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# **OPEN** Retrogradation enthalpy does not always reflect the retrogradation behavior of gelatinized starch

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Starch retrogradation is a term used to define the process in which gelatinized starch undergoes a disorder-to-order transition. A thorough understanding of starch retrogradation behavior plays an important role in maintaining the quality of starchy foods during storage. By means of DSC, we have demonstrated for the first time that at low water contents, the enthalpy change of retrograded starch is higher than that of native starch. In terms of FTIR and Raman spectroscopic results, we showed that the molecular order of reheated retrograded starch samples is lower than that of DSC gelatinized starch. These findings have led us to conclude that enthalpy change of retrograded starch at low water contents involves the melting of recrystallized starch during storage and residual starch crystallites after DSC gelatinization, and that the endothermic transition of retrograded starch gels at low water contents does not fully represent the retrogradation behavior of starch. Very low or high water contents do not favor the occurrence of starch retrogradation.

Rice and wheat are two of the most economically important crops, contributing food for more than 90% of the world's population<sup>1</sup>. The quality of food products made from these staple grains is largely dependent on the properties of their major component starch, which is the most abundant storage carbohydrate in plants. Starch is used widely in food and non-food industries and, due to its low cost and renewability, it is also used to fabricate biodegradable materials. Before consumption by humans or for its application in materials, starch is usually gelatinized, by undergoing an order-to-disorder transition brought about by heating in the presence of water. On storage after gelatinization, the disordered starch chains gradually reassociate into a different ordered structure in a process termed retrogradation. Starch retrogradation is accompanied by a series of physical changes such as increased viscosity and turbidity of pastes, gel formation, exudation of water and increased degree of crystallinity with the appearance of B-type crystalline polymorphs<sup>2</sup>. The changes that starch undergoes during gelatinization and retrogradation are irreversible and are major determinants of its functional properties for food processing, during digestion, and in industrial applications. These properties determine the quality, acceptability, nutritional value, and shelf-life of finished, starch-rich foods<sup>3</sup>.

Gelatinization of starch causes loss of molecular order in both the amylose and amylopectin components. Subsequent retrogradation is the process in which the disaggregated starch chains reassociate into an ordered structure, which is different from the initial state<sup>4</sup>. Some of the partial crystallinity of amylopectin in native starch is restored during retrogradation<sup>5</sup>. Starch gelatinization and retrogradation are often characterized by differential scanning calorimentry (DSC), in combination with other structural or functional techniques<sup>3,6</sup>. The amount of water in the DSC pans plays an important role in the extent of starch gelatinization, which leads to the differential effects on subsequent retrogradation behavior. The effects of water availability on starch gelatinization have been investigated extensively, as reviewed recently by Wang and Copeland3. At low water content, the DSC endothermic transition only represents the limited swelling or incomplete gelatinization of starch granules<sup>7,8</sup>. Even at high water content (water:starch = 1.5:1 or higher), the DSC endotherm does not always represent the complete gelatinization of granules, as shown in studies with cereal and legume starches<sup>7–10</sup>. In comparison to gelatinization, the effect of water content on starch retrogradation has attracted much less attention<sup>6</sup>. A comprehensive study on starch retrogradation over a wide range of water content is lacking despite its major significance for the quality

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of foods during storage<sup>6</sup>. As most food systems are processed under limited water conditions<sup>5</sup> and then stored at 4°C, an understanding of the changes that starch undergoes during gelatinization and retrogradation in such conditions is important to manipulate the textural and nutritional qualities of starchy foods. Under water-limited conditions, starch undergoes only partial gelatinization on DSC heating and will retrograde in a different way during subsequent storage compared with fully gelatinized starch<sup>8</sup>.

Starch retrogradation may be characterized by determining the enthalpy change of retrograded starch gels after storage at 4 °C. The degree of retrogradation (DR) is often described by a parameter obtained by dividing the gelatinization enthalpy change of starch granules by the enthalpy change of reheated retrograded starch gels<sup>11,12</sup>. In the present study, the retrogradation behavior of wheat and rice starches over a wide range of water content was examined. To the best of our knowledge, this is the first study to investigate starch retrogradation over a wide range of water content by DSC in combination with ATR-FTIR and Raman spectroscopy. This study is aimed at gaining a better understanding of the effect of water content on starch retrogradation behavior in food and non-food applications.

### **Experimental Section**

**Materials.** The wheat (Zhoumai 18) flour was kindly provided by the Institute of Crop Science, Chinese Academy of Agricultural Science. The rice (*Oryza sativa*) sample was purchased from a supermarket in Tianjin, China. Amylose (A0512) and amylopectin (A8515) from potato starch were purchased from Sigma Chemical Co. (St. Louis, MO, USA). Other chemical reagents were of all analytical grade.

**Isolation of Starch.** Rice starch (RS) was isolated according to the method of Spigno and De Faveri<sup>13</sup>, and wheat starch (WS) according to a dough ball method<sup>8,14</sup>.

Chemical Analysis of Starch. The amount of damaged starch in the isolated starches was determined using the Megazyme Starch Damage Kit (Megazyme International Ireland Ltd. (Bray Co., Wicklow, Ireland). Amylose content was determined by iodine binding according to Chrastil<sup>15</sup> using a standard curve of 10%, 20%, 25%, 30%, and 35% potato amylose mixed with potato amylopectin. The crude lipid content of starch granules was determined gravimetrically by Soxhlet extraction using petroleum ether. The nitrogen content of wheat grains and starch granules were determined by standard Kjeldahl methodology. Crude protein content was estimated by multiplying the nitrogen content by a conversion factor of 6.25. Moisture content was determined by drying to constant weight at 105 °C and ash content was determined using a muffle furnace at 550 °C.

