



# Article Designing of $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$ (0.0 $\leq x \leq 1.0$ ) Microspheres via Hydrothermal Approach and Their Selective Inhibition on the Growth of Cancerous and Fungal Cells

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**Abstract:** The current study offers an efficient design of novel nanoparticle microspheres (MCs) using a hydrothermal approach. The  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  ( $0.0 \le x \le 1.0$ ) MCs were prepared by engineering the elements, such as cobalt (Co), nickel (Ni), iron (Fe), and gallium (Ga). There was a significant variation in MCs' physical structure and surface morphology, which was evaluated using energy dispersive X-ray analysis (EDX), X-ray diffractometer (XRD), high-resolution transmission electron microscopy (HR-TEM), and scanning electron microscope (SEM). The anti-proliferative activity of MCs was examined by MTT assay and DAPI staining using human colorectal carcinoma cells (HCT-116), human cervical cancer cells (HeLa), and a non-cancerous cell line—human embryonic kidney cells (HEK-293). Post 72 h treatment, MCs caused a dose dependent inhibition of growth and proliferation of HCT-116 and HeLa cells. Conversely, no cytotoxic effect was observed on HEK-293 cells. The anti-fungal action was assessed by the colony forming units (CFU) technique and SEM, resulting in the survival rate of *Candida albicans* as 20%, with severe morphogenesis, on treatment with MCs x = 1.0. These findings suggest that newly engineered microspheres have the potential for pharmaceutical importance, in terms of infectious diseases and anti-cancer therapy.

Keywords: nanomaterial synthesis; anti-cancer agents; anti-fungal; microspheres



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# 1. Introduction

Nanomaterials are promising materials for various therapeutic applications, especially in diagnosing and treating diverse types of cancers and infectious diseases. Metal-based treatments are known to be an attractive research field in medicinal chemistry. Although platinum-based therapy is currently used, it is accompanied by resistance, deterrent side effects and a deficit of selectivity, forcing researchers to find more safe metals [1]. For example, complexes of nano-inorganic metals, such as Ru, Ir, Cu, Ni, Zn, Co, etc., were found to have better anti-cancer properties than cisplatin [2,3]. These nanomaterials possess unique properties, like high surface area and better cell penetration capability [4]. Among them, spinel ferrite nanomaterials are the most preferred materials for biomedicine, magnetic resonance imaging, pharmaceuticals, sensors, and drug delivery [5–8]. The ecofriendly transition metal oxides, such as nickel (Ni), cobalt (Co), and manganese (Mn), are known to exhibit superior redox activity, with natural richness, and simplistic and scalable synthesis [9–12]. There are reports that suggest that treatment with nickel nanoparticles can induce anticancer activities in different types of cancer cells [13–16]. Cobalt nanoparticles also possess anti-cancer activities, as shown in several studies [17–20]. Besides, gallium nanoparticles also show anti-cancer drug delivery and cancer cell imaging capabilities [21–23]. Iron oxide nanoparticles are also applied for cancer cell imaging and treatments [24–26].

Previously, it has been shown that combining two or more nanoparticles is an effective strategy to synthesize nanocomposites for targeted drug delivery and anti-cancer treatment [27–30]. These data support the evidence that combining two or more nanoparticles enhances the anti-cancer activities in colon and breast cancer cells.

The microsphere structure has attracted more attention due to low density, good dispersion, high specific surface area, high surface activity, strong permeability, good stability, super paramagnetic, etc. [31–34]. Some researchers reported that the CoFe<sub>2</sub>O<sub>4</sub> microspheres have the capability to attach to negatively charged bacterial pathogens (e.g., *E. coli*) [35]. Others found that the CoFe<sub>2</sub>O4 microspheres in many samples actively target *E. coli*, and immobilize it on their surface with an efficiency of 90% [36] (Huang et al. 2016). Li et al. investigated the anti-bacterial effectiveness and found that CoFe<sub>2</sub>O4 microspheres can act on gram-negative bacteria at lower concentrations within 1 h [37]. Taking into consideration the advantage of CoFe2O4 microspheres for its ability to attach to negatively charged bacterial cells, Muruganantham et al. fabricated positively charged CoFe2O4 microspheres as biocompatible anode in microbial cells for power generation [38]. Likewise, Ping Chen et al. produced M<sub>x</sub>Fe<sub>3-x</sub>O<sub>4</sub> (M = Mg, Mn, Fe, Co, Ni, Cu, Zn) microspheres for magnetic targeting and microwave heating, that can therefore be used for targeted and controllable drug delivery [39].

