



REVIEW

Recent advances in modulating the microbiome [version 1; peer review: 2 approved]

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Abstract

We are in the midst of “the microbiome revolution”—not a day goes by without some new revelation on the potential role of the gut microbiome in some disease or disorder. From an ever-increasing recognition of the many roles of the gut microbiome in health and disease comes the expectation that its modulation could treat or prevent these very same diseases. A variety of interventions could, at least in theory, be employed to alter the composition or functional capacity of the microbiome, ranging from diet to fecal microbiota transplantation (FMT). For some, such as antibiotics, prebiotics, and probiotics, an extensive, albeit far from consistent, literature already exists; for others, such as other dietary supplements and FMT, high-quality clinical studies are still relatively few in number. Not surprisingly, researchers have turned to the microbiome itself as a source for new entities that could be used therapeutically to manipulate the microbiome; for example, some probiotic strains currently in use were sourced from the gastrointestinal tract of healthy humans. From all of the extant studies of interventions targeted at the gut microbiome, a number of important themes have emerged. First, with relatively few exceptions, we are still a long way from a precise definition of the role of the gut microbiome in many of the diseases where a disturbed microbiome has been described—association does not prove causation. Second, while animal models can provide fascinating insights into microbiota–host interactions, they rarely recapitulate the complete human phenotype. Third, studies of several interventions have been difficult to interpret because of variations in study population, test product, and outcome measures, not to mention limitations in study design. The goal of microbiome modulation is a laudable one, but we need to define our targets, refine our interventions, and agree on outcomes.

Keywords

microbiome, microbiota, antibiotic, probiotic, prebiotic, fecal microbiota transplantation, diet, pharmabiotic

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Introduction: an overview of the gut microbiome

Strictly speaking, the term “microbiome” refers to the collection of genomes from all micro-organisms in a given environment whereas the term “microbiota” refers to all the micro-organisms found in the environment. In practice, these terms are often used interchangeably. The term microbiota has replaced “flora” in order to emphasize the diversity of microbiota and, in particular, that the human intestinal microbiota normally consists not just of bacteria but also of archaea, viruses, fungi, and multicellular parasites. We now know that it represents a highly evolved and complex ecosystem that plays an important role in the development and maintenance of homeostasis^{1,2}. Our understanding of the composition and functions of the gut microbiome has been permitted by dramatic and ever-evolving technologies that identify micro-organisms and describe their genetic makeup and metabolism³. We can now annotate, to an ever-increasing depth of detail, what a given microbiome contains, what its constituents are capable of doing (through an interrogation of their genomes), and what they actually produce (employing metabolomics and other techniques). These rapid advances have been facilitated by equally important advances in informatics which allow us to make sense of the enormous databases that microbiota studies generate; techniques such as network analysis and machine learning help to provide meaningful interpretations^{4,5}. From a variety of laboratory models, such as germ-free and humanized, as well as selective knockout, animal (mostly mouse) models⁶, insights have been gained into the many interactions between the gut microbiome and the host^{1,2,7}. For example, studies on germ-free mice have clearly demonstrated the negative impact of the absence of microbiota on the development and maturation of the immune system^{2,8,9}. Now we learn, again from animal models, of the role of the gut microbiome in the development and ongoing functionality of the central nervous system¹⁰. Although such studies enable considerable flexibility in terms of manipulation of genotype and phenotype, permit a wide range of possible interventions, and facilitate the collection of various biological samples, they are not without their shortcomings^{11–13}, and extrapolations to the human condition must be cautious.

What do we know of the human gut microbiome? There certainly has been no shortage of studies on the composition and, to a lesser extent, on the function of the gut microbiome in humans. There are obvious limitations to the scope of studies that can be performed in human subjects in contrast to animal and *in vitro* models, limitations that the reader must be aware of in perusing the literature. It must be remembered that this is a new field and, although progress is being made¹⁴, there is still a lack of standardization on many of the technical details of microbiome analysis of human samples¹⁵. Although various studies have described links between an altered microbiota and not only gastrointestinal disorders but also diseases as diverse as obesity, diabetes, non-alcoholic fatty liver disease, cancer, and Parkinson’s disease^{16,17}, these are, at best, associations and do not define causation¹⁸. Furthermore, many limitations in patient selection and study design limit the interpretation of many of these studies¹⁸.

Although there are some divergent studies¹⁹, it is generally agreed that the human gut is relatively sterile at birth^{20–24} and acquires its commensal gut microbiome during birth from the mother’s birth canal and thereafter from its oral intake and immediate environment^{25,26}. Microbial diversity rapidly increases over the first three years of life and then stabilizes at a composition that resembles that of an adult^{25,26}; this early and critical phase in the development of the microbiome may be especially vulnerable to modulations both beneficial and detrimental²⁷. The origins of diseases that become manifest in adulthood may well be found in the infant microbiome.

Although it is possible that a host of factors influence the adult microbiome, age, geography, diet, and medications have emerged as the principal drivers of inter-individual variation^{28–36}. However, large population studies revealed that only a small proportion of the variation in the microbiome between individuals could be explained by these and other identifiable factors^{32,33}—we have much to learn.

Needless to say, there has, of late, been considerable interest in strategies that modulate the gut microbiota as well as in microbiota as sources of novel biologically active molecules^{37–39} and predictors of response to various interventions^{40,41}. Our focus will be on the former: an exploration of strategies to modulate the microbiome. Here, we will consider the range of currently available approaches (Table 1), explore their impacts, and assess the potential for novel interventions.

Modulating the microbiome

In considering any intervention that seeks to successfully and, we assume, beneficially modulate the microbiome, one needs to be ever mindful of the complex and dynamic milieu (discussed in the Introduction) which awaits. Simplistic concepts of how a given supplement or medication might influence the microbiota–host interface have generated much hype and even more disappointment. An appreciation of the range of possible interactions between an intervention and host diet, genome, immune system as well as with resident commensals should alert one to the challenges that lie ahead.

Table 1. Range of interventions that may modulate the microbiome.

- Lifestyle modification
 1. Nutritional intervention and modification; the importance of diet in the short and the long term
 2. Caloric restriction
 3. Exercise
 4. Other lifestyle factors
- Clinical interventions
 1. Fecal microbiota transfer
 2. Antibiotics
 3. Prebiotic and probiotics
 4. Pharmabiotics
 5. Impact of non-antibiotic drugs on the microbiome

We limit the term “modulation” to refer to the manipulation of one or more of the following targets: first, the relative distribution of bacterial species or strains; second, the actual number of bacteria; third, their metabolic activity; fourth, their interactions with the host. There may well be other targets that could be modified—virulence, bacterial antigens, and biofilms, for example—but we have chosen to limit our scope to the aforementioned. In theory, the goal of modulation could be to restore a disrupted or depleted microbiota or transform the existing status quo to induce a “healthier” bacterial community. It must be emphasized that these goals are for now overly simplistic and, despite the claims of various commercial entities that offer microbiome analysis, we are still some way from fully understanding what constitutes a healthy microbiota throughout the gastrointestinal tract. When the microbiome is manipulated, attention must always be paid to the potential for negative outcomes such as the inadvertent introduction or promotion of pathogenic species, transference of antibiotic resistance, or induction of deleterious host responses.

Lifestyle modification

Diet and the microbiome

It is now abundantly evident that diet is a major modifier, both in the short and in the long term, of the gut microbiome; this makes absolute sense as, for the most part, microbiota depend for their sustenance on what we ingest. Evidence for the long-term effects of diet comes from studies comparing communities²⁸ or individuals^{29–33,42–44} with very different dietary habits. These differences reflect lifelong or, at the very least, very long-term dietary practices. In the shorter term, very significant changes in diet, such as reducing fiber intake^{45,46}, excluding gluten^{47,48} or fermentable oligo-, di-, or monosaccharides and polyols (FODMAPs)⁴⁹, or dramatically increasing protein intake⁵⁰, can also impact microbiome composition.

Other dietary components have also been shown to influence microbiota composition⁴². High-carbohydrate diets promote the growth of *Clostridium* cluster XVIII, *Lachnospiraceae*, and *Ruminococcaceae* at the expense of *Bacteroides*, *Bifidobacteria*, and *Enterobacteriaceae*, whereas diets high in fat promote bile-tolerant genera such as *Alistipes*, *Bacteroides*, and *Bilophila* and high-protein diets favor butyrate-producing bacteria such as *Roseburia*, *Eubacterium rectale*, *Faecalibacterium prausnitzii*, *Lactobacilli*, and *Bacteroides*^{42–44}.

The role of dietary fiber in the development and sustenance of the colonic microbiota has been recognized for decades. Effects of dietary fiber on colonic transit have been linked with the preventative effects of fiber in relation to a variety of diseases as well as in the treatment of disorders such as chronic constipation³⁵. In turn, these beneficial effects may be related to interactions between fiber and colonic bacteria.