Differential Scanning Calorimetry. Differential scanning calorimetry (DSC) measurements were performed using a Differential Scanning Calorimeter (200 F3, Netzsch, Germany) equipped with a thermal analysis data station. Starch (approximately 3 mg wet weight) was weighed accurately into an aluminum sample pan. Distilled water was added with a pipette to obtain starch:water ratios of 1:0.5, 1:0.75, 1:1, 1:1.5, 1:2.5 and 1:4 (w/v) in the DSC pans, corresponding to the water content of 33, 43, 50, 60, 71 and 80%, respectively. The pans were sealed and allowed to stand overnight at room temperature before analysis. The pans were heated from 20 to  $100\,^{\circ}$ C at a rate of  $10\,^{\circ}$ C/min. An empty aluminum pan was used as the reference. Starch retrogradation was determined on the same gelatinized samples after storage at  $4\,^{\circ}$ C for 7 days. The retrograded starch samples were re-scanned using the heating profiles described for starch gelatinization. Gelatinization enthalpy change of native starch ( $\Delta$ H $_{\rm G}$ ) and enthalpy change on reheating of retrograded starch gels ( $\Delta$ H $_{\rm R}$ ) were obtained using data recording software. All measurements were performed in triplicate. Degree of retrogradation (%DR) was calculated according to the formula:

$$%DR = \Delta H_R / \Delta H_G \times 100\%$$

After DSC measurements, the sample pans were cooled to room temperature and reweighed. Those pans without any weight loss were collected and starch samples were freeze-dried and used for Fourier-transform infra-red (FTIR) and Raman spectroscopic analysis.

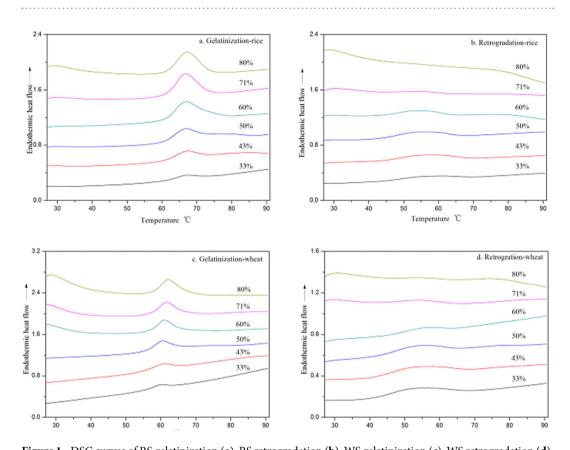
To obtain sufficient material to gain a better understanding of starch retrogradation behavior, retrograded starches were also prepared by simulating the DSC heating conditions without stirring using the RVA to prepare starch gels followed by storage at 4 °C for 7 days. The retrograded starch samples prepared in this way were freeze-dried, ground using a mortar and pestle, and passed through a  $100\,\mu m$  sieve. The resulting retrograded starch powders were mixed with water in ratios of 1:0.5, 1:0.75, 1:1, and 1:1.5 (w/v) in the DSC pans and analyzed using the same procedures for DSC measurements as described above.

**Attenuated Total Reflectance (ATR)-FTIR Spectroscopy.** The residual molecular order of native starch after DSC heating and of retrograded starch gels after DSC reheating was determined directly using a Thermo Scientific Nicolet IS50 spectrometer (Thermo Fisher Scientific, USA). The ratio of absorbance at 1047/1022 cm<sup>-1</sup> was used to estimate the short-range molecular order of starch.

**Laser Confocal Micro-Raman (LCM-Raman) Spectroscopy.** The molecular order of native starch after DSC heating and of retrograded starch gels after DSC reheating was also determined by using a Renishaw Invia Raman microscope system (Renishaw, Gloucestershire, United Kingdom), which was equipped with a Leica microscope (Leica Biosystems, Wetzlar, Germany) and a 785 nm green diode laser source. Spectra were collected on at least five different spots of gelatinized and retrograded starch samples in the range of 3200–100 cm<sup>-1</sup>, with a resolution of approximately 7 cm<sup>-1</sup>. The full width at half height (FWHH) of the band at 480 cm<sup>-1</sup>, which can be used to characterize the molecular order of starch<sup>6</sup>, was calculated using the WiRE 2.0 software.

Starch	Amylose (%)	DS (%)	Water (%)	Protein (%)	Lipid (%)	Ash (%)
Rice	$11.5 \pm 0.3$	$0.89 \pm 0.07$	$11.72\pm0.07$	$0.29 \pm 0.04$	$0.07\pm0.01$	$0.24 \pm 0.09$
Wheat	$25.0 \pm 0.6$	$1.30 \pm 0.02$	$10.62 \pm 0.21$	$0.36 \pm 0.02$	$0.11\pm0.01$	$0.15\pm0.04$

**Table 1. Basic composition of wheat and rice starches.** Values are means  $\pm$  SD. DS: damaged starch.



 $\textbf{Figure 1.} \ \ \mathsf{DSC} \ \mathsf{curves} \ \mathsf{of} \ \mathsf{RS} \ \mathsf{gelatinization} \ (\mathbf{a}), \ \mathsf{RS} \ \mathsf{retrogradation} \ (\mathbf{b}), \ \mathsf{WS} \ \mathsf{gelatinization} \ (\mathbf{c}), \ \mathsf{WS} \ \mathsf{retrogradation} \ (\mathbf{d}).$ 

**Statistical Analysis.** Results are reported as the mean values and standard deviations of at least duplicate measurements. Analyses of variance (ANOVA) by Duncan's test (p < 0.05) were conducted using the SPSS 17.0 Statistical Software Program (SPSS Inc. Chicago, IL, USA).

