

# Metal Nanoparticles: Advanced and Promising Technology in Diabetic Wound Therapy

Qinzhou Zheng<sup>1,\*</sup>, Cuimin Chen<sup>2,\*</sup>, Yong Liu<sup>3</sup>, Jie Gao<sup>2,\*</sup>, Luxin Li<sup>1</sup>, Chuan Yin<sup>4</sup>, Xiaohuan Yuan<sup>1</sup>

<sup>1</sup>College of Life Science, Mudanjiang Medical University, Mudanjiang, People's Republic of China; <sup>2</sup>Changhai Clinical Research Unit, Shanghai Changhai Hospital, Naval Medical University, Shanghai, 200433, People's Republic of China; <sup>3</sup>Center for Comparative Medicine, Mudanjiang Medical University, Mudanjiang, People's Republic of China; <sup>4</sup>Department of Gastroenterology, Shanghai Changzheng Hospital, Naval Medical University, Shanghai, 200003, People's Republic of China

\*These authors contributed equally to this work

Correspondence: Chuan Yin, Department of Gastroenterology, Shanghai Changzheng Hospital, Naval Medical University, Shanghai, 200003, People's Republic of China, Email [ilse1225@163.com](mailto:ilse1225@163.com); Xiaohuan Yuan, College of Life Science, Mudanjiang Medical University, Mudanjiang, 157001, Heilongjiang, People's Republic of China, Tel/Fax +86-453-6984647, Email [yuanxiaohuan@mdjmu.edu.cn](mailto:yuanxiaohuan@mdjmu.edu.cn)

**Abstract:** Diabetic wounds pose a significant challenge to public health, primarily due to insufficient blood vessel supply, bacterial infection, excessive oxidative stress, and impaired antioxidant defenses. The aforementioned condition not only places a significant physical burden on patients' prognosis, but also amplifies the economic strain on the medical system in treating diabetic wounds. Currently, the effectiveness of available treatments for diabetic wounds is limited. However, there is hope in the potential of metal nanoparticles (MNPs) to address these issues. MNPs exhibit excellent anti-inflammatory, antioxidant, antibacterial and pro-angiogenic properties, making them a promising solution for diabetic wounds. In addition, MNPs stimulate the expression of proteins that promote wound healing and serve as drug delivery systems for small-molecule drugs. By combining MNPs with other biomaterials such as hydrogels and chitosan, novel dressings can be developed and revolutionize the treatment of diabetic wounds. The present article provides a comprehensive overview of the research progress on the utilization of MNPs for treating diabetic wounds. Building upon this foundation, we summarize the underlying mechanisms involved in diabetic wound healing and discuss the potential application of MNPs as biomaterials for drug delivery. Furthermore, we provide an extensive analysis and discussion on the clinical implementation of dressings, while also highlighting future prospects for utilizing MNPs in diabetic wound management. In conclusion, MNPs represent a promising strategy for the treatment of diabetic wound healing. Future directions include combining other biological nanomaterials to synthesize new biological dressings or utilizing the other physicochemical properties of MNPs to promote wound healing. Synthetic biomaterials that contain MNPs not only play a role in all stages of diabetic wound healing, but also provide a stable physiological environment for the wound-healing process.

**Keywords:** metal nanoparticles, wound healing, diabetic wound, drug delivery, antibacterial activity

## Introduction

Diabetes mellitus (DM) is a prevalent endocrine disorder characterized primarily by elevated blood glucose levels resulting from either insulin resistance or diminished insulin secretion.<sup>1</sup> The occurrence of DM in the adult population is expected to grow by 69% in developing nations and 20% in affluent countries by 2030 in comparison to 2010.<sup>2</sup> The number of Americans with diabetes is expected to triple.<sup>3</sup>

Prolonged hyperglycemia can cause a variety of organ lesions in diabetic patients, including diabetic nephropathy, skin ulcers, retinopathy, and neuropathy.<sup>4</sup> In diabetes, the failure of wound healing is attributed to macrovascular and microvascular lesions, leading to complications that are a major cause of increased diabetes-related morbidity and mortality.<sup>5</sup> Moreover, the high glucose environment of the wound makes the wound susceptible to infection, which also leads to slow and worse wound recovery.

The current treatment methods mainly include blood glucose control, surgical debridement, skin transplantation, wound dressing change and other conventional treatment methods. However, conventional treatments only clean the wound and do not effectively promote the healing of diabetic wounds.

Therefore, advancing treatment of diabetic wounds requires the addition of innovative biomaterials and bioactive substances. These novel interventions are designed to accelerate wound healing and minimize unattractive scar formation by means of anti-inflammatory, antioxidant, and antimicrobial agents.<sup>6</sup> Among the various emerging technologies, MNPs have shown great potential in promoting diabetic wound healing.<sup>7</sup> MNPs effectively promote diabetic wound healing due to their wide range of antimicrobial activities and other biological properties. Moreover, the special physical and chemical properties of MNPs, including their acoustic, optical, and electrical properties, also play an important role in the wound healing measurement process. MNPs also possess properties that differ from nanoparticles, including a large specific surface area, uniform distribution, and the ability to modify surface morphology. These remarkable properties propel MNPs as an important component in the synthesis of biomaterials for diabetic wound healing.<sup>8</sup> In the realm of therapeutic drug delivery, MNPs offer multiple avenues for effective drug administration. They can be dispersed or encapsulated within metal shells, or covalently attached to their surfaces, enabling efficient drug delivery to targeted diseases. Moreover, MNPs also serve as effective multimodal contrast agents, facilitating disease diagnosis through various modalities such as fluorescence and magnetic resonance imaging. These versatile properties of MNPs render them invaluable in both therapeutic and diagnostic applications.<sup>9,10</sup> MNPs also facilitate intelligent drug release, such as the ability to be externally triggered by infrared radiation or magnetic fields, thereby achieving multi-stimulus-responsive drug release.<sup>11,12</sup> This property provides effective targeting to the complex healing process of diabetic wounds, allowing progress toward accelerated healing at different stages of wound healing.<sup>13</sup>

Recently, the unique physicochemical properties of MNPs have attracted more and more attention in the study of diabetic wound healing. There are numerous reviews mainly focused on a certain application (anti-inflammatory, antibacterial, etc.) of single MNPs (AgNPs, CuNPs) or summaries of the new preparation technology of certain MNPs (green synthesis), but a comprehensive and systematic review is lacking. We hope to comprehensively and systematically elaborate the role, mechanism and clinical application of MNPs in the treatment of diabetic wounds. To comprehensively elucidate the mechanism of MNPs in diabetic wound healing, we carefully selected MNPs that have been extensively studied and utilized, and systematically summarized the direct pro-healing effects of these MNPs on diabetic wounds as well as their indirect effects when combined with other biomaterials.<sup>14–17</sup> Based on these premises, we present a comprehensive review of the latest advancements in MNPs research and elucidate their potential role in enhancing diabetic wound healing through angiogenesis, drug delivery, and intelligent release mechanisms. Furthermore, we summarize and discuss the clinical applications of MNPs along with their associated adverse effects. Finally, propose avenues to deepen the potential of MNPs in the study of diabetic wounds in the future, and make reasonable assumptions.

## Wound Healing Mechanisms

### Structure and Function of the Skin

The skin is composed of the epidermis and dermis, which sit above subcutaneous tissue. The skin is the body's primary defense against external pathogens and other factors, making it an important organ. The epidermis, consisting of numerous keratinocytes, serves as a barrier to maintain water balance in the body. The dermis lies between the epidermis and subcutaneous tissue and is composed of connective tissue made of extracellular matrix (ECM) and fibroblasts. Collagen and elastin secretion maintains the mechanical properties and elasticity of the skin. Additionally, the dermis contains blood vessels, nerve endings, hair follicles, and other tissues. The subcutaneous tissue below the dermis is made up of loose connective tissue and fat providing a myxoid nature that permits fluid absorption.<sup>18</sup>

### Wound Healing Process

A skin wound is the destruction of or damage to the normal anatomical structure and function of the skin. The destruction of the integrity of the epithelial cells or the destruction of the full thickness of the skin may extend to deeper structures, such as blood vessels, muscles, etc.<sup>19</sup>

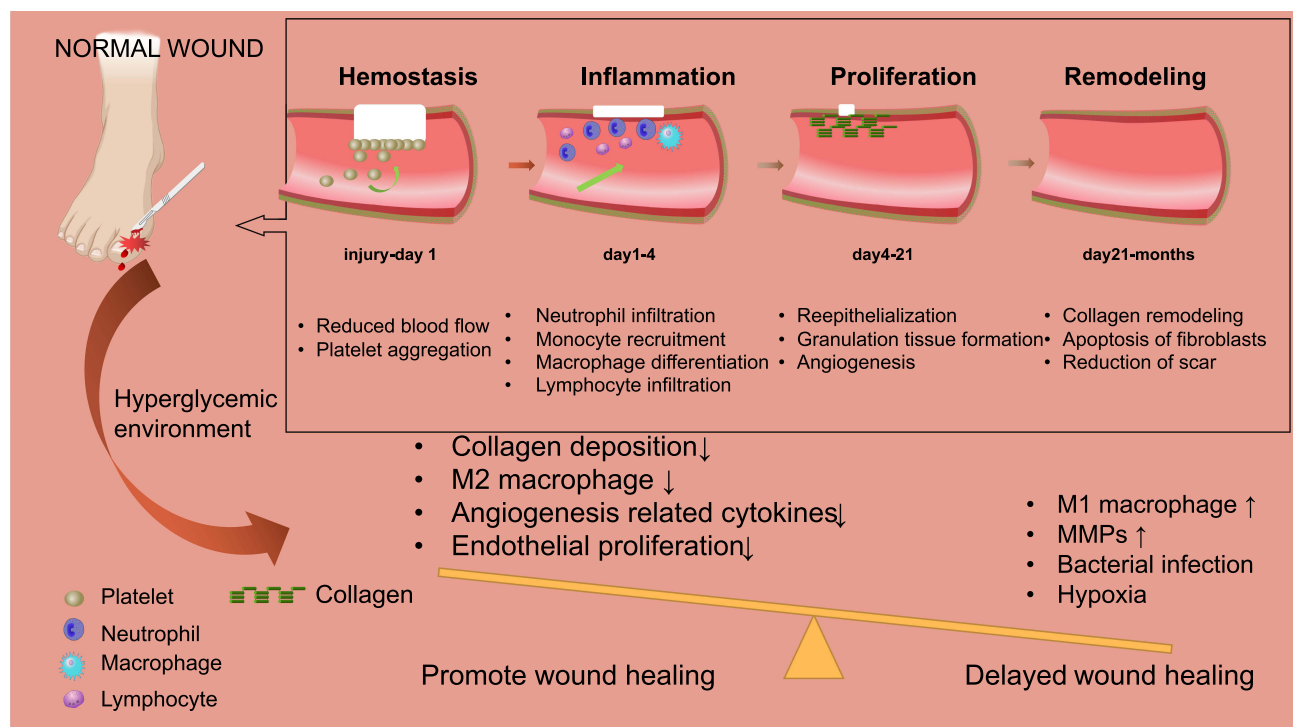
The key steps in the healing process are shown in Figure 1. Platelet aggregation and inflammation start immediately after injury, and neutrophil infiltrates and macrophage differentiation can be seen at the injury site between days 1 and 4 after injury. On days 4 to 21, granulation tissue and neovascularization are apparent. Collagen and skin remodeling and scar tissue removal occur at the wound site after 21 days. The process can be roughly summarized as the four stages of wound healing, closely linked but overlapping: (1) hemostasis, (2) inflammation, (3) proliferation, and (4) remodeling.<sup>20,21</sup>

Platelets form fibrin clots at the injury site immediately after an injury,<sup>22</sup> inducing the synthesis of growth factors and promoting the proliferation and migration of cells.<sup>23</sup> Mast cells release serotonin and histamine, triggering endothelial cells to open up their membrane junctions for unrestricted migration of immune cells.<sup>24</sup> Platelets release cytokines that contribute to localized inflammation and facilitate the formation of granulation tissue, including platelet-derived growth factor (PDGF), transforming growth factor- $\beta$  (TGF- $\beta$ ), epidermal growth factor (EGF), and fibroblast growth factor (FGF).<sup>25</sup> Macrophages phagocytose microbial debris, secrete various cytokines and growth factors, and stimulate the formation of granulation tissue.<sup>26–28</sup> Angiogenesis, fibroblast migration, and ECM production lead to wound healing.<sup>24,27</sup>

In the proliferative phase, new blood vessels grow from existing vessels, collagen is deposited, granulation tissue forms, cells grow, and the wound heals.<sup>29</sup> IL-4 promotes fibroblast differentiation and induces matrix synthesis.<sup>30</sup> When the remodeling period begins, the wound begins to contract. The induction of ECM production begins with tissue granulation and collagen degradation and continues with the production of new collagen. Collagen is remodeled and rearranged along the lines of tension. Finally, old fibroblasts are removed by apoptosis, and new fibroblasts and ECM are generated.<sup>31</sup>

## The Process of Diabetic Wound Healing

In normal physiological conditions, that the injured body follows the natural course of wound healing. However, certain circumstances or abnormal factors can disrupt this process, leading to the development of chronic wounds, such as diabetic wounds. Several factors contribute to the transformation of diabetic wounds into chronic wounds that pose



**Figure 1** Physiological process of wound healing. Comparison of factors promoting wound healing and delaying wound healing.

challenges for effective healing, including. (1) excessive proinflammatory macrophages, (2) microbial interference, and (3) blocked angiogenesis.<sup>32</sup>

Macrophages play a key role in wound healing, and abnormal macrophage polarization in diabetic wounds leads to prolonged wound inflammation. Upon traumatic stimulation, the number of macrophages involved in the inflammatory response increases rapidly, and these cells polarize into classically activated M1 (proinflammatory) subtypes or activated M2 (healing promotion) subtypes.<sup>33</sup> This heterogeneity in the cell population is due to their different responses to microenvironmental signals, described as macrophage polarization.<sup>34</sup> During the initial stages of inflammation, the predominant M1 subtype of macrophages emerges. These M1 macrophages play a vital role in combating bacterial infections, inducing the phagocytosis of pathogens, eliminating neutrophils, clearing tissue debris, and releasing cytokines like tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) and interleukin-6 (IL-6). As tissue remodeling progresses, there is a rise in the M2 macrophages populations.<sup>35</sup> These M2 macrophages are primarily responsible for releasing growth factors such as TGF- $\beta$  and insulin-like growth factor, which actively contribute to tissue regeneration and remodeling.<sup>36</sup> Furthermore, studies have revealed that the impaired recruitment of M2 macrophages hampers diabetic wound healing, while the extraction of TGF- $\beta$  from these macrophages promotes both wound healing and angiogenesis. The impaired activation of macrophages by the nuclear receptor PPAR $\gamma$  has been reported in diabetic wounds, leading to the inability of PPAR $\gamma$  to facilitate wound healing through downregulation of pro-inflammatory factors.<sup>37</sup> Macrophages isolated from peritoneal exudates in streptozotocin-induced diabetic mice demonstrate reduced TNF- $\alpha$  release and an increased production of nitric oxide (NO).<sup>38</sup> These results suggest that hyperglycemia increases the production of M1 macrophages and decreases the generation of the M2 macrophages resulting in a prolonged inflammatory stage and wound healing failure.

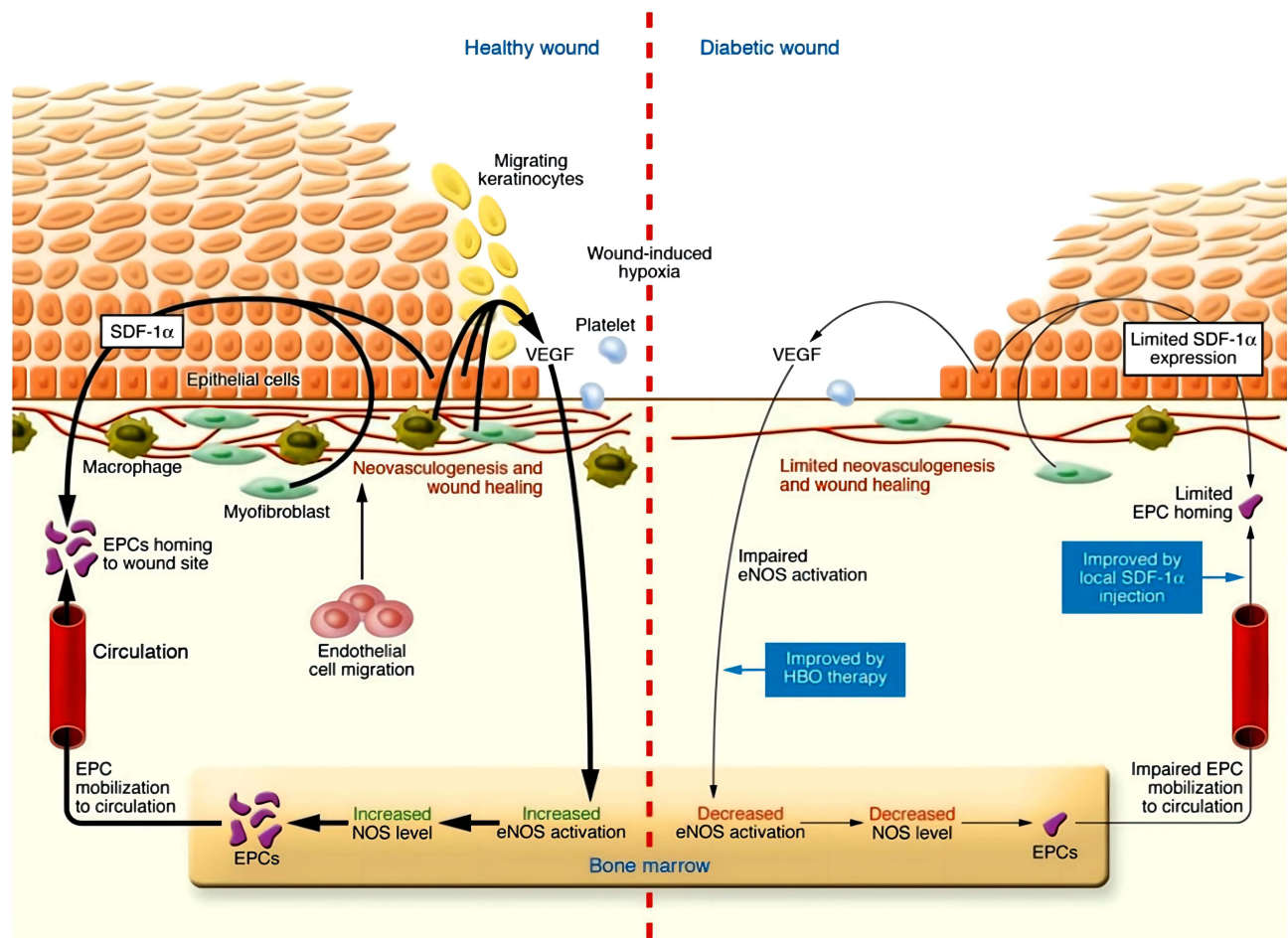
Angiogenesis is an important step in wound healing, and the interaction between related cells and cytokines during angiogenesis is the key to successful blood vessel formation.<sup>39</sup> Therefore, diabetes-induced angiogenesis impairment is another obstacle to regenerating cells at the wound site. Insufficient wound angiogenesis in people with diabetes is manifested by reduced endothelial cell proliferation and a reduced reaction between cells and growth factors.<sup>40</sup> When a wound causes hypoxia, VEGF generated by macrophages, fibroblasts, and epithelial cells activates and phosphorylates eNOS in the bone marrow. This boost in NO levels causes endothelial progenitor cells (EPCs) to mobilize into the bloodstream. The EPCs assist in cellular homing to the site of injury by the chemokine SDF-1 $\alpha$ , where they take part in neovasculation.<sup>41</sup> Gallagher et al demonstrated that impaired eNOS phosphorylation in the bone marrow and reduced SDF-1 $\alpha$  expression in epithelial cells and fibroblasts, observed in a diabetic mouse model (right), resulted in the inability of VEGF to induce eNOS phosphorylation and activation, thereby limiting EPC mobilization from the bone marrow into circulation and reaching the wound site<sup>42</sup> (Figure 2). One study revealed that fibroblasts extracted from diabetic mice have a lower migration rate and decreased VEGF expression,<sup>43</sup> indicating that hyperglycemia damages cell function and inhibits blood vessel formation. In contrast, impaired angiogenesis in diabetic mice with increased VEGF expression is due to decreased VEGF signal transduction and increased sVEGFR-1 expression, which inhibits angiogenesis.<sup>44</sup> Both studies highlight the importance of proper VEGF levels for successful wound healing.<sup>45</sup> When VEGF does not operate properly (by decreased expression or decreased signaling), angiogenesis is impaired, and wound healing is delayed.

