

Research Article

Intravitreal Injection of Hydrogen Peroxide Induces Acute Retinal Degeneration, Apoptosis, and Oxidative Stress in Mice

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Purpose. Oxidative stress is a common pathological condition for multiple retinal diseases. Hydrogen peroxide (H₂O₂) has been applied as an oxidative stress inducer for the *in vitro* studies. Here, we report the *in vivo* effect of H₂O₂ exposure to the mouse retina and its underlying mechanism. **Methods.** The H₂O₂ or saline solution was intravitreally injected into the eyes of female C57BL/6J mice for two consecutive days. The retinal structure was evaluated by *in vivo* imaging using spectral domain optical coherence tomography (OCT) and validated by histological assessment as well as retinal marker expression. In addition, retinal stress, cell apoptosis, and antioxidant enzyme expression were also determined. **Results.** Retinal and outer nuclear layer thickness thinning was observed at days 7 and 14 by OCT imaging with the treatment of 10 μg H₂O₂, which was confirmed by the histopathological analysis. The expressions of photoreceptor (*Rho*, *Rora*, *Rorb*, and *Rcvrn*), bipolar cell (*Chat* and *Calb2*), and retinal pigment epithelial (*Rpe65*) markers were reduced in the H₂O₂-treated group, whereas the expression of retinal ganglion cell marker (*Tubb3*) was increased. TUNEL-positive cells were obviously found in the outer nuclear layer and inner nuclear layer of H₂O₂-treated mice but sparsely found in the ganglion cell layer. Coherently, apoptotic gene expressions (*Casp3*, *Casp9*, *Bax*, and *Parp8*) were significantly increased in the retina with increasing dosages of H₂O₂, while *Bcl2* expression was mildly decreased. In addition, the expressions of Gfap and antioxidant enzyme genes (*Txn2*, *Sod2*, and *Gpx4*) were significantly upregulated in the retina after the H₂O₂ treatment, compared to the vehicle control group. **Conclusions.** This study revealed that intravitreal injection of H₂O₂ induces acute retinal damage by increasing oxidative stress and cell apoptosis in the retina. This acute retinal degeneration mouse model could provide a platform for drug screening against oxidative stress and retinal diseases.

1. Introduction

Retinal diseases, including age-related macular degeneration (AMD), glaucoma, diabetic retinopathy (DR), and retinitis pigmentosa (RP), are the leading cause of irreversible blindness and visual impairment in most developed countries [1, 2], affecting more than 300 million people worldwide. Although intraocular pressure lowering can slow down the progression of glaucoma and photodynamic therapy and antivascular endothelial growth factor treatments are effective in neovascular AMD and DR, the retinal

diseases still cannot be cured. Elucidating the disease mechanisms can facilitate the development of new treatments against the retinal diseases [3].

The common pathology in the retinal diseases is the retinal degeneration mediated by cell apoptosis [4, 5]. Antiapoptotic treatments have been proven to prevent the retinal cells from degeneration [6, 7]. Reactive oxygen species (ROS) induces oxidative stress through lipid peroxidation, disruption of normal mitochondrial function, and DNA damage, all of which can initiate the caspase-mediated apoptosis pathway [8]. Hydrogen peroxide (H₂O₂), as one

of the ROS, has widely been used to induce cellular oxidative stress in different cell lines, including retinal pigment epithelial (RPE) and 611W cells [9, 10]. The *in vitro* cell culture can mimic the oxidative disease mechanisms for initial high-throughput drug screening; yet, the *in vivo* model would be more suitable for the development of antioxidative treatments before clinical trials [11, 12]. Intracameral injection of H_2O_2 has been shown to cause edematous ciliary process edema and deterioration as well as corneal endothelial damage [13]. However, the *in vivo* effect and mechanism of H_2O_2 on mammalian retina in experimental models have yet to be determined.

In the current study, we aimed to investigate the effect and mechanism of intravitreal injection of H_2O_2 in mice so as to establish an *in vivo* platform for drug screening against oxidative stress-related retinal diseases. The retinal structure and cell integrity were evaluated by *in vivo* imaging, histological assessment, and gene expression analysis. In addition, cell apoptosis and oxidative stress status in the H_2O_2 -treated retina were also determined.

2. Material and Methods

2.1. Animals. Female C57BL/6J mice (7-week-old, about 20 grams) were purchased from Beijing Vital River Laboratory Animal Technology Co. Ltd., China. The mice were housed under standard conditions of 12:12 hour dark-light cycle with access to standard rodent chow and water *ad libitum*. For each experimental group, 3–4 mice were used for the experiments. All mice were treated according to the guidelines of the Association for Research in Vision and Ophthalmology (ARVO) Statement on Use of Animals in Ophthalmic and Vision Research. The study protocol was approved by the Animal Experimentation Ethics Committee of the Joint Shantou International Eye Center of Shantou University and the Chinese University of Hong Kong.

2.2. Intravitreal Injection of Hydrogen Peroxide. The H_2O_2 solution (catalog number: 88597; Sigma-Aldrich, St. Louis, MO) was diluted with saline to the final concentration of 5, 8, and 10 $\mu\text{g}/\mu\text{l}$. The mice were anesthetized with 1:1 mixture (1.5 ml/kg) of ketamine (100 mg/ml) and xylazine (20 mg/ml), and intravitreal injection with a posterior approach behind the corneoscleral limbus of the eyeball was conducted under a stereomicroscope (MZ 9.5; Leica, Germany) without damaging the lens. A prepulled glass pipette connected to a Hamilton syringe (Reno, NV) and prefilled with mineral oil (Sigma-Aldrich) was used for the H_2O_2 injection. On the starting day (day 0) and the day after (day 1), 1 μl per eyeball of H_2O_2 solution or equal volume of saline control was intravitreally injected with a period over 2 minutes. At postinjection days 2, 7, and 14, the mice were sacrificed for further histological and gene expression analyses.