#### **Results and Discussion**

**Basic Composition of Rice and Wheat starches.** Amylose contents of the rice and wheat starches were 11.5% and 25.0%, respectively (Table 1). The amylose content of starch from non-waxy rice varieties varies between 7% and 33% <sup>16-18</sup>, whereas non-waxy wheat starch usually contains 20–30% amylose <sup>19</sup>. Damaged starch contents of the rice and wheat starches were 0.89% and 1.30%, respectively, whereas ash, protein, lipid and water contents of the two starches were all within the range of values reported in the literature<sup>20</sup>.

**Thermal Properties of Native and Retrograded Starches.** The DSC curves of starch gelatinization and from reheated retrograded starch gels are presented in Fig. 1. Rice and wheat starches displayed a typical gelatinization endothermic transition in the temperature ranges of 61.2–73.9 °C and 55.1–67.4 °C, respectively. With increasing water content, the gelatinization endothermic transition became progressively more pronounced and symmetrical (Fig. 1a,c). The area of this gelatinization endotherm increased with increasing water content. On reheating the retrograded starch, much broader but shallower endothermic transitions ranging from 42.9–68.9 °C and from 41.9–65.3 °C were noted for rice and wheat starch, respectively (Fig. 1b,d). The area of this retrogradation endotherm increased initially and then decreased with increasing water content.

Thermal transition temperatures of native starch and retrograded starch gels at different water contents are listed in Table 2. There were small differences in the gelatinization transition temperatures of rice starch as water content increased from 31% to 80%. The  $T_o$ ,  $T_p$  and  $T_c$  of rice starch were in the range of 61.2–62.1 °C, 66.9–68.3 °C and 71.0–73.9°, respectively. The transition temperature range ( $T_c$ – $T_o$ ) was from 9.8 to 11.7 °C. The  $T_o$ ,  $T_p$  and  $T_c$  of wheat starch ranged from 55.1, 60.8 and 64.6 °C to 56.8, 62.1 and 67.4 °C, respectively. The thermal transition

	Water content/%	Rices	starch	Wheat starch		
Temperatures		G/°C	R/°C	G/°C	R/°C	
T <sub>o</sub>	33	61.4±0.6ab	$44.1\pm0.8a$	$55.1 \pm 0.3a$	42.8 ± 1.3ab	
	43	61.2 ± 0.3a	43.5 ± 1.1a	$55.2 \pm 0.5a$	$41.9 \pm 0.4a$	
	50	61.2 ± 0.2a	$42.9 \pm 0.6a$	$55.3 \pm 1.3a$	$43.1 \pm 0.7 ab$	
	60	$61.8 \pm 0.1$ abc	$44.2 \pm 0.2a$	$55.5 \pm 0.1a$	43.8 ± 1.9ab	
	71	$62.1 \pm 0.5c$	$44.2 \pm 1.1a$	$56.0 \pm 0.2a$	43.1 ± 2.1ab	
	80	$62.0 \pm 0.3$ bc	nd	$56.1 \pm 0.4a$	45.4 ± 0.9b	
T <sub>p</sub>	33	$67.1 \pm 0.7a$	$56.8 \pm 0.5 ab$	$61.0\pm0.4a$	58.3 ± 1.9c	
	43	$67.3 \pm 0.4a$	58.2 ± 0.6b	$60.8 \pm 0.3a$	57.6 ± 1.6bc	
	50	67.2 ± 0.3a	55.2 ± 0.9a	$61.0 \pm 0.2a$	$54.7 \pm 1.5 ab$	
	60	$66.9 \pm 0.2a$	53.9 ± 0.3a	$61.1 \pm 0.16a$	55.6 ± 1.8abc	
	71	$67.6 \pm 0.4$ ab	$54.7 \pm 2.8a$	$61.8 \pm 0.3b$	53.6 ± 0.8a	
	80	$68.3 \pm 0.1b$	nd	$62.1 \pm 0.0$ b	$54.3 \pm 0.1a$	
$T_c$	33	$71.3 \pm 0.7a$	$67.8 \pm 1.8 bc$	$64.6 \pm 0.3$ b	65.3 ± 0.2d	
	43	$71.0 \pm 0.4a$	$68.9 \pm 2.0c$	$64.8\pm0.2ab$	64.8 ± 0.2cd	
	50	$71.3 \pm 0.3a$	$65.9 \pm 0.8 b$	$65.6 \pm 0.4$ b	$64.5 \pm 0.2$ cd	
	60	$71.8 \pm 0.6a$	$63.2 \pm 0.4a$	$66.2 \pm 0.2 b$	$64.1 \pm 0.9 bc$	
	71	$73.9 \pm 0.1b$	$62.5 \pm 0.2a$	$67.0 \pm 0.2c$	63.4 ± 0.8ab	
	80	$73.3 \pm 0.3b$	nd	$67.4 \pm 0.4c$	$62.7 \pm 0.1a$	
$T_c$ - $T_o$	33	9.9 ± 0.1a	$23.2 \pm 1.4 b$	$9.5 \pm 0.5a$	22.5 ± 1.1b	
	43	9.8 ± 0.1a	25.4 ± 2.2b	$9.6 \pm 0.6a$	$22.9 \pm 0.2b$	
	50	$10.1 \pm 0.4$ a	23.0 ± 1.0b	$10.3 \pm 1.2a$	21.4 ± 0.6b	
	60	$10.0 \pm 0.5a$	$19.0 \pm 0.5a$	$10.7 \pm 0.3$ ab	20.3 ± 2.2b	
	71	$11.7 \pm 0.6$ b	$18.3 \pm 0.9a$	$11.0\pm0.2ab$	20.3 ± 1.3b	
	80	11.3 ± 0.6b	nd	11.3 ± 0.3b	$17.3 \pm 0.9a$	