Common wound treatments that promote rapid healing include controlling infection, promoting skin perfusion recovery, and minimizing inflammation. However, due to the complexity of diabetes, treatment for diabetic wounds is limited. Traditional dressings remain the primary treatment for diabetic wounds, but the treatment process is long, and the prognosis is not optimal. Thus, researchers have aimed to improve diabetic wound healing by introducing MNPs into wound treatment methods. The subsequent section delineates the pivotal role of diverse MNPs in facilitating wound healing, particularly in the context of diabetic wound healing.

## Research and Application of MNPs in Wound Healing

The repair process of diabetic wounds is characterized by slower and more intricate healing compared to normal wounds, thus necessitating the use of materials that can effectively stimulate rapid and complete healing to promote diabetic wound healing. The materials should additionally demonstrate excellent biocompatibility, be non-toxic to living organisms, and promote a moist wound environment. With the advancement of nanotechnology, MNPs have been





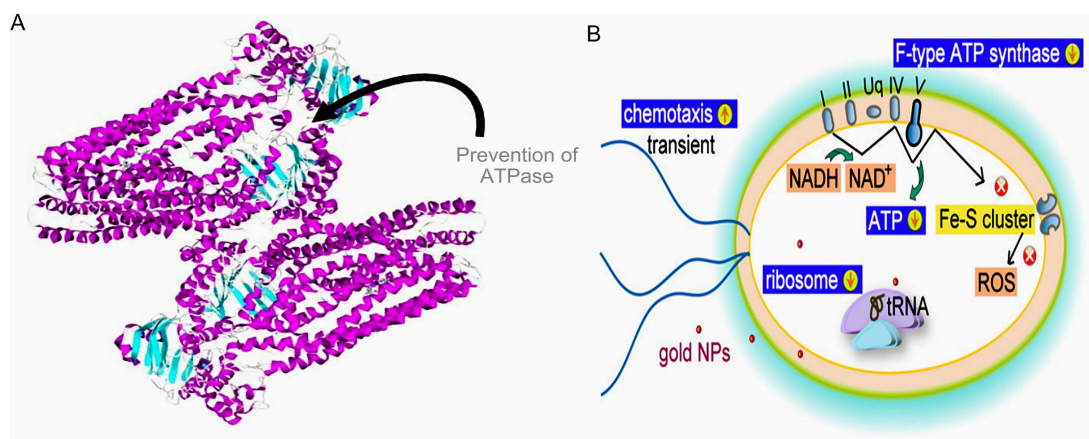
**Figure 2** Comparison of angiogenesis during wound healing in healthy versus diabetic subjects. Used with permission of American Society for Clinical Investigation, from Brem H, Tomic-Canic M. Cellular and molecular basis of wound healing in diabetes. *J Clin Invest*. 2007;117(5):1219–1222; permission conveyed through Copyright Clearance Center, Inc.<sup>41</sup>

demonstrated to possess unique physical and chemical properties that exhibit significant effects on diabetic wound healing. These MNPs directly stimulate wound healing. They also function as indirect carriers in the wound-healing process. Thus, we focus on some of the most commonly studied and clinically used MNPs and their oxides and summarize their effects at the critical stage of wound healing while exploring the potential application of MNPs in combination with other biomaterials as nanomedicine and nanocarriers for treating diabetes wounds.

## AuNPs

Because of their plasmon resonance, biocompatibility, and antioxidant and antibacterial capabilities, AuNPs play a unique role in biomedical applications.<sup>46,47</sup> The antibacterial properties of AuNPs have been exploited to reduce the chance of diabetic wound infection in the study of materials for enhancing diabetic wound healing. AuNPs are also used as carriers to load genes or growth factors to the cells or even the nucleus of the wound site to create a normal wound-healing environment.

The main antimicrobial mechanism of AuNPs is ATP depletion via membrane potential changes and ATP synthase inhibition, which blocks energy metabolism and leads to bacterial death (Figure 3A).<sup>48</sup> According to the study conducted by Cui et al, the presence of AuNPs led to the suppression of oxidative phosphorylation pathways, specifically affecting F-type ATP synthase and ATP levels. The researchers observed a significant reduction in the activity of F-type ATP synthase and the expression of ATP in the membrane proteins of *Escherichia coli* when exposed to AuNPs. Additionally, other studies have demonstrated that bactericidal antibiotics induce cell death by causing an accumulation of excessive



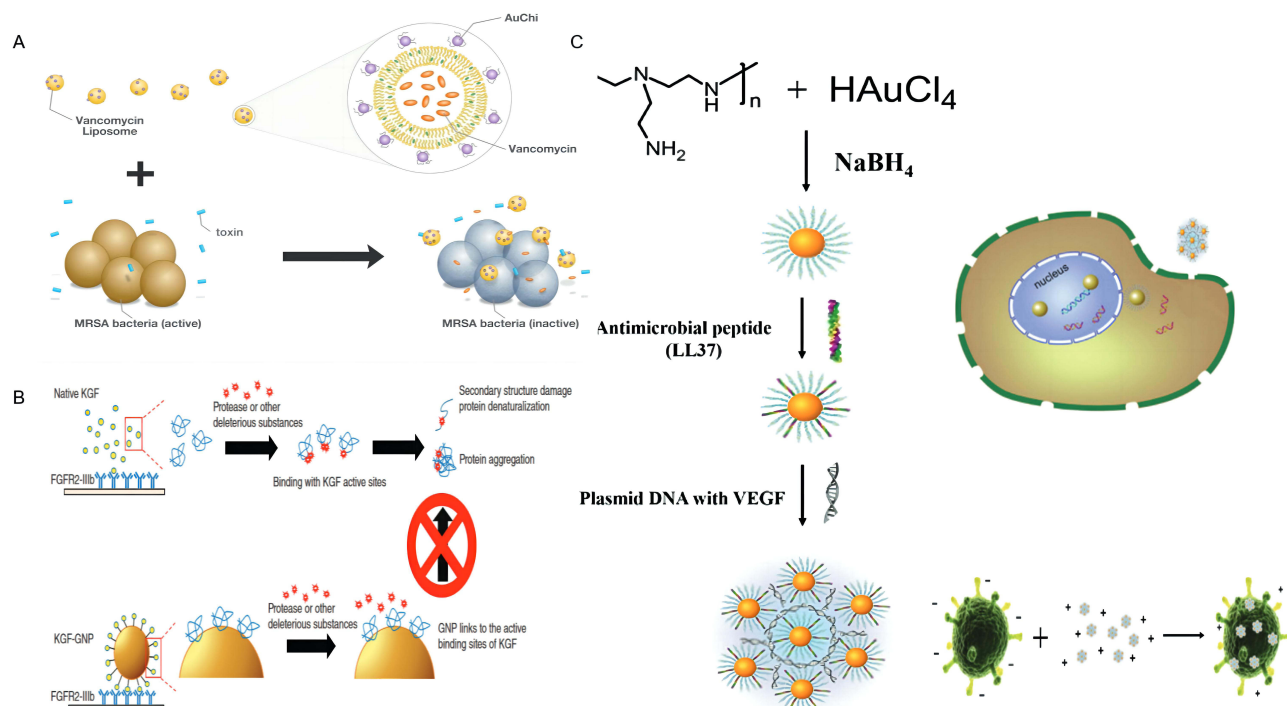
**Figure 3** (A) Changes in membrane potential and inhibition of ATPase activity led to a decrease in cellular metabolism. (B) AuNPs kill bacteria by inhibiting the enzyme ATP synthase.

**Notes:** (A) Reprinted with permission from Dove Medical Press. Vimbela G, Ngo SM, Frazee C, Yang L, Stout DA. Antibacterial properties and toxicity from metallic nanomaterials. *IJN*. 2017;12:3941–3965. Creative Commons.<sup>48</sup> (B) Reprinted from . 33(7). Cui Y, Zhao Y, Tian Y, Zhang W, X L, Jiang X. The molecular mechanism of action of bactericidal gold nanoparticles on *Escherichia coli*. 2327–2333, Copyright (2012), with permission from Elsevier.<sup>50</sup>

hydroxyl radicals. This pathway is closely associated with oxidative damage and results in the depletion of NADH and the tricarboxylic acid cycle.<sup>49</sup> We performed NAD [+] cycling assays and determined ROS levels. The NAD [+] /NADH ratio of AuNP-treated *E. coli* was almost unchanged from that of untreated *E. coli*, suggesting that AuNPs kill bacteria without inducing oxidative stress. Oxidative stress can lead to the death of normal cells in the body (Figure 3B). These results indicate that the low cytotoxicity of AuNPs is because AuNPs may kill bacteria through nonoxidative stress pathways rather than oxidative stress pathways.<sup>50</sup>

The incorporation of exogenous reducing agents, capping agents and stabilizing agents during the synthesis of conventional AuNPs introduces potential toxicities or adverse effects that may compromise their utility. Recently, researchers developed a green synthesis method, which permits AuNPs to be synthesized at mild pressures, temperatures, and pH to avoid many harmful properties. The green AuNPs synthesized by Boomi et al, using an aqueous extract of *Acalypha indica* as a carrier accelerated the inflammatory response and promoted collagen remodeling. The authors also found that the AuNPs synthesized by this method interacted with bacterial cell membranes and exhibited a high antibacterial activity and a high antioxidant activity at a concentration of 100 µg/mL. The combined effect of these factors greatly diminishes the likelihood of wound infection and facilitates the process of wound healing.<sup>51</sup> Annamalai et al employed a green synthesis method to create AuNPs using an extract derived from the leaves of the medicinal plant *Euphorbia hirta*. The AuNPs exhibited excellent antibacterial properties against *Escherichia coli*, *Pseudomonas aeruginosa*, and *Klebsiella pneumoniae*.<sup>52</sup> In similar studies, AuNPs synthesized from the extract of the leaves of the insulin-like plant *Astragalus membranaceus* demonstrated similarly good antioxidant activities, and the antioxidant and anti-inflammatory properties increased over time.<sup>53</sup> The augmented antibacterial efficacy of AuNPs may be attributed to the synergistic effect between the plant extracts and AuNPs.<sup>48</sup> AuNPs synthesized from plant extracts not only contribute to the antibacterial action of AuNPs, but also play different functions in different plant extracts. Due to the diversity of plant extracts, AuNPs synthesized from plant extracts may have different roles. AuNPs can also selectively inhibit lipopolysaccharide-induced (LPS-induced) cytokine production by, downregulating IL-1β-induced epithelial cell proliferation.<sup>54</sup> Barathmanikant created AuNPs using biosynthesis techniques and found that AuNPs (2.5 mg/kg in diabetic mice) created conditions for controlling hyperglycemia by inhibiting ROS formation and lipid peroxidation and scavenging free radicals. The AuNPs synthesized from the extract promoted glucose consumption and produced insulin-like results. Furthermore, the AuNPs demonstrated no toxicity to vital organs and sustained control of blood glucose and lipids.<sup>55</sup>

Due to the abundance of positive charges on their surface, AuNPs serve as a versatile drug delivery platform, as their surface can be readily modified with various medications. Pornpattananangkul et al developed chitosan-coated AuNPs-stabilized phospholipid liposomes (Vancomycin AuChi-Liposome) as a drug delivery platform for antibiotics



**Figure 4 (A)** Vancomycin-loaded liposomes protect them by adsorbing chitosan-coated AuNPs on their surfaces to prevent them from fusing with each other or with bacterial membranes. When bacterial toxins bind to liposomes, the toxins create pores in the liposomes and release vancomycin to kill bacteria. Reprinted with permission from Pornpattananagkul D, Zhang L, Olson S, et al. Bacterial toxin-triggered drug release from gold nanoparticle-stabilized liposomes for the treatment of bacterial infection. *J Am Chem Soc.* 2011;133(11):4132–4139. Copyright © 2011, American Chemical Society.<sup>57</sup> **(B)** Hypothesized mechanism of KGF-GNPs on protein stability. Used with permission of Future Medicine Ltd. Li S, Tang Q, Xu H, et al. Improved stability of KGF by conjugation with gold nanoparticles for diabetic wound therapy. *Nanomedicine.* 2019;14(22):2909–2923.<sup>60</sup> Permission conveyed through Copyright Clearance Center, Inc. **(C)** AuNPs@LL37/pDNAs preparation and antibacterial diagram; AuNPs@LL37/pDNAs cell uptake and cell-to-cell gene delivery to optimize the therapeutic effect of bacterial infected DFUs. Used with permission of Royal Society of Chemistry. Wang S, Yan C, Zhang X, et al. Antimicrobial peptide modification enhances the gene delivery and bactericidal efficiency of gold nanoparticles for accelerating diabetic wound healing. *Biomater Sci.* 2018;6(10):2757–2772.<sup>62</sup> Permission conveyed through Copyright Clearance Center, Inc.

(Figure 4A). They combined positively charged AuNPs with negatively charged liposomes to create a stable liposomal composite that prevents them from fusing and reduces the rate at which the encapsulated antibiotics are released. The stability of AuNPs to liposomes leads to the sustained release of vancomycin, thus vancomycin shows a sustained and highly toxic inhibitory effect on bacteria. The  $\alpha$ -toxin secreted by *S. aureus* specifically binds to the outer membrane of susceptible cells.<sup>56</sup> The rapid poring of liposomes by the binding of the toxin causes liposome cleavage and triggers the release of the antibiotic (vancomycin) from AuNP-stable liposomes to inhibit the growth of *S. aureus*.<sup>57</sup>

Li et al used 4,6-diamino-2-pyrimidinethiol (DAPT)-coated AuNPs (D-Au NPs) to treat intestinal *E. Coli*-induced bacterial infection.<sup>58,59</sup> Due to the negative charge on the surface of proteins, AuNPs can be modified by proteins, stabilize the structure and function of proteins, and be delivered to the site of action. KGF binds to 60 nm gold nanoparticles (GNPs) and poly (ethylene glycol) thiolates to produce KGF-GNPs, which means that KGF can be stably applied to wounds with high efficiency.<sup>60</sup> CD spectroscopy has been widely recognized and used as a method to measure changes in protein secondary structure.<sup>61</sup> In this study, the KGF secondary structure did not change after KGF-GNP crosslinking, as measured by CD. In addition, KGF molecules are tightly clustered on the surface of GNPs, making it easier to simultaneously target nearby KGF receptors (Figure 4B). The secondary structure of KGF-GNPs exhibits exceptional resilience against harsh conditions, including elevated temperatures, acidic environments, and enzymatic degradation. The slow release rate of KGF, as demonstrated in vitro, showcases its stability and biological activity. When KGF-GNPs were utilized on wounds in diabetic mice, they expedited the healing process and promoted re-epithelialization. The expression of key factors like COL-I,  $\alpha$ -SMA, and TGF- $\beta$ 1 were significantly augmented, indicating the positive impact of KGF-GNPs. Wang et al developed a gene delivery system (AuNPs@LL37) by grafting AuNPs onto antimicrobial peptides LL37 as carriers, which acted as both an antimicrobial agent and a nuclear

translocation facilitator<sup>62</sup> (Figure 4C). The cellular uptake, intracellular distribution, and in vitro bactericidal efficacy of AuNPs@LL37 were investigated to elucidate its mechanism of gene delivery. Incorporating AuNPs into LL37 enhanced the composite stability and antibacterial properties while reducing toxicity. The negatively charged surface of MRSA attracted the complex formed by AuNPs@LL37, leading to observed lesions and collapse on MRSA surfaces. AuNPs@LL37 was also bound with plasmid DNA expressing VEGF forming an effective complex termed as AuNPs@LL37/pDNA. In vivo studies using a diabetic wound healing model showed that the complex formed by AuNPs@LL37/pDNA successfully enhanced wound healing efficacy by promoting increased VEGF expression within nuclei along with accelerated angiogenesis. Similarly, Chen et al demonstrated that AuNPs could be used to promote angiogenesis and for gene delivery. AuNPs instantly open the skin's cuticle and increase the absorption of specific compounds through the skin. The authors discovered that the combination of EGCG and alpha lipoic acid antioxidants with AuNPs effectively regulated inappropriate inflammation, oxidative stress, and vascular injury. This synergistic activity significantly expedited diabetic wound healing through precise anti-inflammatory and angiogenic modulation, wherein AuNPs functioned as an adjuvant to augment cutaneous absorption and enhance antioxidant efficacy.<sup>63</sup>

In the aforementioned scenarios, AuNPs were demonstrated to serve a dual role in promoting wound healing. Firstly, they facilitate the efficient healing of infected wounds through their inherent antibacterial properties. Secondly, green synthesis methods employed to synthesize AuNPs using various plant extracts, not only augment their antibacterial activity but also expedite the resolution of inflammation during diabetic wound healing via anti-inflammatory and antioxidant mechanisms. Consequently, this broadens the scope of AuNPs beyond their conventional application as metal-based antimicrobials.