2.3. In Vivo Imaging. The combination of confocal scanning laser ophthalmoscope (cSLO) with spectral domain optical coherence tomography (OCT; RETImap animal, Roland, Germany) was used for *in vivo* imaging of the retina before

and after the H_2O_2 injection. A superluminescent diode with a wavelength of 830 ± 50 nm was used as the laser source, and a maximum scan speed of 25,000 A-scan/sec was applied. A micrometer-resolution, three-dimensional imaging with infrared cSLO provides a planar visualization of the retina. The digital image depth of the cSLO was 16 frames per second with software module eye-tracking activated. The fundus photographs and OCT images were simultaneously captured on the exact retinal locus in a 30° circle surrounding the optic nerve head. These images were averaged automatically by the built-in software to augment the signal-to-noise ratio. The thicknesses of the retina, defined as the distance between the inner limiting membrane and Bruch's membrane, and the outer nuclear layer (ONL) were manually measured from each averaged OCT image at 600 μm away from the edge of the optic disc. Two images were measured for each experimental mouse.

2.4. Histological Assessment of the Retina. The H_2O_2 -treated mice were anesthetized and sacrificed by the perfusion with 0.9% saline followed by 4.0% paraformaldehyde in 0.1 M $\text{Na}_2\text{HPO}_4/\text{NaH}_2\text{PO}_4$ buffer (pH 7.4). The eyeballs were enucleated, post-fixed in 4.0% paraformaldehyde for 4 hours, and cryoprotected with 30% sucrose/PBS for 2 days. The eyeball slices (10 μm) were sectioned using the vibratome (Leica), stained with hematoxylin and eosin, and imaged with the pupil-optic nerve position using a light microscope (Nikon, Japan). At 600 μm from the edge of the optic nerve cup, the thicknesses of the retina, ONL, inner nuclear layer (INL), and inner plexiform layer (IPL) were measured, and the cell densities of each retinal layer were also counted [14]. Six sections were measured for each experimental mouse.

2.5. Retinal Stress and Apoptosis Analyses. Retinal stress was evaluated by the expression of glial fibrillary acidic protein (Gfap) by the immunofluorescence analysis with rabbit anti-Gfap antibody (1:400; Abcam, the United Kingdom) on the vibratome-sectioned eyeball slices (10 μm). Apoptosis was evaluated using the TUNEL method coupled with fluorescein (DeadEnd™ Fluorometric TUNEL System kit, Promega, Madison, WI) and DAPI counterstain (1:2000; Sigma-Aldrich). Apoptotic cells were visualized under the Leica TCS SP5-II fluorescence confocal microscope and counted at 600 μm from the edge of the optic nerve cup. Six sections were counted for each experimental mouse.

2.6. Gene Expression Analysis. The retina was dissected from the eyeball in the RNAlater (Invitrogen, Carlsbad, CA) immediately after enucleation. Total RNA was extracted from the retina using the TRIzol® reagent (Invitrogen) and reverse transcribed into complementary DNA with random primers using the Superscript First-Strand Synthesis System (TaKaRa, Japan). Gene expression analysis was performed using the Power SYBR Green PCR Master Mix (TaKaRa) with specific primers (Supplementary Table 1). The *Gapdh* gene was used as housekeeping gene for normalization.

2.7. Statistical Analysis. The data was represented as mean \pm standard error of mean (SEM) and analyzed using Stata 14.

The data distribution was analyzed by the Kolmogorov-Smirnov test. The means were compared using the independent *t*-test or one-way or repeated two-way analysis of variance (ANOVA) with Bonferroni post hoc test. Statistical significance was defined as $p < 0.05$.

3. Results

3.1. Hydrogen Peroxide Induced Acute Retinal Damages. *In vivo* OCT images were taken at day 0 (baseline) before the intravitreal injection of H_2O_2 for two consecutive days and at postinjection days 2, 7, and 14 (Figure 1(a)). The OCT images at baseline and day 2 of the H_2O_2 -treated mice showed the typical laminar structure of the retina (Figures 1(b) and 1(c)) without obvious changes in the retina (day 0: $267.99 \pm 7.88 \mu m$ and day 2: $269.32 \pm 3.19 \mu m$; Figure 1(j)) and ONL thicknesses (day 0: $84.24 \pm 1.71 \mu m$ and day 2: $96.29 \pm 0.76 \mu m$; Figure 1(k)), which were similar to the morphologies (Figures 1(f) and 1(g)) and thicknesses in the saline-treated group (retina: $275.998 \pm 7.85 \mu m$ at day 0 and $266.66 \pm 0.83 \mu m$ at day 2, Figure 1(j); ONL: $88.45 \pm 3.29 \mu m$ at day 0 and $96.30 \pm 2.80 \mu m$ at day 2, Figure 1(k)). Yet, the thickness of the photoreceptor inner segment ellipsoid zone (PR) was reduced at day 7 (Figure 1(d)) and more severely at day 14 (Figure 1(e)). Moreover, significant decrease in the thickness of the retina (day 7: $169.69 \pm 15.76 \mu m$, $p = 0.003$ and day 14: $150.74 \pm 4.72 \mu m$, $p < 0.001$; Figure 1(j)) and ONL (day 7: $32.78 \pm 2.47 \mu m$, $p < 0.001$ and day 14: $26.72 \pm 6.06 \mu m$, $p < 0.001$; Figure 1(k)) was also identified in the H_2O_2 -treated mice, compared to that in the saline-treated control (retina: $276.58 \pm 6.20 \mu m$ at day 7 and $280.41 \pm 7.93 \mu m$ at day 14; ONL: $90.98 \pm 0.71 \mu m$ at day 7 and $85.28 \pm 3.07 \mu m$ at day 14, Figures 1(h) and 1(i)).