**Table 2.** Thermal transition temperatures of rice and wheat starch samples. Values are means  $\pm$  SD. Values with the same letters in the same column for each starch are not significantly different (p < 0.05). G: gelatinization; R: retrogradation nd: not determined.

broadened with  $T_c-T_o$  ranging from 9.5 to 11.3 °C. These observations are consistent with previous results, which showed that the conclusion temperature of pea and wheat starches increased gradually with increasing water content<sup>7,8</sup>.

After storage for 7 days, the transition temperatures of reheated retrograded starch gels, especially  $T_o$  and  $T_p$ , were much lower than those of native starch granules (Table 2), indicating that the melting of crystallites in retrograded starch gels occurred more readily than for native starch crystallites. The  $T_o$  of retrograded starch gels did not vary greatly with different water content, suggesting similarities in the onset of melting behavior of crystallites in retrograded starch gels. However, the  $T_p$  and  $T_c$  of retrograded rice starch gels decreased from 56.8 to 53.9 °C and from 67.8 to 62.5 °C, respectively, as the water content increased from 33 to 71%. Similarly, the  $T_p$  and  $T_c$  of retrograded wheat starch gels decreased from 58.3 to 53.6 °C and from 65.3 to 62.7 °C, respectively. With increasing water content,  $T_c$ – $T_o$  decreased from 25.4 and 22.9 °C to 18.3 and 17.3 °C for rice and wheat starch, respectively.

Gelatinization enthalpy change increased from 2.6 to 14.4 J/g and from 2.9 to 10.7 J/g for rice and wheat starch, respectively, as the water content increased from 33 to 71%, above which the enthalpy change remained essentially the same (Table 3). This observation was consistent with previous results<sup>8</sup>. Rice starch presented a higher maximum enthalpy change of 14.4 J/g than did wheat starch (10.7 J/g). The maximum enthalpy change of native starch granules has been shown to be variety dependent<sup>8</sup>.

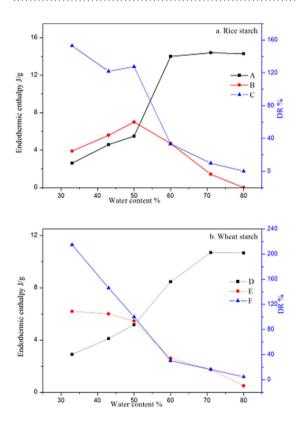
After retrogradation, the enthalpy change of rice and wheat starch gels varied according to water content (Table 3). Enthalpy change of rice starch gels increased from 3.9 J/g at a water content of 33% to a maximum value of 7.0 J/g at a water content of 50%, and then decreased to zero at a water content of 80%. In comparison, the retrogradation enthalpy change of wheat starch gels reached a maximum value of 6.2 J/g at a water content of 33%, and then decreased progressively to 0.5 J/g at a water content of 80%. When the water content was above 80%, no retrogradation of wheat starch gel was observed by DSC (data not shown), consistent with previous reports<sup>21–23</sup>.

Interestingly, the enthalpy change of retrograded starch gels was greater than that of native starch granules when the water content was between 33 and 50% (Table 3), resulting in a calculated value for the degree of retrogradation of the starch gels greater than 100% (Table 3, Fig. 2). This suggests that the broad endothermic transition of retrograded starch gels involves not only the melting of recrystallized starch formed during retrogradation, but the further melting of residual crystallites that remained in the retrograded starch gels. As proposed<sup>8</sup>, the DSC endothermic transition does not represent the full gelatinization of starch granules at DSC water-limited conditions.

To further substantiate the above hypothesis, the thermal transition of freeze-dried retrograded starch powders was determined (Fig. 3). Two separated endothermic transitions were noted in the range of 40~65 °C and