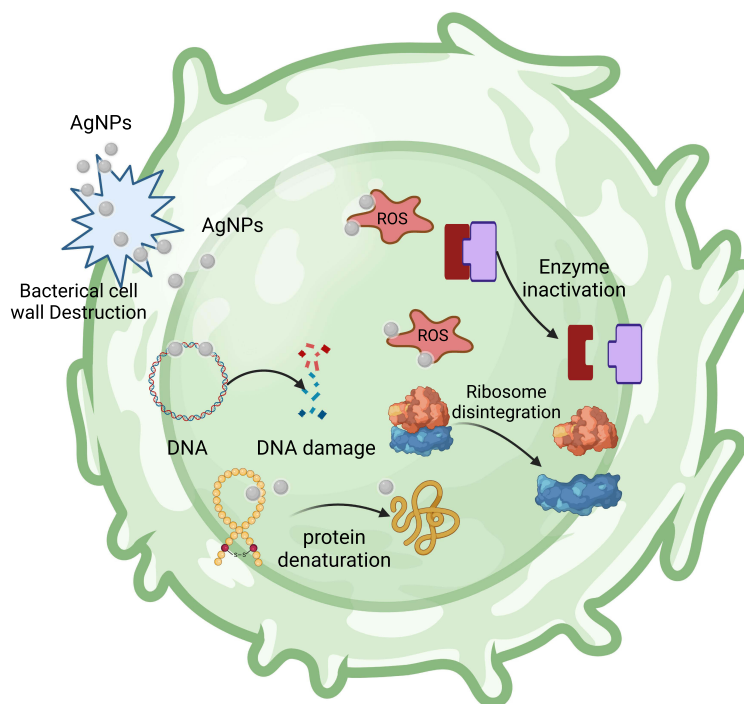
## AgNPs

AgNPs have become the most widely studied and used MNPs due to their obvious anti-inflammatory properties and effective inhibition of drug-resistant bacteria.<sup>64</sup> Numerous studies have described the effect of AgNPs or AgNPs combined with other composite materials on diabetic wound healing. As shown in Table 1, some of these dressings containing AgNPs have been widely used in clinical practice.<sup>65</sup> The antibacterial mechanisms of AgNPs encompass the

**Table 1** The Mechanism Underlying the Diabetic Wound Healing of MNPs

Nanoparticles	Antibacterial Mechanism	Ref.	Anti-Inflammatory Mechanism	Ref.	Promote Angiogenesis	Ref.
AuNPs	Change bacterial cell membrane potential and inhibit ATP synthase activity; Antibiotic delivery platform		Decreasing LPS induced cytokine production	[54]		
AgNPs	Destroy cell membranes; Oxidative stress; Protein binding and inactivation; Inhibit DNA replication	[63,68,69]	Suppressing pro-inflammatory cytokine production; Reduced macrophage inflammatory response	[70–73]		
CuNPs/ CuONPs	Depletion of cell membrane potential resulting from the formation of cell filaments; Induction of intracellular reactive oxygen species Bacterial membrane rupture	[74–76]	PTT accelerates the transition of inflammatory phase; Accelerate the transition of wound inflammation	[73,77]	Stimulate VEGF expression	[78,79]
TiO <sub>2</sub> NPs	Production and sustained release of hydroxyl radical superoxide ions;	[80,81]	Reducing platelet numbers	[82,83]		
ZnONPs	Disruption of cell membrane; Oxidative stress	[84,85]	Blocking pro-inflammatory cytokines; Suppressing LPS induced COX-2 expression; Regulation of macrophage polarization	[86–89]	Stimulates angiogenesis related growth factors	[90]
FeNPs CeO <sub>2</sub> NPs	Induce bacteria to produce ROS	[91]	Generation of ROS Reduce ROS production levels	[92] [93]	Stimulating endothelial cells to up-regulate HIF-1 $\alpha$ expression	[94]
Y <sub>2</sub> ONPs			Limiting ROS production	[94]		





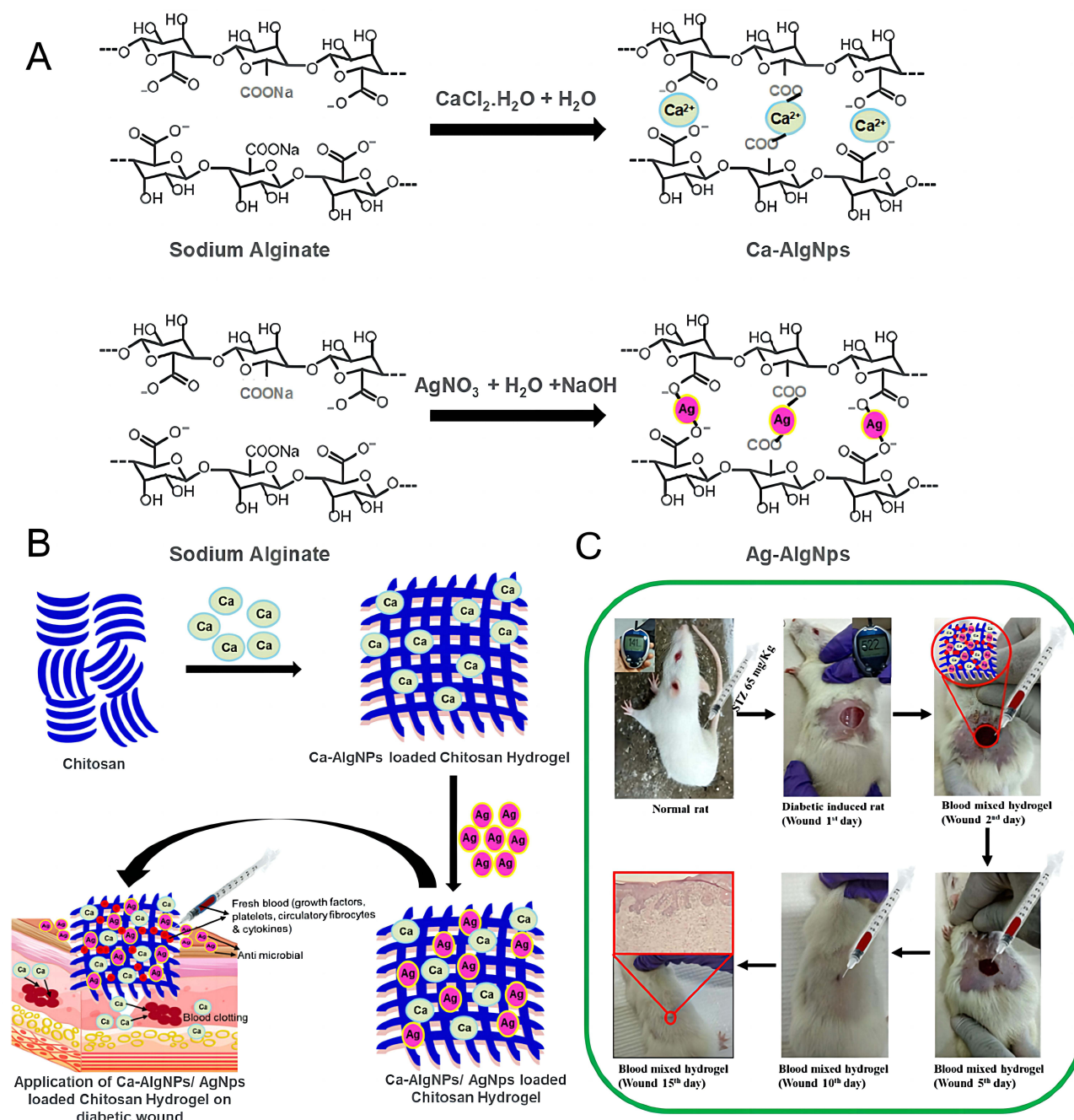
**Figure 5** Schematic representation of the antimicrobial mechanism of AgNPs, describing the ROS-dependent pathway, DNA damage, protein denaturation and enzymatic inactivation of the antimicrobial action of AgNPs. Adapted from Xu L, Wang YY, Huang J, Chen CY, Wang ZX, Xie H. Silver nanoparticles: synthesis, medical applications and biosafety. *Theranostics*. 2020;10(20):8996–9031. Creative Commons.<sup>17</sup>

ROS-dependent pathway, protein binding and inactivation, as well as inhibition of DNA replication (Figure 5).<sup>17,66</sup> The  $\text{Ag}^+$  released by AgNPs binds mercaptan to proteins and enzyme sets (-SH) found on the cell surface, leading to cell membrane instability and a disruption of ATP synthesis pathways.<sup>67</sup> AgNPs subsequently adhere to the membrane wall, leading to the formation of pores through which they penetrate the bacteria and interact with intracellular components or sulfur-containing proteins.<sup>66</sup> The collapse of the proton motive force and inhibition of ATP generation occur upon disruption of the bacterial cell wall and membrane.<sup>68</sup>

The  $\text{Ag}^+$  released by AgNPs causes the intracellular formation of a wide spectrum of membrane-reactive ROS, which eventually causes cell death due to oxidative stress.<sup>48</sup> During Ag treatment,  $\text{Ag}^+$  may produce superoxide radicals (the primary form of ROS) by damaging enzymes in the respiratory chain. This damage may be caused by  $\text{Ag}^+$ -mercaptan interactions.<sup>95</sup> The observed mechanism demonstrates the broad-spectrum antibacterial activity of AgNPs against diverse bacterial strains, encompassing both gram-positive and gram-negative bacteria, which possess distinct cell structures.<sup>2</sup> The antibacterial activity of AgNPs is attributed to the simultaneous involvement of multiple mechanisms of action.  $\text{Ag}^+$  demonstrates a remarkable synergistic effect, exhibiting promising antimicrobial activity against a wide range of drug-resistant bacteria.<sup>96</sup>

To regulate the abrupt discharge of AgNPs, which leads to an elevated local concentration of  $\text{Ag}^+$  and cytotoxicity, we utilized the gradual degradation characteristics of the hydrogel to achieve a controlled release effect. The chitosan and dextran within the AgNP@CNDM hydrogel create an internal network channel for absorbing wound exudates, while also allowing for the gradual degradation the chitosan to release  $\text{Ag}^+$  ions that can act on bacterial cell membranes, inhibiting inflammatory responses and accelerating wound healing in diabetic infections.<sup>97</sup> Similarly, the chitosan/Ca-AIcNP/AgNP hydrogel also utilizes the biodegradation function of chitosan to gradually release  $\text{Ca}^{2+}$  from AgNPs and Ca alginate nanoparticles within the hydrogel, thereby facilitating hemostasis acceleration and achieving continuous sterilization.<sup>98</sup> In further investigation, the sustained release properties of the hydrogel were utilized to introduce fresh blood into the chitosan/Ca-AIcNP/AgNP hydrogel. By harnessing the growth factors present in the blood, an artificial enrichment of these growth factors within the hydrogel was achieved, thereby enhancing wound healing<sup>99</sup> (Figure 6A–C). Wang et al





**Figure 6 (A–C)** Illustration of the synthesis of calcium alginate nanoparticles (Ca-AlgNPs) and AgNPs and the promotion of wound healing by combining Ca-AlgNPs and AgNPs on a gel and injecting them into wounds of diabetic mice. Reprinted from *Int J Biol Macromol*. Volume 178. Choudhary M, Chhabra P, Tyagi A, Singh H. Scar free healing of full thickness diabetic wounds: a unique combination of silver nanoparticles as antimicrobial agent, calcium alginate nanoparticles as hemostatic agent, fresh blood as nutrient/growth factor supplier and chitosan as base matrix. 41–52. Copyright © 2021, with permission from Elsevier B.V.<sup>99</sup>

utilized alginate-based hydrogels containing hemin (protoporphyrin IX with a ferric iron ion) and AgNPs to enhance antibacterial activity and ROS scavenging, while also promoting cell attachment and growth.<sup>100</sup> Similarly, AgNPs were synthesized from *aloe leaf* extracts using biosynthetic techniques. These green MNPs showed satisfactory inhibition of *Staphylococcus aureus*. A mixture of green AgNPs and photobiomodulation (PBM) encourages cell migration to diabetic wounds and exerts antibacterial effects.<sup>70</sup> PBM leads to the proliferation of cell migration activity and increases collagen production in diabetic human skin fibroblasts.<sup>101</sup> By activating cellular processes, PBM assists in treating diabetic wounds.<sup>72</sup> The biogenic AgNPs also exhibit potent antibacterial activity against the most prevalent wound infection pathogens, namely *Pseudomonas aeruginosa* and *Staphylococcus aureus*.

AgNPs facilitate the transition from the inflammatory phase to the proliferative phase of diabetic wounds, thereby promoting wound healing. This is achieved through the suppression of proinflammatory cytokine secretion and expression. Singla and colleagues utilized *bamboo cellulose nanocrystals* loaded with AgNPs to develop nanobiocomposite (NC) hydrogels, which demonstrated a remarkable ability to expedite wound healing in diabetic individuals. These NCs effectively inhibit the expression of proinflammatory cytokines and growth factors, further enhancing the healing process.<sup>71</sup> The acceleration of the healing process can be attributed to a reduction in inflammation, early proliferation, and enhanced collagen formation. In a study conducted by Frankova et al, the effects of AgNPs on normal dermal fibroblasts (NHDFs) and normal epidermal keratinocytes (NHEKs) were examined. The findings revealed that AgNPs have the potential to promote wound healing by suppressing the secretion of proinflammatory cytokines such as IL-8, IL-6, IL-12, and TNF- $\alpha$  by NHDFs and NHEKs, particularly when the wound is not infected.<sup>102</sup> Rangasamy et al prepared nanoparticles (AgPyNPs) composed of silver-pyridoxine complexes using the antioxidant properties of pyridoxine.<sup>103</sup> They observed that AgPyNPs induced interleukin-8 (IL-8) secretion to enhance the immune response of macrophages to eliminate microorganisms. At the same time, the antioxidant properties of pyridoxine reduce cytotoxicity by reducing the production of ROS in macrophages. In addition, AgPyNPs induce IL-8 to increase wound reepithelialization by stimulating keratinocyte proliferation and migration.<sup>72</sup> Therefore, we speculate that AgNPs accelerate the wound healing rate in a hyperglycemic environment where the wound site poorly heals due to a prolonged inflammatory period.

Compared with other MNPs, AgNPs are the most widely used in wound healing and entered clinical use earlier. Their excellent antibacterial activity, especially their ability to inhibit drug-resistant bacteria, makes AgNPs widely popular. However, with the deepening of other MNP studies, AgNPs play a single role in diabetic wound healing. However, Fathil et al successfully formed AgLTF by complexing AgNPs with LTF and utilized this for the targeted therapy of diabetic wounds by loading DsiRNA.<sup>104</sup> The negatively charged DsiRNA was adsorbed onto AgLTF to silence the prostaglandin transporter (PGT) and promote wound healing. Although this study did not employ AgNPs as a drug carrier, the successful adsorption of DsiRNA onto AgLTF demonstrates the potential use of AgNPs as carriers in future applications beyond their antibacterial properties. In the future, AgNPs have the potential to serve as highly efficient antibacterial agents and carriers for protein or nucleic acid adsorption, thereby modifying the wound microenvironment and facilitating diabetic wound healing.

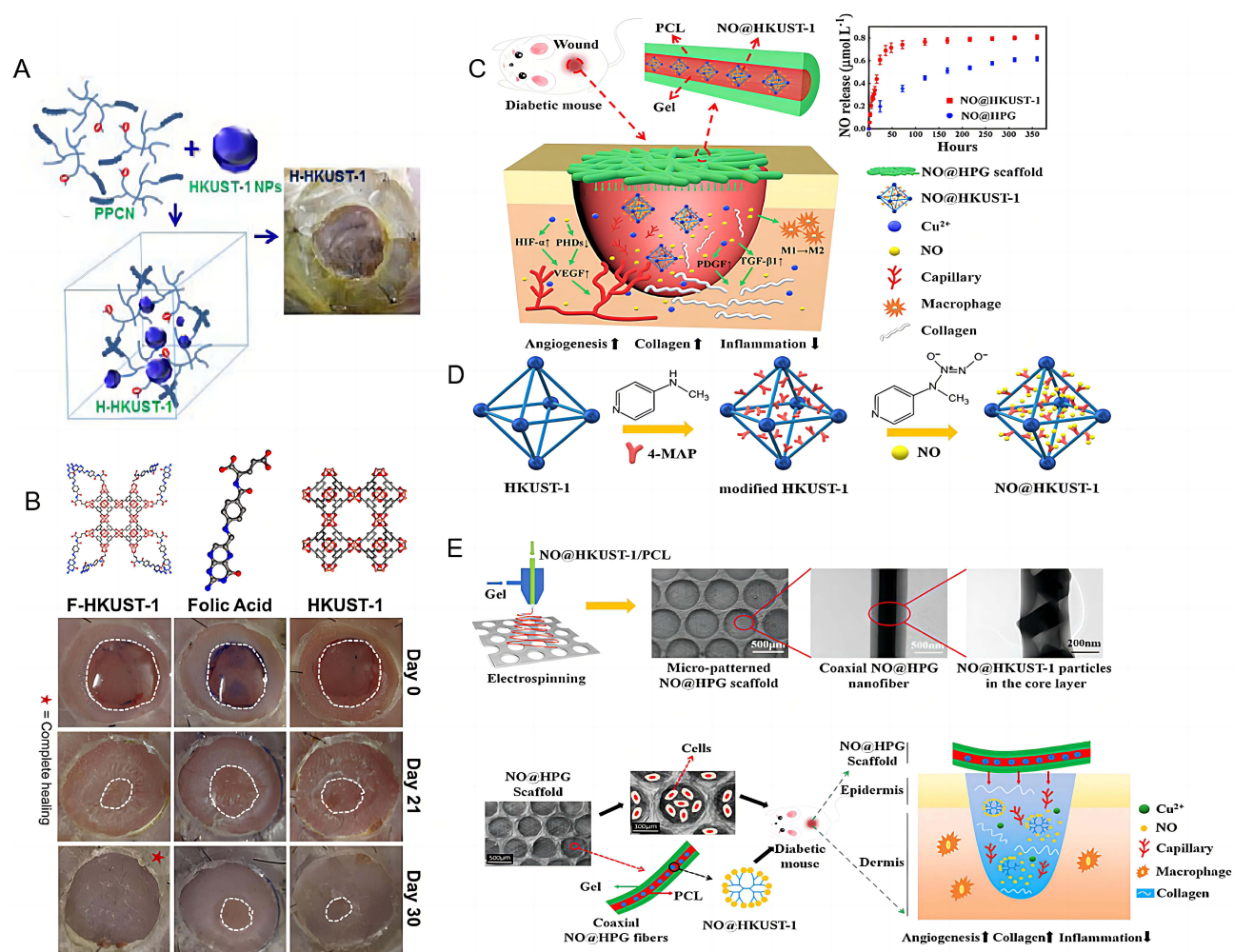
## CuNPs

The application of Cu<sup>2+</sup> in biomaterials has a long history, beginning with contraceptive IUDs. Cu<sup>2+</sup> demonstrates antibacterial properties similar to those of many other metal ions,<sup>105</sup> making them widely applicable. It can be argued that rapid control of wound infection can also promote faster healing. Chatterjee reported a novel method for synthesizing stable CuNPs that are highly efficient at bacterial cell filamentation and cell killing. In addition to inducing membrane lysis and intracellular production of reactive ROS that inhibit bacterial growth and division, CuNPs contribute to the remodeling of bacterial lipopolysaccharides and promote antibacterial membrane activity by reducing cell–cell aggregation and matrix instability.<sup>74</sup> Different CuNP synthetic methods and conditions produce NPs with variable antibacterial effects. Furthermore, the antibacterial efficacy of CuNPs was significantly augmented by the photothermal response of Cu.

Veerapandian et al successfully synthesized mixed glucosamine-functionalized CuNPs (GLCN-CuNPs) and observed that the antibacterial activity of GLCN-CuNPs surpassed that of conventional CuNPs. The in vitro antibacterial activity of GLCN-CuNPs under UV irradiation was higher than that of GLCN-CuNPs and kanamycin without UV exposure. These results may be due to the modification of GLCN-CuNPs surfactant metal nanometers, promoting the production of oxygen-free radicals and their derived structural activities.<sup>75</sup> Similarly, the concentration and size of the crystal structure of CuNPs also impact the antibacterial activity. Hassan et al observed increased bacterial inhibition with increasing concentrations of CuO nanocrystals, which are known to produce ROS. These ROS interact with the cell membrane to produce free radicals, which enter the cell and damage its internal contents. Eventually, this ROS disruption causes bacterial cells to leak.<sup>106</sup> The antibacterial effects of CuONPs and CuNPs are also quite different. The antibacterial activity of CuNPs surpasses that of CuONPs, potentially due to the enhanced electron transfer between bacteria and CuNPs. The interaction between MN bacteria and positively charged MNPs facilitates electron transfer and disruption of bacterial membranes. Akhavan et al compared the interaction between CuONPs and CuNPs on bacteria and found that CuNPs were more successful in inhibiting bacterial growth. More direct interactions between CuNPs and bacterial strains may lead to greater infiltration and rupture of bacterial membranes. Cell membrane rupture can lead to cellular enzyme

dysfunction and, ultimately, cell death.<sup>107</sup> CuONPs synthesized via a green method using *Ficus religiosa* leaf extract show long-lasting inhibitory activity against human pathogens. CuONPs also inhibit the survival of pathogenic bacteria in albino rat wounds and enhance wound healing activity.<sup>108</sup> Studies on the antibacterial properties of CuNPs reveal that CuNPs induce bacterial membrane lysis and promote ROS production in bacteria through the action of positively charged CuNPs and negatively charged bacterial membranes. The photothermal responsiveness of Cu may also catalyze the surface activity of CuNPs under UV light to enhance their antimicrobial activity.