In the present study, the elements such as cobalt, nickel, gallium, and iron oxide were combined, and gallium was substituted with cobalt and nickel, to prepare  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  ( $0.0 \le x \le 1.0$ ) MCs by hydrothermal approach. The MCs structure and morphology were evaluated by using XRD, EDX, TEM, and SEM techniques. The impact of  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  ( $0.0 \le x \le 1.0$ ) MCs was examined using colon cancer HCT-116 and cervical cancer HeLa by using MTT assay. In addition, the anti-fungal effects of  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  ( $0.0 \le x \le 1.0$ ) MCs were also evaluated on *Candida* cells.

#### 2. Materials and Methods

# 2.1. Synthesis and Characterizations of MCs

Carbon microspheres were prepared hydrothermally using 1M glucose aqueous solution in a Teflon-lined autoclave, at 180 °C for 10 h. The black solid was obtained by centrifuging, washed many times with deionized H<sub>2</sub>O and C<sub>2</sub>H<sub>5</sub>OH (ethanol), respectively, and then dried at 60 °C. For the synthesis of Co<sub>0.5</sub>Ni<sub>0.5</sub>Ga<sub>x</sub>Fe<sub>2-x</sub>O<sub>4</sub> ( $0.0 \le x \le 1.0$ ) MCs, the stoichiometric amount of carbon sphere template and nitrate salts of Fe, Co, Ni, and Ga were dissolved in 50 mL of deionized water under continuous stirring, followed by sonication for approximately 30 min. Afterwards, the homogeneous solution was transferred into a Teflon-lined stainless steel autoclave and sealed to heat at 180 °C for 10 h. The final

products were washed with distilled water and ethanol and dried at 60 °C. Finally, the solid product was heated at 500 °C for 4 h to get the  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  ( $0.0 \le x \le 1.0$ ) MCs. The characterization of synthesized nanomaterials was done as per the protocol by Almessiere et al. [30].

# 2.2. Anti-Cancer Activity

# 2.2.1. Cell Culture and Testing of Nanoparticles Using MTT Method

Human colorectal carcinoma (HCT-116) and human cervical cells (HeLa) were taken for the study of the impact of  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  ( $0.0 \le x \le 1.0$ ) MCs on cell viability and cell proliferation. A non-cancer cell line, such as embryonic kidney cells (HEK-293), was used as control cell line and to examine the specificity of the MCs. As per the previously described method [40,41], the cells were cultured to 75–80% confluency and further processed using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) assay (Invitrogen, Waltham, MA, USA). The MTT assay was done where HCT-116, HeLa, and HEK-293 cells were treated with MCs doses, ranging from 2.0 to 80 µg/mL for 72 h. In the control group, no MCs were added. Both the control and sample-treated cells were exposed to 10 µL of MTT (5 mg/mL) and were incubated in a CO<sub>2</sub> incubator for 4 h. Later, the cell culture media was replaced with DMSO (1%), and the plate was examined at a wavelength of 570 nm by using a Plate reader (BioTek Instruments, Winooski, VT, USA). The percentage of cell viability was calculated for the statistical analysis [42,43].

## 2.2.2. DAPI Staining for DNA Analysis

DAPI (4',6-diamidino-2-phenylindole) staining assay was done to examine cancer cell DNA. The cells were divided into two groups: the control group, in which no MCs were added, and the experimental group where  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  ( $0.0 \le x \le 1.0$ ) MCs (50 µg/mL) were added. Seventy-two hours post-treatment, both groups were exposed to paraformaldehyde and then washed with phosphate-buffered saline (PBS). Cells were then stained with DAPI for 5 min, and examined by using confocal scanning microscope (Zeiss, Jena, Germany) [42,43]. The data are presented as mean standard deviation obtained from triplicates and one-way ANOVA followed by Dunnett's post hoc test with GraphPad Prism Software for final statistical analysis.