Accumulating evidence indicates that effects of fiber on microbiota may be more complex and this should come as no surprise given the heterogeneity of the molecular structures that are found under the umbrella of the term “fiber”⁵¹. For example, in the American Gut project, it was found that the number

of unique plant species consumed, rather than being a vegan or omnivore, was the best predictor of microbial diversity⁵². Very specific effects may be linked to the intake of certain fibers – for example, in a randomized clinical trial from China among subjects with type II diabetes mellitus it was found that fibers that promoted the growth of strains that produced short-chain fatty acids resulted a greater amelioration of hemoglobin A1c levels than those that did not⁵³. Clearly, there is much to be learned about the effects of fibers on gut microbiota.

In addition to fiber, dietary ingredients and food additives have been shown to have a substantial impact on the gut microbiota⁴⁴. Suez and colleagues, for example, found that mice (and, in limited data, humans) consuming non-caloric artificial sweeteners were prone to the development of glucose intolerance, possibly mediated by changes to the intestinal microbiota⁵⁴. With regard to other supplements and additives, recent research has revealed the role of vitamin D in determining microbiota composition⁵⁵. Curcumin, which has attracted much interest of late for potential anti-inflammatory and anti-cancer properties, also appears to exert anti-bacterial effects. These include the inhibition of biofilm production and the down-regulation of quorum-sensing virulence factors such as alginate, swarming, and motility^{56–58}.

It stands to reason that, though less studied, dietary strategies that involve the exclusion of individual but commonly consumed food items or even whole food groups are likely to alter the composition of microbiota. Some of these approaches may, at least in theory, pose problems for the microbiota; the exclusion of FODMAPs, gluten, and fiber, for example, has the potential to deprive important members of the colonic microbiome, such as *Bifidobacteria*, *Prevotella*, and *Bacteroides*, of key nutritional factors such as oligosaccharides and fiber. Although such effects have been demonstrated in the short term⁴⁹, the longer-term implications are unknown. Changes in the fecal microbiome have indeed been described in relation to this diet; a reduction in *Bifidobacteria* being most notable^{59–61}. The clinical impact of these and other dietary changes in the long term, in particular, remains unclear⁶². The Mediterranean diet, for example, has been much lauded for its potential to reduce risk for cardiovascular disease and colon cancer; yet, when formally tested, it did not impact on one microbial metabolite, trimethylamine N-oxide (TMAO), that has been linked with risks for both atherosclerosis and colon cancer⁶³.

What is abundantly clear from all of the above observations is that the impact of diet must be accounted and corrected for in any study of the microbiome in humans. It is also evident that the microbiome contains considerable functional redundancy which allows it to maintain stability in the face of dietary shifts⁶⁴; this was exemplified by the work of Reichardt and colleagues on short-chain fatty acid production⁶⁵.

Caloric restriction

The challenges that dietary studies face are illustrated by an extreme dietary strategy: fasting. Although changes in microbiota diversity and composition have been described in anorexia

nervosa and related eating disorders, it has proven difficult to disentangle cause from effect. It would be surprising if fasting, if prolonged, did not impact the gut microbiome⁶⁶; what remains to be defined is whether there are microbiota signatures specific for eating disorders that might play a role in the pathogenesis of these disorders⁶⁷⁻⁶⁹. Given the interest that surrounds the potential role of the gut microbiome in obesity, the participation of the microbiome in various calorie-reducing strategies has been the subject of some study. Fasting-induced changes in the microbiota have not only been associated with beneficial metabolic effects⁷⁰ but also have demonstrated positive effects on intestinal inflammation⁷¹ and even central nervous system disorders^{72,73}. The possible contribution of the microbiome to weight loss and the beneficial metabolic impacts of bariatric surgery have also been explored. A variety of changes in the fecal microbiome have been demonstrated following gastric bypass and other bariatric procedures and were summarized in a recent systematic review⁷⁴. Guo and colleagues concluded, on the basis of 12 animal experiments and nine clinical studies, that four phyla—*Bacteroidetes*, *Fusobacteria*, *Verrucomicrobia*, and *Proteobacteria*—increased following bariatric surgery but that *Firmicutes*, *Clostridiales*, *Clostridiaceae*, *Blautia*, and *Dorea* were reduced⁷⁴.

One potentially detrimental consequence of a limited or inadequate dietary intake is that bacteria may turn to host glycans in the mucus layer as substitutes for dietary glycans, thereby upsetting the integrity of the mucus layer which is maintained, in health, through specific bacteria–nutrient interactions^{75,76}. This disruption of the mucus layer seems to be especially likely to occur in fiber-deprived diets and may render the host more susceptible to pathogens⁷⁷. This is not to say that the degradation of host glycans is inevitably deleterious, as exemplified by the associations between *Akkermansia muciniphilia* and positive health status⁷⁸.

Exercise

Similar challenges are found in attempting to assess the impact of exercise on the microbiome given the almost universal linkage between physical exercise and dietary habit as part of what is referred to as a “healthy lifestyle”⁷⁹. At one extreme, professional athletes commonly consume much higher amounts of protein which also impact on the composition of the microbiome⁸⁰. Other interactions also complicate the effects of exercise; for example, body habitus (lean versus obese) significantly affected the impact of 6 weeks of endurance exercise on microbial diversity and short-chain fatty acid concentration⁸⁰; this finding may reflect diet-related changes in the pre-exercise microbiota. Nevertheless, accumulating evidence indicates that exercise has an independent effect on the microbiome⁸¹. For example, an increase in members of the genus *Veillonella* has been identified among marathon runners, and inoculation of these same bacterial taxa into mice was shown to promote endurance by converting exercise-induced lactate into propionate⁸².

Other lifestyle factors

Other lifestyle factors, such as cigarette smoking⁸³⁻⁸⁵, alcohol consumption^{85,86}, and recreational drug use⁸⁷, have also been

linked to changes in the microbiota. With respect to the first of these, the oral microbiota has been of special interest⁸⁸ given the known relationships between cigarette smoking and oral cancer.

Clinical interventions

Fecal microbiota transfer

It has only been in the last decade or so that fecal microbiota transplantation (FMT), though apparently employed on an empiric basis for centuries (if not millennia), has achieved some degree of scientific respectability. Most impressive have been results in recurrent *Clostridioides difficile*–associated disease (CDAD), where cure rates up to and in excess of 90% have been reported⁸⁹. Various preparations and delivery protocols have been employed with some differences in apparent efficacy⁹⁰; what remains to be determined is what components of the transplanted fecal microbiota are truly essential for efficacy. FMT has been widely used on an empiric basis in a host of other indications, and instructions for the performance of FMT at home can even be found on the internet. This practice is ill advised: recent reports of severe systemic infections and even death following FMT remind us of the potential hazards of this therapy⁹¹. It is notable that results from the use of FMT in other indications are far less impressive than those reported in CDAD. This should come as no surprise as one has now strayed from a disorder caused by a single organism to ones of varying phenotype where, despite considerable efforts, the precise role of the microbiome in etiology remains unclear. Thus, although systematic reviews and some individual trials suggest efficacy for FMT in ulcerative colitis⁹²⁻⁹⁴ and irritable bowel syndrome (IBS)^{95,96}, results from individual studies provide a far from clear-cut picture; some report either no benefit or even inferior outcomes for FMT⁹⁷⁻⁹⁹. FMT is clearly a powerful tool but a very blunt instrument; it is a technology in need of considerable refinement once one strays from CDAD. Results in more complex polygenic diseases will undoubtedly require a much more tailored and personalized approach which ultimately should involve the definition of the microbial cocktail that is most effective for each phenotype¹⁰⁰. The small intestinal microbiome has long been recognized to play a pivotal role in the pathogenesis of the symptomatology of hepatic encephalopathy; limited clinical trial data suggest that FMT may also have a role here¹⁰¹. The microbiome may play a more fundamental role in the etiology of non-alcoholic fatty liver disease (NAFLD) and its more advanced manifestation, non-alcoholic steatohepatitis¹⁰²; here, microbiome modulation, including FMT, holds promise^{103,104}; clinical trials are awaited. It must also be remembered that the gut microbiome includes organisms other than bacteria, such as viruses¹⁰⁵, which therefore may be transmitted via or influenced by FMT^{106,107}.

FMT is not without risk. Not only can infectious agents be transmitted (as illustrated by recent instances of transmittal of extended-spectrum beta-lactamase–producing *Escherichia coli* which proved fatal in one instance)¹⁰⁸ but it is also theoretically possible that the transfer of microbial signatures linked to disease states leads to the future emergence of these disorders in the recipient, hence the call for greater regulation of FMT¹⁰⁹.

Another barrier to progress is our lack of understanding of how exactly FMT works or how it might work in different clinical situations¹¹⁰. Clues are beginning to emerge, especially in relation to efficacy in CDAD^{111,112}, but the exact bacterial recipe required for benefits in CDAD or other potential indications has yet to be defined. It seems likely that the composition of the donated material will differ substantially between disease states.

Antibiotics

A detailed discussion of the indications for and efficacy of antibiotics in human health is beyond the scope of this review. Inevitably, antibiotics that are given orally or that undergo biliary excretion and enterohepatic circulation, regardless of the route of administration, will impact on the gut microbiome to a greater or lesser extent. These “innocent bystander” effects may impair host resistance to pathogens, setting the stage for CDAD or fungal overgrowth, and certain populations are especially at risk¹¹³. Antibiotic resistance is a global public health issue; global trends in resistance to one common pathogen, *Helicobacter pylori*, are described as “alarming”¹¹⁴. The human gut microbiota has been described as “a reservoir of antibiotic resistance genes”¹¹⁵; in one study 1,093 antibiotic resistance genes were identified among apparently healthy Chinese individuals¹¹⁵; the potential for horizontal and vertical transfer of such resistance is a source of great concern.