Metal-organic frameworks (MOFs) have emerged as promising drug delivery carriers in biomedicine, thanks to their remarkably porous internal structure. However, their potential application is hindered by their inherent instability in protein solutions, posing challenges to their effective utilization.<sup>109</sup> Zhang et al used the high porosity of HKUST-1 to adsorb NO and created a micropatterned scaffold containing NO@HKUST-1 (NO@HPG scaffold) in the core layer of the nanofibers to stably load NO onto HKUST-1 (Figure 7C and D). They found that NO@HPG scaffolds released Cu<sup>2+</sup> and NO at sustained low concentrations, synergistically promoting hematopoiesis and collagen deposition.

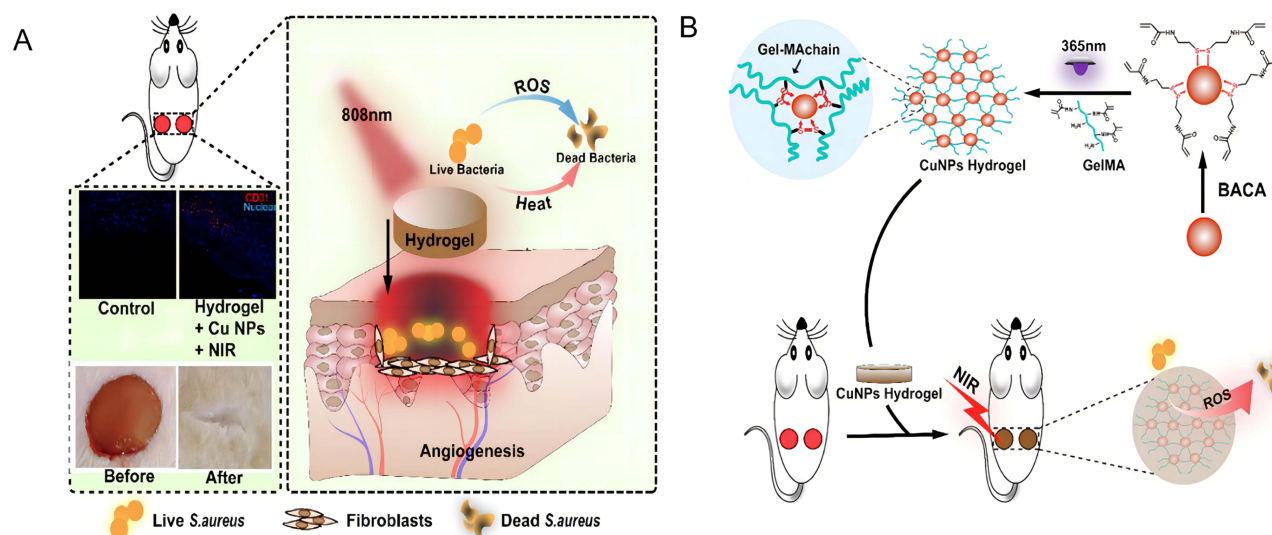


**Figure 7** (A) Schematic illustration of PPCN binding to H-HKUST-1 and its effect on the wound. Adapted from Xiao J, Chen S, Yi J, Zhang HF, Ameer GA. A cooperative copper metal-organic framework-hydrogel system improves wound healing in diabetes. *Adv Funct Mater.* 2017;27(1):1604872<sup>113</sup> © 2016, WILEY - VCH Verlag GmbH & Co. KGaA, Weinheim. Permission conveyed through Copyright Clearance Center, Inc. (B) Comparison of wound healing between folic acid modified HKUST-1 and the other two groups. Reprinted with permission from Xiao J, Zhu Y, Huddleston S, et al. Copper metal-organic framework nanoparticles stabilized with folic acid improve wound healing in diabetes. *ACS Nano.* 2018;12(2):1023–1032. Copyright © 2018, American Chemical Society.<sup>115</sup> (C and D) The NO@HPG scaffold promotes angiogenesis, collagen deposition and inhibition of the inflammatory response in diabetic mice through the slow release of Cu<sup>2+</sup> and NO. A NO sustained-release system with core-shell structure was designed by electrospinning using a Cu-based organic-organic framework (HKUST-1) loaded with NO. To promote wound healing in diabetic mice. (E) Synthesis of NO@HKUST-1 and synthesis of NO@HKUST-1/PCL/Gel (NO@HPG) scaffolds acting on wounds. Figures 7C-E were Reprinted with permission from Zhang P, Li Y, Tang Y, et al. Copper-based metal-organic framework as a controllable nitric oxide-releasing vehicle for enhanced diabetic wound healing. *ACS Appl Mater Interfaces.* 2020;12(16):18319–18331. Copyright © 2020, American Chemical Society.<sup>110</sup>



Simultaneously, the release of  $\text{Cu}^{2+}$  and NO from the NO@HPG scaffold attenuated the inflammatory response and facilitated the transition of wounds from the inflammatory phase to the proliferative phase (Figure 7E).<sup>77,110</sup> Hyperglycemia in diabetic patients can impede endogenous NO synthesis, thereby contributing to the delayed healing of diabetic wounds.<sup>111</sup> Therefore, augmenting endogenous NO synthesis or administering exogenous NO can hinder inflammation, foster angiogenesis and stimulate collagen deposition during the process of diabetic wound healing.<sup>112</sup> Xiao et al synthesized HKUST-1 NPs embedded in PPCN (H-HKUST-1) by embedding Cu metal-organic frameworks (HKUST-1 NPs) into an antioxidant thermoresponsive citrate-based hydrogel (PPCN). This hydrogel effectively reduced the abrupt release of  $\text{Cu}^{2+}$  and cytotoxicity. Hydrogels also increase the stability of HKUST-1 NPs in physiological protein solutions, allowing them to be applied without losing their function due to premature degradation. In addition, the slow release of  $\text{Cu}^{2+}$  from H-HKUST-1 stimulated wound angiogenesis and promoted collagen synthesis and deposition in diabetic mice, thereby increasing epidermal regeneration (Figure 7A).<sup>113</sup> Zhang et al demonstrated that surface coating of Cu-based MOF effectively retarded the release of  $\text{Cu}^{2+}$  and enhanced the in vivo stability of Cu-based MOF within a microenvironment.<sup>114</sup> In a subsequent study, surface modification techniques were used to increase the hydrophobicity of Cu-based metal-organic frameworks (Cu-MOFs) to ensure the stability of Cu-MOFs in protein solutions. They therefore used folic acid to modify the surface of Cu-MOFs to increase the hydrophobicity of Cu-MOFs and synthesized folic acid-modified HKUST-1 (F-HKUST-1). The study demonstrated that modification with folic acid not only achieved a slow release of  $\text{Cu}^{2+}$  but was also less cytotoxic and safer than the conventional fluorine-containing hydrophobic coating F-HKUST-1. Based on the performance of the HKUST-1 NPs described above, F-HKUST-1 also promotes angiogenesis and collagen synthesis in wounds (Figure 7B).<sup>115</sup> To facilitate angiogenesis during the early stages of wound healing, HKUST-1 was engineered by incorporating biotin and combining it with an acellular dermal matrix (ADM) scaffold to create a specialized biological niche for cellular attachment and proliferation (known as ADM-B-HKUST-1), which serves as a conducive environment for hosting mesenchymal stem cells (MSCs). By releasing  $\text{Cu}^{2+}$  and promoting MSC adhesion, ADM-B-HKUST-1 demonstrates potential in addressing angiogenic disorders associated with diabetic wounds.<sup>116</sup> Stabilizing the CuNPs structure by embedding it in hydrogels or modifying the surface of MOFs to release  $\text{Cu}^{2+}$  slowly stimulates collagen deposition at the wound site at a more suitable concentration than traditional CuNPs. Therefore, the Cu-based organic framework serves as a dual-functional platform for wound healing; not only does it act as a drug carrier to promote wound healing, but the released  $\text{Cu}^{2+}$  participates in treating the refractory wounds associated with diabetes mellitus (Figure 7C). In future studies, this vector can be used to load proteins, growth factors, genes or even living cells to increase the range of applications of Cu-based MOFs.

$\text{Cu}^{2+}$  exhibits the potential to stimulate angiogenesis<sup>117</sup> and augment the expression of skin-related proteins in diabetic wounds.<sup>118</sup> Huang et al utilized electrospinning to fabricate a radially structured fibrous membrane composed of a blend of polycaprolactone (PCL) and polyvinylpyrrolidone (PVP), incorporating copper peroxide nanoparticles (CPs) as an agent for chemo-kinetic therapy (CDT).<sup>119</sup> The most innovative aspect lies in the in-situ generation of  $\text{H}_2\text{O}_2$  within the acidic environment of diabetic wounds for the Fenton reaction, which effectively eliminates wound bacteria. Simultaneously,  $\text{Cu}^{2+}$  released enhances in vitro angiogenesis by modulating human umbilical vein endothelial cell (HUVEC) behavior. Alizadeh et al demonstrated that at non-toxic concentrations, CuNPs promoted endothelial cell proliferation and migration.<sup>79</sup> As previously stated, the thermal sensitivity of Cu enhances its antibacterial properties and facilitates the healing process of wounds. Could the photothermal responsiveness of Cu also stimulate wound proliferation and angiogenesis by stimulating the rapid release of  $\text{Cu}^{2+}$ ? Tao et al developed a composite hydrogel composed of methacrylate-modified gelatin and N, N-bis (allyl) cysteamine (BACA)-chelated CuNPs synthesized via radical polymerization and photosensitization. Using the local surface plasmon resonance (LSPR) effect, CuNPs effectively converted energy from near-infrared laser irradiation (808 nm) into local heat for photothermal treatment (Figure 8A). The photothermal properties of CuNPs permit the rapid release of antibacterial  $\text{Cu}^{2+}$ . In addition, the  $\text{Cu}^{2+}$  that is released has the potential to promote the proliferation of NIH-3T3 fibroblasts and facilitate neovascularization, while minimizing any inflammatory reactions<sup>73</sup> (Figure 8B). Exploiting thermal properties to improve wound healing is an active area of research. For example, Wang et al created a tissue-engineered membrane to treat tumors and regenerate skin tissue. Using an improved electrostatic spinning method, the authors incorporated  $\text{Cu}_2\text{S}$  nanoflowers into biopolymer fibers and successfully prepared micropatterned nanocomposite films. The  $\text{Cu}^{2+}$  released after the degradation of polymer scaffolds



**Figure 8 (A)** The CuNPs embedded in the hydrogel (BACA/Cu NPs/Gel-MA hydrogel) generate ROS combined with photothermal treatment to kill bacteria and promote wound healing. **(B)** Schematic diagram of CuNPs embedded in hydrogels and synergistic antibacterial activity by photothermal therapy and ROS production under visible light irradiation at 808 nm. Used with permission of Royal Society of Chemistry from Tao B, Lin C, Deng Y, et al. Copper-nanoparticle-embedded hydrogel for killing bacteria and promoting wound healing with photothermal therapy. *J Mater Chem B*. 2019;7(15):2534–2548; permission conveyed through Copyright Clearance Center, Inc.<sup>73</sup>

promoted skin cell growth and capillary formation.<sup>120</sup> VEGF expression is known to be inhibited in diabetic wounds.<sup>121</sup> Enhancing the  $\text{Cu}^{2+}$  concentration at the site of injury stimulates VEGF expression to facilitate angiogenesis and promote connexin expression during the process of wound healing.<sup>76</sup> For example, Borkow et al prepared wound-healing dressing in CuO that induced the production of VEGF and HIF-1 $\alpha$ . VEGF plays a significant role in promoting angiogenesis, while HIF-1 aids in the induction of protein expression related to the formation of the vascular network and wound recovery.<sup>122</sup> Cu also contributes to wound healing wounds by promoting the stability of ECM protein synthesis, including fibrinogen and collagen.<sup>78</sup>

In comparison to other MNPs, CuNPs exhibit the ability to promote angiogenesis, reduce wound susceptibility, and significantly enhance diabetic wound healing. Unlike AuNPs and AgNPs, which facilitate wound healing by eradicating bacteria, CuNPs not only possess antibacterial properties but also directly stimulate VEGF expression to expedite wound healing. The utilization of Cu-based MOFs as drug delivery carriers or photothermal therapy for augmenting the antibacterial activity of CuNPs can expand their application to promote wound healing.

## ZnNPs

Zinc oxide nanoparticles (ZnONPs) influence diabetic wound healing by at least two mechanisms. They exert hypoglycemic effects and demonstrate the same antibacterial, antioxidant, and anti-inflammatory effects as other MNPs.

Zn promotes liver sugar production by acting on the insulin signaling pathway, thereby enhancing glucose utilization. Furthermore, it inhibits intestinal glucose absorption, thus reducing blood glucose and inhibiting glucagon secretion,<sup>123</sup> reducing gluconeogenesis and glycogenolysis. Umrani et al developed a drug containing ZnONPs that had a significant antidiabetic effect, improving glucose tolerance, increasing serum insulin, and lowering blood glucose.<sup>124</sup> Research has indicated that the promotion of wound angiogenesis, re-epithelialization, and keratinocyte migration can be facilitated by Zn.<sup>90</sup>

Yadav et al created ZnONPs via a green synthesis method using *Portulaca Oleracea* extract and studied their effect on wound healing. ZnONPs showed strong antioxidant and anti-inflammatory activities in vitro. Because of the reduced oxidative stress and inflammatory response, the topical administration of ZnONPs to damaged dermal tissue promoted the rapid synthesis and deposition of collagen fibers, as well as the restoration of tissue granulation and epithelial lining, resulting in faster wound closure and healing.<sup>125</sup> The size of ZnONPs may affect their antibacterial activity<sup>84</sup> by altering their ability to disrupt the bacterial cell membrane and promote oxidative stress in bacteria.<sup>126</sup> Synthesized ZnONPs (with an average particle diameter of approximately 12 nm) inhibit 100% of bacterial growth at a concentration of  $1.0 \times 10^{-2}$  to



$3.0 \times 10^{-3}$  M. This is a result of the *E. coli* membrane destruction, causing an increase in permeability and resulting in the accumulation of NPs within both the bacterial membrane and cytoplasmic regions.<sup>84</sup> ZnONPs also inhibit and inactivate *Campylobacter jejuni* cell growth, possibly via the destruction of the *Campylobacter* cell membrane and oxidative stress.<sup>85</sup> The ZnONPs-loaded sodium alginate-gum acacia hydrogels (SAGA-ZnONPs), formed by crosslinking the hydroxyl groups of sodium alginate and gum acacia polymers with glutaraldehyde's aldehyde group, exhibit potent antibacterial activity and facilitate fibroblast migration at the appropriate concentrations.<sup>127</sup>

ZnONPs may also demonstrate anti-inflammatory properties during infection or after chemical exposure.<sup>86</sup> They exert antibacterial activity via adsorption-induced membrane damage and ROS-mediated cytotoxicity.<sup>128</sup> The anti-inflammatory activities of ZnONPs include decreasing mast cell proliferation and LPS-induced COX-2 production and blocking the expression of proinflammatory factors. ZnONPs also inhibit caspase-1, an IL-1 converting enzyme that promotes mast cell activity.<sup>87</sup> The generation of malondialdehyde, a well-known indicator of oxidative stress, is also inhibited by ZnONPs in a dose-dependent manner.<sup>86</sup> ZnNPs inhibit mast cell proliferation by regulating p53 protein levels. Prolonged p53 activation is demonstrated to induce tumorigenic inflammation through the release of high-mobility histone 1.<sup>88</sup> ZnONPs exhibit a dose-dependent inhibition of COX-2 activation in LPS-treated macrophages. LPS promotes the release of prostaglandin E2 (PGE2), a promoter of inflammation, by activating COX-2 gene expression. However, ZnONPs demonstrate no direct effect on PGE2 expression. Likewise, ZnONPs effectively suppress the production of macrophage-derived nitric oxide (NO) by downregulating the expression of inducible nitric oxide synthase, thus mitigating the potentially harmful effects of inflammatory responses on cells. In a study conducted by Nagajyothiet et al, ZnONPs were synthesized using the root extract of *Paeonia tenuifolia*, revealing that ZnONPs not only enhanced the generation of oxygen free radicals but also inhibited the expression of proinflammatory factors. These findings indicate that ZnONPs possess remarkable anti-inflammatory and antioxidant properties, thereby promoting the cellular activities associated with wound healing.<sup>128</sup>

Compared to other types of MNPs, ZnNPs not only effectively eliminate bacteria to facilitate wound healing but also expedite the inflammatory phase by inhibiting the expression of pro-inflammatory factors, thus accelerating the overall wound-healing process. Moreover, ZnNPs also play a vital role in biomaterial synthesis. Qian et al engineered a hydrogel using glycyrrhizic acid (GA) and induced the self-assembly of ZnNPs, combined with photo-crosslinked methyl acrylated silk fibroin (SF) to create a promising biomaterial.<sup>89</sup> This hybrid hydrogel enhances mechanical properties and injectability, and is notable for its non-cytotoxic formulation achieved by incorporating ZnNPs. The addition of ZnNPs further allows for leveraging their anti-inflammatory properties to regulate macrophage polarization. Hu et al via a carrier-based approach, utilized a zinc-based nanoscale metal-organic framework (NMOF) to develop an ROS-responsive hydrogel. This hydrogel encapsulated BR@Zn-BTB nanoparticles, resulting in an improved drug loading capacity and controlled release of both the drug and  $\text{Zn}^{2+}$ .<sup>59</sup> This approach synergistically enhances the antibacterial and anti-inflammatory effects of ZnNPs and BR.

Based on the studies mentioned above, the incorporation of ZnNPs not only enhances the safety and immunomodulatory functions of biomaterials but also promotes wound healing through synergistic drug action. Thus, ZnNPs promote diabetic wound healing. In addition, because of the special role of Zn in the insulin pathway, ZnNPs can be used to change the blood glucose environment at the site of diabetes to promote wound healing. This is a method that should be developed in future research.

## TiNPs

The photocatalytic properties of TiNPs, specifically  $\text{TiO}_2$ , are responsible for its antibacterial effects. When exposed to ultraviolet light,  $\text{TiO}_2$  generates ROS that effectively eliminates 100% of microorganisms. Additionally, the growth and viability of bacteria are influenced by positively charged nanomaterials through alterations in their permeability. The bactericidal action is further facilitated by the highly reactive ROS produced during the photocatalytic processes involving  $\text{TiO}_2$ .<sup>80</sup>  $\text{nTiO}_2$ /PU nanocomposite films produce hydroxyl radicals when exposed to the photocatalytic activity of sunlight. Hydroxyl free radicals kill bacteria via CoA reduction, but  $\text{nTiO}_2$ /PU nanocomposite films do not show antibacterial activity in the absence of illumination.<sup>81</sup> The investigation of nanocatalysts represents a novel and promising research area in the field of wound healing and remodeling. Lopez-goerne et al synthesized  $\text{TiO}_2$ - $\text{SiO}_2$  submicron

mesoporous particles with varying sizes and shapes via the sol-gel method, which contained ultras mall platinum nanoparticles possessing catalytic properties. These nanoparticles stimulated althy cell proliferation while simultaneously combating multiple bacterial infections. In addition, the catalytic nanoparticles speed up wound healing, which could reduce the risk of delayed diabetic wound healing leading to amputation.<sup>129</sup>

Upon encountering foreign bacteria or pathogens, Toll-like receptors (TLRs) present on the surface of titanium dioxide nanoparticle (TiO<sub>2</sub>NP) platelets are triggered. TLR stimulation enhanced the proinflammatory response of platelets and the expression of p-selectin.<sup>82</sup> PSG1 (P-selectin glycol protein) is produced by neutrophils and monocytes and interacts with p-selectin to create O<sub>2</sub> (a reactive oxygen species) in macrophages, resulting in oxidative stress. Studies reveal that blood platelet counts decrease after incubation with TiO<sub>2</sub>NPs.<sup>82,83</sup> In contrast to other MNPs, the antibacterial action of TiNPs is via photocatalysis and promotes the proinflammatory response of platelets to kill bacteria. TiNPs may therefore have low biotoxicity in normal cells. Thus, TiNPs should be more widely used in the study of promoting wound healing.