#### 2.3. Anti-Candidal Activity

#### 2.3.1. Preparation of Inoculum and Nanomaterial

*Candida albicans* ATCC14053 (yeast) was chosen for the anti-fungal assay of the prepared  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  ( $0.0 \le x \le 1.0$ ) MCs. *C. albicans* was aerobically grown using Sabouraud broth (SDB), at  $28 \pm 2$  °C for 24 h with shaking. Subsequently, the cell density was adjusted to approximately  $10^7$  CFU/mL. For the nanomaterial sample preparation, a concentration of 20 mg/mL of synthesized MCs was used after sonication for 10 to 15 min, to get a suspended solution.

# 2.3.2. Study of CFU and Morphogenesis of C. albicans

To demonstrate the anti-candida activity of MCs, colony-forming units (CFU) were used to evaluate the viable microbial cell concentrations in a sample, post treatment. Briefly, sterile tubes containing different concentrations of nanomaterial in SDB, were sonicated for 10 to 15 min to get the suspended solution. An adjusted inoculum of *C. albicans* was added to the MCs solution and incubated in a rotary shaker for 48 h at  $28 \pm 2$  °C. Untreated *C. albicans* was used as a control. On the completion of the incubation period, 100 µL of culture was plated out onto the SDA plates and incubated as mentioned above. On the other day, the plates were observed and recorded for the number of colonies on each plate. Each colony counted is taken to have emerged from a single viable *Candida* cell. The plates having colonies above 400 were considered as a lawn culture. The survival percentage of *C. albicans* was calculated by the formula S% = (A/B) × 100 (where, A is the number of CFU in the control) [44].

The fungal morphogenesis caused by the treatment of MCs was also studied by using SEM, following the protocol proposed by Aldakheel et al. [45]. The data are presented as mean standard deviation obtained from triplicates and one-way ANOVA followed by Dunnett's post hoc test with GraphPad Prism Software, USA for final statistical analysis.

#### 3. Results and Discussion

3.1. Microstructural Analysis of  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  ( $0.0 \le x \le 1.0$ ) MCs

The structure analysis of  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  ( $0.0 \le x \le 1.0$ ) MCs was offered through X-ray diffractometer, as seen in Figure 1. All MCs pattern denoted the index peaks of cubic spinel phase. Besides, the XRD patterns disclosed a pure spinel structure and slightly unsolicited phase. The cell parameters a, b, and c, were evaluated by full-proof software. It was found that the raise was with the growing amount of Ga from 5.883 (8) to 8.353 (5) (Å). The average crystallite size was measured by Scherrer's equation and found to be around  $20 \pm 6$  nm. Figure 2 exhibits the FESEM images of  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  (x = 0.2, 0.6 and 1.0) MCs. The samples showed an accumulation of the cubic shape particles. The elemental composition (EDX) of  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  (x = 0.2) MCs is shown in Figure 3. It approved the formation of MCs without any impurity. PDI was measured via zeta potential and the values were around 0.344, which indicated polydispersed particle size distribution. The TEM images demonstrated the agglomerated state of nanoparticles of MCs as seen in Figure 4.



**Figure 1.** XRD powder patterns of  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  ( $0.0 \le x \le 1.0$ ) MCs.



Figure 2. FESEM and particles size distribution of  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  (x = 0.2, 0.6 and 1.0) MCs.



**Figure 3.** EDX of  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  (x = 0.2) MCs.



**Figure 4.** TEM images of  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  ( $0.0 \le x \le 1.0$ ) MCs.

# 3.2. Biological Activities

3.2.1. Impact of  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  ( $0.0 \le x \le 1.0$ ) MCs on Cancer Cell Viability

The anti-proliferative impact of  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  ( $0.0 \le x \le 1.0$ ) MCs on both colon cancer (HCT-116) and cervical cancer (HeLa) cells was examined. The cell viability assay confirmed a significant decrease in the cell viability after the treatments of  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  ( $0.0 \le x \le 1.0$ ) MCs. The treatments of  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  MCs ( $0.0 \le x \le 1.0$ ) showed inhibitory action on cancer cell growth and proliferation. The calculated inhibitory concentration (IC<sub>50</sub>) of MCs was 25 µg/mL to 59 µg/mL for HCT-116 and 35 µg/mL to 60 µg/mL for HeLa cells. The impact of MCs on non-cancerous cells (HEK-293) was also tested. There was a decrease in the cancer viability, but the percentage of the decrease was not statistically significant (Table 1). Based on these observations, it may be suggested that the synthesized  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  ( $0.0 \le x \le 1.0$ ) MCs possess an inhibitory effect on HCT-116 and HeLa cells compared to HEK-293 cells. This is the first study that demonstrates the inhibitory effect on cell viability of synthesized  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  ( $0.0 \le x \le 1.0$ ) MCs against HCT-116 and HeLa cells.