Antibiotics mediate other actions through effects on the microbiota, including effects on inflammation, metabolism, and tumorigenesis; the net impact, whether detrimental or beneficial, is determined by antibiotic, microbial, and host factors^{116–122}. The long-term implications for human health of these antibiotic effects are only now being appreciated^{122,123}. Infants seem to be especially vulnerable; accumulating evidence indicates that early and repeated exposure to antibiotics in infancy, even in the very small doses that we ingest through the food chain as a consequence of their use in animal husbandry, may predispose to the development of inflammatory and metabolic diseases in later life^{121–128}. A call to arms to address the global use of antibiotics is certainly appropriate¹²⁹.

Probiotics and prebiotics

The role of prebiotics and probiotics in gastrointestinal health and disease has been the subject of a recent review by one of the authors of this review¹³⁰ and has also been the subject of a very recent review in this journal¹³¹ and therefore will not be repeated in detail here. The International Scientific Association for Probiotics and Prebiotics defines a prebiotic as “a substrate that is selectively utilized by host microorganisms conferring health benefit”¹³². Probiotics are most commonly defined as “live microorganisms which when administered in adequate amounts confer a health benefit on the host”¹³³. Therefore, in their simplest terms, prebiotics are substances that act as substrates for bacterial digestion and thus proliferation and lead to the generation of metabolites, such as short-chain fatty acids, that are beneficial to the host¹³⁴, whereas probiotics are live organisms that engage in beneficial interactions with the host.

Substances with prebiotic effects may be found in cereals as well as in plants such as onions, garlic, bananas, chicory

root, and Jerusalem artichokes but typically are present at low levels and may not exert prebiotic effects in these forms¹³⁵. Fructo-oligosaccharides (FOSs) are known to be present in about 36,000 varieties of plants, and wheat is a major source of fructans. More biologically active and selective prebiotics include galacto-oligosaccharides (GOSs), FOSs, oligofructose (OF), chicory fiber, and inulin. Human milk oligosaccharides are important prebiotics provided in breast milk to infants and promote the proliferation of *Bifidobacteria* and, in this manner, have been linked to a number of health benefits¹³⁶. Other components of breast milk also exert beneficial impacts on the infant’s microbiome and immune system^{137,138}. Research on prebiotics currently includes a focus on the development of highly selective “designer” prebiotics targeted to impact on specific taxa within gut microbiota; others question the wisdom of the importance or, indeed, the feasibility of the selectivity approach given the functional redundancy that is inherent to the gut microbiome^{139,140}.

Of late, clinical studies on prebiotics have a strong emphasis on metabolic outcomes such as blood glucose regulation, calcium homeostasis, and weight loss^{132,135,141}. Immunological (for example, enhancing antibody responses to vaccines, promoting anti-inflammatory cytokine profiles)^{135,142}, neurological (some benefits in terms of mood and cognition)¹³⁴, cardiovascular (improvement in lipid profiles)¹³⁵, and gastrointestinal effects have also been studied; notable examples are effects on colon transit and benefits in IBS⁶¹.

Probiotics have been lauded for centuries for a host of beneficial effects; most await confirmation in high-quality clinical trials^{130,131}. Nevertheless, a considerable volume of basic science research attests to the ability of various probiotic strains to engage with the mucosal immune system, modulate host metabolism, and even influence gut neuromuscular function^{130,143}. More remote effects on the liver and central nervous system have also been demonstrated for orally ingested probiotics^{144,145}. We now have a considerable understanding of how probiotics interact with the host to generate these effects; for example, the molecular basis of the anti-inflammatory effects of certain species of *Bifidobacteria* have been described in great detail in elegant *in vitro* and animal studies¹⁴⁶.

As ever, the situation in humans is less straightforward and recent studies emphasize the complexity of the interactions between the administered probiotic, commensal microbiota, and the host that determine the ability of the probiotic to gain a foothold in the gut, colonize, and exert its effects^{147,148}. A vision of a probiotic as simply displacing “bad” bacteria is clearly very naïve.

Of the myriad clinical claims that have been made for probiotics, a few pass muster and have been detailed in several recent reviews and meta-analyses^{130,131}. The most consistent benefits have been described in the prevention or treatment (or both) of diarrhea in children^{149,150}, antibiotic-associated diarrhea^{151,152}, necrotizing enterocolitis¹⁵³, IBS¹⁵⁴, and some phenotypes of inflammatory bowel disease^{155–157}. It must be stressed that these are aggregate results; though stating that probiotics, in general, have an effect, conclusions are unable to provide direction to

the clinician on a specific preparation for a given indication¹⁵⁷. Results from individual studies are inconsistent, and major deficits in study design often limit interpretability. Given the tremendous inter-individual variability in the composition of the gut microbiome, it may be unrealistic to expect consistent results from a given microbial strategy in any disease state. Efforts to define what microbial or host factors determine responses could pave the way toward “personalized bacteriotherapy”. There is much to be done.

Probiotics and prebiotics may also be combined as synbiotics; although this concept is attractive in theory and synbiotic preparations have enjoyed some notable successes¹⁵⁸, synergy is not inevitable¹⁵⁹ nor is it always possible to tease out the relative contributions of probiotic or prebiotic to any observed benefit.

Pharmabiotics

The term “pharmabiotic” has been coined to encompass any material with potential health benefit that can be mined from microbiota, microbiota–host, or microbiota–dietary interactions in the gut^{160,161}; therefore, it includes not just live organisms but dead or altered organisms as well as bacterial products or metabolites. Some concrete examples include bacterially produced natural antibiotics, bacteriocins¹⁶², genetically modified organisms^{163,164}, bacteriophages^{165,166}, and short-chain fatty acids¹⁶⁷. *E. coli* has been engineered to exert a variety of effects, including overproducing AI-2 signaling molecules and thereby beneficially tilting the Firmicutes/Bacteroidetes ratio in a mouse model of streptomycin-induced dysbiosis¹⁶⁸. Vaccination against *Vibrio cholerae* infection in the gut has been achieved with *E. coli* overexpressing both AI-2 and the genus-specific autoinducer-1, CA-1¹⁶⁹; in another example, an engineered *E. coli* seeks and kills *Pseudomonas aeruginosa* via quorum sensing and expression of antimicrobial peptides¹⁷⁰. In another approach, the exonuclease Cas3—clustered regularly interspaced short palindromic repeats (CRISPR)-associated protein 3—from type I systems was engineered into a probiotic to selectively and efficiently kill pathogenic bacteria with specific genetic properties¹⁷¹.

Though still rather new in terms of clinical application, these and other technologies offer exciting possibilities for microbiota modulation in the future and may be vital to

the resolution of the antibiotic crisis that we currently face. Evolving approaches such as CRISPR-based technologies have revolutionized genome editing and have already been applied to the development of novel antimicrobial strategies^{172–174}.

Impact of non-antibiotic drugs on the microbiome

Interventions that modulate intrinsic defense mechanisms against bacterial colonization can be predicted to alter microbiota composition. Acid suppression induced by proton pump inhibitors (PPIs) has been variably but not consistently linked to a predisposition to *Clostridioides difficile* infection, enteric infections, and small intestinal bacterial overgrowth^{175,176}. Studies of human feces have indeed demonstrated a decrease in *Clostridiales* and an increase in *Actinomycetales*, *Micrococcaceae*, and *Streptococcaceae* among PPI users; these changes were previously associated with an increased susceptibility to this feared complication of antibiotic use^{177–179}. Similarly, drugs that alter motility and intestinal transit, of which there are many, may also alter microbiota composition¹⁸⁰. It is likely that many other drugs engage with the microbiota with resultant enhancement or reduction in efficacy or induction of side effects^{177,181}, yet another fertile field for future microbiome research.

Conclusions

The study of the human gut microbiome has emerged as one of the hottest areas of biology and biomedicine and continues to yield tantalizing insights into the contributions of our microbial fellow travelers to health and disease. Accordingly, the modulation of the microbiome to prevent or treat disease has attracted considerable attention, and various strategies have emerged. In most instances, however, progress has been hampered by a lack of clarity on the precise role of the microbiome in a given disorder, variations in human disease phenotype, and variability in formulation and delivery of putative therapies. Progress on all fronts is required to move microbiota modulation to the forefront of medical practice.

