OPEN ACCESS **MOLECULES** ISSN 1420-3049 www.mdpi.com/journal/molecules

Article

Hepatoprotective Action of *Radix Paeoniae Rubra* Aqueous Extract against CCl₄-Induced Hepatic Damage

Ruidong Li[†], Wenyuan Guo[†], Zhiren Fu, Guoshan Ding, You Zou and Zhengxin Wang *

Department of Organ Transplantation, Changzheng Hospital, The Second Military Medical University, Shanghai 200003, China; E-Mails: rdliczsh@yahoo.com.cn (R.L.); wyguoczsh@yahoo.com.cn (W.G.); zrfuczsh@yahoo.com.cn (Z.F.); gsdingczsh@yahoo.com.cn (G.D.); yzczsh@yahoo.com.cn (Y.Z.)

- [†] These authors contributed equally to this work.
- * Author to whom correspondence should be addressed; E-Mail: zxwangczsh@sina.com; Tel.: +86-021-81885745; Fax: +86-021-63276788.

Received: 30 August 2011; in revised form: 7 October 2011 / Accepted: 11 October 2011 / Published: 17 October 2011

Abstract: In the present study the capacity of *Radix Paeoniae Rubra* aqueous extract (RPRAE) as an antioxidant to protect against carbon tetrachloride (CCl₄)-induced oxidative stress and hepatotoxicity in Wistar rats was investigated. Six groups of rats were used. *Radix Paeoniae Rubra aqueous extract* (100 or 200 or 300 mg/kg of bw) or bifendate (100 mg/kg of bw) were given daily by gavage to the animals on 28 consecutive days to elucidate the protective effects against CCl₄-induced hepatotoxicity. The 20% CCl₄/olive oil was gavage of gastric tube twice a week (on the third and seventh days of each week). The animals of normal control group were given only vehicle. The animals of CCl₄-treated group were administered with CCl₄ twice a week (on the third and seventh days of each week) and with vehicle on rest of the days. The test materials were found effective as hepatoprotective agents, as evidenced by plasma and liver biochemical parameters. Therefore, the results of this study show that *Radix Paeoniae Rubra aqueous extract* can protect the liver against CCl₄-induced oxidative damage in rats, and the hepatoprotective effects might be correlated with its antioxidant and free radical scavenger effects.

Keywords: hepatoprotection; rat; carbon tetrachloride; Radix Paeoniae Rubra; antioxidant

1. Introduction

Oxidative stress contributes a decisive generating factor in the pathogenesis of acute and chronic liver diseases [1-3]. Carbon tetrachloride (CCl₄) has been extensively used in animal models to explore chemical toxin-induced hepatic injury [4-6]. The metabolism of CCl₄ catalyzed by liver microsomal cytochrome P450 rapidly overproduces free radicals that deplete hepatic glutathione and initiate a chain lipid peroxidation of the hepatocyte membrane. This ultimately results in the overproduction of reactive oxygen species (ROS) and hepatocyte injuries [7-9].

Radix Paeoniae Rubra ("Chi-shao" in Chinese) is one of the most widely used Chinese herbal drugs. *Radix Paeoniae Rubra* (RPR) has been introduced into the treatment of neural insult, after extensive pharmacological research and clinical trials [10]. The active compounds in RPR include paeoniflorin, benzoylpaeoniflorin, albiflorin, lactiflorin, oxypaeoniflorin, paeonin, paeoniflorigenone, paeonoside, paeonolide, paeonol, galloylpaeoniflorin, gallotannin. Natural remedies from medicinal plants are considered to be effective and safe alternative treatments for hepatotoxicity. Some previous studies have confirmed that RPR can prevent D-galactosamine induced mouse liver injury [11], BCG and LPS induced mouse immunological liver injury [12], CCl₄-induced hepatic fibrosis [13,14], promote postoperative hepatocyte reproduction [15]. In this study, we tested the hepatoprotective action of *Radix Paeoniae Rubra aqueous extract* against the CCl₄-induced hepatic damage.

2. Results

The effects of aqueous extract of *Radix Paeoniae Rubra* at three dose levels (100, 200 and 300 mg/kg, b.w.) on serum marker enzymes are shown in Table 1. Hepatic injury induced by CCl₄ caused significant rise in marker enzymes AST, ALT, ALP activities and decrease in serum TP, Alb, G levels (P < 0.01). Administration of aqueous extract of *Radix Paeoniae Rubra* at three different dose levels attenuated the increased levels of the serum enzymes, produced by CCl₄, and caused a subsequent recovery towards normalization almost like that of bifendate treatment.