x	IC <sub>50</sub> (HCT-116) Cells (µg/mL)	IC <sub>50</sub> (HeLa) Cells (μg/mL)	IC <sub>50</sub> (HEK-293) Cells
0.0	$25\pm2$	$35\pm4$	No inhibition
0.2	$46\pm3$	$56\pm4$	No inhibition
0.4	$42\pm4$	$48\pm5$	No inhibition
0.6	$29\pm2$	$50\pm5$	No inhibition
0.8	$50\pm5$	$49\pm4$	No inhibition
1.0	$59\pm5$	$60\pm5$	No inhibition

**Table 1.** Impact of  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  ( $0.0 \le x \le 1.0$ ) MCs on cancerous and normal cell lines.

Inhibitory concentration (IC).

We previously reported the impact of different nanomaterials on colon and breast cancer cells [45–47]. The presence of cobalt complexes in the nanocomposites may change the count and length of aliphatic chains, within the coordinated ligands. This significantly promotes its interaction with biological molecules, as well as its anti-proliferative effect on

cancer cells [48]. Also, the apoptosis seen in the cancer cells may be attributed to the mode of action by Co II and Ni II, which was found with high anti-tumor effect by stimulating mitochondria-intermediated apoptosis, along with arresting the S-phase of the cell cycle [3].

## 3.2.2. Disintegration of Cancer Cell DNA

The treatment of  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  ( $0.0 \le x \le 1.0$ ) MCs (50 µg/mL) caused a significant decrease in the number (35,000 to 45,000 cells/well) (Figure 5B–G) of HeLa cancer cells, compared to control cells (200,000 to 210,000 cells/well) (Figure 5A). The decrease in cancer cells was due to the mechanism of programmed cell death (known as apoptosis). The uptake of nanospheres by the treated cells also depends on the nanospheres concentrations, time of incubation, and other stress factors present [49]. The obtained results showed that the engineered microspheres have a cytotoxic effect on cancerous cells, through its magnetic effect, as MCs are supposed to localize into the target cell with the support of magnetic microspheres. These results agree with other studies where drugloaded targeted magnetic microspheres are used for cancer therapy [50]. The mechanism of action of nanocomposites on cancer cells may due to ionic strength, magnetism, and other effects [51]. These factors may lead to changes in the permeability of cell membrane, dissolving the lipids, and fusion with the cell lysosomes. Such factors may cause cell death, leading to cell cytotoxicity.



**Figure 5.** Cancer cell death via treatment of  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  MCs: shows the impact of  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  ( $0.0 \le x \le 1.0$ ) MCs on cervical (HeLa) cancer cells stained with DAPI post 72 h treatment. Figure (**A**): control (without MCs treatment), (**B**): (0.0), (**C**): (0.02), (**D**): (0.04), (**E**): (0.06), (**F**): (0.08), and (**G**): (0.01).

# 3.2.3. Anti-Fungal Activity

The anti-candida action of MCs was assessed by CFU technique. The varying concentration of test material was taken into consideration. The survival percentage of *Candida* in the inoculated media added with nanomaterial was investigated as the potential of the synthesized microspheres. After plating the harvested treated culture on SDA plates for 24 h, the existing colonies of *Candida* were observed. The survival rate of *Candida* is shown in Figure 6a. It was observed that the survival rate of *Candida* was reduced with the increasing ratio of microspheres, i.e., the maximum effect was shown by x = 1.0 with a survival rate of 20%, followed by 0.8 and 0.6, with 57 and 75%, respectively. Other ratios were not found to be significantly useful and showed an approximately 100% survival rate,

that was similar to control, i.e., the untreated *Candida*. As shown in Figure 6b, the number of cells on the agar plates has significantly decreased when compared to the control plate (untreated *Candida*). This indicates that the increased ratios of microspheres had an impact on the growth of the organism, where maximum growth reduction was seen with samples x = 1.0, x = 0.8, and x = 0.6. Using one-way ANOVA, the variation in the survival rates of *C. albicans* when using a different concentration of nanomaterial was found to be significant (p < 0.001).