## FeNPs

The antibacterial mechanism of iron oxide nanoparticles (FeONPs) may be related to the ROS production induced by FeO. The vitamin B2-functionalized FeO nanozyme trio functions to decrease bacterial viability while also scavenging ROS to protect cells from oxidative damage.<sup>92</sup> These properties are extremely beneficial in controlling bacterial infections during wound healing. Arakha et al investigated the impact of the FeONP-bacterial interface interaction mode on antimicrobial susceptibility to FeONPs, and found that chitosan-coated FeONPs exhibited significantly enhanced antibacterial activity. It was also revealed that the FeONP-chitosan coating enhanced ROS production at the interface, thus providing antibacterial activity.<sup>91</sup>

FeONPs have also demonstrated magnetic and catalytic capabilities.<sup>130</sup> Exosomes derived from stem cells have been identified as potent mediators that enhance wound healing by promoting angiogenesis and re-epithelialization through the modulation of miR-21-3p.<sup>131</sup> Moreover, MNPs such as Fe<sub>3</sub>O<sub>4</sub> nanoparticles and Fe<sub>2</sub>O<sub>3</sub> nanoparticles are magnetic, exerting a weak magnetic force on cells under a static magnetic field and promoting tissue regeneration.<sup>132</sup> Wu et al used mesenchymal stem cells from bone marrow to create novel exosomes and confirmed that the amplification of miR-21-5p increased angiogenesis and glial cell migration in the presence of Fe<sub>3</sub>O<sub>4</sub> nanoparticles and a static magnetic field.<sup>133</sup>

## CeO<sub>2</sub>NPs

Cesium oxide NPs (CeO<sub>2</sub>NPs) has the same broad-spectrum antibacterial activity as other MNPs. Positively charged CeO<sub>2</sub>NPs adsorb well to negatively charged bacterial cells and penetrates into them and inactivates bacterial enzymes, resulting in the production of ROS. ROS further damages nucleic acids and proteins, eventually leading to bacterial cell death.<sup>134</sup> When encountering polysaccharide-encapsulated bacteria, CeO<sub>2</sub>NPs interacts with the extracellular bacterial environment and produces ROS, which directly destroys the cell wall and enters the cell to inactivate nucleic acids and proteins for bactericidal purposes.<sup>135</sup>

CeO<sub>2</sub>NPs are known as free radical scavengers.<sup>93</sup> Ghaznavi et al discovered that CeO<sub>2</sub>NPs protect against the oxidative stress caused by high glucose levels.<sup>136</sup> Chigurupati found that a local application of water-soluble CeO<sub>2</sub>NPs increased the proliferation and migration of fibroblasts and keratinocytes in mice, expediting entire cutaneous wound healing. Water-soluble CeO<sub>2</sub>NPs also infiltrated the wound tissue and reduced oxidative damage to cell membrane proteins.<sup>94</sup> Hirst et al revealed that CeO<sub>2</sub>NPs scoured ROS and inhibited the production of inducible nitric oxide synthase (iNOS), an inflammatory mediator, in J774A.1 mouse macrophages.<sup>137</sup> We speculate that the antioxidant mechanism of CeO<sub>2</sub>NPs occurs because the nanoparticles act as direct antioxidants, limiting ROS production, or that these nanoparticles may directly reduce ROS production levels.

Similar to CuNPs, CeO<sub>2</sub>NPs can also induce the formation of blood vessels in wounds. Das et al demonstrated that the small size of CeO<sub>2</sub>NPs and their higher surface concentration of Ce<sup>3+</sup>/Ce<sup>4+</sup> promoted the development of tubular structures in endothelial cells. They also observed that CeO<sub>2</sub>NPs decreased intracellular oxygen levels while temporarily stimulating HIF-1 $\alpha$  to translocate into the nucleus and up-regulate the expression of angiogenic proteins.<sup>94</sup> Augustine et al developed electrospun poly (3-hydroxybutyrate co-3-hydroxyvalerate) PHBV membranes containing different

concentrations of CeO<sub>2</sub>NPs (0.5%, 1%, 2%, or 4%w/w) for the treatment of diabetic wounds. They found that CeO<sub>2</sub>NPs in PHBV/nCeO<sub>2</sub> significantly increased angiogenesis and facilitated keratinocyte migration.<sup>138</sup>

## Adverse Effects of MNPs

One challenge remaining in MNP research is biotoxicity. If the concentration of metal ions is too high or there is a sudden release of many metal ions, there will be high cytotoxicity and irreversible damage to the liver and kidney. During the treatment of diabetic wounds, long-term skin contact may cause MNPs to enter the blood circulation and eventually deposit in the liver and kidney organs. A large accumulation of MNPs can trigger the inflammatory response and oxidative stress of normal liver and kidney cells, leading to cell death.<sup>139,140</sup> Chen et al observed elevated serum aspartate aminotransferase (AST) and alanine transaminase (ALT) levels in mice treated with TiO<sub>2</sub>NPs at high doses, leading to liver cell necrosis, apoptosis, and liver fibrosis.<sup>141</sup> The serum ALT level in mice treated with ZnNPs increased significantly, indicating liver necrosis.<sup>142</sup> Jain et al reported that 24 hours after injecting FeO into male Sprague–Dawley rats, serum AST levels significantly increased, leading to adverse liver reactions after acute exposure.<sup>143</sup> Nosrati et al identified a potential nephrotoxic risk following repeated oral administration of AgNPs in mice.<sup>144</sup> Subsequent staining of the kidneys from mice exposed to varying doses of AgNPs for 28 consecutive days revealed infiltration of inflammation into the capillaries within the kidney tubules, which was accompanied by an increased expression of genes associated with apoptosis in renal interstitial and epithelial cells. These findings suggest that the accumulation of high levels of AgNPs may induce inflammatory reactions with normal cells, leading to nephrotoxicity. This underscores the importance of strict dosage control when utilizing MNPs. There are also many studies evaluating the toxicity of CeO<sub>2</sub> NPs. These studies reveal that long-term exposure to high concentrations of CeO<sub>2</sub>NPs resulted in genetic damage (DNA damage to white blood cells and hepatocytes) and biochemical changes (changes in serum lactate dehydrogenase and reduced glutathione levels in the brain) in rats.<sup>145,146</sup>

## Application of Metal-Based Nanoparticles to Biomaterials

As highlighted earlier, the delayed healing of diabetic wounds is attributed to the multiple stages involved in wound healing. Therefore, a dressing for diabetic wound healing needs to fulfill multiple functions simultaneously and prevent excessive accumulation of MNPs. Numerous studies have reported that hydrogels and microneedles can achieve controlled and sustained release of drugs and MNPs, thereby reducing the concentration of metal ions at the wound site. Furthermore, we have comprehensively summarized the utilization of MNPs as nanocarriers for the treatment of diabetic wounds. MNPs exhibit the ability to encapsulate drugs and synergistically collaborate with therapeutic agents, thereby enabling precise drug delivery and multifunctionality in wound dressings.

## Dressings Containing Metal-Based Nanomaterials

### Hydrogels

Hydrogels, which are formed by the physical or chemical cross-linking of 3D polymer networks, exhibit high absorptivity and adjustable properties, rendering them optimal carriers for drug delivery systems.<sup>147</sup> By modifying the properties of hydrogels, the rate of drug release can be effectively controlled, enabling the release of MNPs at non-cytotoxic concentrations. Hydrogels are also one of the most commonly used dressings for wound healing. The cytotoxicity caused by the sudden release of MNPs is a major challenge in the treatment of diabetic wounds with MNPs. To control the sudden release of MNPs, which leads to the increase of local MNP concentration and cell poisoning death, we use the slow degradation characteristics of hydrogels to MNP achieve the effect of slow release.

The aforementioned p(NIPAM-co-hemin)/ALG-EDA hydrogel,<sup>100</sup> AgNP@CNDM hydrogel,<sup>97</sup> the SF/GA/Zn hybrid hydrogel<sup>89</sup> and chitosan/Ca-AlgNP/AgNP hydrogel<sup>98</sup> demonstrate the capability of sustained release, which ensures a suitable concentration of metal ions for bactericidal activity in wounds. Simultaneously, these hydrogels exhibit an enhanced mechanical strength and adhesion rate, enabling optimal wound coverage and continuous moisturization to promote wound healing. Moreover, these hydrogels not only facilitate the controlled release of metal ions but also safeguard the structural integrity of MNPs. The discussions on HKUST-1 NPs<sup>113</sup> and BR@Zn-BTB/Gel<sup>59</sup> reveal that the hydrogel not only preserves the structural integrity of MNPs but also facilitates MOFs to accommodate more drugs,

thereby improving the drug loading efficiency and enabling the controlled release of metal ions. Furthermore, the hydrogel enhances the therapeutic efficacy of diabetic wounds treated with MNPs and drugs.

### Microneedles

Diabetic wounds are susceptible to infection, and the prolonged infectious state facilitates bacterial proliferation and the formation of bacterial biofilms within the wound.<sup>148</sup> Bacterial biofilms are self-synthesizing substrates of hydrated extracellular polymeric substances (EPS) produced by bacteria.<sup>149</sup> The presence of biofilm hinders the efficacy of conventional antibacterial methods, such as gels, fiber membranes, and creams, to eradicate bacteria residing within the biofilm.<sup>150</sup> Therefore, we chose to take advantage of a new penetrating drug delivery method, namely the physical penetration of biofilms, to deliver antibacterial agents to the bacterial community to achieve the bactericidal effect.<sup>151</sup>

Bacteria-sensitive nanoparticles (NPs) and doxycycline (DOX) loaded in soluble microneedles (MNs) have been used to treat wound infections.<sup>152</sup> The MNs penetrate the biofilm, and as the MNs dissolve, the internal NPs and DOX are released. Inspired by this, other researchers have combined the antibacterial properties of MNPs with microneedles, using microneedles to precisely deliver MNPs to wound infections. For instance, Permana et al synthesized AgNPs using chitosan-modified green tea extract and assembled them into soluble microneedles. Compared with the control cream containing AgNPs, the number of bacteria in the infected wound was significantly reduced, which effectively achieved the purpose of sterilization and promoting wound recovery.<sup>148</sup> Yang et al utilized hyaluronic acid (HA) microneedles incorporating cerium/zinc-based nanomaterials (ZCO) for the treatment of diabetic wounds. Upon penetration of the biofilm, the  $\text{Zn}^{2+}$  and  $\text{Ce}^{3+/4+}$  ions within the microneedle are gradually released through the action of hyaluronidase in the wound, thereby exerting bactericidal effects while also exhibiting scavenging properties against ROS. This dual functionality effectively mitigates inflammatory reactions within the wound, facilitating wound remodeling and cellular proliferation, ultimately leading to a significantly accelerated healing rate in diabetic wounds.<sup>153</sup> Zeng et al similarly used HA microneedles as a tool for penetrating infected wound biofilms, which were loaded with dimethylglyoxylglycine (DMOG) encapsulated in PCN-224 MOF nanoparticles (DMOG@PCN-224 NPs) and co-loaded with antibiotic meropenem (MEM). The laser-activated PCN-224 NPs (Zr6) converted  $\text{O}_2$  to a single oxygen state ( $^1\text{O}_2$ ) and acted together with MEM to sterilize the bacteria, greatly reducing the antibiotic dose without reducing the bacterial killing effect. Over time, DMOG@PCN-224 NPs gradually degrade to release DMOG, which competitively inhibits prolyl hydroxylase and promotes wound tissue remodeling and angiogenesis.<sup>154</sup> The use of microneedles might increase the penetration of MNPs into the wound. Due to the safety of microneedles, there is no need to worry about the biotoxicity of MNPs to the user.<sup>151</sup>

### Biomaterials for MNPs Combined Drugs

The remarkable attributes of high surface area, photothermal responsiveness, and versatility enable the physical or chemical cross-linking of MNPs with other biomaterials to form drug-delivery carriers.<sup>155</sup> MNPs deliver the drug to the wound after loading the drug, and play a synergistic role with the drug to promote wound healing when the drug is released. MNPs are useful for the delivery of drugs, small molecular proteins, nucleic acids, etc.<sup>156</sup>

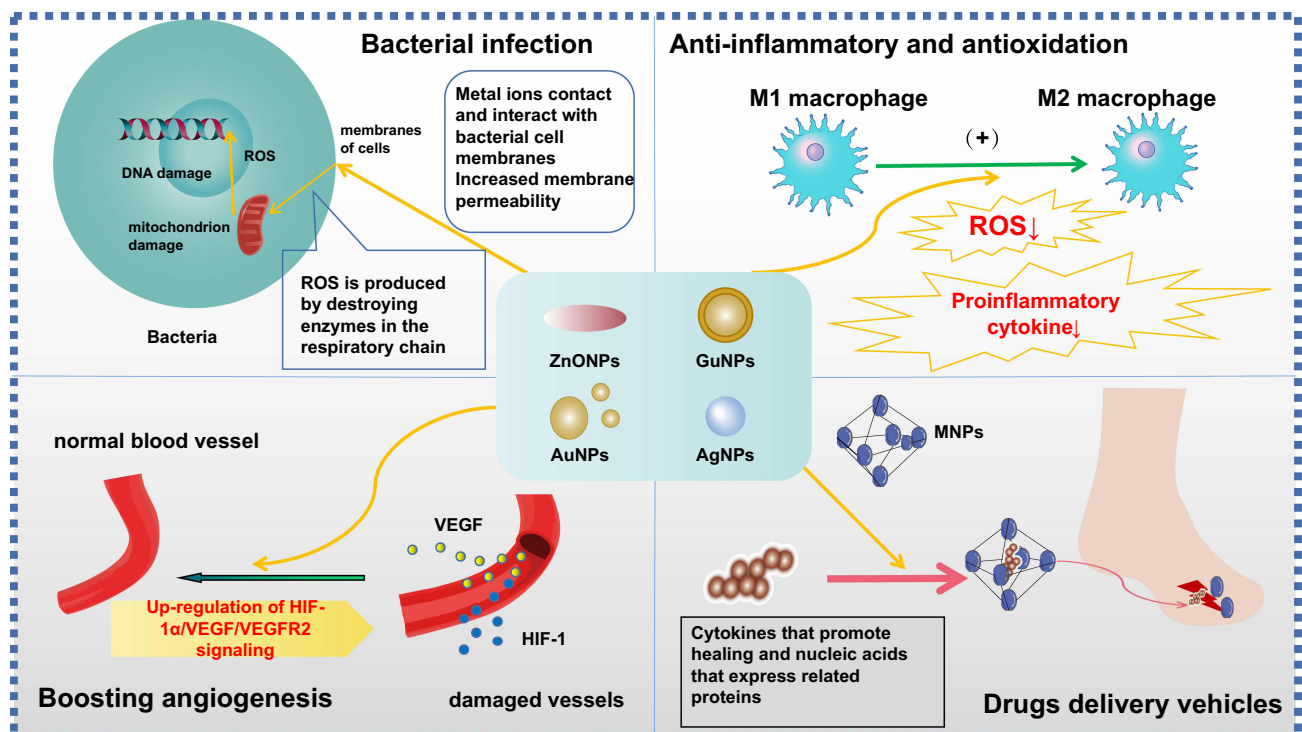
The growth factor KGF was loaded with AuNPs as a carrier to produce KGF-GNPs nanoparticles,<sup>61</sup> which enabled KGF to maintain biological activity under a harsh environment and significantly accelerated the reepithelialization of diabetic wounds. AuNPs@LL37 binds to plasmid DNA to form AuNPs@LL37/pDNAs,<sup>62</sup> and VEGF is successfully transfected, while AuNPs still play the role of the antibacterial agent. More typical are MOF applications, such as NO@HKUST-1,<sup>77,110</sup> ADM-B-HKUST-1,<sup>116</sup> and BR@Zn-BTB.<sup>59</sup> By modifying the surface of MOFs, large pores in MOFs can be loaded with NO, drugs and even living cells. MOFs serve as a carrier platform to deliver these agents to the corresponding location in the wound and release the agent in a controlled manner to enhance the healing ability of the wound. Yu et al constructed a nanocarrier using Haase and 3-aminophenylboronic acid (APBA), which can be loaded with gentamicin (Gen) to form HB/Ag/g nanoparticles after the addition of AgNPs. AgNPs and Gen are simultaneously released at the wound site, acting synergistically on the infected wound. The simultaneous use of AgNPs and Gen not only reduces the amount of AgNPs but also enhances the antibacterial activity of this nanoparticle.<sup>157</sup>

# The Mechanism Underlying the Diabetic Wound Healing Ability of MNPs

MNPs have several therapeutic effects on diabetic wounds, including promoting wound angiogenesis and exhibiting antibacterial, anti-inflammatory, and anti-oxidative properties. Numerous studies have demonstrated the exceptional performance of metal-based nanomaterials in these aspects through various means (Table 1) (Figure 9).

## Antioxidant Activity

Bacteria can cause wound infections and delay healing by increasing proinflammatory cytokines.<sup>16,158</sup> MNPs can come in contact with bacteria and cause death.<sup>74,159</sup> MNPs, in particular, have advantageous physicochemical properties that result in high antibacterial activity.<sup>48</sup> According to the above, we summarized the antibacterial function of biomaterials containing MNPs. Examples include HPAMAM-N(CH<sub>3</sub>)<sub>2</sub>/AuNP,<sup>69</sup> vancomycin AuChi-liposome,<sup>56</sup> chitosan/Ca-AlgNP /AgNP hydrogel,<sup>99</sup> Ag/PAAc nanogel,<sup>160</sup> GLCN-CuNPs,<sup>75</sup> CuONPs,<sup>107</sup> ZnONPs,<sup>84</sup> SAGA-ZnONPs,<sup>127</sup> nTiO<sub>2</sub>/PU nanocomposite films,<sup>129</sup> and FeONPs.<sup>92</sup> These contain MNPs that interact with bacterial cell walls, significantly increasing the permeability of bacterial cell membranes. Eventually, the bacteria die because they cannot regulate membrane transport properly.<sup>161</sup> Some researchers have innovatively applied photothermal therapy (PTT) to effectively eliminate bacteria and promote wound healing.<sup>120</sup> He et al, utilized polyhexamethylenediamine (PHMB) to modify AuNPs. In the near-infrared light, the formation of bacterial biofilm was synergistically inhibited, and the bacteria were quickly removed to heal the wound.<sup>59</sup> Molybdenum bronze nanoparticles (SMB NPs) with sonodynamic therapeutic effects were used in combination with a second near-infrared (NIR-II) photothermal effect.<sup>162</sup> NIR-II stimulation of SMB NPs produced an extremely strong oxidase mimetic activity and photothermal conversion efficiency, and promoted bacteria to produce excessive reactive oxygen species to eliminate bacteria in deep infected wounds. The same monolayer HE MXenes prepared from transition metals with high entropy and low Gibbs free energy showed extremely strong bactericidal effects and minimal side effects after irradiation through NIR-II.<sup>163</sup> The implementation of external stimuli, such as light, heat, and sound, to modify the microstructure of MNPs significantly amplifies their bactericidal effect. These remarkable



**Figure 9** The main therapeutic methods of MNPs for diabetic wounds are depicted in a mechanism diagram. Inclusion (bacterial infection, anti-inflammatory and antioxidant effects, angiogenesis promotion, drug carrier delivery).



advancements have unveiled the extensive clinical potential of MNPs in combating drug-resistant bacterial infections and facilitating the healing process of infected wounds.