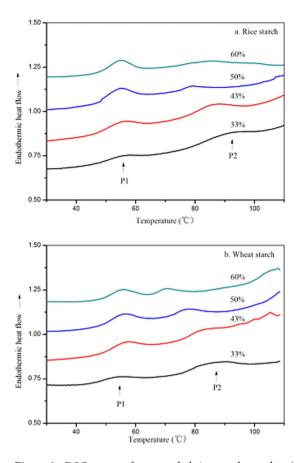
Starch	Water content/%	$\Delta H_G J/g$	$^{a}\Delta H_{R} J/g$	<sup>b</sup> ΔH <sub>R</sub> J/g	DR%
Rice starch	33	2.6 ± 0.1a	$3.9 \pm 0.0c$	4.2 ± 0.2a	153.1 ± 5.7f
	43	4.6 ± 0.1b	5.6 ± 0.2e	$5.2\pm0.4b$	121.9 ± 3.4e
	50	5.5 ± 0.1c	$7.0 \pm 0.4 f$	6.6 ± 0.0b	82.8 ± 5.0d
	60	14.0 ± 0.2d	$4.7\pm0.4\mathrm{d}$	4.6 ± 0.1a	33.3 ± 2.7c
	71	14.4 ± 0.7d	1.4 ± 0.2b	nm	9.9 ± 0.6b
	80	14.3 ± 0.2d	0a	nm	0a
Wheat starch	33	2.9 ± 0.0a	$6.2 \pm 0.2e$	4.4 ± 0.5b	214.8 ± 5.0e
	43	4.1 ± 0.1b	$6.0 \pm 0.0e$	$4.6\pm0.7b$	146.1 ± 2.5d
	50	5.5 ± 0.0c	5.5 ± 0.1d	$4.1 \pm 0.3b$	$100.1 \pm 2.3c$
	60	8.5 ± 0.1d	$2.6\pm0.4c$	$3.3 \pm 0.1a$	$30.5 \pm 4.4 b$
	71	$10.7 \pm 0.4e$	1.8 ± 0.1b	nm	16.4 ± 0.3a
	80	10.7 ± 1.1e	$0.5 \pm 0.0a$	nm	$4.7 \pm 0.3a$

**Table 3.** Enthalpy change of native and retrograded starches. Values are means  $\pm$  SD. Values with the same letters in the same column for each starch are not significantly different (p < 0.05). a: reheating retrograded starch gels; b: reheating retrograded starch powders/water mixtures; DR% was calculated based on the enthalpy change of regrograded starch gels. nm: not measured.



**Figure 2.** Gelatinization enthalpy change of RS (**A**), retrogradation enthalpy change of RS (**B**), DR of RS (**C**), Gelatinization enthalpy change of WS (**D**), retrogradation enthalpy change of WS (**E**), DR of WS (**F**).

70~100 °C. The lower temperature endotherms are proposed to be the melting of retrograded starch crystallites, whereas the ones at higher temperature are proposed to be due to the melting of residual crystallites remaining after DSC gelatinization. The observation of two separated endotherms indicated that melting of recrystallized starch formed during retrogradation and residual crystallites after DSC gelatinization occurred separately rather than simultaneously. The second endothermic transition is unlikely to be due to the melting of amylose-lipid complexes, which occurs at higher temperatures. Moreover, the lipid content of the starches was very low. The broad endothermic transitions noted for retrograded starch gels are consistent with the consecutive melting of recrystallized starch and residual starch crystallites. However, unlike the freeze-dried material, the endotherms of the reheated starch gels in the DSC pans may overlap because water is likely to migrate more readily from recrystallized starch to residual starch crystallites in gels than in rehydrated powders. Interestingly, the enthalpy change of freeze-dried retrograded starch powers was lower than that of retrograded starch gels (Table 3), consistent with



**Figure 3.** DSC curves of retrograded rice starch powders (a), retrograded wheat starch powders (b). p1: represents the melting of recrystallized starch during retrogradation; p2: represents the melting of residual starch crystallites after DSC gelatinization.

the findings that freeze drying can disrupt the crystalline and molecular order of  $starch^{24}$ . At higher water content of 60-80%, the degree of retrogradation was between 0-32.3% and between 4.7-34.1% for rice and wheat starch, respectively.

Short-range Molecular Order of Starch by ATR-FTIR and Raman Spectroscopy. The short-range molecular order of double helices in starch can be characterized by the ratio of absorbances at 1047/1022 cm<sup>-1</sup> obtained from FTIR spectroscopy of starch<sup>25–28</sup>. To verify the further melting of starch granule crystallites remaining in retrograded starch on DSC reheating, the molecular order of native starch after DSC heating and retrograded starch gels after DSC reheating was determined by ATR-FTIR and Raman spectroscopy. To identify the differences in the molecular order of these starch samples, the deconvoluted FTIR spectra of wheat starch in the range of 1200–800 cm<sup>-1</sup> were obtained (Fig. 4). The ratios of 1047/1022 cm<sup>-1</sup> of starch after gelatinization decreased with increasing water content (Table 4), indicating that the degree of disruption of molecular order in starch after DSC heating increased with increasing water content. Similar results were also observed with retrograded starch gels after DSC reheating, although in some cases no significant differences were observed. The ratios of 1047/1022 cm<sup>-1</sup><sub>R</sub> were lower than those of 1047/1022 cm<sup>-1</sup><sub>G</sub> over the whole range of water content, although the difference was small in some cases. This result showed that the molecular order of the retrograded starch gels after DSC reheating was lower than that of gelatinized starch, providing evidence to support the conclusion from the DSC data that there was further melting of starch crystallites remaining in retrograded starch gels.

As the LCM-Raman spectra of starch samples were similar, only those of wheat starch are presented (Fig. 5). Several clear bands can be seen at 480, 865, 943, 1264 and 2900 cm<sup>-1</sup>, which are related to  $\delta$  (CH<sub>2</sub>),  $\nu s$  (C1-O-C4),  $\nu s$  (C1-O-C5), skeletal (C-C-O), and  $\nu$  (C-H) modes, respectively²9-31. Of these bands, the ones at 480 and 2900 cm<sup>-1</sup> are often used to characterize the molecular order of native starch granules or the changes in molecular order of starch samples during gelatinization or retrogradation6.29,32-34. Full width at half maximum height (FWHM) of the strong band at 480 cm<sup>-1</sup> is often used to characterize the relative crystallinity of starch samples; this parameter is most responsive to changes in crystallinity²9. Values for FWHM<sub>R</sub> were higher than those for FWHM<sub>G</sub> over the whole range of water content (although the difference was small at high water content), indicating that the relative crystallinity of retrograded starch gels after DSC reheating was lower than that of gelatinized starch samples, consistent with the ATR-FTIR results.