Abbreviations

CDAD, *Clostridioides difficile*-associated disease; CRISPR, clustered regularly interspaced short palindromic repeats; FMT, fecal microbiota transplantation; FODMAPs, fermentable oligo-, di-, or mono-saccharides and polyols; FOS, fructo-oligosaccharide; IBS, irritable bowel syndrome; PPI, proton pump inhibitor

References

- Victor DW 3rd, Quigley EM: **The Microbiome and the Liver: The Basics**. *Semin Liver Dis.* 2016; **36**(4): 299–305.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Dominguez-Bello MG, Godoy-Vitorino F, Knight R, et al.: **Role of the microbiome in human development**. *Gut.* 2019; **68**(6): 1108–14.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Claesson MJ, Clooney AG, O'Toole PW: **A clinician's guide to microbiome analysis**. *Nat Rev Gastroenterol Hepatol.* 2017; **14**(10): 585–95.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Camacho DM, Collins KM, Powers RK, et al.: **Next-Generation Machine Learning for Biological Networks**. *Cell.* 2018; **173**(7): 1581–92.
[PubMed Abstract](#) | [Publisher Full Text](#) | **F1000 Recommendation**
- Mallick H, Ma S, Franzosa EA, et al.: **Experimental design and quantitative analysis of microbial community multiomics**. *Genome Biol.* 2017; **18**(1): 228.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Ito R, Takahashi T, Ito M: **Humanized mouse models: Application to human diseases**. *J Cell Physiol.* 2018; **233**(5): 3723–8.
[PubMed Abstract](#) | [Publisher Full Text](#)



7. Adak A, Khan MR: **An insight into gut microbiota and its functionalities.** *Cell Mol Life Sci.* 2019; **76**(3): 473–93.
[PubMed Abstract](#) | [Publisher Full Text](#)
8. Bhattarai Y, Kashyap PC: **Germ-Free Mice Model for Studying Host-Microbial Interactions.** *Methods Mol Biol.* 2016; **1438**: 123–35.
[PubMed Abstract](#) | [Publisher Full Text](#)
9. Uzbay T: **Germ-free animal experiments in the gut microbiota studies.** *Curr Opin Pharmacol.* 2019; **49**: 6–10.
[PubMed Abstract](#) | [Publisher Full Text](#)
10. **F** Dinan TG, Cryan JF: **The Microbiome-Gut-Brain Axis in Health and Disease.** *Gastroenterol Clin North Am.* 2017; **46**(1): 77–89.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
11. Arrieta MC, Walter J, Finlay BB: **Human Microbiota-Associated Mice: A Model with Challenges.** *Cell Host Microbe.* 2016; **19**(5): 575–8.
[PubMed Abstract](#) | [Publisher Full Text](#)
12. McCoy KD, Geuking MB, Ronchi F: **Gut Microbiome Standardization in Control and Experimental Mice.** *Curr Protoc Immunol.* 2017; **117**(1): 23.1.1–23.1.13.
[PubMed Abstract](#) | [Publisher Full Text](#)
13. Mooser C, Gomez de Agüero M, Ganal-Vonarburg SC: **Standardization in host-microbiota interaction studies: challenges, gnotobiology as a tool, and perspective.** *Curr Opin Microbiol.* 2018; **44**: 50–60.
[PubMed Abstract](#) | [Publisher Full Text](#)
14. Costea PI, Zeller G, Sunagawa S, *et al.*: **Towards standards for human fecal sample processing in metagenomic studies.** *Nat Biotechnol.* 2017; **35**(11): 1069–76.
[PubMed Abstract](#) | [Publisher Full Text](#)
15. Song EJ, Lee ES, Nam YD: **Progress of analytical tools and techniques for human gut microbiome research.** *J Microbiol.* 2018; **56**(10): 693–705.
[PubMed Abstract](#) | [Publisher Full Text](#)
16. Quigley EMM: **Microbiota-Brain-Gut Axis and Neurodegenerative Diseases.** *Curr Neurol Neurosci Rep.* 2017; **17**(12): 94.
[PubMed Abstract](#) | [Publisher Full Text](#)
17. **F** Noce A, Marrone G, Di Daniele F, *et al.*: **Impact of Gut Microbiota Composition on Onset and Progression of Chronic Non-Communicable Diseases.** *Nutrients.* 2019; **11**(5): pii: E1073.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
18. Quigley EMM: **Gut microbiome as a clinical tool in gastrointestinal disease management: are we there yet?** *Nat Rev Gastroenterol Hepatol.* 2017; **14**(5): 315–20.
[PubMed Abstract](#) | [Publisher Full Text](#)
19. Willyard C: **Could baby's first bacteria take root before birth?** *Nature.* 2018; **553**(7688): 264–6.
[PubMed Abstract](#) | [Publisher Full Text](#)
20. Perez-Muñoz ME, Arrieta MC, Ramer-Tait AE, *et al.*: **A critical assessment of the "sterile womb" and "in utero colonization" hypotheses: implications for research on the pioneer infant microbiome.** *Microbiome.* 2017; **5**(1): 48.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
21. Walker WA: **Bacterial Colonization of the Newborn Gut, Immune Development, and Prevention of Disease.** *Nestle Nutr Inst Workshop Ser.* 2017; **88**: 23–33.
[PubMed Abstract](#) | [Publisher Full Text](#)
22. Leiby JS, McCormick K, Sherrill-Mix S, *et al.*: **Lack of detection of a human placenta microbiome in samples from preterm and term deliveries.** *Microbiome.* 2018; **6**(1): 196.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
23. Reh binder EM, Ledrup Carlsen KC, Staff AC, *et al.*: **Is amniotic fluid of women with uncomplicated term pregnancies free of bacteria?** *Am J Obstet Gynecol.* 2018; **219**(3): 289.e1–289.e12.
[PubMed Abstract](#) | [Publisher Full Text](#)
24. Theis KR, Romero R, Winters AD, *et al.*: **Does the human placenta delivered at term have a microbiota? Results of cultivation, quantitative real-time PCR, 16S rRNA gene sequencing, and metagenomics.** *Am J Obstet Gynecol.* 2019; **220**(3): 267.e1–267.e39.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
25. Ryan PM, Stanton C, Ross RP, *et al.*: **Paediatrician's perspective of infant gut microbiome research: current status and challenges.** *Arch Dis Child.* 2019; **104**(7): 701–5.
[PubMed Abstract](#) | [Publisher Full Text](#)
26. Zhuang L, Chen H, Zhang S, *et al.*: **Intestinal Microbiota in Early Life and Its Implications on Childhood Health.** *Genomics Proteomics Bioinformatics.* 2019; **17**(1): 13–25.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