## Anti-Inflammatory Mechanism

Some MNPs enhance the transition of the inflammatory phase by promoting macrophages to M2 polarization and eliminating the ROS production induced by high glucose environment. Some even inhibit cellular pro-inflammatory factor expression, reducing inflammation and accelerating wound healing.<sup>164</sup> Because of their small size, MNPs easily enter the blood, interact with proteins in the plasma, and are surrounded by these cellular elements.<sup>165</sup> Some small MNPs enter the cell through pores or ion channels in the cell membrane and activate M2-type macrophages.<sup>75</sup> At higher concentrations, the smallest MNPs are easily entoxified by most cell vesicles.<sup>166</sup> M2 macrophages limit the inflammatory response and promote the transition from the inflammatory phase to the proliferative phase in diabetic wounds. We found that reducing ROS production and blocking proinflammatory cytokine production were the main mechanisms by which almost all MNPs inhibited inflammation and accelerated wound healing. MNPs block the build-up of inflammation in cells. For example, AuNPs,<sup>63</sup> FeONPs,<sup>92</sup> Y<sub>2</sub>ONPs and CeO<sub>2</sub>NPs<sup>93,136</sup> enhance the anti-inflammatory and antioxidant capacity of diabetic wounds by reducing ROS production and blocking the production of proinflammatory cytokines.

## An Angiogenesis Promotion

MNPs induce up-regulation of pro-angiogenic growth factors in diabetic wounds, thereby stimulating cell proliferation. Due to the dysregulated release of cytokines, including EGF, VEGF, TNF- $\alpha$ , and IL-6, in individuals with diabetes, angiogenesis and cell migration are hindered. Several MNPs have demonstrated remarkable efficacy in promoting angiogenesis, a critical process for enhancing wound healing in diabetic patients. For instance, ZnONPs induce endothelial cell migration by producing NO via the MAPK/Akt/eNOS pathway.<sup>90,167</sup> AuNPs,<sup>168</sup> CuONPs<sup>122</sup> and HKUST-1NPs<sup>113</sup> release metal ions to stimulate the production of angiogenic related cytokines. The surface valence of CeO<sub>2</sub>NPs stimulates endothelial cells to form tubular structures and also promotes angiogenesis by stabilizing the expression of HIF-1 $\alpha$  in endothelial cells.<sup>168</sup> In addition to angiogenesis, MNPs, such as KGF-GNPs,<sup>60</sup> AuNPs@LL37,<sup>62</sup> and F-HKUST-1,<sup>115</sup> may also promote diabetic wound healing by carrying cytokines or genes that stimulate cell migration and collagen deposition.

Recently, the focus on promoting diabetic wound healing has shifted towards enhancing angiogenesis in these wounds, coupled with antibacterial, anti-inflammatory, and anti-oxidation properties. However, the research on metal-based nanomaterials has surpassed the exploration of individual MNPs' functions. Presently, researchers are delving into various strategies to functionalize MNPs, cross-link them with biomaterials, employ external stimuli (eg, light, magnetic, ultrasound) to activate MNPs, or combine them with bimetallic nanoparticles. Consequently, the application of MNPs in the treatment of diabetic wounds extends beyond the enhancement of their antibacterial properties. Numerous studies have significantly advanced our understanding of the antioxidant and angiogenesis functions of metal-based nanomaterials. Additionally, other unexplored or challenging MNPs, such as Pd or cerium nanoparticles, are being investigated for their remarkable healing effects in the treatment of diabetic wounds.

## Clinical Application of MNPs

Current treatments for chronic nonhealing wounds, such as debridement, negative-pressure wound therapy, and skin grafting, are often costly, burdensome, and associated with discomfort. In contrast, dressings containing MNPs provide a superior and more convenient solution for promoting wound healing compared to traditional treatments.

Currently, multiple MNPs-containing dressings utilized in clinical treatment have undergone clinical validation. Below, we discuss and summarize some of the dressings in clinical trials and already on the market (Table 2). Mehdi Teimouri conducted clinical trials on the use of an Agicoat silver Nano-Crystalline Dressing for wound healing caused by fractures. Patients treated with this dressing experienced less pain and a significantly shorter wound healing time compared to those treated with Gaz Vaseline.<sup>169</sup> In a clinical trial assessing the efficacy of CuO-impregnated foam polymer dressings in preventing postoperative wound infection, patients using the Cu dressings had significantly lower rates of wound infection compared to those using non-Cu dressings.<sup>170</sup> In the controlled clinical trial using the SilvrSTAT Gel, a significant reduction in wound size was observed after

**Table 2** Clinical Application and Clinical Trials of Metal in Wound Healing

Clinical Application/ Clinical Trials	Product	Description	Clinical Uses	Mechanism of Action/Promotes Burn Wound Healing	Ref.
Clinical trials	CuO impregnated foam polymer dressings	Wound dressing impregnated with CuO particles	Preventing postoperative wound infection	Reduce the infection of resistant bacteria	[170]
Clinical trials	Agicoat	Agicoat silver Nano-Crystalline Dressing	Wound healing caused by fractures	A significant effect on wound healing and cell proliferation, decreased pain	[169]
Clinical trials	SilvrSTAT	Gel containing AgNPs	Promotes burn wound healing	After 2 weeks of use and almost complete healing was achieved after 8 weeks	[171]
Clinical application	Acticoat™	Nanocrystalline Ag wound dressing	Burn Venous ulceration Diabetic ulcer	When the dressing is wetted with sterile water or wound exudate, it activates the release of silver ions into the solution, thereby providing antibacterial activity within the dressing	[172]
Clinical application	MedCu Technologies	Wound dressing impregnated with CuO particles	Diabetic wounds Leg and foot ulcers Pressure ulcers Burns and Surgical wounds	Increased blood vessel formation, Angiogenic factor gene and in situ upregulation	[174]
Clinical application	Biatain® Ag	Ag alginate dressing	Fill the cavity Absorb and lock the seepage Continuous broad-spectrum antibacterial Promote Coagulation	Silver particles can directly enter the thallus and bind with oxygen metabolism enzyme (-SH), so that the thallus suffocates and dies	[175]
Clinical application	AQUACEL® Ag	Hydrophilic fiber Ag dressing	Form an antimicrobial barrier to protect the wound bed Absorb a lot of wound seepage and bacteria Helps remove nonviable tissue from the wound	Hydrophilic fibers absorb and lock bacteria in the seepage; Silver ions in the fibers are activated by exudation to kill bacteria in the dressing	[176]

2 weeks of use and almost complete healing was achieved after 8 weeks.<sup>171</sup> This study effectively demonstrated the beneficial impact of MNPs in the process of wound healing in diabetic patients. The clinical application of commercially available dressings containing MNPs has also been reported. Lee et al conducted a comparative study on the application of various dressings for wounds infected with methicillin-resistant *Staphylococcus aureus* (MRSA). Acticoat™. The dressings released Ag<sup>+</sup> for a longer period of time and caused less inflammation at the wound site. As a result, the frequency of dressing change is greatly reduced and wound healing is accelerated.<sup>172</sup> Acticoat™ on diabetic wounds in a randomized clinical trial effectively promoted wound healing without any observed adverse effects, thereby demonstrating the efficacy and safety of Acticoat™ for treating diabetic wounds.<sup>173</sup> In addition, MedCu Technologies is a sterile wound dressing impregnated internally with CuO particles. In studies of noninfected diabetic wounds, this dressing reduced inflammation at the wound site and stimulate wound angiogenesis and epithelial formation.<sup>174</sup> Moreover, both Biatain® Ag<sup>175</sup> and AQUACEL® Ag<sup>176</sup> effectively killed bacteria to protect wounds from infection and reduce patient pain during dressing changes, which is beneficial for wound prognosis and patient experience.

At the same time, we also summarized the latest patents on dressings containing MNPs (Table 3). The AMP and CeONs were encapsulated in GelMA-DOPA hydrogels by Cheng et al. The synergistic antibacterial activity of AMP and CeONs was harnessed, while the CeONs effectively eliminated ROS generated during wound inflammation upon release. This dual action resulted in bacterial eradication and suppression of inflammation within the wound.<sup>177</sup> Similarly, a multi-enzyme-like activity exhibiting multifunctional hydrogel made up of mussel-inspired carbon dot reduced-Ag (CDs/AgNPs) and Cu/Fe-nitrogen-doped carbon (Cu,Fe-NC) was designed. This hydrogel promotes wound healing by utilizing the synergistic antibacterial properties of Cu and AgNPs and nanozyme in the hydrogel to remove ROS.<sup>178</sup> Au-AgNCs@ GSH and Cu-Ag nanocluster composite hydrogel are the latest designed dressings. Their common feature is the use of double MNPs to achieve the effect of antibacterial activity enhancement.

**Table 3** Patents on Dressings Containing MNPs

Title of Invention	Publication Number	Mechanism of Action
A spray hydrogel multifunctional wound dressing	CN114569785A	The sprayable hydrogels loaded with CeO <sub>2</sub> NPs and antimicrobial peptides have broad-spectrum antibacterial and reactive oxygen scavenging properties
Preparation method and application of nanoenzyme antibacterial hydrogel	CN114129766B	Quasi-peroxidase activity and Cu,Fe-N-C nanoenzymes can produce ROS to kill bacteria
A method for preparing Cu-Ag nanocluster composite hydrogel	CN107375993A	The hydrogel is stable and moisturizing, and the combination of CuNPs and AgNPs enhances the antibacterial activity
A rare earth metal-based dressing for wound surface sterilization and a preparation method thereof	CN116251223A	The dressing contained rare earth Eu loaded Au-AgNCs@ GSH, the MNPs were evenly dispersed on the surface of the particles and <i>E. coli</i> and <i>S. aureus</i> had strong inhibitory and killing effects.

Clinical trials and practical applications demonstrate the effectiveness of MNP-containing dressings in absorbing wound secretions and releasing metal ions for antimicrobial therapy. No accumulation of MNPs was found in the wound. These findings provide strong evidence for the safety and efficacy of these dressings in practical applications. By promoting wound healing and reducing patient pain, these dressings play a vital role in improving patient outcomes.

## Challenges and Future Prospects

In summary, our findings demonstrate that MNPs possess antibacterial, antioxidant, anti-inflammatory, and pro-angiogenic activities, thereby accelerating the healing process of diabetic wounds. However, it is important to note that MNPs are easily released into body fluids in large quantities, leading to direct tissue damage caused by a metal ion attack. This presents a significant challenge for the application of MNPs. Through further research and improvement in the preparation technology of MNPs, we aim to achieve non-biologically toxic concentrations in vivo. These advancements will enable the safe and efficient release of MNPs into the body while maximizing their precise effects. Nevertheless, it should be acknowledged that long-term accumulation of MNPs in vivo may still result in a certain degree of cytotoxicity. Therefore, we are faced with the challenge of enhancing the therapeutic efficacy of MNPs using current technologies while simultaneously expediting their elimination from the body and maintaining a balance between the therapeutic benefits and production costs.<sup>166,172,174</sup>

Therefore, in the future, it is imperative to not only expand the application range of MNPs but also streamline and optimize their production and utilization in biomaterials. Additionally, exploring novel MNPs with multifaceted effects should be a prospective avenue for research. Furthermore, synergistic combinations of multiple MNPs or integrating MNPs with diverse biomaterials can be employed at different stages of diabetic wound healing to ensure enhanced functionality while minimizing potential complications.

## Conclusion

Compared to normal wounds, the complex pathophysiological environment of diabetic wounds presents greater challenges to the healing process. Here, we not only provide a comprehensive overview of the roles played by different types of MNPs in diabetic wound healing but also explore investigations into their significant effects on the intricate mechanisms involved, with a particular focus on their anti-inflammatory, anti-oxidative, anti-bacterial, and pro-angiogenic properties. Additionally, considering the high surface area and adsorbability of MNPs, we summarize their potential as drug carriers for efficient delivery and their synergistic combination with other biomaterials. The utilization of MNPs not only enhances biocompatibility but also ensures sustained and safe therapeutic effects on diabetic wounds through controlled release mechanisms. These innovative investigations deepen our understanding of the potential applications of MNPs in treating diabetic wounds while providing novel research directions for exploring therapeutic mechanisms involving these nanoparticles.

In the future, with the synthesis of an increasing number of MNPs and their integration with biomaterials in diabetic wound research, not only will the healing rate of diabetic wounds be enhanced, but also more MNPs-containing bio-materials will transition from research to clinical application, thereby illuminating the healing process of diabetic wounds.

## Funding

This work was supported by the Natural Science Foundation of China (82072051), Central Finance supports Local Colleges and Universities Talent Development Funding from Heilongjiang Provincial Department of Finance (2020GSP09), and Torch Program of Mudanjiang Medical College Science Foundation (2022-MYHJ-006).

## Disclosure

The authors report no conflicts of interest in this work.

## References

1. Ezhilarasu H, Vishalli D, Dheen ST, Bay BH, Srinivasan DK. Nanoparticle-based therapeutic approach for diabetic wound healing. *Nanomaterials (Basel)*. 2020;10(6):E1234. doi:10.3390/nano10061234
2. Vijayakumar V, Samal SK, Mohanty S, Nayak SK. Recent advancements in biopolymer and metal nanoparticle-based materials in diabetic wound healing management. *Int J Biol Macromol*. 2019;122:137–148. doi:10.1016/j.ijbiomac.2018.10.120
3. He K, Hemmila MR, Cain-Nielsen AH, Machado-Aranda DA, Frydrych LM, Delano MJ. Complications and resource utilization in trauma patients with diabetes. *PLoS One*. 2019;14(8):e0221414. doi:10.1371/journal.pone.0221414
4. Miyajima K, Hui Teoh S, Yamashiro H, et al. Effects on glycemic control in impaired wound healing in spontaneously diabetic Torii (SDT) fatty rats. *Med Arch*. 2018;72(1):4–8. doi:10.5455/medarch.2018.72.4-8
5. Tam JCW, Lau KM, Liu CL, et al. The in vivo and in vitro diabetic wound healing effects of a 2-herb formula and its mechanisms of action. *J Ethnopharmacol*. 2011;134(3):831–838. doi:10.1016/j.jep.2011.01.032
6. Shu B, Xie JL, Xu YB, et al. Effects of skin-derived precursors on wound healing of denervated skin in a nude mouse model. *Int J Clin Exp Pathol*. 2015;8(3):2660–2669.
7. Korzeniewska E, Szczepny A, Lipiński P, Drózd T, Kielbasa P, Miernik A. Prototype of a textronic sensor created with a physical vacuum deposition process for staphylococcus aureus detection. *Sensors (Basel)*. 2020;21(1):E183. doi:10.3390/s21010183
8. Wang J, Li J, Wang Y, Gao M, Zhang X, Yang P. Development of versatile metal–organic framework functionalized magnetic graphene core–shell biocomposite for highly specific recognition of glycopeptides. *ACS Appl Mater Interfaces*. 2016;8(41):27482–27489. doi:10.1021/acsami.6b08218
9. Sun Y, Wang D, Xu L, et al. Fluorescent small Au nanodots prepared from large Ag nanoparticles for targeting and imaging cancer cells. *RSC Adv*. 2015;5(64):52088–52094.
10. Wang C, Yao Y, Song Q. Gold nanoclusters decorated with magnetic iron oxide nanoparticles for potential multimodal optical/magnetic resonance imaging. *J Mater Chem C*. 2015;3(23):5910–5917. doi:10.1039/C5TC00290G
11. Feng Q, Zhang Y, Zhang W, et al. Tumor-targeted and multi-stimuli responsive drug delivery system for near-infrared light induced chemo-phototherapy and photoacoustic tomography. *Acta Biomater*. 2016;38:129–142. doi:10.1016/j.actbio.2016.04.024
12. Feng Q, Zhang Y, Zhang W, et al. Programmed near-infrared light-responsive drug delivery system for combined magnetic tumor-targeting magnetic resonance imaging and chemo-phototherapy. *Acta Biomater*. 2017;49:402–413. doi:10.1016/j.actbio.2016.11.035
13. Wang F, Zhang W, Li H, Chen X, Feng S, Mei Z. How effective are nano-based dressings in diabetic wound healing? A comprehensive review of literature. *Int J Nanomed*. 2022;17:2097–2119. doi:10.2147/IJN.S361282
14. Ermini ML, Voliani V. Antimicrobial nano-agents: the copper age. *ACS Nano*. 2021;15(4):6008–6029. doi:10.1021/acsnano.0c10756
15. Kalantari K, Mostafaei E, Afifi AM, et al. Wound dressings functionalized with silver nanoparticles: promises and pitfalls. *Nanoscale*. 2020;12(4):2268–2291. doi:10.1039/c9nr08234d
16. Wang L, Hu C, Shao L. The antimicrobial activity of nanoparticles: present situation and prospects for the future. *IJN*. 2017;12:1227–1249. doi:10.2147/IJN.S121956
17. Xu L, Wang YY, Huang J, Chen CY, Wang ZX, Xie H. Silver nanoparticles: synthesis, medical applications and biosafety. *Theranostics*. 2020;10(20):8996–9031. doi:10.7150/thno.45413
18. Nosrati H, Khodaei M, Alizadeh Z, Banitalebi-Dehkordi M. Cationic, anionic and neutral polysaccharides for skin tissue engineering and wound healing applications. *Int J Biol Macromol*. 2021;192:298–322. doi:10.1016/j.ijbiomac.2021.10.013
19. Wang J, Windbergs M. Functional electrospun fibers for the treatment of human skin wounds. *Eur J Pharm Biopharm*. 2017;119:283–299. doi:10.1016/j.ejpb.2017.07.001
20. Stadelmann WK, Digenis AG, Tobin GR. Physiology and healing dynamics of chronic cutaneous wounds. *Am J Surg*. 1998;176(2A Suppl):26S–38S. doi:10.1016/s0002-9610(98)00183-4
21. Ho J, Hantash B. The principles of wound healing. *Exp Rev Dermatol*. 2014;8:639–658. doi:10.1586/17469872.2013.857161
22. Pastar I, Stojadinovic O, Yin NC, et al. Epithelialization in wound healing: a comprehensive review. *Adv Wound Care (New Rochelle)*. 2014;3(7):445–464. doi:10.1089/wound.2013.0473
23. Evans ND, Oreffo ROC, Healy E, Thurner PJ, Man YH. Epithelial mechanobiology, skin wound healing, and the stem cell niche. *J Mech Behav Biomed Mater*. 2013;28:397–409. doi:10.1016/j.jmbbm.2013.04.023
24. Rajendran NK, Kumar SSD, Hourel NN, Abrahamse H. A review on nanoparticle based treatment for wound healing. *J Drug Delivery Sci Technol*. 2018;44:421–430. doi:10.1016/j.jddst.2018.01.009
25. Guo S, Dipietro LA. Factors affecting wound healing. *J Dent Res*. 2010;89(3):219–229. doi:10.1177/0022034509359125