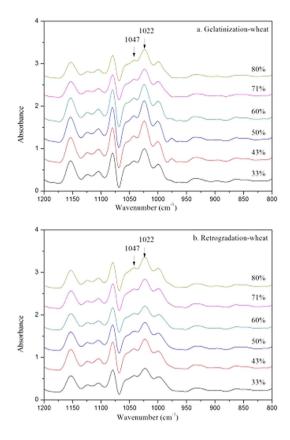


Figure 4. The deconvoluted FTIR spectra of gelatinized WS after DSC heating (a), reheated retrograded WS (b).

Starch	Water content/%	1047/1022cm <sup>-1</sup> <sub>G</sub>	1047/1022cm <sup>-1</sup> <sub>R</sub>	$FWHM_G$	FWHM <sub>R</sub>
Rice starch	33	$0.695 \pm 0.017c$	$0.649 \pm 0.023c$	$15.47 \pm 0.08a$	15.54 ± 0.09a
	43	$0.647 \pm 0.008$ b	$0.625 \pm 0.007 \mathrm{abc}$	$15.45 \pm 0.54a$	16.38 ± 0.44b
	50	$0.657 \pm 0.006$ b	0.635 ± 0.005bc	$15.59 \pm 0.05a$	15.84 ± 0.36ab
	60	$0.646 \pm 0.009$ b	$0.645 \pm 0.007c$	$15.73 \pm 0.01a$	$15.77 \pm 0.07$ ab
	71	$0.595 \pm 0.014$ a	$0.595 \pm 0.032ab$	$15.31 \pm 0.09a$	15.36 ± 0.58a
	80	0.604 ± 0.012a	$0.586 \pm 0.011a$	$15.66 \pm 0.23a$	16.38 ± 0.25ab
Wheat starch	33	0.694 ± 0.015b	$0.652 \pm 0.006$ b	$15.94 \pm 0.35 ab$	16.32 ± 0.08b
	43	$0.687 \pm 0.010$ b	$0.655 \pm 0.007$ b	$16.53 \pm 0.02b$	$16.99 \pm 0.24c$
	50	$0.680 \pm 0.001$ b	$0.664 \pm 0.008$ b	$15.92 \pm 0.04$ ab	16.23 ± 0.24b
	60	$0.689 \pm 0.017$ b	$0.660 \pm 0.016$ b	$15.67 \pm 0.35a$	$15.78 \pm 0.25a$
	71	$0.662 \pm 0.014$ a	$0.658 \pm 0.014b$	$16.32 \pm 0.41$ b	$16.40 \pm 0.04$ b
	80	$0.647 \pm 0.037a$	$0.623 \pm 0.005$ a	15.53 ± 0.50a	16.44 ± 0.28b

Table 4. The ratios of  $1047/1022 \, \mathrm{cm}^{-1}$  and FWHMs of the band at  $480 \, \mathrm{cm}^{-1}$  of starch samples. Values are means  $\pm$  SD. Values with the same letters in the same column for each starch are not significantly different (p < 0.05). G represents native starch after DSC heating; R represents retrograded starch gels after DSC reheating.

The maximum enthalpy change of retrograded starch gels was observed at water content around 50%, which does not correspond to the maximum degree of starch retrogradation. The crystallites formed during retrogradation are less ordered and less stable than crystallites in native starch granules. The observed higher enthalpy change of retrograded starch gels at low water content indicated that the enthalpy change of retrograded starch gels involves not only the melting of new starch crystallites formed on storage, but also further melting of starch crystallites remaining after gelatinization. This conclusion was supported by the ATR-FTIR and Raman spectroscopy analysis of molecular order of starch after gelatinization and after reheating of retrograded starch gels. Several studies investigated the effect of water content on retrogradation of starch<sup>21–23,35–37</sup>, but none reported an apparent value greater than 100% for the degree of retrogradation. The effect of water content on starch retrogradation, as determined by measuring DSC enthalpy change of recrystallized amylopectin, displayed a parabolic shape, with maximum retrogradation occurring in starch gels at 40–45% water content<sup>21,22,37</sup>.

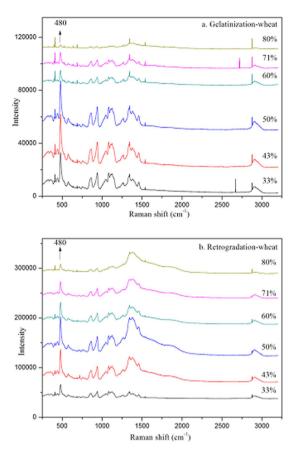


Figure 5. LCM-Raman spectra of gelatinized WS after DSC heating (a), reheated retrograded WS (b).