Group	AST (U/L)	ALT (U/L)	ALP (U/L)
NC	193.21 ± 12.65	39.75 ± 2.09	152.77 ± 10.65
CCl ₄ -treatment	401.43 ± 29.85 ^b	307.47 ± 19.65 ^b	518.36 ± 36.27 ^b
CCl ₄ +RPRAE (100 mg/kg b.w.)	314.06 ± 22.14 ^d	263.16 ± 13.61 ^c	403.75 ± 33.32 ^d
CCl ₄ +RPRAE (200 mg/kg b.w.)	251.63 ± 15.07 ^d	194.09 ± 13.69 ^d	284.18 ± 14.09 ^d
CCl ₄ +RPRAE (300 mg/kg b.w.)	192.86 ± 12.54 ^d	74.03 ± 4.53 ^d	170.39 ± 12.57 ^d
CCl ₄ +Bifendate (100 mg/kg b.w.)	217.53 ± 15.06 ^d	121.47 ± 8.64 ^d	206.41 ± 18.54 ^d

Table 1. Effect of aqueous extract of Radix Paeoniae Rubra on serum AST, ALT and ALP activities.

^b P < 0.01, compared with NC group; ^c P < 0.05, ^d P < 0.01, compared with CCl₄-treated group. Note: NC (normal control).

The hepatic cells in the CCl₄-inducted group were found to suffer obvious fatty degeneration and necrosis and vacuole formation after CCl₄ treatment. Administration of aqueous extract of *Radix Paeoniae Rubra* at doses of 100, 200 and 300 mg/kg for 28 days could reduce the fatty degeneration and necrosis hepatic injury score. This finding was consistent with the levels of the enzymes markers.

Group	TP (mg/L)	Alb (mg/L)	G (mg/L)
NC	88.32 ± 5.06	44.16 ± 2.08	35.16 ± 1.43
CCl ₄ -treatment	42.07 ± 2.75 ^b	30.15 ± 1.66 ^b	13.65 ± 1.09 ^b
CCl ₄ +RPRAE (100 mg/kg b.w.)	69.35 ± 4.53 ^d	37.12 ± 1.53 ^d	22.41 ± 1.35 ^d
CCl ₄ +RPRAE (200 mg/kg b.w.)	$74.62 \pm 5.13^{\text{ d}}$	43.62 ± 2.47 ^d	28.94 ± 1.84 ^d
CCl ₄ +RPRAE (300 mg/kg b.w.)	86.01 ± 4.76^{d}	50.49 ± 3.09 ^d	34.27 ± 2.53 ^d
CCl ₄ +Bifendate (100 mg/kg b.w.)	$71.54 \pm 5.32^{\text{ d}}$	$47.53 \pm 3.11^{\text{ d}}$	$32.61 \pm 2.05^{\text{d}}$

Table 2. Effect of aqueous extract of Radix Paeoniae Rubra on serum TP, Alb and G levels.

^b P < 0.01, compared with NC group; ^d P < 0.01, compared with CCl₄-treated group. Note: NC (normal control).

Liver index and hydroxyproline level in CCl₄ treated rats were significantly higher than in control (liver index: $4.73 \pm 0.24 vs. 3.15 \pm 0.12$; hydroxyproline: $481.63 \pm 23.11 vs. 108.54 \pm 8.54$) as shown in Table 3. Liver index and hydroxyproline level in rats treated with aqueous extract of *Radix Paeoniae Rubra* and bifendate were significantly decreased. The effect was dose-dependent.

Table 3. Effect of aqueous extract of Radix Paeoniae Rubra on liver index and hydroxyproline levels.

Group	Liver index	Hydroxyproline (µg·g ⁻¹)
NC	3.15 ± 0.12	108.54 ± 8.54
CCl ₄ -treatment	4.73 ± 0.24 ^b	481.63 ± 23.11 ^b
CCl ₄ +RPRAE (100 mg/kg b.w.)	4.31 ± 0.25 °	361.57 ± 23.07 ^d
CCl ₄ +RPRAE (200 mg/kg b.w.)	$3.91 \pm 0.19^{\text{ d}}$	239.11 ± 16.74 ^d
CCl ₄ +RPRAE (300 mg/kg b.w.)	$3.43 \pm 0.12^{\text{ d}}$	156.37 ± 12.14 ^d
CCl ₄ +Bifendate (100 mg/kg b.w.)	3.57 ± 0.19^{d}	194.08 ± 18.62 ^d

^b P < 0.01, compared with NC group; ^c P < 0.05, ^d P < 0.01, compared with CCl₄-treated group. Note: NC (normal control).