To verify the results of *in vivo* imaging, the structure of the H_2O_2 -treated retina was resolved by the hematoxylin and eosin staining (Figures 2(a) and 2(b)). At day 2, no alteration of retinal thickness was observed (retina: $177.20 \pm 16.93 \mu m$, $p = 0.841$; ONL: $55.40 \pm 5.33 \mu m$, $p = 0.956$; INL: $41.98 \pm 3.84 \mu m$, $p = 0.344$; and IPL: $56.39 \pm 12.24 \mu m$, $p = 0.559$, Figures 2(c)–2(f)). The retinal degeneration observed in the OCT imaging was confirmed by the histological assessment that, at days 7 and 14, the thicknesses of the retina (day 7: $105.01 \pm 17.85 \mu m$, $p = 0.004$; day 14: $102.25 \pm 17.45 \mu m$, $p = 0.003$, Figure 2(c)), ONL (day 7: $33.76 \pm 9.12 \mu m$, $p = 0.030$; day 14: $40.74 \pm 6.10 \mu m$, $p = 0.032$, Figure 2(d)), INL (day 7: $22.90 \pm 4.06 \mu m$, $p = 0.009$; day 14: $27.16 \pm 3.62 \mu m$, $p = 0.032$, Figure 2(e)), and IPL (day 7: $34.30 \pm 5.27 \mu m$, $p < 0.001$; day 14: $32.17 \pm 5.34 \mu m$, $p < 0.001$, Figure 2(f)) were significantly reduced after the H_2O_2 treatment when compared to the saline control (retina: $173.71 \pm 6.19 \mu m$, ONL: $55.70 \pm 1.89 \mu m$, and INL: $37.68 \pm 2.31 \mu m$). The OCT and histology results indicated that intravitreal injection of H_2O_2 induces the thinning of the laminar structure in the retina. Furthermore, immunofluorescence analysis demonstrated a persistent increase in Gfap expression from days 2 to 14 after the H_2O_2 treatment (Figure 2(g)), indicating the induction of retinal stress by the H_2O_2 treatment.

To confirm the retinal cell damage induced by H_2O_2 , gene expressions of retinal markers were determined after H_2O_2 application. For the photoreceptor cells, rhodopsin gene (*Rho*) expression decreased significantly with increasing doses of H_2O_2 ($5 \mu g$: 0.67 ± 0.03 folds, $8 \mu g$: 0.60 ± 0.03 folds, and $10 \mu g$: 0.38 ± 0.05 folds, $p < 0.001$; Figure 3(b)), compared to the saline control. Similarly, the expressions of other photoreceptor genes [15], recoverin (*Rcvrn*; $8 \mu g$: 0.65 ± 0.05 folds, $p < 0.01$ and $10 \mu g$: 0.47 ± 0.06 folds, $p < 0.001$, Figure 3(c)), nuclear receptor ROR-alpha (*Rora*; $10 \mu g$: 0.62 ± 0.06 folds, $p < 0.001$, Figure 3(d)), and nuclear receptor ROR-beta (*Rorb*; $10 \mu g$: 0.59 ± 0.08 folds, $p < 0.01$, Figure 3(e)) genes, were also dose-dependently downregulated in the H_2O_2 -treated retina, compared to the saline-treated retina. For the amacrine cells, the expression of choline acetyltransferase (*Chat*) was significantly reduced in all doses of H_2O_2 applications ($5 \mu g$: 0.58 ± 0.01 folds, $8 \mu g$: 0.58 ± 0.06 folds, and $10 \mu g$: 0.52 ± 0.09 folds, $p < 0.01$; Figure 3(f)), compared to the saline control. For the retinal ganglion cells (RGCs) [16], the expression of calretinin gene (*Calb2*) was significantly lower in the $8 \mu g$ H_2O_2 -treated group than that of the saline control group by 0.46 folds ($p < 0.01$; Figure 3(g)); yet, the expression of the β III-tubulin gene (*Tubb3*) was significantly higher in the $10 \mu g$ H_2O_2 group than that of the saline control group by 2.45 folds ($p < 0.01$; Figure 3(h)). Nonetheless, the expression of retinal pigment epithelium 65 gene (*Rpe65*) did not show significant changes among the treatment groups (Figure 3(i)). The gene expression results confirmed that the retinal degeneration begins immediately after the H_2O_2 application in mice.

3.2. Hydrogen Peroxide Induced Retinal Cell Apoptosis in a Dose-Dependent Manner. To delineate the mechanisms of the H_2O_2 -induced retinal cell damage, cell apoptosis in the retina was evaluated by the TUNEL assay. No TUNEL-positive cell was found in all retinal layers of the saline-injected mice (Figure 4(b)). In contrast, the TUNEL-positive cells were increased dose-dependently in all retinal layers. In the $10 \mu g$ H_2O_2 -induced group, 11689.15 ± 498.87 cells/ mm^2 of TUNEL-positive cells were found in the ONL ($p = 0.003$), 6108.11 ± 858.73 cells/ mm^2 in the INL ($p = 0.121$), and 2889.52 ± 834.59 cells/ mm^2 ($p = 0.487$) in the ganglion cell layer (GCL; Figure 4(c)). Comparatively, less TUNEL-positive cells were observed in the 5 and $8 \mu g$ H_2O_2 injection ($5 \mu g$: 3085.22 ± 1314.75 cells/ mm^2 in the ONL, $p = 0.297$; 1029.08 ± 289.01 cells/ mm^2 in the INL, $p = 0.904$; and 183.37 ± 99.51 cells/ mm^2 in the GCL, $p = 1.000$; $8 \mu g$: 6773.00 ± 698.94 cells/ mm^2 in the ONL, $p = 0.006$; 3743.66 ± 919.53 cells/ mm^2 in the INL, $p = 0.072$; and 437.08 ± 259.99 cells/ mm^2 in the GCL, $p = 1.000$).

To verify the results of TUNEL assay on the H_2O_2 -treated retina, the expression of apoptosis-related genes was examined. Coherent to the TUNEL results, the expressions of caspase-3 (*Casp3*; $5 \mu g$: 1.84 ± 0.14 folds, $8 \mu g$: 2.39 ± 0.10 folds, and $10 \mu g$: 3.01 ± 0.13 folds, $p < 0.001$; Figure 5(b)) and Bcl-2-associated X (*Bax*; $5 \mu g$: 2.29 ± 0.13 folds, $p < 0.01$, $8 \mu g$: 3.21 ± 0.19 folds, and $10 \mu g$: 4.12 ± 0.35 folds, $p < 0.001$; Figure 5(d)) genes were significantly increased in the H_2O_2 -treated retina in a dose-dependent manner, compared to