27. **F** Codagnone MG, Spichak S, O'Mahony SM, *et al.*: **Programming Bugs: Microbiota and the Developmental Origins of Brain Health and Disease.** *Biol Psychiatry.* 2019; **85**(2): 150–63.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
28. **F** Arumugam M, Raes J, Pelletier E, *et al.*: **Enterotypes of the human gut microbiome.** *Nature.* 2011; **473**(7346): 174–80.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
29. Claesson MJ, Jeffery IB, Conde S, *et al.*: **Gut microbiota composition correlates with diet and health in the elderly.** *Nature.* 2012; **488**(7410): 178–84.
[PubMed Abstract](#) | [Publisher Full Text](#)
30. **F** Yatsunenkov T, Rey FE, Manary MJ, *et al.*: **Human gut microbiome viewed across age and geography.** *Nature.* 2012; **486**(7402): 222–7.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
31. Martínez I, Stegen JC, Maldonado-Gómez MX, *et al.*: **The gut microbiota of rural papua new guineans: composition, diversity patterns, and ecological processes.** *Cell Rep.* 2015; **11**(4): 527–38.
[PubMed Abstract](#) | [Publisher Full Text](#)
32. Zhernakova A, Kurilshikov A, Bonder MJ, *et al.*: **Population-based metagenomics analysis reveals markers for gut microbiome composition and diversity.** *Science.* 2016; **352**(6285): 565–9.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
33. Falony G, Joossens M, Vieira-Silva S, *et al.*: **Population-level analysis of gut microbiome variation.** *Science.* 2016; **352**(6285): 560–4.
[PubMed Abstract](#) | [Publisher Full Text](#)
34. Valdes AM, Walter J, Segal E, *et al.*: **Role of the gut microbiota in nutrition and health.** *BMJ.* 2018; **361**: k2179.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
35. **F** Makki K, Deehan EC, Walter J, *et al.*: **The Impact of Dietary Fiber on Gut Microbiota in Host Health and Disease.** *Cell Host Microbe.* 2018; **23**(6): 705–15.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
36. Johnson AJ, Vangay P, Al-Ghalith GA, *et al.*: **Daily Sampling Reveals Personalized Diet-Microbiome Associations in Humans.** *Cell Host Microbe.* 2019; **25**(6): 789–802.e5.
[PubMed Abstract](#) | [Publisher Full Text](#)
37. Wang J, Jia H: **Metagenome-wide association studies: fine-mining the microbiome.** *Nat Rev Microbiol.* 2016; **14**(8): 508–22.
[PubMed Abstract](#) | [Publisher Full Text](#)
38. Garcia-Gutierrez E, Mayer MJ, Cotter PD, *et al.*: **Gut microbiota as a source of novel antimicrobials.** *Gut Microbes.* 2019; **10**(1): 1–21.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
39. Skelly M, Sato Y, Kearney S, *et al.*: **Mining the microbiota for microbial and metabolite-based immunotherapies.** *Nat Rev Immunol.* 2019; **19**(5): 305–23.
[PubMed Abstract](#) | [Publisher Full Text](#)
40. **F** Routy B, Le Chatelier E, Derosa L, *et al.*: **Gut microbiome influences efficacy of PD-1-based immunotherapy against epithelial tumors.** *Science.* 2018; **359**(6371): 91–7.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
41. Kashyap PC, Quigley EM: **Therapeutic implications of the gastrointestinal microbiome.** *Curr Opin Pharmacol.* 2018; **38**: 90–6.
[PubMed Abstract](#) | [Publisher Full Text](#)
42. **F** Wu GD, Chen J, Hoffmann C, *et al.*: **Linking long-term dietary patterns with gut microbial enterotypes.** *Science.* 2011; **334**(6052): 105–8.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
43. Tidjani Alou M, Lagier JC, Raoult D: **Diet influence on the gut microbiota and dysbiosis related to nutritional disorders.** *Human Microbiome Journal.* 2016; **1**: 3–11.
[Publisher Full Text](#)
44. Zhang N, Ju Z, Zuo T: **Time for food: The impact of diet on gut microbiota and human health.** *Nutrition.* 2018; **51–52**: 80–5.
[PubMed Abstract](#) | [Publisher Full Text](#)
45. **F** Saffouri GB, Shields-Cutler RR, Chen J, *et al.*: **Small intestinal microbial dysbiosis underlies symptoms associated with functional gastrointestinal disorders.** *Nat Commun.* 2019; **10**(1): 2012.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
46. Deehan EC, Walter J: **The Fiber Gap and the Disappearing Gut Microbiome: Implications for Human Nutrition.** *Trends Endocrinol Metab.* 2016; **27**(5): 239–42.
[PubMed Abstract](#) | [Publisher Full Text](#)
47. Sanz Y: **Effects of a gluten-free diet on gut microbiota and immune function in healthy adult humans.** *Gut Microbes.* 2010; **1**(3): 135–7.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
48. **F** Hansen LBS, Roager HM, Søndergaard NB, *et al.*: **A low-gluten diet induces changes in the intestinal microbiome of healthy Danish adults.** *Nat Commun.* 2018; **9**(1): 4630.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
49. **F** Sloan TJ, Jalanka J, Major GAD, *et al.*: **A low FODMAP diet is associated with changes in the microbiota and reduction in breath hydrogen but not colonic volume in healthy subjects.** *PLoS One.* 2018; **13**(7): e0201410.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
50. Clarke SF, Murphy EF, O'Sullivan O, *et al.*: **Exercise and associated dietary extremes impact on gut microbial diversity.** *Gut.* 2014; **63**(12): 1913–20.
[PubMed Abstract](#) | [Publisher Full Text](#)
51. Holscher HD: **Dietary fiber and prebiotics and the gastrointestinal microbiota.** *Gut Microbes.* 2017; **8**(2): 172–84.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
52. **F** McDonald D, Hyde E, Debelius JW, *et al.*: **American Gut: an Open Platform for Citizen Science Microbiome Research.** *mSystems.* 2018; **3**(3): pii: e00031-18.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
53. **F** Zhao L, Zhang F, Ding X, *et al.*: **Gut bacteria selectively promoted by dietary fibers alleviate type 2 diabetes.** *Science.* 2018; **359**(6380): 1151–6.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)