MDA level is widely used as a marker of free radical mediated lipid peroxidation injury. We measured MDA levels in the livers and the results are shown in Table 4. MDA levels in the CCl₄-treated group were significantly higher than that in the normal control group (P < 0.01). Consistent with the serum levels of ALT, AST, and ALP, administration with aqueous extract of *Radix Paeoniae Rubra* significantly decreased CCl₄-induced hepatic lipid peroxidation. The percentages of MDA levels in three doses of RPRAE-treated group (100, 200, 300 mg/kg) was significantly lower than that in the CCl₄-treated group (P < 0.01). Bifendate also inhibited the elevating MDA levels upon CCl₄ administration (Table 4). In addition, treatment with CCl₄ also significantly decreased the GSH levels in the liver as compared to the normal control group. In contrast with CCl₄-treated group, mice treated with aqueous extract of *Radix Paeoniae Rubra* (100, 200, 300 mg/kg) showed a significantly increase of GSH levels. These findings indicated that the free radicals being released in the liver were effectively scavenged by aqueous extract of *Radix Paeoniae Rubra*.

CCl₄ treatment also resulted in the depletion of the hepatic antioxidant enzymes. The activities of SOD, CAT, GSH-Px and TAOC were depleted to 127.39 ± 10.54 , 32.08 ± 0.14 , 36.19 ± 0.22 and 2.13 ± 0.12 respectively of the control (Table 4). The treatment of rats with aqueous extract of *Radix Paeoniae Rubra* provided protection against the depletion in activities of these enzymes (Table 4).

This shows aqueous extract of *Radix Paeoniae Rubra* to protect rats against oxidative injury by maintaining the levels of these enzymes even after CCl₄ treatment. Likewise, bifendate treatment also enhanced liver antioxidant enzymes activities in CCl₄-trated rats. CCl₄ induced inhibition in SOD, CAT, GSH-Px and TAOC may probably be due to their inactivation during the catalytic cycle. Under these conditions, aqueous extract of *Radix Paeoniae Rubra*, possessing potent free radical scavenging activity, may ameliorate the levels of H₂O₂ and O₂⁻, consequently restoring enzyme activity. The components of aqueous extract of *Radix Paeoniae Rubra* may also induce the de novo synthesis of the antioxidant enzymes.

Group	TBARS (nmol MDA/mg Prot)	GSH (mg/g Prot)	SOD (U/mg prot)	CAT (U/mg prot)	GSH-Px (U/mg prot)	TAOC (U/mg prot)
NC	234.42 ± 0.21	23.24 ± 1.33	363.14 ± 24.76	65.32 ± 0.43	52.11 ± 0.34	4.74 ± 0.32
CCl ₄ -treatment	$8.93\pm0.65\ ^{b}$	10.26 ± 1.07 ^b	$127.39 \pm 10.54 \ ^{b}$	$32.08 \pm 0.14 \ ^{b}$	36.19 ± 0.22 ^b	$2.13\pm0.12^{\ b}$
CCl ₄ +RPRAE (100 mg/kg b.w.)	6.85 ± 0.35 ^d	16.53 ± 1.25 ^d	184.64 ± 13.29 ^d	$44.08\pm0.25~^d$	42.13 ± 0.27 ^c	2.89 ± 0.19 ^c
CCl ₄ +RPRAE (200 mg/kg b.w.)	$5.42 \pm 0.24^{\ d}$	19.62 ± 1.66^{d}	295.1 ± 21.64 ^d	53.17 ± 0.31 ^d	46.27 ± 0.21 ^d	$3.74 \pm 0.14^{\ d}$
CCl ₄ +RPRAE (300 mg/kg b.w.)	4.63 ± 0.3^{d}	$25.13 \pm 1.72^{\text{ d}}$	342.72 ± 22.54 ^d	62.07 ± 0.36 ^d	54.37 ± 0.33 ^d	$4.83 \pm 0.31^{\ d}$
CCl ₄ +Bifendate (100 mg/kg b.w.)	4.98 ± 0.37 ^d	26.09 ± 2.01 ^d	315.54 ± 25.01 ^d	57.53 ± 0.4 ^d	50.39 ± 0.38 ^d	5.35 ± 0.29 ^d

Table 4. Effect of aqueous extract of *Radix Paeoniae Rubra* on MDA and GSH levels and SOD, CAT, GSH-Px, TAOC activities.