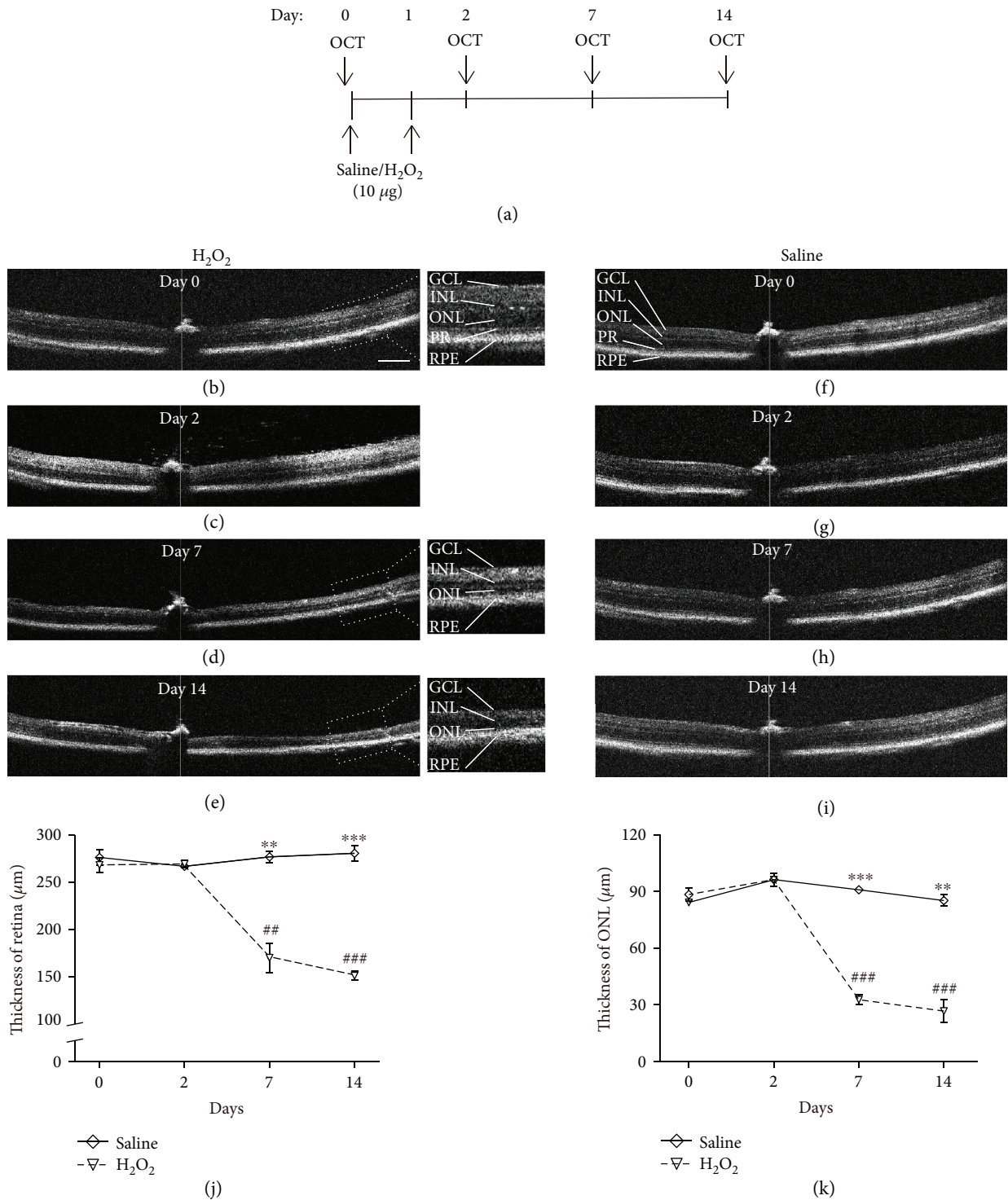


FIGURE 1: OCT analysis of the retina after the H₂O₂ treatment. (a) The schematic diagram of OCT analysis along the H₂O₂ treatment. (b–e) OCT images of the retina at days 0, 2, 7, and 14 of the H₂O₂ group. (f–i) OCT images of the retina at days 0, 2, 7, and 14 of the saline group. Retinal lesion was showed by a decreased in thickness of ONL and disruption of the PR both at days 7 and 14 after H₂O₂ exposure (d, e). (j) The total thickness of the retina showed a significant decrease after 7 days of H₂O₂ treatment and became more severe after 14 days. (k) The local thickness of the ONL also showed a significant decrease after 7 days of H₂O₂ treatment and more severe after 14 days ($n=6$ from 3 mice). ** $p < 0.01$, *** $p < 0.001$ compared to H₂O₂; ## $p < 0.01$, ### $p < 0.001$ compared to day 0 within the H₂O₂ group. Scale bar: 200 μm.

