. Since vAh is one of the most important bacterial pathogens that causes persistent outbreaks of motile Aeromonas septicemia (MAS) in warm-water fishes (9, 10), approaches are sought to suppress the pathogen and control the MAS disease. To explore the potential of P. mosselii Gil3 as a probiotic against vAh, the genome of the strain was sequenced in this study.

Genome sequencing was performed on the Illumina MiSeq sequencer using a Nextera XT kit (Illumina, San Diego, CA, USA) and a 2 \times 250-bp paired-end sequencing kit. Resulting sequences were trimmed with a quality cutoff of 0.01 using CLC Genomics Workbench (CLCBio, Cambridge, MA, USA). A total of 101 contigs were *de novo* generated using the SPAdes Genome Assembler version 3.5.0 with an average coverage of 43.6-fold. The average contig size was 57,550 bp with an N_{50} value of 135,596 bp. The assembled genome size was 5,812,302 bp with a G+C content of 64.2%. The draft genome was annotated using RASTtk (http://rast .nmpdr.org) and the NCBI Prokaryotic Genome Annotation Pipeline (https://www.ncbi.nlm.nih.gov/genome/annotation_prok). A total of 5,286 coding genes were identified, including four copies of 5S rRNA, two copies of 16S rRNA, and one copy of 23S rRNA. Among coding genes, 5,207 coding sequences were associated with 470 metabolic subsystems.

Several aromatic metabolism pathways were identified in *P.* mosselii Gil3, including salicylate and gentisate catabolic pathways, a hydroxyphenylacetic acid catabolic pathway and a β -ketoadipate pathway. Additionally, *P.* mosselii Gil3 contains gene clusters predicted for the synthesis of antimicrobial compound xantholysin (11) and a ppyS homologous gene for synthesis of antibiotic pseudopyronines (12). Phylogenetic analysis of concatenated multilocus genes (gyrB, recA, ropB, ropD, and NuoD) showed that *P.* mosselii Gil3 forms a clade with 100% bootstrap support with *Pseudomonas* sp. 250J (accession no. JHEE00000000), *P.* putida 1A00316 (CP014343), *P.* mosselii DSM 17497 (JHYW0000000), and *P.* mosselii BW11M1 (LSLE00000000).

Accession number(s). This whole-genome shotgun project has been deposited in DDBJ/ENA/GenBank under the accession number LZCV00000000. The version described in this paper is the first version, LZCV01000000.

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