54. **F** Suez J, Korem T, Zeevi D, *et al.*: **Artificial sweeteners induce glucose intolerance by altering the gut microbiota.** *Nature.* 2014; 514(7521): 181–6. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
55. Tabatabaeizadeh SA, Tafazoli N, Ferns G, *et al.*: **Vitamin D, the gut microbiome and inflammatory bowel disease.** *J Res Med Sci.* 2018; 23: 75. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
56. Kali A, Bhuvaneshwar D, Charles PM, *et al.*: **Antibacterial synergy of curcumin with antibiotics against biofilm producing clinical bacterial isolates.** *J Basic Clin Pharm.* 2016; 7(3): 93–6. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
57. Packiavathy IA, Sasikumar P, Pandian SK, *et al.*: **Prevention of quorum-sensing-mediated biofilm development and virulence factors production in *Vibrio* spp. by curcumin.** *Appl Microbiol Biotechnol.* 2013; 97(23): 10177–87. [PubMed Abstract](#) | [Publisher Full Text](#)
58. Packiavathy IA, Priya S, Pandian SK, *et al.*: **Inhibition of biofilm development of uropathogens by curcumin - an anti-quorum sensing agent from *Curcuma longa*.** *Food Chem.* 2014; 148: 453–60. [PubMed Abstract](#) | [Publisher Full Text](#)
59. **F** Halmos EP, Christophersen CT, Bird AR, *et al.*: **Diets that differ in their FODMAP content alter the colonic luminal microenvironment.** *Gut.* 2015; 64(1): 93–100. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
60. **F** McIntosh K, Reed DE, Schneider T, *et al.*: **FODMAPs alter symptoms and the metabolome of patients with IBS: a randomised controlled trial.** *Gut.* 2017; 66(7): 1241–51. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
61. **F** Huaman JW, Meگو M, Manichanh C, *et al.*: **Effects of Prebiotics vs a Diet Low in FODMAPs in Patients With Functional Gut Disorders.** *Gastroenterology.* 2018; 155(4): 1004–7. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
62. Staudacher HM, Lomer MCE, Farquharson FM, *et al.*: **A Diet Low in FODMAPs Reduces Symptoms in Patients With Irritable Bowel Syndrome and A Probiotic Restores Bifidobacterium Species: A Randomized Controlled Trial.** *Gastroenterology.* 2017; 153(4): 936–47. [PubMed Abstract](#) | [Publisher Full Text](#)
63. Griffin LE, Djuric Z, Angioletta CJ, *et al.*: **A Mediterranean diet does not alter plasma trimethylamine N-oxide concentrations in healthy adults at risk for colon cancer.** *Food Funct.* 2019; 10(4): 2138–47. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
64. Moya A, Ferrer M: **Functional Redundancy-Induced Stability of Gut Microbiota Subjected to Disturbance.** *Trends Microbiol.* 2016; 24(5): 402–13. [PubMed Abstract](#) | [Publisher Full Text](#)
65. **F** Reichardt N, Vollmer M, Holtrop G, *et al.*: **Specific substrate-driven changes in human faecal microbiota composition contrast with functional redundancy in short-chain fatty acid production.** *ISME J.* 2018; 12(2): 610–22. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
66. Zheng X, Zhou K, Zhang Y, *et al.*: **Food withdrawal alters the gut microbiota and metabolome in mice.** *FASEB J.* 2018; 32(9): 4878–88. [PubMed Abstract](#) | [Publisher Full Text](#)
67. Mack I, Penders J, Cook J, *et al.*: **Is the Impact of Starvation on the Gut Microbiota Specific or Unspecific to Anorexia Nervosa? A Narrative Review Based on a Systematic Literature Search.** *Curr Neuropsychopharmacol.* 2018; 16(8): 1131–49. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
68. Seitz J, Trinh S, Herpertz-Dahlmann B: **The Microbiome and Eating Disorders.** *Psychiatr Clin North Am.* 2019; 42(1): 93–103. [PubMed Abstract](#) | [Publisher Full Text](#)
69. Herpertz-Dahlmann B, Seitz J, Baines J: **Food matters: how the microbiome and gut-brain interaction might impact the development and course of anorexia nervosa.** *Eur Child Adolesc Psychiatry.* 2017; 26(9): 1031–41. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
70. Li G, Xie C, Lu S, *et al.*: **Intermittent Fasting Promotes White Adipose Browning and Decreases Obesity by Shaping the Gut Microbiota.** *Cell Metab.* 2017; 26(4): 672–685.e4. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
71. Rangan P, Choi I, Wei M, *et al.*: **Fasting-Mimicking Diet Modulates Microbiota and Promotes Intestinal Regeneration to Reduce Inflammatory Bowel Disease Pathology.** *Cell Rep.* 2019; 26(10): 2704–2719.e6. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
72. Cignarella F, Cantoni C, Ghezzi L, *et al.*: **Intermittent Fasting Confers Protection in CNS Autoimmunity by Altering the Gut Microbiota.** *Cell Metab.* 2018; 27(6): 1222–1235.e6. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
73. Zhou ZL, Jia XB, Sun MF, *et al.*: **Neuroprotection of Fasting Mimicking Diet on MPTP-Induced Parkinson's Disease Mice via Gut Microbiota and Metabolites.** *Neurotherapeutics.* 2019; 16(3): 741–60. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
74. **F** Guo Y, Huang ZP, Liu CQ, *et al.*: **Modulation of the gut microbiome: A systematic review of the effect of bariatric surgery.** *Eur J Endocrinol.* 2018; 178(1): 43–56. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
75. Schroeder BO, Birchenough GMH, Ståhlman M, *et al.*: **Bifidobacteria or Fiber Protects against Diet-Induced Microbiota-Mediated Colonic Mucus Deterioration.** *Cell Host Microbe.* 2018; 23(1): 27–40.e7. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
76. Birchenough G, Schroeder BO, Bäckhed F, *et al.*: **Dietary destabilisation of the balance between the microbiota and the colonic mucus barrier.** *Gut Microbes.* 2019; 10(2): 246–50. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
77. **F** Desai MS, Seekatz AM, Koropatkin NM, *et al.*: **A Dietary Fiber-Deprived Gut Microbiota Degrades the Colonic Mucus Barrier and Enhances Pathogen Susceptibility.** *Cell.* 2016; 167(5): 1339–1353.e21. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
78. Ottman N, Geerlings SY, Aalvink S, *et al.*: **Action and function of *Akkermansia muciniphila* in microbiome ecology, health and disease.** *Best Pract Res Clin Gastroenterol.* 2017; 31(6): 637–42. [PubMed Abstract](#) | [Publisher Full Text](#)
79. O'Sullivan O, Cronin O, Clarke SF, *et al.*: **Exercise and the microbiota.** *Gut Microbes.* 2015; 6(2): 131–6. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
80. **F** Allen JM, Mailing LJ, Niemi GM, *et al.*: **Exercise Alters Gut Microbiota Composition and Function in Lean and Obese Humans.** *Med Sci Sports Exerc.* 2018; 50(4): 747–57. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
81. Mailing LJ, Allen JM, Buford TW, *et al.*: **Exercise and the Gut Microbiome: A Review of the Evidence, Potential Mechanisms, and Implications for Human Health.** *Exerc Sport Sci Rev.* 2019; 47(2): 75–85. [PubMed Abstract](#) | [Publisher Full Text](#)
82. **F** Scheiman J, Lubner JM, Chavkin TA, *et al.*: **Meta-omics analysis of elite athletes identifies a performance-enhancing microbe that functions via lactate metabolism.** *Nat Med.* 2019; 25(7): 1104–9. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
83. Lee SH, Yun Y, Kim SJ, *et al.*: **Association between Cigarette Smoking Status and Composition of Gut Microbiota: Population-Based Cross-Sectional Study.** *J Clin Med.* 2018; 7(9): pii: E282. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
84. Shanahan ER, Shah A, Koloski N, *et al.*: **Influence of cigarette smoking on the human duodenal mucosa-associated microbiota.** *Microbiome.* 2018; 6(1): 150. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
85. Capurso G, Lahner E: **The interaction between smoking, alcohol and the gut microbiome.** *Best Pract Res Clin Gastroenterol.* 2017; 31(5): 579–88. [PubMed Abstract](#) | [Publisher Full Text](#)
86. Hernández-Quiroz F, Nirmalkar K, Villalobos-Flores LE, *et al.*: **Influence of moderate beer consumption on human gut microbiota and its impact on fasting glucose and β -cell function.** *Alcohol.* 2019; pii: S0741-8329(19)30068-0. [PubMed Abstract](#) | [Publisher Full Text](#)
87. Martínez S, Campa A, Narasimhan G, *et al.*: **Pilot Study on the Effect of Cocaine Use on the Intestinal Microbiome and Metabolome and Inflammation in HIV-Infected Adults in the Miami Adult Studies in HIV (MASH) Cohort (P13-027-19).** *Curr Dev Nutr.* 2019; 3(Supplement_1). [Publisher Full Text](#)
88. Wu J, Peters BA, Dominianni C, *et al.*: **Cigarette smoking and the oral microbiome in a large study of American adults.** *ISME J.* 2016; 10(10): 2435–46. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
89. Cammarota G, Ianiro G, Gasbarrini A: **Faecal microbiota transplantation in clinical practice.** *Gut.* 2018; 67(1): 196–197. [PubMed Abstract](#) | [Publisher Full Text](#)
90. **F** Ianiro G, Maida M, Burisich J, *et al.*: **Efficacy of different faecal microbiota transplantation protocols for *Clostridium difficile* infection: A systematic review and meta-analysis.** *United European Gastroenterol J.* 2018; 6(8): 1232–44. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
91. **F** DeFillipp Z, Bloom PP, Torres Soto M, *et al.*: **Drug-Resistant *E. coli* Bacteremia Transmitted by Fecal Microbiota Transplant.** *N Engl J Med.* 2019; 381(21): 2043–50. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
92. Costello SP, Hughes PA, Waters O, *et al.*: **Effect of Fecal Microbiota Transplantation on 8-Week Remission in Patients With Ulcerative Colitis: A Randomized Clinical Trial.** *JAMA.* 2019; 321(2): 156–164. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
93. Lam WC, Zhao C, Ma WJ, *et al.*: **The Clinical and Steroid-Free Remission of Fecal Microbiota Transplantation to Patients with Ulcerative Colitis: A Meta-Analysis.** *Gastroenterol Res Pract.* 2019; 2019: 1287493. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
94. **F** Imdad A, Nicholson MR, Tanner-Smith EE, *et al.*: **Fecal transplantation for treatment of inflammatory bowel disease.** *Cochrane Database Syst Rev.* 2018; 11: S23. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
95. **F** Ianiro G, Eusebi LH, Black CJ, *et al.*: **Systematic review with meta-analysis: efficacy of faecal microbiota transplantation for the treatment of irritable bowel syndrome.** *Aliment Pharmacol Ther.* 2019; 50(3): 240–8. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
96. Xu D, Chen VL, Steiner CA, *et al.*: **Efficacy of Fecal Microbiota Transplantation**