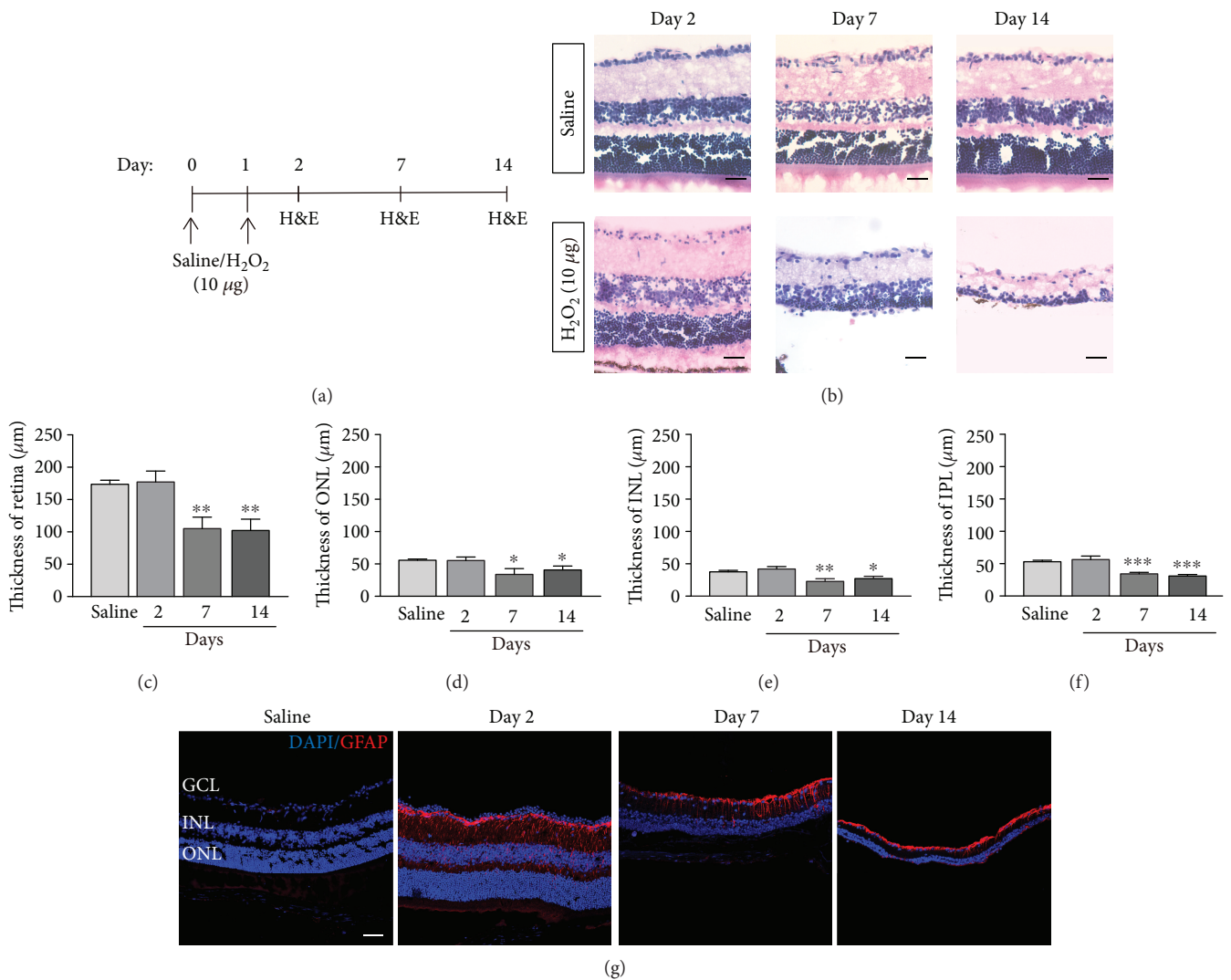


FIGURE 2: Histological assessment of the retina after the H₂O₂ treatment. (a) The schematic diagram of histological analysis after the H₂O₂ treatment. (b) Representative of the retinae stained by hematoxylin and eosin. (c–f) Thickness of the retina, ONL, INL, and IPL after the H₂O₂ treatment. (c) Thickness of the retina was decreased at postinjection days 7 and 14 but not day 2. (d) Thickness of the ONL was decreased at postinjection days 7 and 14 but not day 2. (e) Thickness of the INL was decreased at postinjection days 7 and 14 but not day 2. (f) Thickness of the IPL was decreased at postinjection days 7 and 14 but not day 2. ($n = 5-6$ from 3 mice). * $p < 0.05$, ** $p < 0.01$, and *** $p < 0.001$ compared to the saline group. Scale bar: 200 μm. (g) Glial fibrillary acidic protein (Gfap; red) expression in the retina after the H₂O₂ treatment.

the saline-treated control (Figure 5(a)). In addition, the expressions of caspase-9 (*Casp9*; 1.54 ± 0.15 folds, $p < 0.05$; Figure 5(c)) and poly(ADP-ribose) polymerase family member 8 (*Parp8*; 1.75 ± 0.019 folds, $p < 0.01$; Figure 5(e)) genes were also significantly increased in the 10 μg H₂O₂ group. Yet, the expression of the antiapoptosis gene *Bcl2* did not show statistically significant changes after the H₂O₂ application (Figure 5(f)). Collectively, the TUNEL assay and gene expression results indicated that the H₂O₂-induced retinal degeneration could be caused by cell apoptosis.

3.3. Hydrogen Peroxide Induced Oxidative Stress in the Retina. In addition to cell apoptosis, the expression of antioxidant genes was determined. Compared to the saline

control, the expression of thioredoxin-2 (*Txn2*) gene was significantly upregulated with increasing doses of H₂O₂ (8 μg: 1.53 ± 0.11 folds, $p < 0.01$ and 10 μg: 1.78 ± 0.10 folds, $p < 0.001$; Figure 6(a)). In addition, the application of 8 μg and 10 μg H₂O₂ also showed a significant elevation in superoxide dismutase 2 (*Sod2*) gene expression by 1.34 ± 0.09 folds ($p < 0.01$) and 10 μg: 1.32 ± 0.02 folds ($p < 0.05$), respectively, compared to the saline control (Figure 6(b)). In contrast, the expression of glutathione peroxidase 4 (*Gpx4*) gene did not show statistically significant changes after the H₂O₂ application (Figure 6(c)). These suggested that the increased expression of antioxidant genes could be responded to the H₂O₂-induced oxidative stress elevation in the retina.