26. Behm B, Babilas P, Landthaler M, Schreml S. Cytokines, chemokines and growth factors in wound healing. *J Eur Acad Dermatol Venereol*. 2012;26(7):812–820. doi:10.1111/j.1468-3083.2011.04415.x
27. Li J, Chen J, Kirsner R. Pathophysiology of acute wound healing. *Clin Dermatol*. 2007;25(1):9–18. doi:10.1016/j.clindermatol.2006.09.007
28. Pakyari M, Farrokhi A, Maharlooie MK, Ghahary A. Critical role of transforming growth factor beta in different phases of wound healing. *Adv Wound Care (New Rochelle)*. 2013;2(5):215–224. doi:10.1089/wound.2012.0406
29. Atala A, Irvine DJ, Moses M, Shaunak S. Wound healing versus regeneration: role of the tissue environment in regenerative medicine. *MRS Bull*. 2010;35(8):597–606. doi:10.1557/mrs2010.528
30. Fertin C, Nicolas JF, Gillery P, Kalis B, Banchereau J, Maquart FX. Interleukin-4 stimulates collagen synthesis by normal and scleroderma fibroblasts in dermal equivalents. *Cell Mol Biol*. 1991;37(8):823–829.
31. Hinz B. Formation and function of the myofibroblast during tissue repair. *J Invest Dermatol*. 2007;127(3):526–537. doi:10.1038/sj.jid.5700613
32. Lazarus GS, Cooper DM, Knighton DR, et al. Definitions and guidelines for assessment of wounds and evaluation of healing. *Arch Dermatol*. 1994;130(4):489–493. doi:10.1001/archderm.1994.01690040093015
33. Basu Mallik S, Jayashree BS, Shenoy RR. Epigenetic modulation of macrophage polarization- perspectives in diabetic wounds. *J diabet complicat*. 2018;32(5):524–530. doi:10.1016/j.jdiacomp.2018.01.015
34. Gordon S, Martinez FO. Alternative activation of macrophages: mechanism and functions. *Immunity*. 2010;32(5):593–604. doi:10.1016/j.immuni.2010.05.007
35. Mantovani A, Biswas SK, Galdiero MR, Sica A, Locati M. Macrophage plasticity and polarization in tissue repair and remodelling. *J Pathol*. 2013;229(2):176–185. doi:10.1002/path.4133
36. Willenborg S, Lucas T, van Loo G, et al. CCR2 recruits an inflammatory macrophage subpopulation critical for angiogenesis in tissue repair. *Blood*. 2012;120(3):613–625. doi:10.1182/blood-2012-01-403386
37. Kim SY, Nair MG. Macrophages in wound healing: activation and plasticity. *Immunol Cell Biol*. 2019;97(3):258–267. doi:10.1111/imcb.12236
38. Han G, Ceilleury R. Chronic wound healing: a review of current management and treatments. *Adv Ther*. 2017;34(3):599–610. doi:10.1007/s12325-017-0478-y
39. Sun C, Sun L, Ma H, et al. The phenotype and functional alterations of macrophages in mice with hyperglycemia for long term. *J Cell Physiol*. 2012;227(4):1670–1679. doi:10.1002/jcp.22891
40. Tahergorabi Z, Khazaei M. Imbalance of angiogenesis in diabetic complications: the mechanisms. *Int J Prev Med*. 2012;3(12):827. doi:10.4103/2008-7802.104853
41. Brem H, Tomic-Canic M. Cellular and molecular basis of wound healing in diabetes. *J Clin Invest*. 2007;117(5):1219–1222. doi:10.1172/JCI32169
42. Gallagher KA, Liu ZJ, Xiao M, et al. Diabetic impairments in NO-mediated endothelial progenitor cell mobilization and homing are reversed by hyperoxia and SDF-1a. *J Clin Invest*. 2007;117(5):1249–1259. doi:10.1172/JCI29710
43. Lerman OZ, Galiano RD, Armour M, Levine JP, Gurtner GC. Cellular dysfunction in the diabetic fibroblast. *Am J Pathol*. 2003;162(1):303–312. doi:10.1016/S0002-9440(10)63821-7
44. Hazarika S, Dokun AO, Li Y, Popel AS, Kontos CD, Annex BH. Impaired angiogenesis after hindlimb ischemia in type 2 diabetes mellitus: differential regulation of vascular endothelial growth factor receptor 1 and soluble vascular endothelial growth factor receptor 1. *Circ Res*. 2007;101(9):948–956. doi:10.1161/CIRCRESAHA.107.160630
45. Kendall RL, Thomas KA. Inhibition of vascular endothelial cell growth factor activity by an endogenously encoded soluble receptor. *Proc Natl Acad Sci USA*. 1993;90(22):10705–10709. doi:10.1073/pnas.90.22.10705
46. Dreaden EC, Alkilany AM, Huang X, Murphy CJ, El-Sayed MA. The golden age: gold nanoparticles for biomedicine. *Chem Soc Rev*. 2012;41(7):2740–2779. doi:10.1039/c1cs15237h
47. Mohamed MM, Fouad SA, Elshoky HA, Mohammed GM, Salaheldin TA. Antibacterial effect of gold nanoparticles against *Corynebacterium pseudotuberculosis*. *Int J Vet Sci Med*. 2017;5(1):23–29. doi:10.1016/j.ijvsm.2017.02.003
48. Vimbela G, Ngo SM, Frazee C, Yang L, Stout DA. Antibacterial properties and toxicity from metallic nanomaterials. *IJN*. 2017;12:3941–3965. doi:10.2147/IJN.S134526
49. Kohanski MA, Dwyer DJ, Hayete B, Lawrence CA, Collins JJ. A Common Mechanism of Cellular Death Induced by Bactericidal Antibiotics. *Cell*. 2007;130(5):797–810. doi:10.1016/j.cell.2007.06.049
50. Cui Y, Zhao Y, Tian Y, Zhang W, X L, Jiang X. The molecular mechanism of action of bactericidal gold nanoparticles on *Escherichia coli*. *Biomaterials*. 2012;33(7):2327–2333. doi:10.1016/j.biomaterials.2011.11.057
51. Boomi P, Ganesan R, Prabu Poorani G, et al. Phyto-engineered gold nanoparticles (AuNPs) with potential antibacterial, antioxidant, and wound healing activities under in vitro and in vivo conditions. *Int J Nanomed*. 2020;15:7553–7568. doi:10.2147/IJN.S257499
52. Annamalai A, Christina VLP, Sudha D, Kalpana M, Lakshmi PTV. Green synthesis, characterization and antimicrobial activity of Au NPs using *Euphorbia hirta* L. leaf extract. *Colloids Surf, B*. 2013;108:60–65. doi:10.1016/j.colsurfb.2013.02.012
53. Wei SC, Chang L, Huang CC, Chang HT. Dual-functional gold nanoparticles with antimicrobial and proangiogenic activities improve the healing of multidrug-resistant bacteria-infected wounds in diabetic mice. *Biomater Sci*. 2019;7(11):4482–4490. doi:10.1039/c9bm00772e
54. Kingston M, Pfau JC, Gilmer J, Brey R. Selective inhibitory effects of 50-nm gold nanoparticles on mouse macrophage and spleen cells. *J Immunotoxicol*. 2016;13(2):198–208. doi:10.3109/1547691X.2015.1035819
55. BarathManiKanth S, Kalishwaralal K, Sriram M, et al. RAesneatrcih-oxidant effect of gold nanoparticles restrains hyperglycemic conditions in diabetic mice. *J Nanobiotechnol*. 2010;8:16. doi:10.1186/1477-3155-8-16
56. Song L, Hobaugh MR, Shustak C, Cheley S, Bayley H, Gouaux JE. Structure of staphylococcal alpha-hemolysin, a heptameric transmembrane pore. *Science*. 1996;274(5294):1859–1866. doi:10.1126/science.274.5294.1859
57. Pornpattananangkul D, Zhang L, Olson S, et al. Bacterial toxin-triggered drug release from gold nanoparticle-stabilized liposomes for the treatment of bacterial infection. *J Am Chem Soc*. 2011;133(11):4132–4139. doi:10.1021/ja111110e
58. Li J, Cha R, Zhao X, et al. Gold Nanoparticles Cure Bacterial Infection with Benefit to Intestinal Microflora. *ACS Nano*. 2019;13(5):5002–5014. doi:10.1021/acsnano.9b01002
59. He X, Dai L, Ye L, et al. A vehicle-free antimicrobial polymer hybrid gold nanoparticle as synergistically therapeutic platforms for *Staphylococcus aureus* infected wound healing. *Adv Sci*. 2022;9(14):2105223. doi:10.1002/advs.202105223



60. Li S, Tang Q, Xu H, et al. Improved stability of KGF by conjugation with gold nanoparticles for diabetic wound therapy. *Nanomedicine*. 2019;14(22):2909–2923. doi:10.2217/nmm-2018-0487
61. Miles AJ, Wallace BA. Circular dichroism spectroscopy of membrane proteins. *Chem Soc Rev*. 2016;45(18):4859–4872. doi:10.1039/C5CS00084J
62. Wang S, Yan C, Zhang X, et al. Antimicrobial peptide modification enhances the gene delivery and bactericidal efficiency of gold nanoparticles for accelerating diabetic wound healing. *Biomater Sci*. 2018;6(10):2757–2772. doi:10.1039/C8BM00807H
63. Chen SA, Chen HM, Yao YD, Hung CF, Tu CS, Liang YJ. Topical treatment with anti-oxidants and Au nanoparticles promote healing of diabetic wound through receptor for advance glycation end-products. *Eur J Pharm Sci*. 2012;47(5):875–883. doi:10.1016/j.ejps.2012.08.018
64. Nosrati H, Heydari M, Tootiaei Z, Ganjbar S, Khodaei M. Delivery of antibacterial agents for wound healing applications using polysaccharide-based scaffolds. *J Drug Delivery Sci Technol*. 2023;84:104516. doi:10.1016/j.jddst.2023.104516
65. Chaloupka K, Malam Y, Seifalian AM. Nanosilver as a new generation of nanoparticle in biomedical applications. *Trends Biotechnol*. 2010;28(11):580–588. doi:10.1016/j.tibtech.2010.07.006
66. Agnihotri S, Mukherji S, Mukherji S. Immobilized silver nanoparticles enhance contact killing and show highest efficacy: elucidation of the mechanism of bactericidal action of silver. *Nanoscale*. 2013;5(16):7328. doi:10.1039/c3nr00024a
67. Dizaj SM, Lotfipour F, Barzegar-Jalali M, Zarrintan MH, Adibkia K. Antimicrobial activity of the metals and metal oxide nanoparticles. *Mater Sci Eng C*. 2014;44:278–284. doi:10.1016/j.msec.2014.08.031
68. Lok CN, Ho CM, Chen R, et al. Proteomic analysis of the mode of antibacterial action of silver nanoparticles. *J Proteome Res*. 2006;5(4):916–924. doi:10.1021/pr0504079
69. Zhang Y, Peng H, Huang W, Zhou Y, Yan D. Facile preparation and characterization of highly antimicrobial colloid Ag or Au nanoparticles. *J Colloid Interface Sci*. 2008;325(2):371–376. doi:10.1016/j.jcis.2008.05.063
70. Ayuk SM, Houreld NN, Abrahamse H. Collagen production in diabetic wounded fibroblasts in response to low-intensity laser irradiation at 660 nm. *Diabetes Technol Ther*. 2012;14(12):1110–1117. doi:10.1089/dia.2012.0125
71. Singla R, Soni S, Patil V, et al. In vivo diabetic wound healing potential of nanobiocomposites containing bamboo cellulose nanocrystals impregnated with silver nanoparticles. *Int J Biol Macromol*. 2017;105(Pt 1):45–55. doi:10.1016/j.ijbiomac.2017.06.109
72. Dhillip Kumar SS, Houreld NN, Abrahamse H. Selective laser efficiency of green-synthesized silver nanoparticles by aloe arborescens and its wound healing activities in normal wounded and diabetic wounded fibroblast cells: in vitro studies. *IJN*. 2020;15:6855–6870. doi:10.2147/IJN.S257204
73. Tao B, Lin C, Deng Y, et al. Copper-nanoparticle-embedded hydrogel for killing bacteria and promoting wound healing with photothermal therapy. *J Mater Chem B*. 2019;7(15):2534–2548. doi:10.1039/c8tb03272f
74. Chatterjee AK, Chakraborty R, Basu T. Mechanism of antibacterial activity of copper nanoparticles. *Nanotechnology*. 2014;25(13):135101. doi:10.1088/0957-4484/25/13/135101
75. Veerapandian M, Sadhasivam S, Choi J, Yun K. Glucosamine functionalized copper nanoparticles: preparation, characterization and enhancement of anti-bacterial activity by ultraviolet irradiation. *Chem Eng J*. 2012;209:558–567. doi:10.1016/j.cej.2012.08.054
76. Das D, Nath BC, Phukon P, Dolui SK. Synthesis and evaluation of antioxidant and antibacterial behavior of CuO nanoparticles. *Colloids Surf, B*. 2013;101:430–433. doi:10.1016/j.colsurfb.2012.07.002
77. Kobayashi Y. The regulatory role of nitric oxide in proinflammatory cytokine expression during the induction and resolution of inflammation. *J Leukoc Biol*. 2010;88(6):1157–1162. doi:10.1189/jlb.0310149
78. Borkow G, Gabbay J, Dardik R, et al. Molecular mechanisms of enhanced wound healing by copper oxide-impregnated dressings. *Wound Repair Regen*. 2010;18(2):266–275. doi:10.1111/j.1524-475X.2010.00573.x
79. Alizadeh S, Seyedalipour B, Shafieyan S, Kheime A, Mohammadi P, Aghdami N. Copper nanoparticles promote rapid wound healing in acute full thickness defect via acceleration of skin cell migration, proliferation, and neovascularization. *Biochem Biophys Res Commun*. 2019;517(4):684–690. doi:10.1016/j.bbrc.2019.07.110
80. Haldorai Y, Shim JJ. Novel chitosan-TiO<sub>2</sub> nanohybrid: preparation, characterization, antibacterial, and photocatalytic properties. *Polym Compos*. 2014;35(2):327–333. doi:10.1002/pc.22665
81. Charpentier PA, Burgess K, Wang L, Chowdhury RR, Lotus AF, Moula G. Nano-TiO<sub>2</sub>/polyurethane composites for antibacterial and self-cleaning coatings. *Nanotechnology*. 2012;23(42):425606. doi:10.1088/0957-4484/23/42/425606
82. Sonmez O, Sonmez M. Role of platelets in immune system and inflammation. *Porto Biomed J*. 2017;2(6):311–314. doi:10.1016/j.pbj.2017.05.005
83. Seisenbaeva GA, Fromell K, Vinogradov VV, et al. Dispersion of TiO<sub>2</sub> nanoparticles improves burn wound healing and tissue regeneration through specific interaction with blood serum proteins. *Sci Rep*. 2017;7(1):15448. doi:10.1038/s41598-017-15792-w
84. Brayner R, Ferrari-Iliou R, Brivois N, Djediat S, Benedetti MF, Fiévet F. Toxicological impact studies based on Escherichia coli bacteria in ultrafine ZnO nanoparticles colloidal medium. *Nano Lett*. 2006;6(4):866–870. doi:10.1021/nl052326h
85. Xie Y, He Y, Irwin PL, Jin T, Shi X. Antibacterial activity and mechanism of action of zinc oxide nanoparticles against Campylobacter jejuni. *Appl Environ Microbiol*. 2011;77(7):2325–2331. doi:10.1128/AEM.02149-10
86. Mittal M, Siddiqui MR, Tran K, Reddy SP, Malik AB. Reactive oxygen species in inflammation and tissue injury. *Antioxid Redox Signal*. 2014;20(7):1126–1167. doi:10.1089/ars.2012.5149
87. Agarwal H, Nakara A, Shanmugam VK. Anti-inflammatory mechanism of various metal and metal oxide nanoparticles synthesized using plant extracts: a review. *Biomed Pharmacother*. 2019;109:2561–2572. doi:10.1016/j.biopha.2018.11.116
88. Kim MH, Seo JH, Kim HM, Jeong HJ. Aluminum-doped zinc oxide nanoparticles attenuate the TSLP levels via suppressing caspase-1 in activated mast cells. *J Biomater Appl*. 2016;30(9):1407–1416. doi:10.1177/0885328216629822
89. Qian Y, Zheng Y, Jin J, et al. Immunoregulation in diabetic wound repair with a photoenhanced glycyrrhizic acid hydrogel scaffold. *Adv Mater*. 2022;34(29):2200521. doi:10.1002/adma.202200521
90. Lin PH, Sermersheim M, Li H, Lee PHU, Steinberg SM, Ma J. Zinc in wound healing modulation. *Nutrients*. 2017;10(1):E16. doi:10.3390/nu10010016
91. Arakha M, Pal S, Samantarai D, et al. Antimicrobial activity of iron oxide nanoparticle upon modulation of nanoparticle-bacteria interface. *Sci Rep*. 2015;5:14813. doi:10.1038/srep14813

92. Gu Y, Huang Y, Qiu Z, et al. Vitamin B2 functionalized iron oxide nanozymes for mouth ulcer healing. *Sci China Life Sci.* **2020**;63(1):68–79. doi:10.1007/s11427-019-9590-6
93. Garzón-Manjón A, Aranda-Ramos A, Melara-Benitez B, et al. Simple synthesis of biocompatible stable CeO<sub>2</sub> Nanoparticles as antioxidant agents. *Bioconjugate Chem.* **2018**;29(7):2325–2331. doi:10.1021/acs.bioconjchem.8b00300
94. Chigurupati S, Mughal MR, Okun E, et al. Effects of cerium oxide nanoparticles on the growth of keratinocytes, fibroblasts and vascular endothelial cells in cutaneous wound healing. *Biomaterials.* **2013**;34(9):2194–2201. doi:10.1016/j.biomaterials.2012.11.061
95. Park HJ, Kim JY, Kim J, et al. Silver-ion-mediated reactive oxygen species generation affecting bactericidal activity. *Water Res.* **2009**;43(4):1027–1032. doi:10.1016/j.watres.2008.12.002
96. Silver S. Bacterial silver resistance: molecular biology and uses and misuses of silver compounds. *FEMS Microbiol Rev.* **2003**;27(2–3):341–353. doi:10.1016/S0168-6445(03)00047-0
97. Shi G, Chen W, Zhang Y, Dai X, Zhang X, Wu Z. An antifouling hydrogel containing silver nanoparticles for modulating the therapeutic immune response in chronic wound healing. *Langmuir.* **2019**;35(5):1837–1845. doi:10.1021/acs.langmuir.8b01834
98. Zielińska D, Struszczyk MH, Madej-Kielbik L, et al. Design of New-generation usable forms of topical haemostatic agents containing Chitosan. *Molecules.* **2017**;22(12):2240. doi:10.3390/molecules22122240
99. Choudhary M, Chhabra P, Tyagi A, Singh H. Scar free healing of full thickness diabetic wounds: a unique combination of silver nanoparticles as antimicrobial agent, calcium alginate nanoparticles as hemostatic agent, fresh blood as nutrient/growth factor supplier and chitosan as base matrix. *Int J Biol Macromol.* **2021**;178:41–52. doi:10.1016/j.ijbiomac.2021.02.133
100. Wang YY, Addisu KD, Gebrie HT, et al. Multifunctional thermosensitive hydrogel based on alginate and P(NIPAM-co-HEMIN) composites for accelerated diabetic wound healing. *Int J Biol Macromol.* **2023**;241:124540. doi:10.1016/j.ijbiomac.2023.124540
101. Masood N, Ahmed R, Tariq M, et al. Silver nanoparticle impregnated chitosan-PEG hydrogel enhances wound healing in diabetes induced rabbits. *Int J Pharm.* **2019**;559:23–36. doi:10.1016/j.ijpharm.2019.01.019
102. Franková J, Pivodová V, Vágnerová H, Juraňová J, Ulrichová J. Effects of silver nanoparticles on primary cell cultures of fibroblasts and keratinocytes in a wound-healing model. *J Appl Biomater Funct Mater.* **2016**;14(2):e137–142. doi:10.5301/jabfm.5000268
103. Paul A, Ju H, Rangasamy S, Shim Y, Song JM. Nanosized silver (II) pyridoxine complex to cause greater inflammatory response and less cytotoxicity to RAW264.7 macrophage cells. *Nanoscale Res Lett.* **2015**;10(1):140. doi:10.1186/s11671-015-0848-9
104. Fathil MAM, Katas H. Antibacterial, Anti-biofilm and pro-migratory effects of double layered hydrogels packaged with lactoferrin-DsiRNA-silver nanoparticles for chronic wound therapy. *Pharmaceutics.* **2023**;15(3):991. doi:10.3390/pharmaceutics15030991
105. Chauhan PS, Shrivastava V, Tomar RS. Biofabrication of copper nanoparticles: a next-generation antibacterial agent against wound-associated pathogens. *Turk J Pharm Sci.* **2018**;15(3):238–247. doi:10.4274/tjps.52724
106. Hassan MS, Amna T, Yang OB, El-Newehy MH, Al-Deyab SS, Khil MS. Smart copper oxide nanocrystals: synthesis, characterization, electrochemical and potent antibacterial activity. *Colloids Surf., B.* **2012**;97:201–206. doi:10.1016/j.colsurfb.2012.04.032
107. Akhavan O, Ghaderi E. Cu and CuO nanoparticles immobilized by silica thin films as antibacterial materials and photocatalysts. *Surf Coat Technol.* **2010**;205(1):219–223. doi:10.1016/j.surfcoat.2010.06.036
108. Sankar R, Baskaran A, Shivashangari KS, Ravikumar V. Inhibition of pathogenic bacterial growth on excision wound by green synthesized copper oxide nanoparticles leads to accelerated wound healing activity in Wistar Albino rats. *J Mater Sci Mater Med.* **2015**;26(7):214. doi:10.1007/s10856-015-5543-y
109. Della Rocca J, Liu D, Lin W. Nanoscale metal-organic frameworks for biomedical imaging and drug delivery. *Acc Chem Res.* **2011**;44(10):957–968. doi:10.1021/ar200028a
110. Zhang P, Li Y, Tang Y, et al. Copper-based metal-organic framework as a controllable nitric oxide-releasing vehicle for enhanced diabetic wound healing. *ACS Appl Mater Interfaces.* **2020**;12(16):18319–18331. doi:10.1021/acsami.0c01792
111. Su CH, Li WP, Tsao LC, et al. Enhancing microcirculation on multitriggering manner facilitates angiogenesis and collagen deposition on wound healing by photoreleased NO from hemin-derivatized colloids. *ACS Nano.* **2019**;13(4):4290–4301. doi:10.1021/acsnano.8b09417
112. Champeau M, Póvoa V, Militão L, et al. Supramolecular poly(acrylic acid)/F127 hydrogel with hydration-controlled nitric oxide release for enhancing wound healing. *Acta Biomater.* **2018**;74:312–325. doi:10.1016/j.actbio.2018.05.025
113. Xiao J, Chen S, Yi J, Zhang HF, Ameer GA. A cooperative copper metal-organic framework-hydrogel system improves wound healing in diabetes. *Adv Funct Mater.* **2017**;27(1):1604872. doi:10.1002/adfm.201604872
114. Zhang W, Hu Y, Ge J, Jiang HL, Yu SH. A facile and general coating approach to moisture/water-resistant metal-organic frameworks with intact porosity. *J Am Chem Soc.* **2014**;136(49):16978–16981. doi:10.1021/ja509960n
115. Xiao J, Zhu Y, Huddleston S, et al. Copper metal-organic framework nanoparticles stabilized with folic acid improve wound healing in diabetes. *ACS Nano.* **2018**;12(2):1023–1032. doi:10.1021/acsnano.7b01850
116. Deng Z. A biotin-stabilized HKUST-1ADM scaffold for facil.pdf; **2023**.
117. Gopal A, Kant V, Gopalakrishnan A, Tandan SK, Kumar D. Chitosan-based copper nanocomposite accelerates healing in excision wound model in rats. *Eur J Pharmacol.* **2014**;731:8–19. doi:10.1016/j.ejphar.2014.02.033
118. Mandinova L, Mandinova A, Kyurkchiev S, et al. Copper chelation represses the vascular response to injury. *Proc Natl Acad Sci U S A.* **2003**;100(11):6700–6705. doi:10.1073/pnas.1231994100
119. Huang Y, Qi L, Liu Z, et al. Radially electrospun fibrous membrane incorporated with copper peroxide nanodots capable of self-catalyzed chemodynamic therapy for angiogenesis and healing acceleration of diabetic wounds. *ACS Appl Mater Interfaces.* **2023**;acsami.3c06703. doi:10.1021/acsami.3c06703
120. Wang X, Lv F, Li T, et al. Electrospun micropatterned nanocomposites incorporated with Cu<sub>2</sub>S nanoflowers for skin tumor therapy and wound healing. *ACS Nano.* **2017**;11(11):11337–11349. doi:10.1021/acsnano.7b05858
121. Romano Di Peppe S, Mangoni A, Zambruno G, et al. Adenovirus-mediated VEGF(165) gene transfer enhances wound healing by promoting angiogenesis in CD1 diabetic mice. *Gene Ther.* **2002**;9(19):1271–1277. doi:10.1038/sj.gt.3301798
122. Imtiyaz HZ, Simon MC. Hypoxia-inducible factors as essential regulators of inflammation. *Curr Top Microbiol Immunol.* **2010**;345:105–120. doi:10.1007/82\_2010\_74
123. Jansen J, Karges W, Rink L. Zinc and diabetes--clinical links and molecular mechanisms. *J Nutr Biochem.* **2009**;20(6):399–417. doi:10.1016/j.jnutbio.2009.01.009