^b P < 0.01, compared with NC group; ^c P < 0.05, ^d P < 0.01, compared with CCl₄-treated group. Note: NC (normal control).

3. Discussion

Since the changes associated with CCl₄-induced liver damage are similar to those of acute viral hepatitis [16], CCl₄-mediated hepatotoxicity was chosen as the experimental model. The ability of a hepatoprotective drug to reduce the injurious effects or to preserve the normal hepatic physiological mechanisms which that have been disturbed by a hepatotoxin, is the index of its protective effects [17]. Liver damage induced by carbon tetrachloride (CCl₄) involves biotransformation of free radical derivatives, increased lipid peroxidation and excessive cell death in liver tissue [18]. The toxic effects of CCl₄ on liver have been extensively studied [19-22]. Serum AST, ALT are the most sensitive biomarkers used in the diagnosis of liver diseases [23]. During hepatocellular damage, varieties of enzymes normally located in the cytosol are released into the blood flow. Their quantification in plasma is useful biomarkers of the extent and type of hepatocellular damage [24]. Hepatocellular necrosis leads to elevation of the serum marker enzymes, which are released from the liver into blood [25]. Increased levels of ALT, AST, ALP, TP, Alb and G are conventional indicators of liver injury [26].

For the therapeutic strategies of liver injury and disease, it is important to find antioxidant compounds that are able to block liver injuries through free radicals generated due to toxic chemicals.

Therefore, the present study speculated that the aqueous extract of *Radix Paeoniae Rubra* protects against diseases that are caused by reactive oxygen species (ROS) because it has radical scavenging ability based on its antioxidant activity against CCl₄ in rats. In this study, rats treated with CCl₄ developed significant hepatic damage as manifested by a significant increase in activities of AST, ALT, ALP, and decrease in levels of TP, Alb and G that are indicators of hepatocyte damage and loss of functional integrity. These findings may reflect that 28 days were enough to detect abnormal AST, ALT, ALP, TP, Alb and G production in serum in this animal model. Centrilobular necrosis, lymphocytes infiltration and steatosis were also apparent in this study. Administration of the aqueous extract of *Radix Paeoniae Rubra* significantly reverse these abnormal indexs. This indicated that aqueous extract of *Radix Paeoniae Rubra* can ameliorate hepatic function of hepatic injury induced by CCl₄. Wang *et al.* reported that the aqueous extract of *Radix Paeoniae Rubra* can ameliorate function of hepatic injury induced by D-galactosamine [11]. These findings are in agreement with our previous results.

Liver hydroxyproline levels were measured as an index of hepatic collagen content. Hepatic stellate cells activated by CCl₄-induced oxidative stress overproduce extracellular matrix components and tissue inhibitors of metalloproteinases (TIMPs). These TIMPs inhibit collagenases, which normally degrade collagen. Thus, the rise in collagen deposition occurs [27]. In our study, liver hydroxyproline level showed a massive increase after CCl₄ long-term administration. Treatment with the aqueous extract of *Radix Paeoniae Rubra* and bifendate decreased its level significantly. Our finding supported other authors' observations that long-term administration of CCl₄ resulted in marked increase in the level of hydroxyproline [28,29]. Co-treatment with antioxidants or plant extracts [29,30] prevented the increase in hepatic hydroxyproline level significantly.

On the other hand, the metabolic transformation of CCl₄ by the NADPH-cytochrome P450 system to form the trichloromethyl radical CCl₃• and trichloromethyl peroxy radical CCl₃OO•, may also play an important role in the process of lipid peroxide which has the major role in the degenerative processes of the tissues [31-33]. Enhanced lipid peroxidation may associated with depletion of antioxidant, GSH in the heart of CCl₄ treated rats which is a characteristic observation in CCl₄-intoxicated rats [34,35]. GSH is important in effecting detoxification of the reactive metabolites of cells; tissue necrosis is initiated when reserves of GSH are markedly depleted [34,36]. Thus, the reduced (relative to normal) levels of GSH observed in the heart tissues of rats (administered CCl₄) might reflect to increased oxidative damage.