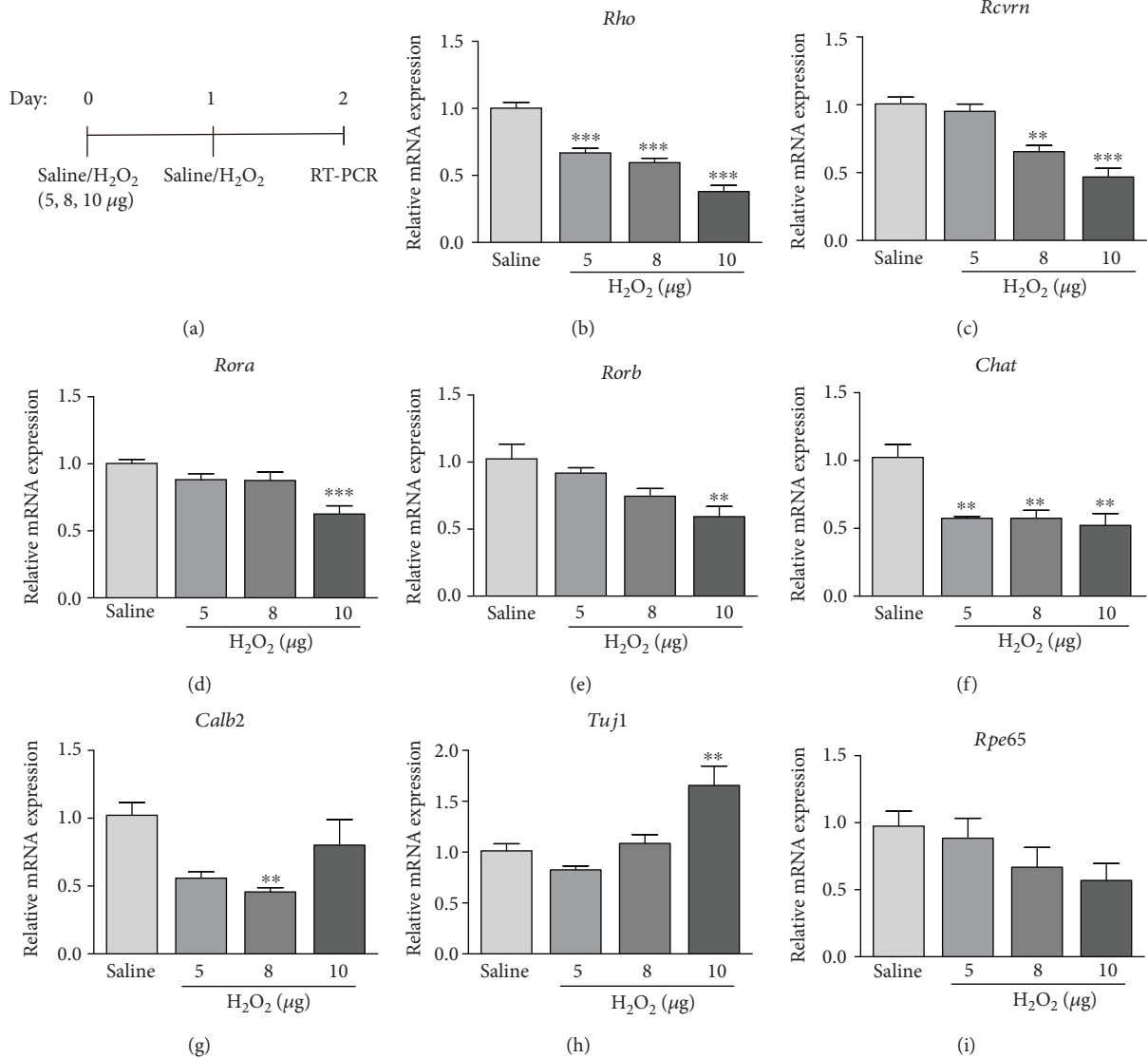


FIGURE 3: Retinal gene expression analysis after the H₂O₂ treatment. (a) The schematic diagram of gene expression analysis after the H₂O₂ treatment. The expression of photoreceptor markers including (b) *Rho*, (c) *Rcvrn*, (d) *Rora*, and (e) *Rorb* genes was decreased significantly with the increasing dose of H₂O₂. (f) The expression of *Chat* gene was decreased significantly after treatment with H₂O₂. (g) The expression of *Calb2* gene was decreased significantly after treatment with 8 µg H₂O₂. (h) The expression of *Tuj1* gene was significantly increased in 10 µg H₂O₂ group. (i) The expression of *RPE65* gene was similar to that in the control group. ***p* < 0.01, ****p* < 0.001, compared to the saline group (*n* = 5–6 from 3 mice).

4. Discussion

In the current study, our results showed that (1) H₂O₂ exposure induces the thinning of the whole retina, ONL, and photoreceptor inner segment ellipsoid zone; (2) H₂O₂ exposure induces retinal cell apoptosis and reduces retinal cell density; and (3) H₂O₂ increases the oxidative stress in the retina.

Retinal cell death is a key pathology of retinal degenerative diseases, including AMD, glaucoma, RP, and DR. The underlying molecular mechanisms still remain elusive. The transgenic models, such as *rd1* or *rd10*, are useful to investigate the pathology and the disease mechanisms of retinal degeneration as they exhibit progressive retinal cell loss

spontaneously [17]; yet, most of them have an early onset of degeneration, and the severity and the onset of the retinal degeneration cannot be modulated [3]. Instead, chemical-induced models can also be applied for the study of disease mechanisms and for the screening of potential treatments. N-Methyl-D-aspartate (NMDA) has been applied to induce RGC apoptosis through the NMDA receptor [18]. In this study, we injected H₂O₂ intravitreally to induce retinal degeneration in mice, and we found that H₂O₂ induces degeneration in all retinal layers (Figures 1 and 2), which the photoreceptor cells are the most severely damaged, followed by the cells in the INL. Interestingly, the expression of β III-tubulin gene (*Tubb3*) showed an increase in the 10 µg H₂O₂ treatment (Figure 3), in which cell apoptosis and