- in Irritable Bowel Syndrome: A Systematic Review and Meta-Analysis. *Am J Gastroenterol*. 2019; 114(7): 1043–50.
PubMed Abstract | Publisher Full Text
97. Gutin L, Piceno Y, Fadrosch D, *et al.*: Fecal microbiota transplant for Crohn disease: A study evaluating safety, efficacy, and microbiome profile. *United European Gastroenterol J*. 2019; 7(6): 807–14.
PubMed Abstract | Publisher Full Text | Free Full Text
98. Aroniadis OC, Brandt LJ, Oneto C, *et al.*: Faecal microbiota transplantation for diarrhoea-predominant irritable bowel syndrome: a double-blind, randomised, placebo-controlled trial. *Lancet Gastroenterol Hepatol*. 2019; 4(9): 675–85.
PubMed Abstract | Publisher Full Text
99. Halkjær SI, Christensen AH, Lo BZS, *et al.*: Faecal microbiota transplantation alters gut microbiota in patients with irritable bowel syndrome: results from a randomised, double-blind placebo-controlled study. *Gut*. 2018; 67(12): 2107–15.
PubMed Abstract | Publisher Full Text | F1000 Recommendation
100. Cammarota G, Ianiro G: FMT for ulcerative colitis: closer to the turning point. *Nat Rev Gastroenterol Hepatol*. 2019; 16(5): 266–8.
PubMed Abstract | Publisher Full Text
101. Bajaj JS, Kassam Z, Fagan A, *et al.*: Fecal microbiota transplant from a rational stool donor improves hepatic encephalopathy: A randomized clinical trial. *Hepatology*. 2017; 66(6): 1727–38.
PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
102. Quigley EMM, Abu-Shanab A, Murphy EF, *et al.*: The Metabolic Role of the Microbiome: Implications for NAFLD and the Metabolic Syndrome. *Semin Liver Dis*. 2016; 36(4): 312–6.
PubMed Abstract | Publisher Full Text
103. Sharpton SR, Maraj B, Harding-Theobald E, *et al.*: Gut microbiome-targeted therapies in nonalcoholic fatty liver disease: a systematic review, meta-analysis, and meta-regression. *Am J Clin Nutr*. 2019; 110(1): 139–49.
PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
104. Nobili V, Mosca A, Alterio T, *et al.*: Fighting Fatty Liver Diseases with Nutritional Interventions, Probiotics, Symbiotics, and Fecal Microbiota Transplantation (FMT). *Adv Exp Med Biol*. 2019; 1125: 85–100.
PubMed Abstract | Publisher Full Text
105. Mukhopadhyay I, Segal JP, Carding SR, *et al.*: The gut virome: the 'missing link' between gut bacteria and host immunity? *Therap Adv Gastroenterol*. 2019; 12: 175628481983662.
PubMed Abstract | Publisher Full Text | Free Full Text
106. Broecker F, Klumpp J, Schuppler M, *et al.*: Long-term changes of bacterial and viral compositions in the intestine of a recovered *Clostridium difficile* patient after fecal microbiota transplantation. *Cold Spring Harb Mol Case Stud*. 2016; 2(1): a000448.
PubMed Abstract | Publisher Full Text | Free Full Text
107. Broecker F, Russo G, Klumpp J, *et al.*: Stable core virome despite variable microbiome after fecal transfer. *Gut Microbes*. 2017; 8(3): 214–20.
PubMed Abstract | Publisher Full Text | Free Full Text
108. <https://www.fda.gov/vaccines-blood-biologics/safety-availability-biologics/important-safety-alert-regarding-use-fecal-microbiota-transplantation-and-risk-serious-adverse>. (accessed August 2nd 2019).
109. Ossorio PN, Zhou Y: Regulating stool for microbiota transplantation. *Gut Microbes*. 2019; 10(2): 105–8.
PubMed Abstract | Publisher Full Text | Free Full Text
110. Khoruts A, Sadowsky MJ: Understanding the mechanisms of faecal microbiota transplantation. *Nat Rev Gastroenterol Hepatol*. 2016; 13(9): 508–16.
PubMed Abstract | Publisher Full Text | Free Full Text
111. McDonald JAK, Mullish BH, Pechlivanis A, *et al.*: Inhibiting Growth of *Clostridioides difficile* by Restoring Valerate, Produced by the Intestinal Microbiota. *Gastroenterology*. 2018; 155(5): 1495–1507.e15.
PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
112. Mullish BH, McDonald JAK, Pechlivanis A, *et al.*: Microbial bile salt hydrolases mediate the efficacy of faecal microbiota transplant in the treatment of recurrent *Clostridioides difficile* infection. *Gut*. 2019; 68(10): 1791–800.
PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
113. Lewis JD, Chen EZ, Baldassano RN, *et al.*: Inflammation, Antibiotics, and Diet as Environmental Stressors of the Gut Microbiome in Pediatric Crohn's Disease. *Cell Host Microbe*. 2015; 18(4): 489–500.
PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
114. Savoldi A, Carrara E, Graham DY, *et al.*: Prevalence of Antibiotic Resistance in *Helicobacter pylori*: A Systematic Review and Meta-analysis in World Health Organization Regions. *Gastroenterology*. 2018; 155(5): 1372–1382.e17.
PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
115. Hu Y, Yang X, Qin J, *et al.*: Metagenome-wide analysis of antibiotic resistance genes in a large cohort of human gut microbiota. *Nat Commun*. 2013; 4: 2151.
PubMed Abstract | Publisher Full Text
116. Pérez-Cobas AE, Gosalbes MJ, Friedrichs A, *et al.*: Gut microbiota disturbance during antibiotic therapy: a multi-omic approach. *Gut*. 2013; 62(11): 1591–601.
PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
117. Fujisaka S, Ussar S, Clish C, *et al.*: Antibiotic effects on gut microbiota and metabolism are host dependent. *J Clin Invest*. 2016; 126(12): 4430–43.
PubMed Abstract | Publisher Full Text | Free Full Text
118. Zackular JP, Baxter NT, Iverson KD, *et al.*: The gut microbiome modulates colon tumorigenesis. *mBio*. 2013; 4(6): 74.
PubMed Abstract | Publisher Full Text | Free Full Text
119. DuPont HL: Therapeutic Effects and Mechanisms of Action of Rifaximin in Gastrointestinal Diseases. *Mayo Clin Proc*. 2015; 90(8): 1116–24.
PubMed Abstract | Publisher Full Text
120. Pérez-Cobas AE, Artacho A, Knecht H, *et al.*: Differential effects of antibiotic therapy on the structure and function of human gut microbiota. *PLoS One*. 2013; 8(11): e80201.
PubMed Abstract | Publisher Full Text | Free Full Text
121. Hernández E, Bargiela R, Diez MS, *et al.*: Functional consequences of microbial shifts in the human gastrointestinal tract linked to antibiotic treatment and obesity. *Gut Microbes*. 2013; 4(4): 306–15.
PubMed Abstract | Publisher Full Text | Free Full Text
122. Modi SR, Collins JJ, Relman DA: Antibiotics and the gut microbiota. *J Clin Invest*. 2014; 124(10): 4212–8.
PubMed Abstract | Publisher Full Text | Free Full Text
123. Blaser MJ: Antibiotic use and its consequences for the normal microbiome. *Science*. 2016; 352(6285): 544–5.
PubMed Abstract | Publisher Full Text | Free Full Text
124. Cho I, Yamanishi S, Cox L, *et al.*: Antibiotics in early life alter the murine colonic microbiome and adiposity. *Nature*. 2012; 488(7413): 621–6.
PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
125. Cox LM, Yamanishi S, Sohn J, *et al.*: Altering the intestinal microbiota during a critical developmental window has lasting metabolic consequences. *Cell*. 2014; 158(4): 705–21.
PubMed Abstract | Publisher Full Text | Free Full Text
126. Livanos AE, Greiner TU, Vangay P, *et al.*: Antibiotic-mediated gut microbiome perturbation accelerates development of type 1 diabetes in mice. *Nat Microbiol*. 2016; 1(11): 16140.
PubMed Abstract | Publisher Full Text | Free Full Text
127. Ruiz VE, Battaglia T, Kurtz ZD, *et al.*: A single early-in-life macrolide course has lasting effects on murine microbial network topology and immunity. *Nat Commun*. 2017; 8(1): 518.
PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
128. Nobel YR, Cox LM, Kirigin FF, *et al.*: Metabolic and metagenomic outcomes from early-life pulsed antibiotic treatment. *Nat Commun*. 2015; 6: 7486.
PubMed Abstract | Publisher Full Text | Free Full Text
129. Blaser MJ: The Past and Future Biology of the Human Microbiome in an Age of Extinctions. *Cell*. 2018; 172(6): 1173–7.
PubMed Abstract | Publisher Full Text
130. Quigley EMM: Prebiotics and Probiotics in Digestive Health. *Clin Gastroenterol Hepatol*. 2019; 17(2): 333–44.
PubMed Abstract | Publisher Full Text
131. Brüssow H: Probiotics and prebiotics in clinical tests: an update [version 1; peer review: 2 approved]. *F1000Res*. 2019; 8: pii: F1000 Faculty Rev-1157.
PubMed Abstract | Publisher Full Text | Free Full Text
132. Gibson GR, Hutkins R, Sanders ME, *et al.*: Expert consensus document: The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of prebiotics. *Nat Rev Gastroenterol Hepatol*. 2017; 14(8): 491–502.
PubMed Abstract | Publisher Full Text
133. http://www.who.int/foodsafety/publications/fs_management/en/probiotics.pdf. accessed August 3rd 2019.
134. Flint HJ, Duncan SH, Scott KP, *et al.*: Links between diet, gut microbiota composition and gut metabolism. *Proc Nutr Soc*. 2015; 74(1): 13–22.
PubMed Abstract | Publisher Full Text
135. Davani-Davari D, Negahdaripour M, Karimzadeh I, *et al.*: Prebiotics: Definition, Types, Sources, Mechanisms, and Clinical Applications. *Foods*. 2019; 8(3): pii: E92.
PubMed Abstract | Publisher Full Text | Free Full Text
136. Bering SB: Human Milk Oligosaccharides to Prevent Gut Dysfunction and Necrotizing Enterocolitis in Preterm Neonates. *Nutrients*. 2018; 10(10): pii: E1461.
PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
137. Le Doare K, Holder B, Bassett A, *et al.*: Mother's Milk: A Purposeful Contribution to the Development of the Infant Microbiota and Immunity. *Front Immunol*. 2018; 9: 361.
PubMed Abstract | Publisher Full Text | Free Full Text
138. Kirmiz N, Robinson RC, Shah IM, *et al.*: Milk Glycans and Their Interaction with the Infant-Gut Microbiota. *Annu Rev Food Sci Technol*. 2018; 9: 429–50.
PubMed Abstract | Publisher Full Text | Free Full Text
139. Louca S, Polz MF, Mazel F, *et al.*: Function and functional redundancy in microbial systems. *Nat Ecol Evol*. 2018; 2(6): 936–43.
PubMed Abstract | Publisher Full Text
140. Bindels LB, Delzenne NM, Cani PD, *et al.*: Towards a more comprehensive concept for prebiotics. *Nat Rev Gastroenterol Hepatol*. 2015; 12(5): 303–10.
PubMed Abstract | Publisher Full Text
141. Nicolucci AC, Hume MP, Martínez I, *et al.*: Prebiotics Reduce Body Fat and Alter Intestinal Microbiota in Children Who Are Overweight or With Obesity.