Antioxidant enzymes activities in liver of CCl₄-treated group rats were significantly lower than those in normal control group. Antioxidant enzymes (SOD, GPx and catalase) represent one protection against oxidative tissue-damage [37]. The cleavage of CCl₄ leads to the formation of highly unstable free radicals (CCl₃• or CCl₃O₂•), to initiate peroxidation [38]. Thus, the inhibition of the generation of free radicals retards CCl₄-induced lipid peroxidation. The administration of the aqueous extract caused an elevation of the levels of CAT, GSH-Px, TAOC and SOD and decreased the level of TBARS in liver, suggesting that it can restore these antioxidant enzymes and/or activate enzyme activity in the damage caused by CCl₄.

4. Experimental

4.1. Material

Roots of *Radix Paeoniae Rubra* was purchased from a herb market in Shanghai city, China in 2010 and identified by Dr. FL Ma, Department of Plant Biology (Shanghai Jiaoatong University, China). A voucher specimen was deposited in the laboratory of the School of Pharmacology, under the number 20100541.

4.2. Preparation of Aqueous Extract of Roots of Radix Paeoniae Rubra

Roots of *Radix Paeoniae Rubra* were dried under shade and pulverized. *Radix Paeoniae Rubra* root powder (600 g) was suspended in distilled water (5,000 mL), heated and boiled under reflux for 120 min. The extracts were filtered through Whatman No. 2 filter paper, and the filtrate was pooled and concentrated using a rotary evaporator at 50 °C and freeze-dried at -70 °C. The yield of dried aqueous extract of roots of *Radix Paeoniae Rubra* (RPRAE) was 13.85% (w/w).

4.3. Treatment of Animals

Male Wister rats (200-220 g/rat) were used in the experiments. These rats were provided with laboratory rodent diet and water ad libitum and randomly divide into six groups (eight rats/group): normal control (NC), CCl₄-treated group, three RPRAE-treated groups and a bifendate-treated group (positive control). To study the protective effect against the CCl₄-induced chronic hepatic injury and oxidative stress, Radix Paeoniae Rubra aqueous extract (100 or 200 or 300 mg/kg of bw) or bifendate (100 mg/kg of bw) were given daily by gavage to the animals on 28 consecutive days to elucidate the protective effect against CCl₄-induced hepatotoxicity. The 20% CCl₄/olive oil was gavage of gastric tube twice a week (on the third and seventh days of each week). The animals of normal control group were given only vehicle. The animals of CCl₄-treated group were administered with CCl₄ twice a week (on the third and seventh days of each week) and with vehicle on rest of the days. In the 29th day, each rat was anesthetized with ethyl ether, then weighed, blood drawn and the organ tissues taken out. Blood samples were collected for the assays of aspartate aminotransferase (GST), alanine aminotransferase (GLT), alkaline phosphatase (ALP) activities and total protein (TP), albumin (Alb), globulin (G) levels. The livers were excised from the animals and assayed for the reduced glutathione (GSH) level, antioxidant enzymes activity, malondialdehyde (MDA) formation and pathological histology, according to the procedures described bellowed. The liver index should be calculated as followings: (liver wet weight/body weight) ×100. All experimental procedures involving animals were conducted in accordance with National Institutes of Health (NIH). This experiment was approved by the Institutional Animal Care and Use Committee of China.

4.4. Histopathological Examination

Hepatic tissues of rats were collected from the same lobe of the livers and fixed in 10% neutral formalin solution. Hepatic tissue was dehydrated and embedded in paraffin. Cross-sections were stained using haematoxylin and eosin (H & E) for photomicroscopic observation.

4.5. Biochemical Analysis

Conventional liver biochemical tests (AST, ALT, ALP) were performed on a clinical chemistry automatic analyzer ADVIA 1650 (Bayer Diagnostics, Tarrytown, NY, USA). AST, ALT, and ALP were measured using commercial AST, ALT and ALP kits (Bayer Diagnostics).

The serum total concentrations of total protein (TP), albumin (ALB), and globulin (G) were measured by using commercial kits (Labtest Diagnóstica, Minas Gerais, Brazil) on the Express Plus biochemical analyzer (Ciba-Corning Diagnostics, Medfield, Massachusetts, USA). Hydroxyproline content in the liver was monitored using a diagnostic kit (Jiancheng Bioengineering Institute, Nanjing, China).