124. Umrani RD, Paknikar KM. Zinc oxide nanoparticles show antidiabetic activity in streptozotocin-induced Type 1 and 2 diabetic rats. *Nanomedicine*. 2014;9(1):89–104. doi:10.2217/nnm.12.205
125. Yadav E, Singh D, Yadav P, Verma A. Ameliorative effect of biofabricated ZnO nanoparticles of *Trianthema portulacastrum* Linn. on dermal wounds via removal of oxidative stress and inflammation. *RSC Adv*. 2018;8(38):21621–21635. doi:10.1039/C8RA03500H
126. Huang Z, Zheng X, Yan D, et al. Toxicological effect of ZnO nanoparticles based on bacteria. *Langmuir*. 2008;24(8):4140–4144. doi:10.1021/la7035949
127. Raguvanan R, Manuja BK, Chopra M, et al. Sodium alginate and gum acacia hydrogels of ZnO nanoparticles show wound healing effect on fibroblast cells. *Int J Biol Macromol*. 2017;96:185–191. doi:10.1016/j.ijbiomac.2016.12.009
128. Nagajyothi PC, Cha SJ, Yang JJ, Sreekanth TVM, Kim KJ, Shin HM. Antioxidant and anti-inflammatory activities of zinc oxide nanoparticles synthesized using *Polygala tenuifolia* root extract. *J Photochem Photobiol B Biol*. 2015;146:10–17. doi:10.1016/j.jphotobiol.2015.02.008
129. López-Goerne T, Ramirez P, Alvarez D, et al. Physicochemical properties and *in vivo* evaluation of Pt/TiO<sub>2</sub>–SiO<sub>2</sub> nanopowders. *Nanomedicine*. 2018;13(17):2171–2185. doi:10.2217/nnm-2018-0078
130. Cormode DP, Gao L, Koo H. Emerging biomedical applications of enzyme-like catalytic nanomaterials. *Trends Biotechnol*. 2018;36(1):15–29. doi:10.1016/j.tibtech.2017.09.006
131. Hu Y, Rao SS, Wang ZX, et al. Exosomes from human umbilical cord blood accelerate cutaneous wound healing through miR-21-3p-mediated promotion of angiogenesis and fibroblast function. *Theranostics*. 2018;8(1):169–184. doi:10.7150/thno.21234
132. Li X, Wei J, Aifantis KE, et al. Current investigations into magnetic nanoparticles for biomedical applications. *J Biomed Mater Res A*. 2016;104(5):1285–1296. doi:10.1002/jbm.a.35654
133. Wu D, Kang L, Tian J, et al. Exosomes derived from bone mesenchymal stem cells with the stimulation of Fe<sub>3</sub>O<sub>4</sub> nanoparticles and static magnetic field enhance wound healing through upregulated miR-21-5p. *Int J Nanomed*. 2020;15:7979–7993. doi:10.2147/IJN.S275650
134. Kannan SK, Sundrarajan M. A green approach for the synthesis of a cerium oxide nanoparticle: characterization and antibacterial activity. *Int J Nanosci*. 2014;13(03):1450018. doi:10.1142/S0219581X14500185
135. Zholobak NM, Ivanov VK, Shcherbakov AB. Chapter 12 - Interaction of nanoceria with microorganisms. In: Grumezescu AM, editor. *Nanobiomaterials in Antimicrobial Therapy*. William Andrew Publishing; 2016:419–450. doi:10.1016/B978-0-323-42864-4.00012-9
136. Ghaznavi H, Najafi R, Mehrzadi S, et al. The neuro-protective effects of cerium and yttrium oxide nanoparticles on high glucose-induced oxidative stress and apoptosis in undifferentiated PC12 cells. *Neurological Res*. 2015;37(7):624–632.
137. Nosrati H, Heydari M, Khodaei M. Cerium oxide nanoparticles: synthesis methods and applications in wound healing. *Mater Today Bio*. 2023;23:100823. doi:10.1016/j.mtbio.2023.100823
138. Augustine R, Hasan A, Patan NK, et al. Cerium oxide nanoparticle incorporated electrospun Poly(3-hydroxybutyrate-co-3-hydroxyvalerate) membranes for diabetic wound healing applications. *ACS Biomater Sci Eng*. 2020;6(1):58–70. doi:10.1021/acsbomaterials.8b01352
139. Yao Y, Zang Y, Qu J, Tang M, Zhang T. The toxicity of metallic nanoparticles on liver: the subcellular damages, mechanisms, and outcomes. *Int J Nanomed*. 2019;14:8787–8804. doi:10.2147/IJN.S212907
140. Sengul AB, Asmatulu E. Toxicity of metal and metal oxide nanoparticles: a review. *Environ Chem Lett*. 2020;18(5):1659–1683. doi:10.1007/s10311-020-01033-6
141. Wang J, Zhou G, Chen C, et al. Acute toxicity and biodistribution of different sized titanium dioxide particles in mice after oral administration. *Toxicol Lett*. 2007;168(2):176–185. doi:10.1016/j.toxlet.2006.12.001
142. Chen J, Dong X, Zhao J, Tang G. In vivo acute toxicity of titanium dioxide nanoparticles to mice after intraperitoneal injection. *J Appl Toxicol*. 2009;29(4):330–337. doi:10.1002/jat.1414
143. Jain TK, Reddy MK, Morales MA, Leslie-Pelecky DL, Labhasetwar V. Biodistribution, clearance, and biocompatibility of iron oxide magnetic nanoparticles in rats. *Mol Pharm*. 2008;5(2):316–327. doi:10.1021/mp7001285
144. Thakur N, Manna P, Das J. Synthesis and biomedical applications of nanoceria, a redox active nanoparticle. *J Nanobiotechnol*. 2019;17(1):84. doi:10.1186/s12951-019-0516-9
145. Kumari M, Kumari SI, Grover P. Genotoxicity analysis of cerium oxide micro and nanoparticles in Wistar rats after 28 days of repeated oral administration. *Mutagenesis*. 2014;29(6):467–479. doi:10.1093/mutage/geu038
146. Ranjbar A, Ghasemi H, Abedian A, Kheiripour N. Cerium oxide nanoparticle modulates hepatic damage, inflammatory and oxidative stress biomarkers in a dose-dependent manner: an *in vivo* study of rat liver. *Nanomed J*. 2018;5(4). doi:10.22038/nmj.2018.05.00009
147. Xing Y, Zeng B, Yang W. Light responsive hydrogels for controlled drug delivery. *Front Bioeng Biotechnol*. 2022;10:1075670. doi:10.3389/fbioe.2022.1075670
148. Permana AD, Anjani QK, Utomo E, et al. Selective delivery of silver nanoparticles for improved treatment of biofilm skin infection using bacteria-responsive microparticles loaded into dissolving microneedles. *Mater Sci Eng C*. 2021;120:111786. doi:10.1016/j.msec.2020.111786
149. Flemming HC, Wingender J. The biofilm matrix. *Nat Rev Microbiol*. 2010;8(9):623–633. doi:10.1038/nrmicro2415
150. Waghule T, Singhvi G, Dubey SK, et al. Microneedles: a smart approach and increasing potential for transdermal drug delivery system. *Biomed Pharmacother*. 2019;109:1249–1258. doi:10.1016/j.biopha.2018.10.078
151. Bariya SH, Gohel MC, Mehta TA, Sharma OP. Microneedles: an emerging transdermal drug delivery system. *J Pharm Pharmacol*. 2012;64(1):11–29. doi:10.1111/j.2042-7158.2011.01369.x
152. Permana AD, Mir M, Utomo E, Donnelly RF. Bacterially sensitive nanoparticle-based dissolving microneedles of doxycycline for enhanced treatment of bacterial biofilm skin infection: a proof of concept study. *Int J Pharm X*. 2020;2:100047. doi:10.1016/j.ijpx.2020.100047
153. Yang J, Chu Z, Jiang Y, et al. Multifunctional hyaluronic acid microneedle patch embedded by cerium/zinc-based composites for accelerating diabetes wound healing. *Adv Healthcare Mater*. 2023;12(24):2300725. doi:10.1002/adhm.202300725
154. Zeng Y, Wang C, Lei K, et al. Multifunctional MOF-based microneedle patch with synergistic chemo-photodynamic antibacterial effect and sustained release of growth factor for chronic wound healing. *Adv Healthcare Mater*. 2023;12(19):2300250. doi:10.1002/adhm.202300250
155. Pang Q, Jiang Z, Wu K, Hou R, Zhu Y. Nanomaterials-based wound dressing for advanced management of infected wound. *Antibiotics (Basel)*. 2023;12(2):351. doi:10.3390/antibiotics12020351
156. Luther DC, Huang R, Jeon T, et al. Delivery of drugs, proteins, and nucleic acids using inorganic nanoparticles. *Adv Drug Deliv Rev*. 2020;156:188–213. doi:10.1016/j.addr.2020.06.020

157. Yu N, Wang X, Qiu L, et al. Bacteria-triggered hyaluronan/AgNPs/gentamicin nanocarrier for synergistic bacteria disinfection and wound healing application. *Chem Eng J*. 2020;380:122582. doi:10.1016/j.cej.2019.122582
158. Konturek PC, Brzozowski T, Konturek SJ, Kwiecien S, Dembinski A, Hahn EG. Influence of bacterial lipopolysaccharide on healing of chronic experimental ulcer in rat. *Scand J Gastroenterol*. 2001;36(12):1239–1247. doi:10.1080/003655201317097065
159. Zhou Y, Kong Y, Kundu S, Cirillo JD, Liang H. Antibacterial activities of gold and silver nanoparticles against *Escherichia coli* and *Bacillus Calmette-Guérin*. *J Nanobiotechnol*. 2012;10(1):19. doi:10.1186/1477-3155-10-19
160. Choi JB, Park JS, Khil MS, et al. Characterization and antimicrobial property of poly(acrylic acid) nanogel containing silver particle prepared by electron beam. *Int J Mol Sci*. 2013;14(6):11011–11023. doi:10.3390/ijms140611011
161. Sondi I, Salopek-Sondi B. Silver nanoparticles as antimicrobial agent: a case study on *E. coli* as a model for Gram-negative bacteria. *J Colloid Interface Sci*. 2004;275(1):177–182. doi:10.1016/j.jcis.2004.02.012
162. He X, Hou J, Sun X, et al. NIR-II photo-amplified sonodynamic therapy using sodium molybdenum bronze nanoplateform against subcutaneous *Staphylococcus Aureus* infection. *Adv Funct Mater*. 2022;32(38):2203964. doi:10.1002/adfm.202203964
163. He X, Qian Y, Wu C, et al. Entropy-mediated high-entropy MXenes nanotherapeutics: NIR-II-enhanced intrinsic oxidase mimic activity to combat methicillin-resistant *Staphylococcus Aureus* infection. *Adv Mater*. 2023;35(26):2211432. doi:10.1002/adma.202211432
164. Kuhn DA, Vanhecke D, Michen B, et al. Different endocytotic uptake mechanisms for nanoparticles in epithelial cells and macrophages. *Beilstein J Nanotechnol*. 2014;5:1625–1636. doi:10.3762/bjnano.5.174
165. Vinluan RD, Zheng J. Serum protein adsorption and excretion pathways of metal nanoparticles. *Nanomedicine*. 2015;10(17):2781–2794. doi:10.2217/nnm.15.97
166. Simko M, Fiedler U, Gazsó A, Nentwich M The impact of nanoparticles on cellular functions; November 9, 2011. <http://hw.oew.ac.at/nanotrust-dossier>. Accessed December 22, 2023.
167. Nosrati H, Aramideh khoury R, Nosrati A, et al. Nanocomposite scaffolds for accelerating chronic wound healing by enhancing angiogenesis. *J Nanobiotechnol*. 2021;19(1):1. doi:10.1186/s12951-020-00755-7
168. Hao D, Zhang G, Gong Y, Ma Z. Development and biological evaluation of cerium oxide loaded polycaprolactone dressing on cutaneous wound healing in nursing care. *Mater Lett*. 2020;265:127401. doi:10.1016/j.matlet.2020.127401
169. Teimouri M, Lalehzar SS. Evaluation of the therapeutic effect of dressing containing Silver (Ag coat) in the process of healing skin blisters caused by limb fractures: a clinical trial study. *BMC Surg*. 2023;23:101. doi:10.1186/s12893-023-02012-8
170. Arendsen LP, Thakar R, Bassett P, Sultan AH. The impact of copper impregnated wound dressings on surgical site infection following caesarean section: a double blind randomised controlled study. *Eur J Obstetrics Gynecol Reprod Biol*. 2020;251:83–88. doi:10.1016/j.ejogrb.2020.05.016
171. Essa MS, Ahmad KS, Zayed ME, Ibrahim SG. Comparative study between silver nanoparticles dressing (SilverSTAT Gel) and conventional dressing in diabetic foot ulcer healing: a prospective randomized study. *Int J Low Extrem Wounds*. 2023;22(1):48–55. doi:10.1177/1534734620988217
172. Erring M, Gaba S, Mohsina S, Tripathy S, Sharma RK. Comparison of efficacy of silver-nanoparticle gel, nano-silver-foam and collagen dressings in treatment of partial thickness burn wounds. *Burns*. 2019;45(8):1888–1894. doi:10.1016/j.burns.2019.07.019
173. Abdollahimajd F, Pourani MR, Mahdavi H, Mirzadeh H, Younespour S, Moravvej H. Efficacy and safety of chitosan-based bio-compatible dressing versus nanosilver (ActicoatTM) dressing in treatment of recalcitrant diabetic wounds: a randomized clinical trial. *Dermatol Ther*. 2022;35(9):e15682. doi:10.1111/dth.15682
174. Melamed E, Rovitsky A, Roth T, Assa L, Borkow G. Stimulation of healing of non-infected stagnated diabetic wounds by copper oxide-impregnated wound dressings. *Medicina*. 2021;57(10):1129. doi:10.3390/medicina57101129
175. Senet P, Bause R, Jørgensen B, Fogh K. Clinical efficacy of a silver-releasing foam dressing in venous leg ulcer healing: a randomised controlled trial: clinical efficacy of a silver-releasing foam dressing in venous leg ulcer healing. *Int Wound J*. 2014;11(6):649–655. doi:10.1111/iwj.12022
176. Kuo FC, Chen B, Lee MS, Yen SH, Wang JW. AQUACEL® Ag surgical dressing reduces surgical site infection and improves patient satisfaction in minimally invasive total knee arthroplasty: a prospective, randomized, controlled study. *Biomed Res Int*. 2017;2017:1–8. doi:10.1155/2017/1262108
177. Cheng H, Shi Z, Yue K, et al. Sprayable hydrogel dressing accelerates wound healing with combined reactive oxygen species-scavenging and antibacterial abilities. *Acta Biomater*. 2021;124:219–232. doi:10.1016/j.actbio.2021.02.002
178. Zhu J, Han Q, Li Q, et al. A multi-enzyme-like activity exhibiting mussel-inspired nanozyme hydrogel for bacteria-infected wound healing. *Biomater Sci*. 2023;11(8):2711–2725. doi:10.1039/D2BM02004A

## International Journal of Nanomedicine

Dovepress

## Publish your work in this journal

The International Journal of Nanomedicine is an international, peer-reviewed journal focusing on the application of nanotechnology in diagnostics, therapeutics, and drug delivery systems throughout the biomedical field. This journal is indexed on PubMed Central, MedLine, CAS, SciSearch®, Current Contents®/Clinical Medicine, Journal Citation Reports/Science Edition, EMBase, Scopus and the Elsevier Bibliographic databases. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/international-journal-of-nanomedicine-journal>