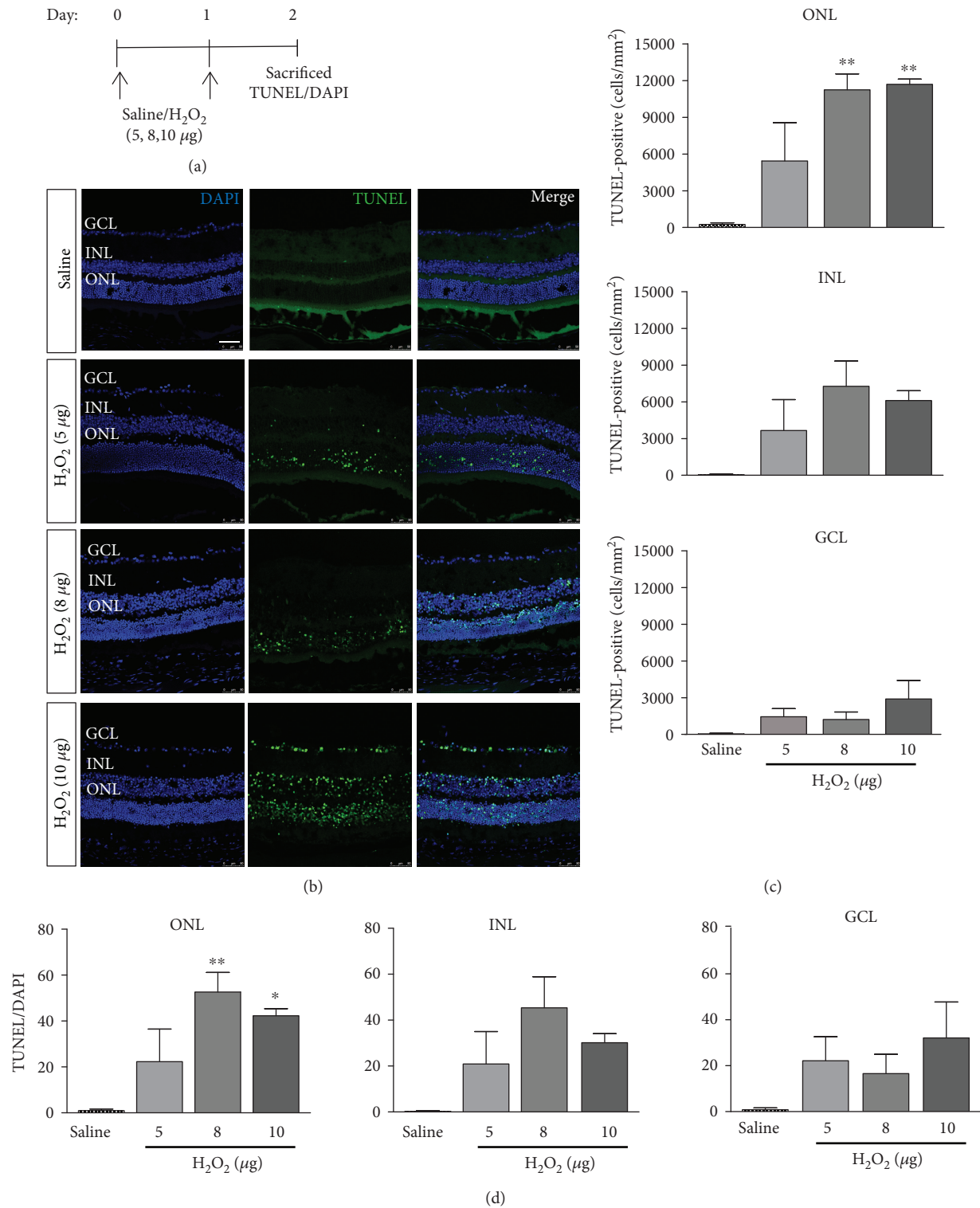


FIGURE 4: Apoptosis analysis in the retina after the H₂O₂ treatment. (a) The schematic diagram of apoptosis analysis after the H₂O₂ treatment. (b) No TUNEL-positive cells were found in the saline group. After H₂O₂ treatment, TUNEL-positive cells were found in both the ONL and INL in each group and sparsely found in GCL in the 10 μg H₂O₂ group. (c) The number of TUNEL-positive cells increased after the H₂O₂ treatment in the ONL, INL, and GCL. (d) The proportions of TUNEL-positive cells showed an increase after H₂O₂ treatment in the ONL, INL, and GCL. **p* < 0.05, ***p* < 0.01 compared to the saline-treated group (*n* = 6 from 3 mice). Scale bar: 50 μm.