Hepatic thiobarbituric acid reactive substances (TBARS) was assayed by the method of Ohkawa *et al.* [39] using the thiobarbituric acid reaction, except that 1.0 mM EDTA was added to the reaction medium. The concentration of hepatic TBARS is expressed as malondialdehyde (MDA) equivalents. Hepatic GSH was assayed by the method of Sedlak and Lindsay [40] using Ellman's reagent. In this assay, GSH was used as a standard.

The catalase (CAT) activity was determined according to the Aebi method [41]. The rate of H_2O_2 decomposition was followed by monitoring absorption at 240 nm. One unit of CAT activity is defined as the amount of enzymes required to decompose 1 µmol of hydrogen peroxide in 1 min. The enzyme activity was expressed as µmol H_2O_2 consumed/min/mg protein. Superoxide dismutase (SOD) activity was estimated according to the method of Beauchamp and Fridovich [42]. The developed blue colour in the reaction was measured at 560 nm. Units of SOD activity were expressed as the amount of enzyme required to inhibit the reduction of NBT by 50% and the activity was expressed as U/mg protein. Glutathione peroxidase (GSH-Px) activity assayed using the method of Chiu *et al.* [43] in liver extracts. The level of total antioxidant capacity (TAOC) was measured by the method of ferric reducing/antioxidant power assay [44].

4.6. Statistical Analysis

The results of hepatoprotective effect were recorded for individuals within all groups. All data were presented as mean \pm SD. Results were statistically analyzed by one-way ANOVA using SPSS10.0 statistical software. A value of *P* < 0.05 was considered significant.

5. Conclusions

Our results demonstrate a very good protective effect of *Radix Paeoniae Rubra* against CCl₄induced liver injury, which is probably due at least partly to its antioxidative properties, scavenging CCl₄-associated free radicals. However, the mechanism of the hepatoprotective activity of *Radix Paeoniae Rubra* remains still to be proven. *Radix Paeoniae Rubra* might be useful as a natural ingredient for the prevention of oxidative damage in liver cells and tissues.

Conflict of Interest

The author declares no conflict of interest.

References and Notes

- 1. Tanikawa, K.; Torimura, T. Studies on oxidative stress in liver diseases: Important future trends in liver research. *Med. Mol. Morphol.* **2006**, *39*, 22-27.
- 2. Cesaratto, L.; Vascotto, C.; Calligaris, S.; Tell, G. The importance of redox state in liver damage. *Ann. Hepatol.* **2004**, *3*, 86-92.
- Tuma, D.J. Role of malondialdehyde-acetaldehyde adducts in liver injury. *Free Radic. Biol. Med.* 2002, *32*, 303-308.
- 4. Naik, S.R.; Panda, V.S. Antioxidant and hepatoprotective effects of Ginkgo biloba phytosomes in carbon tetrachlorideinduced liver injury in rodents. *Liver Int.* **2007**, *27*, 393-399.
- 5. Lee, C.P.; Shih, P.H.; Hsu, C.L.; Yen, G.C. Hepatoprotection of tea seed oil (Camellia oleifera Abel.) against CCl4-induced oxidative damage in rats. *Food Chem. Toxicol.* **2007**, *45*, 888-895.
- Wu, Z.M.; Wen, T.; Tan, Y.F.; Liu, Y.; Ren, F.; Wu, H. Effects of salvianolic acid A on oxidative stress and liver injury induced by carbon tetrachloride in rats. *Basic Clin. Pharmacol. Toxicol.* 2007, *100*, 115-120.
- 7. Weber, L.W.; Boll, M.; Stampfl, A. Hepatotoxicity and mechanism of action of haloalkanes: Carbon tetrachloride as a toxicological model. *Crit. Rev. Toxicol.* **2003**, *33*, 105-136.
- 8. Poli, G. Liver damage due to free radicals. Br. Med. Bull. 1993, 49, 604-620.
- 9. Basu, S. Carbon tetrachloride-induced lipid peroxidation: Eicosanoid formation and their regulation by antioxidant nutrients. *Toxicology* **2003**, *189*, 113-127.
- 10. Gong, X.D.; Sucher, N.J. Stroke therapy in traditional Chinese medicine (TCM): Prospects for drug discovery and development. *Trends Pharmacol. Sci.* **1999**, *20*, 191-196.
- Wang, D.-S.; Gao, J.-J.; Zhang, C.-G.; Xiang, L.; Yang, Q.-Y.; Xin, Y.-Q.; Cen, D.-Y.; Dong, L.-Y.; Chen, Z.-W. Protective effects of Chishaogancaochongji on chemical liver injury induced by D-galactosamine in mice. *Chin. J. Clin. Pharmacol. Ther.* 2006, 11, 1026-1029.
- Lu, Y.-S.; Xing, J.; Chen, Y.-L.; Yu, Y. Experimental studies on anti-oxidation capacity of astragalus and paeoniae rubra misture to mouse of immunological liver injury. *Chin. Arch. Tradit. Chin. Med.* 2008, 26, 302-303.
- 13. Lv, Y.-S.; Xing, J.; Chen, Y.-L.; Zhang, X.; Zhong, X.-L. Experimental studies of Astragalus and Paeoniae rubra misture on anti-hepatic fibrosis. *Zhong Guo Xian Dai Yi Xue Za Zhi* **2008**, *18*, 1997-2018.
- 14. Wang, Y.-P.; Cheng, M.-L.; Zhang, B.-F.; Mu, M.; Wu, J. Effects of blueberry on hepatic fibrosis and transcription factor Nrf2 in rats. *World J. Gastroenterol.* **2010**, *16*, 2657-2663.
- Mao, D.W.; Zhang, R.Z.; Cheng, W.L.; Wang, L.; Qiu, H. Effect of rheum officinale and Radix Paeoniae Rubra on hepatocyte reproduction blood clotting in postoperative liver failure rats. *J. Changchun Univ. Tradit. Chin. Med.* 2010, *26*, 641-643.
- 16. Suja, S.R.; Latha, P.G.; Pushpangadan, P.; Rajasekharan, S. Evaluation of hepatoprotective effect of Helminthostachys zeylanica (L.) Hook against carbontetrachloride induced liver damage in wistar rats. *J. Ethnopharmacol.* **2004**, *92*, 61-66.
- 17. Yadav, N.P.; Dixit, V.K. Hepatoprotective activity of leaves of Khalanchoe pinnata pers. *J. Ethanopharmacol.* **2003**, *86*, 197-202.