**Figure 6.** (a) Survival rate of *C. albicans* after the treatment with  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  ( $0.0 \le x \le 1.0$ ) MCs (p < 0.001); (b) Agar plates showing the CFU count of *C. albicans* after the treatment with  $Co_{0.5}Ni_{0.5}Ga_xFe_{2-x}O_4$  MCs: (A) untreated; (B) x = 0.0; (C) x = 0.20; (D) x = 0.40; (E) x = 0.60; (F) x = 0.80; (G) x = 1.0.

Morphogenesis of *C. albicans* caused by the treatment of MCs was also studied by using SEM. The image (Figure 7) presents the morphological disruption caused by the MCs treatment. Figure 7B depicts the attachment and interaction of MCs and cellular surfaces, leading to disruption of *Candida* cell surface. The image clearly shows the membrane with attached MCs, creating perforations, whereas in Figure 7A, the untreated cells appear normal, with a smooth cell surface. The direct physical association of MCs and cells have been viewed, and the MCs have been observed to anchor onto cell walls and directly incorporate onto them. An apparent enhancement of anti-fungal activity was achieved on

the different organizations of the metal ions. It is evident from the obtained results that the synthesized nanomaterial activity was also enhanced with further metal coordination. This elevation in the anti-fungal activity can be justified based on structures, carrying additional carbon, and nitrogen bonds. Additionally, chelation may have reduced the polarity of the metal ion by sharing positive charge partially with the donor groups. Therefore, this phenomenon enhances the lipophilic nature of the metal atom, hence favoring greater penetration through the cell wall of the *Candida* and thus inhibiting them significantly. Moreover, the dipole moment, solubility, and conductivity are also affected by several metal ions, which could also be the additional factors responsible for increasing the nanomaterial's liposolubility, hence elevating the biological activity [29,52–54].



Figure 7. SEM micrograph presenting the morphogenesis of *C. albicans.* (A) Untreated cells (control); (B) MCs Treated cells.

# 4. Conclusions

The Co<sub>0.5</sub>Ni<sub>0.5</sub>Ga<sub>x</sub>Fe<sub>2-x</sub>O<sub>4</sub> (0.0  $\leq x \leq 1.0$ ) microspheres (MCs) were prepared using the hydrothermal approach, and the structures were confirmed using XRD, EDX, HR-TEM, TEM, and SEM techniques. The anti-proliferative impact of Co<sub>0.5</sub>Ni<sub>0.5</sub>Ga<sub>x</sub>Fe<sub>2-x</sub>O<sub>4</sub> (0.0  $\leq x \leq 1.0$ ) MCs was examined by using MTT assay that showed a dose-dependent inhibition of cancer cells (HCT-116, and HeLa). Conversely, no cytotoxic effect was observed on the normal cell line (HEK-293). The treatment of Co<sub>0.5</sub>Ni<sub>0.5</sub>Ga<sub>x</sub>Fe<sub>2-x</sub>O<sub>4</sub> (0.0  $\leq x \leq 1.0$ ) MCs also caused nuclear DNA disintegration in the cancer cells, as revealed by DAPI staining. Besides, the anti-fungal action was assessed by the CFU technique, and the survival rate of *C. albicans* was found to be reduced with the increasing ratio of Co<sub>0.5</sub>Ni<sub>0.5</sub>Ga<sub>x</sub>Fe<sub>2-x</sub>O<sub>4</sub> (0.0  $\leq x \leq 1.0$ ) MCs. These findings suggest that the synthesized Co<sub>0.5</sub>Ni<sub>0.5</sub>Ga<sub>x</sub>Fe<sub>2-x</sub>O<sub>4</sub> (0.0  $\leq x \leq 1.0$ ) MCs possess potential anti-fungal and anti-cancer capabilities, to be considered for potential pharmaceutical applications.

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