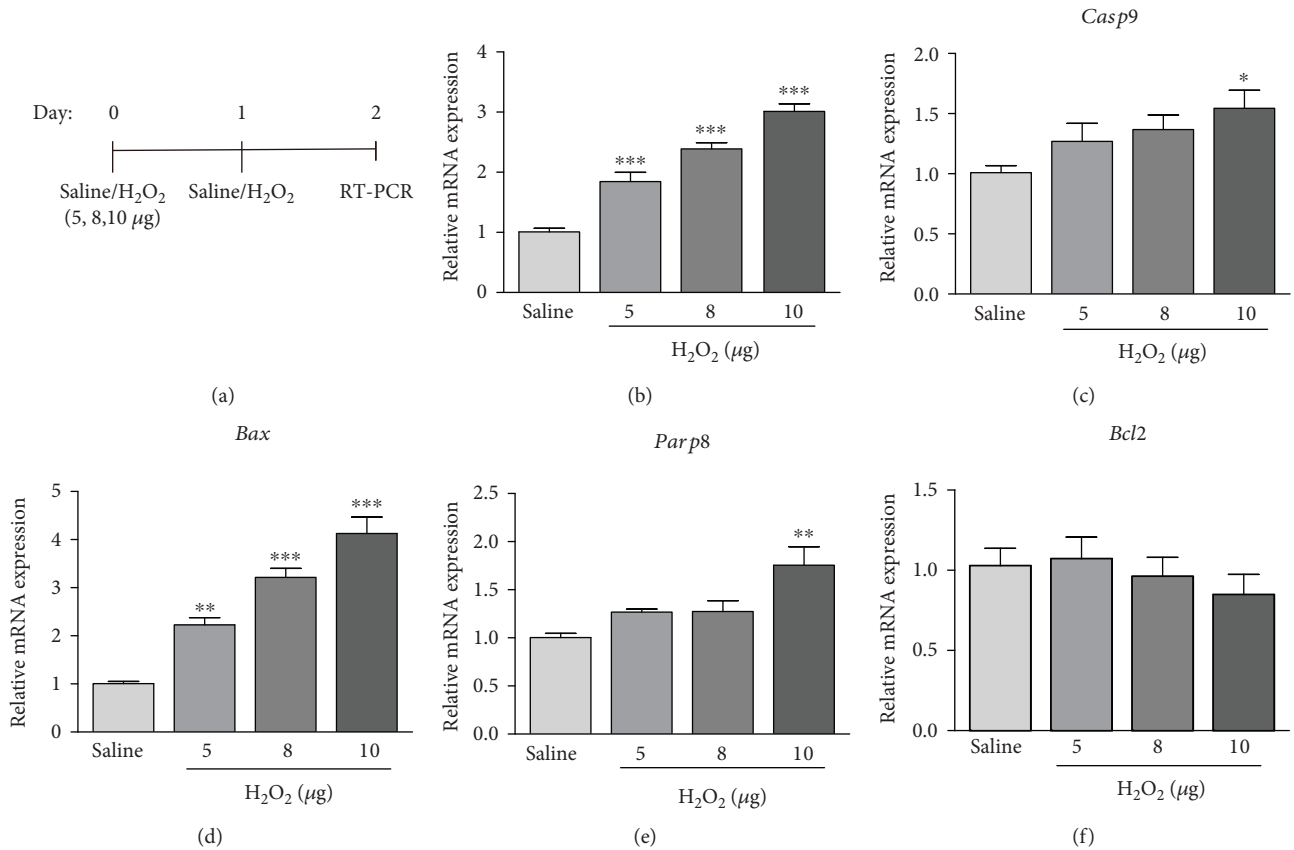


FIGURE 5: Expression analysis of apoptotic genes after the H₂O₂ treatment. (a) The schematic diagram of gene expression analysis after the H₂O₂ treatment. The expression of (b) *Casp3*, (c) *Casp9*, (d) *Bax*, and (e) *Parp8* genes was increased with increasing doses of H₂O₂. (f) The expression of *Bcl2* gene did not show a significant change after the H₂O₂ treatment. **p* < 0.05, ***p* < 0.01, and ****p* < 0.001, compared to the saline-treated group.

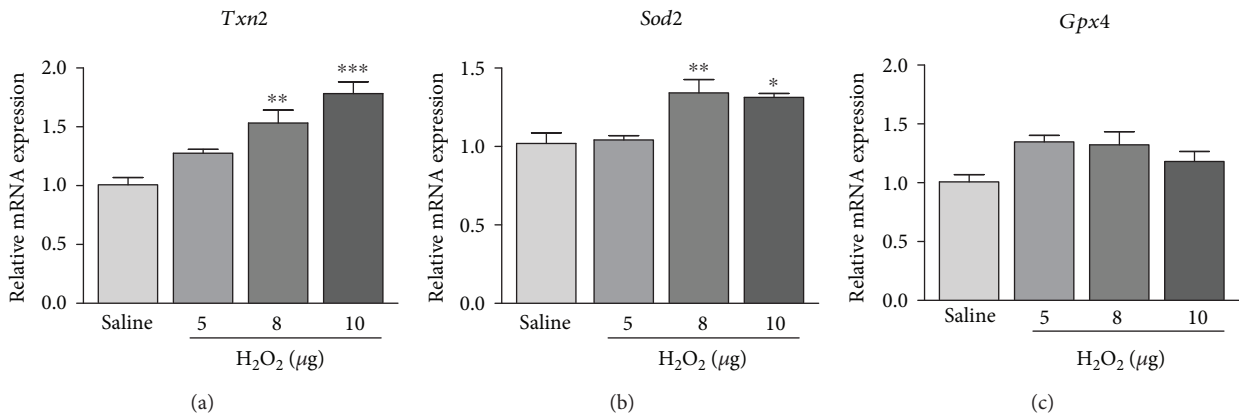


FIGURE 6: Expression analysis of oxidative stress-related genes after the H₂O₂ treatment. After H₂O₂ treatment, the expression of antioxidant genes (a) *Txn2*, (b) *Sod2*, and (c) *Gpx4* increased significantly between groups in the retina. **p* < 0.05, ***p* < 0.01, and ****p* < 0.001, compared to the saline-treated group.

oxidative stress are significantly found. This could be explained by previous reports that overexpression of βIII-tubulin represents a major mechanism of drug resistance to microtubule-interacting agents, such as taxanes and Vinca alkaloids, and βIII-tubulin is conditionally expressed in

A2780 cells after hypoxia [19]. Nevertheless, we demonstrated a dose-dependent retinal damage for the H₂O₂-induced model (Figures 3 and 4), indicating that the severity of retinal degeneration can be modulated and controlled. Besides, the reduction in IPL thickness and the increased

Gfap expression (Figure 2) could resemble retinal stress and ischemia [20], which could be induced by the H₂O₂-mediated oxidative stress.

Previous studies reported that H₂O₂ induces apoptosis in RGCs *in vitro* and inhibits the phosphorylation of p38 and extracellular signal-regulated kinases1/2 [21]. The rabbit study also showed that intracameral injection of H₂O₂ causes morphological changes in the ciliary processes with upregulation of 3-aminotriazole, an inhibitor of catalase [13]. H₂O₂ has been found to be toxic to both the lens and cornea in high concentrations [22]. However, the *in vivo* effect of H₂O₂ exposure to the retina is still poorly understood. This study, for the first time, reported the effect of intravitreal injection of H₂O₂ in mouse retina. Based on the *in vivo* imaging and histological assessment, we confirmed that H₂O₂ induces acute retinal degeneration in mice, in which cell apoptosis begins right after the H₂O₂ application although the retina structure is still maintained (Figures 4 and 5).

Mitochondrial ROS triggers the release of cytochrome c and other proapoptotic proteins, which can trigger caspase activation and apoptosis [23]. H₂O₂ has been found to induce apoptosis in rat nucleus pulposus cells as well as PC12 cells through the mitochondria-mediated pathway [24, 25]. Coherently, we found that H₂O₂ exposure enhances the expression of the mitochondrial antioxidant genes (*Txn2* and *Sod2*) in the retina (Figure 6). Simultaneously, the mitochondrial apoptosis-related genes (*Casp3*, *Bax*, and *Parp8*) are upregulated in the H₂O₂-treated retina (Figure 5). Herein, we postulate that H₂O₂ induces mitochondrial oxidative stress, which in turn triggers the mitochondrial apoptotic pathway and leads to the retinal cell death and the structural disruption. Further investigations could focus on the mitochondrial oxidative stress-targeted pharmacological agents against the H₂O₂-induced retinal degeneration.

In summary, this study revealed that intravitreal injection of H₂O₂ induces acute retinal degeneration in mice by increasing oxidative stress and cell apoptosis. This acute retinal degeneration mouse model could provide a drug screening platform for oxidative stress and retinal diseases.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors indicated no potential conflicts of interest.

Authors' Contributions

BH and HC did the conception and design. HC provided financial support. BH, JLL, XZ, and SWC helped in the collection and/or assembly of data. BH contributed in data analysis and interpretation. BH, TKN, and HC contributed in manuscript writing. TKN and HC helped in the final approval of the manuscript.

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Supplementary Materials

Supplementary Table 1: “the primers used in real-time PCR.” (*Supplementary Materials*)

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