- 18. Recknagel, R.O.; Glende, E.A.; Dolak, J.A.; Waller, R.L. Mechanisms of carbon tetrachloride toxicity. *Pharmacol. Ther.* **1989**, *43*, 139-154.
- 19. Junnila, M.; Rahko, T.; Sukura, A.; Lindberg, L.A. Reduction of carbon tetrachloride-induced hepatotoxic effects by oral administration of betaine in male Han-Wistar rats: A morphometric histological study. *Vet. Pathol.* **2000**, *37*, 231-238.
- Amin, A.; Mahmoud-Ghoneim, D. Zizyphus spina-christi protects against carbon tetrachlorideinduced liver fibrosis in rats. *Food Chem. Toxicol.* 2009, 47, 2111-2119.
- Cui, C.P.; Wei, P.; Liu, Y.; Zhang, D.J.; Wang, L.S.; Wu, C.T. The protective role of Hepatopoietin Cn on liver injury induced by carbon tetrachloride in rats. *Hepatol. Res.* 2009, *39*, 200-206.
- Kim, H.Y.; Kim, J.K.; Choi, J.H.; Jung, J.Y.; Oh, W.Y.; Kim, D.C.; Lee, H.S.; Kim, Y.S.; Kang, S.S.; Lee S.H.; Lee, S.M. Hepatoprotective effect of pinoresinol on carbon tetrachlorideinduced hepatic damage in mice. *J. Pharmacol. Sci.* 2000, *112*, 105-112.
- 23. Pari, L.; Kumar, A.N. Hepatoprotective activity of Moringa Oleifera on antitubercular drug induced liver damage in rats. *J. Med.* **2002**, *5*, 171-177.
- 24. Pari, L.; Murugan, P. Protective role of tetrahydrocurcumin against erythromycin estolate-induced hepatotoxicity. *Pharmacol. Res.* **2004**, *49*, 481-486.
- 25. Ashok Shenoy, K.; Somayaji, S.N.; Bairy, K.L. Evaluation of hepatoprotective activity of Gingo biloba in rats. *Indian J. Pharmacol.* **2002**, *46*, 167-174.
- 26. Achiliya, G.S.; Wadodkar, S.O.; Dorle, A.K. Evaluation of hepatoprotective effect of Amakadi Ghrita against carbon tetrachloride induced hepatic damage in rats. *J. Ethnopharmacol.* **2004**, *90*, 229-232.
- Das, D.; Pemberton, P.W.; Burrows, P.C.; Gordon, C.; Smith, A.; McMahon, R.F.; Warnes, T.W. Antioxidant properties of colchicine in acute carbon tetrachloride induced rat liver injury and its role in the resolution of established cirrhosis. *Biochim. Biophys. Acta* 2000, *1502*, 351-362.
- 28. Sun, H.; Che, Q.-M.; Zhao, X.; Pu, X.-P. Antifibrotic effects of chronic baicalein administration in a CCl₄ liver fibrosis model in rats. *Eur. J. Pharmacol.* **2010**, *631*, 53-60.
- Lai, J.-T.; Hsieh, W.-T.; Fang, H.-L.; Lin, W.-C. The protective effects of a fermented substance from Saccharomyces cerevisiae on carbon tetrachloride-induced liver damage in rats. *Clin. Nutr.* 2009, 28, 338-345.
- Domitrović, R.; Jakovac, H.; Romić, Ž.; Rahelić, D.; Tadić, Ž. Antifibrotic activity of Taraxacum officinale root in carbon tetrachloride-induced liver damage in mice. *J. Ethnopharmacol.* 2010, 130, 569-577.
- Jayakumara, T.; Sakthivel, M.; Thomasb, P.A.; Geraldinea, P. Pleurotus ostreatus, an oyster mushroom, decreases the oxidative stress induced by carbon tetrachloride in rat kidneys, heart and brain. *Chem. Biol. Interact.* 2008, 176, 108-120.
- Kaurinovic, B.; Popovic, M.; Vlaisavljevic, S.; Raseta, M. Antioxidant Activities of Melittis melissophyllum L. (Lamiaceae), *Molecules* 2011, *16*, 3152-3167.
- 33. Pithayanukul, P.; Nithitanakool, S.; Bavovada, R. Hepatoprotective potential of extracts from seeds of areca catechu and nutgalls of quercus infectoria. *Molecules* **2009**, *14*, 4987-5000.
- 34. Wu, J.; Danielsson, A.; Zern, M.A. Toxicity of hepatotoxins: New insights into mechanisms and therapy. *Expert Opin. Investig. Drugs* **1999**, *8*, 585-607.