Another interesting finding from the present study is the decreasing  $T_p$  and  $T_c$  of retrograded starch gels with increasing water content. Two hypotheses, not mutually exclusive, are proposed to interpret this observation. The crystallites formed during retrogradation could become progressively less stable with increasing water content, leading to lower  $T_p$  and  $T_c$  of retrograded starch gels. Another explanation is that the higher  $T_p$  and  $T_c$  of retrograded starch gels at low water content is due to further melting of residual starch crystallites that remained in retrograded starch gels. The melting of starch crystallites is not complete at the end of DSC heating, especially at low water content  $^{38,39}$ . This interpretation is supported by the observation that the  $T_c$  of retrograded rice and wheat starch gels was slightly lower than those of native starch granules at low water content, and that the transition temperature range of retrograded starch gels decreased with increasing water content (Table 2). Taken together, we can conclude that the endothermic transition of retrograded starch gels at low water content does not reflect accurately the retrogradation behavior of gelatinized starch.

It is also interesting to note that the enthalpy change of starch gelatinization remained essentially unchanged at a water content above 71%, whereas the enthalpy change of starch retrogradation decreased progressively up to a water content of 80%. This result indicated that although the enthalpy change reached a maximum value, presumably corresponding to the complete gelatinization of starch, the retrogradation behavior of fully gelatinized starch was different at different water contents. The high water content brought about low degree of retrogradation, indicating that with increasing water content, the coming together and realignment of dispersed starch chains becomes progressively more difficult.

#### **Conclusions**

Differential scanning calorimetry studies over a wide range of water content have shown that water strongly influences retrogradation behavior of rice and wheat starches. The DSC thermal transition parameters have demonstrated for the first time that the enthalpy change of retrograded starch gels was greater than that of native starch at water content of 33 to 50%. In combination with ATR-FTIR and Raman results, we conclude that the DSC endothermic transition of retrograded starch gels at low water content does not truly reflect the retrogradation behavior of gelatinized starch. The enthalpy change of retrograded starch gels at low water contents represents the melting of starch crystallites formed by retrogradation and of residual crystallites remaining after gelatinization. At high water content, when complete gelatinization of starch may occur, the degree of retrogradation is influenced by water content, whereas at very low and very high water content, the DSC enthalpy change indicated that little retrogradation of starch occurred.