- Gastroenterology*. 2017; **153**(3): 711–22.
PubMed Abstract | Publisher Full Text
142. F Shokryazdan P, Faseleh Jahromi M, Navidshad B, et al.: **Effects of prebiotics on immune system and cytokine expression**. *Med Microbiol Immunol*. 2017; **206**(1): 1–9.
PubMed Abstract | Publisher Full Text | F1000 Recommendation
 143. Miller LE, Zimmermann AK, Ouwehand AC: **Contemporary meta-analysis of short-term probiotic consumption on gastrointestinal transit**. *World J Gastroenterol*. 2016; **22**(21): 5122–31.
PubMed Abstract | Publisher Full Text | Free Full Text
 144. Koopman N, Molinaro A, Nieuwdorp M, et al.: **Review article: can bugs be drugs? The potential of probiotics and prebiotics as treatment for non-alcoholic fatty liver disease**. *Aliment Pharmacol Ther*. 2019; **50**(6): 628–39.
PubMed Abstract | Publisher Full Text
 145. Kim N, Yun M, Oh YJ, et al.: **Mind-altering with the gut: Modulation of the gut-brain axis with probiotics**. *J Microbiol*. 2018; **56**(3): 172–82.
PubMed Abstract | Publisher Full Text
 146. Smolinska S, Groeger D, O'Mahony L: **Biology of the Microbiome 1: Interactions with the Host Immune Response**. *Gastroenterol Clin North Am*. 2017; **46**(1): 19–35.
PubMed Abstract | Publisher Full Text
 147. Maldonado-Gómez MX, Martínez I, Bottacini F, et al.: **Stable Engraftment of *Bifidobacterium longum* AH1206 in the Human Gut Depends on Individualized Features of the Resident Microbiome**. *Cell Host Microbe*. 2016; **20**(4): 515–26.
PubMed Abstract | Publisher Full Text
 148. F Zmora N, Zilberman-Schapira G, Suez J, et al.: **Personalized Gut Mucosal Colonization Resistance to Empiric Probiotics Is Associated with Unique Host and Microbiome Features**. *Cell*. 2018; **174**(6): 1388–1405.e21.
PubMed Abstract | Publisher Full Text | F1000 Recommendation
 149. <http://www.worldgastroenterology.org/UserFiles/file/guidelines/probiotics-and-prebiotics-english-2017.pdf>. accessed August 3rd 2019.
 150. Florez ID, Veroniki AA, Al Khalifah R, et al.: **Comparative effectiveness and safety of interventions for acute diarrhea and gastroenteritis in children: A systematic review and network meta-analysis**. *PLoS One*. 2018; **13**(12): e0207701.
PubMed Abstract | Publisher Full Text | Free Full Text
 151. Guo Q, Goldenberg JZ, Humphrey C, et al.: **Probiotics for the prevention of pediatric antibiotic-associated diarrhea**. *Cochrane Database Syst Rev*. 2019; **4**: CD004827.
PubMed Abstract | Publisher Full Text | Free Full Text
 152. F Goldenberg JZ, Yap C, Lytvyn L, et al.: **Probiotics for the prevention of *Clostridium difficile*-associated diarrhea in adults and children**. *Cochrane Database Syst Rev*. 2017; **12**: CD006095.
PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
 153. F Sun J, Marwah G, Westgarth M, et al.: **Effects of Probiotics on Necrotizing Enterocolitis, Sepsis, Intraventricular Hemorrhage, Mortality, Length of Hospital Stay, and Weight Gain in Very Preterm Infants: A Meta-Analysis**. *Adv Nutr*. 2017; **8**(5): 749–63.
PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
 154. F Ford AC, Harris LA, Lacy BE, et al.: **Systematic review with meta-analysis: the efficacy of probiotics, prebiotics, synbiotics and antibiotics in irritable bowel syndrome**. *Aliment Pharmacol Ther*. 2018; **48**(10): 1044–60.
PubMed Abstract | Publisher Full Text | F1000 Recommendation
 155. F Derwa Y, Gracie DJ, Hamlin PJ, et al.: **Systematic review with meta-analysis: the efficacy of probiotics in inflammatory bowel disease**. *Aliment Pharmacol Ther*. 2017; **46**(4): 389–400.
PubMed Abstract | Publisher Full Text | F1000 Recommendation
 156. Jia K, Tong X, Wang R, et al.: **The clinical effects of probiotics for inflammatory bowel disease: A meta-analysis**. *Medicine (Baltimore)*. 2018; **97**(51): e13792.
PubMed Abstract | Publisher Full Text | Free Full Text
 157. F Astó E, Méndez I, Audivert S, et al.: **The Efficacy of Probiotics, Prebiotic Inulin-Type Fructans, and Synbiotics in Human Ulcerative Colitis: A Systematic Review and Meta-Analysis**. *Nutrients*. 2019; **11**(2): pii: E293.
PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
 158. F Panigrahi P, Parida S, Nanda NC, et al.: **A randomized synbiotic trial to prevent sepsis among infants in rural India**. *Nature*. 2017; **548**(7668): 407–12.
PubMed Abstract | Publisher Full Text | F1000 Recommendation
 159. F Krumbek JA, Rasmussen HE, Hutkins RW, et al.: **Probiotic *Bifidobacterium* strains and galactooligosaccharides improve intestinal barrier function in obese adults but show no synergism when used together as synbiotics**. *Microbiome*. 2018; **6**(1): 121.
PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
 160. Shanahan F: **Therapeutic implications of manipulating and mining the microbiota**. *J Physiol*. 2009; **587**(Pt 17): 4175–9.
PubMed Abstract | Publisher Full Text | Free Full Text
 161. Shanahan F, Collins SM: **Pharmabiotic manipulation of the microbiota in gastrointestinal disorders, from rationale to reality**. *Gastroenterol Clin North Am*. 2010; **39**(3): 721–6.
PubMed Abstract | Publisher Full Text
 162. Hegarty JW, Guinane CM, Ross RP, et al.: **Bacteriocin production: a relatively unharmed probiotic trait? [version 1; peer review: 2 approved]**. *F1000Res*. 2016; **5**: 2587.
PubMed Abstract | Publisher Full Text | Free Full Text
 163. Vo P, Lee HM, Na D: **Synthetic Bacteria for Therapeutics**. *J Microbiol Biotechnol*. 2019; **29**(6): 845–55.
PubMed Abstract | Publisher Full Text
 164. Ferreira AK, Mambelli LI, Pillai SY: **Intervening in disease through genetically-modified bacteria**. *Best Pract Res Clin Gastroenterol*. 2017; **31**(6): 693–7.
PubMed Abstract | Publisher Full Text
 165. Sagona AP, Grigonyte AM, MacDonald PR, et al.: **Genetically modified bacteriophages**. *Integr Biol (Camb)*. 2016; **8**(4): 465–74.
PubMed Abstract | Publisher Full Text
 166. Rehman S, Ali Z, Khan M, et al.: **The dawn of phage therapy**. *Rev Med Virol*. 2019; **29**(4): e2041.
PubMed Abstract | Publisher Full Text
 167. Daille B, van Oudenhove L, Vervliet B, et al.: **The role of short-chain fatty acids in microbiota-gut-brain communication**. *Nat Rev Gastroenterol Hepatol*. 2019; **16**(8): 461–78.
PubMed Abstract | Publisher Full Text
 168. Thompson JA, Oliveira RA, Djukovic A, et al.: **Manipulation of the quorum sensing signal AI-2 affects the antibiotic-treated gut microbiota**. *Cell Rep*. 2015; **10**(11): 1861–71.
PubMed Abstract | Publisher Full Text
 169. F Duan F, March JC: **Engineered bacterial communication prevents *Vibrio cholerae* virulence in an infant mouse model**. *Proc Natl Acad Sci U S A*. 2010; **107**(25): 11260–4.
PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
 170. F Saeidi N, Wong CK, Lo TM, et al.: **Engineering microbes to sense and eradicate *Pseudomonas aeruginosa*, a human pathogen**. *Mol Syst Biol*. 2011; **7**: 521.
PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
 171. Hwang IY, Tan MH, Koh E, et al.: **Reprogramming microbes to be pathogen-seeking killers**. *ACS Synth Biol*. 2014; **3**(4): 228–37.
PubMed Abstract | Publisher Full Text
 172. Barrangou R, Doudna JA: **Applications of CRISPR technologies in research and beyond**. *Nat Biotechnol*. 2016; **34**(9): 933–41.
PubMed Abstract | Publisher Full Text
 173. Knott GJ, Doudna JA: **CRISPR-Cas guides the future of genetic engineering**. *Science*. 2018; **361**(6405): 866–9.
PubMed Abstract | Publisher Full Text | Free Full Text
 174. F Barrangou R, Notebaart RA: **CRISPR-Directed Microbiome Manipulation across the Food Supply Chain**. *Trends Microbiol*. 2019; **27**(6): 489–96.
PubMed Abstract | Publisher Full Text | F1000 Recommendation
 175. Malfertheiner P, Kandulski A, Venerito M: **Proton-pump inhibitors: understanding the complications and risks**. *Nat Rev Gastroenterol Hepatol*. 2017; **14**(12): 697–710.
PubMed Abstract | Publisher Full Text
 176. F Singh A, Cresci GA, Kirby DF: **Proton Pump Inhibitors: Risks and Rewards and Emerging Consequences to the Gut Microbiome**. *Nutr Clin Pract*. 2018; **33**(5): 614–24.
PubMed Abstract | Publisher Full Text | F1000 Recommendation
 177. F Le Bastard Q, Al-Ghalith GA, Grégoire M, et al.: **Systematic review: human gut dysbiosis induced by non-antibiotic prescription medications**. *Aliment Pharmacol Ther*. 2018; **47**(3): 332–45.
PubMed Abstract | Publisher Full Text | F1000 Recommendation
 178. F Freedberg DE, Toussaint NC, Chen SP, et al.: **Proton Pump Inhibitors Alter Specific Taxa in the Human Gastrointestinal Microbiome: A Crossover Trial**. *Gastroenterology*. 2015; **149**(4): 883–5.e9.
PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation
 179. Imhann F, Bonder MJ, Vich Vila A, et al.: **Proton pump inhibitors affect the gut microbiome**. *Gut*. 2016; **65**(5): 740–8.
PubMed Abstract | Publisher Full Text | Free Full Text
 180. Roager HM, Hansen LB, Bahl MI, et al.: **Colonic transit time is related to bacterial metabolism and mucosal turnover in the gut**. *Nat Microbiol*. 2016; **1**(9): 16093.
PubMed Abstract | Publisher Full Text
 181. F Guthrie L, Kelly L: **Bringing microbiome-drug interaction research into the clinic**. *EBioMedicine*. 2019; **44**: 708–15.
PubMed Abstract | Publisher Full Text | Free Full Text | F1000 Recommendation

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