- Rop, O.; Reznicek, V.; Valsikova, M.; Jurikova, T.; Mlcek, J.; Kramarova, D. Antioxidant Properties of European Cranberrybush Fruit (*Viburnum opulus* var. edule) *Molecules* 2010, *15*, 4467-4477.
- Azizi, J.; Ismail, S.; Mordi, M.N.; Ramanathan, S.; Said, M.I.M.; Mansor, S.M. In Vitro and in Vivo Effects of Three Different Mitragyna speciosa Korth Leaf Extracts on Phase II Drug Metabolizing Enzymes—Glutathione Transferases (GSTs) *Molecules* 2010, 15, 432-441.
- 37. Halliwell, B.; Gutteridge, J.M.C. Role of free radicals and catalytic metal irons in human disease: An overview. *Meth. Enzymol.* **1990**, *186*, 59-85.
- 38. Recknagel, R.O.; Glende, E.A.; Dolak, J.A.; Waller, R.L. Mechanism of carbon tetrachloride toxicity. *Pharmacol. Ther.* **1989**, *43*, 139-154.
- 39. Ohkawa, H.; Ohishi, N.; Yagi, K. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Anal. Biochem.* **1979**, *142*, 290-296.
- 40. Sedlak, J.; Lindsay, R.H. Estimation of total, protein-bound, and nonprotein sulfhydryl groups in tissue with Ellman's reagent. *Anal. Biochem.* **1968**, *25*, 192-205.
- 41. Aebi, H. Catalase. In *Methods of Enzymatic Analysis*, Bergmeyer, H.U., Ed.; Academic Press: New York, NY, USA, 1974; Volume 2, pp. 673-678.
- 42. Beauchamp, C.; Fridovich, I. Superoxide dismutase: improved assay and assay applicable to acrylamide gels. *Anal. Biochem.* **1971**, *44*, 276-287.
- 43. Chiu, D.T.Y.; Stults, F.H.; Tappel, A.L. Purification and properties of rat lung soluble glutathione peroxidase. *Biochim. Biophys. Acta* **1976**, *445*, 558-566.
- 44. Benzie, I.F.F.; Stain, J.J. The ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power", the FRAP assay. *Anal. Biochem.* **1996**, *239*, 70-76.

Sample Availability: Not available.

 \bigcirc 2011 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (http://creativecommons.org/licenses/by/3.0/).