#### References

- 1. Fasahat, P., Rahman, S. & Ratnam, W. Genetic controls on starch amylose content in wheat and rice grains. *J. Genet.* **93,** 279–292 (2014).
- Hoover, R., Hughes, T., Chung, H. J. & Liu, Q. Composition, molecular structure, properties, and modification of pulse starches: A review. Food Res. Int. 43, 399–413 (2010).
- 3. Wang, S. & Copeland, L. Molecular disassembly of starch granules during gelatinization and its effect on starch digestibility: a review. *Food Funct.* **4**, 1564–1580 (2013).
- 4. Atwell, W. A., Hood, L. F., Lineback, D. R., Varriano-Marston, E. & Zobel, H. F. The terminology and methodology associated with basic starch phenomena, *Cereal Foods World* 33, 306–311 (1988).
- 5. Delcour J. A. & Hoseney, R. C. Principles of Cereal Science and Technology. (3rd ed.). St. Paul: MN, USA: AACC International, 23–51 (2010).
- Wang, S., Li, C., Copeland, L., Niu, Q. & Wang, S. Starch retrogradation: A comprehensive review. Compr. Rev. Food Sci. F. 14, 568–585 (2015).
- 7. Wang, S. & Copeland, L. Phase Transitions of Pea Starch over a wide range of water content. J. Agric. Food Chem. 60, 6439-6446 (2012).
- 8. Wang, S., Li, C., Yu, J., Copeland, L. & Wang, S. Phase transition and swelling behaviour of different starch granules over a wide range of water content. LWT-Food Sci. Tech. 59, 597–604 (2014).
- 9. Jenkins, P. J. & Donald, A. M. Gelatinisation of starch: a combined SAXS/WAXS/DSC and SANS study. *Carbohydr. Res.* **308**, 133–147 (1998)
- 10. Vermeylen, R. et al. Gelatinization of starch in excess water: beyond the melting of lamellar crystallites. A combined wide- and small-angle X-ray scattering study. Biomacromolecules. 7, 2624–2630 (2006).
- 11. Wu, Y., Chen, Z. X., Li, X. X. & Li, M. Effect of tea polyphenols on the retrogradation of rice starch. Food Res. Int. 42, 221-225 (2009).
- 12. Vandeputte, G. E., Vermeylen, R., Geeroms, J. & Delcour, J. A. Rice starches. III. Structural aspects provide insight in amylopectin retrogradation properties and gel texture. *J. Cereal Sci.* 38, 61–68 (2003).
- 13. Spigno, G. & De, F. D. M. Gelatinization kinetics of rice starch studied by non-isothermal calorimetric technique: influence of extraction method, water concentration and heating rate. *J. Food Eng.* **62**, 337–344 (2004).
- Yu, J. et al. Effect of laboratory milling on properties of starches isolated from different flour millstreams of hard and soft wheat. Food Chem. 172, 504–514 (2015).
- 15. Chrastil, J. Improved colorimetric determination of amylose in starches or flours. Carbohydr. Res. 159, 154-158 (1987).
- 16. Kong, X., Zhu, P., Sui, Z. & Bao, J. Physicochemical properties of starches from diverse rice cultivars varying in apparent amylose content and gelatinisation temperature combinations. *Food Chem.* **172**, 433–440 (2015).
- Vandeputte, G. E., Vermeylen, R., Geeroms, J. & Delcour, J. A. Rice starches. I. Structural aspects provide insight into crystallinity characteristics and gelatinisation behaviour of granular starch. J. Cereal Sci. 38, 43–52 (2003).
- 18. Singh, N., Kaur, L., Sandhu, K. S., Kaur, J. & Nishinari, K. Relationships between physicochemical, morphological, thermal, rheological properties of rice starches. *Food Hydrocolloid*. **20**, 532–542 (2006).
- Singh, N., Singh, J., Kaur, L., Sodhi, N. S. & Gill, B. S. Morphological, thermal and rheological properties of starches from different botanical sources. Food Chem. 81, 219–231 (2003).
- 20. Waterschoot, J., Gomand, S. V., Fierens, E. & Delcour, J. A. Production, structure, physicochemical and functional properties of maize, cassava, wheat, potato and rice starches. *Starch-Starke*. 67, 14–29 (2015).
- 21. Longton, J. & Legrys, G. A. Differential scanning calorimetry studies on the crystallinity of ageing wheat starch gels. *Starch-Starke*. 33, 410–414 (1981).
- Zeleznak, K. J. & Hoseney, R. C. The role of water in the retrogradation of wheat starch gels and bread crumb. Cereal Chem. 63, 407-411 (1986).
- 23. Zhou, X., Wang, R., Yoo, S. H. & Lim, S. T. Water effect on the interaction between amylose and amylopectin during retrogradation. *Carbohydr. Polym.* **86**, 1671–1674 (2011).
- 24. Zhang, B. et al. Freeze-drying changes the structure and digestibility of B-polymorphic starches. J. Agric. Food Chem. 62, 1482–1491 (2014).
- 25. Soest, J. J. G. V., Tournois, H., Wit, D. D. & Vliegenthart, J. F. G. Short-range structure in (partially) crystalline potato starch determined with attenuated total reflectance Fourier-transform IR spectroscopy. *Carbohyd. Res.* 279, 201–214 (1995).
- Kizil, R., Irudayaraj, J. & Seetharaman, K. Characterization of Irradiated Starches by Using FT-Raman and FTIR Spectroscopy. J. Agric. Food Chem. 50, 3912–3918 (2002).
- 27. Hu, X. P. et al. Effects of continuous and intermittent retrogradation treatments on in vitro digestibility and structural properties of waxy wheat starch. Food Chem. 174, 31–36 (2015).
- 28. Chung, H.-J., Liu, Q. & Hoover, R. Effect of single and dual hydrothermal treatments on the crystalline structure, thermal properties, and nutritional fractions of pea, lentil, and navy bean starches. Food Res. Int. 43, 501–508 (2010).
- Mutungi, C., Passauer, L., Onyango, C., Jaros, D. & Rohm, H. Debranched cassava starch crystallinity determination by Raman spectroscopy: correlation of features in Raman spectra with X-ray diffraction and <sup>13</sup>C CP/MAS NMR spectroscopy. Carbohydr. Polym. 87, 598–606 (2012).
- 30. Piccinini, M. et al. The Application of NIR FT-Raman spectroscopy to monitor starch rRetrogradation and crumb firmness in Semolina bread. Food Anal. Method. 5, 1145–1149 (2012).
- 31. Labanowska, M., Weselucha-Birczynska, A., Kurdziel, M. & Puch, P. Thermal effects on the structure of cereal starches. EPR and Raman spectroscopy studies. *Carbohydr. Polym.* 92, 842–848 (2013).
- 32. Bulkin, B. J., Kwak, Y. & Dea, I. C. M. Retrogradation kinetics of waxy-corn and potato starches; a rapid, Raman-spectroscopic study. *Carbohydr. Res.* 160, 95–112 (1987).
- 33. Flores-Morales, A., Jiménez-Estrada, M. & Mora-Escobedo, R. Determination Of The structural changes By FT-IR, Raman, and CP/ MAS <sup>13</sup>C NMR spectroscopy on retrograded starch of maize Tortillas. *Carbohydr. Polym.* **87,61**–68 (2012).
- 34. Fechner, P. M., Wartewig, S., Kleinebudde, P. & Neubert, R. H. Studies of the retrogradation process for various starch gels using Raman spectroscopy. *Carbohydr. Res.* **340**, 2563–2568 (2005).
- 35. Jouppila, K., Kansikas, J. & Roos, Y. H. Factors affecting crystallization and crystallization kinetics in amorphous corn starch.
- Carbohydr. Polym. 36, 143–149 (1998).
  36. Lee, E.-J., Lim, S.-T. & Chung, H.-J. Retrogradation behavior of rice flour/starch gel using response surface methodology. Food Sci. Biotechnol. 24, 891–894 (2015).
- 37. Liu, Q. & Thompson, D. B. Effects of moisture content and different gelatinization heating temperatures on retrogradation of waxy-type maize starches. *Carbohydr. Res.* 314, 221–235 (1998).
- 38. Derycke, V. *et al.* Starch gelatinization and amylose-lipid interactions during rice parboiling investigated by temperature resolved wide angle X-ray scattering and differential scanning calorimetry. *J. Cereal Sci.* **42**, 334–343 (2005).
- 39. Vermeylen, R. et al. Structural transformations during gelatinization of starches in limited water: Combined wide- and small-angle X-ray scattering study. Biomacromolecules. 7, 1231–1238 (2006).

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#### **Author Contributions**

Shujun Wang and Shuo Wang conceived and designed the study. C.L. and X. Z. conducted the experiments and data analysis. Shujun Wang and C.L. contributed to the earlier draft of the manuscript. Shujun Wang and L.C. revised and edited the manuscript.

## **Additional Information**

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