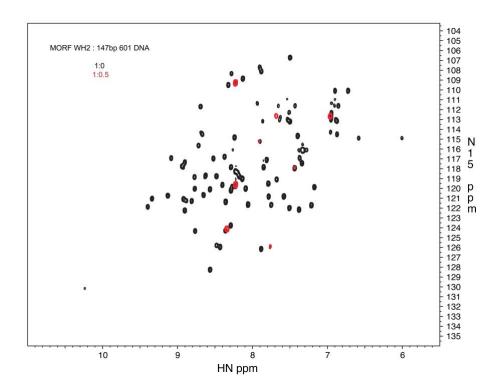
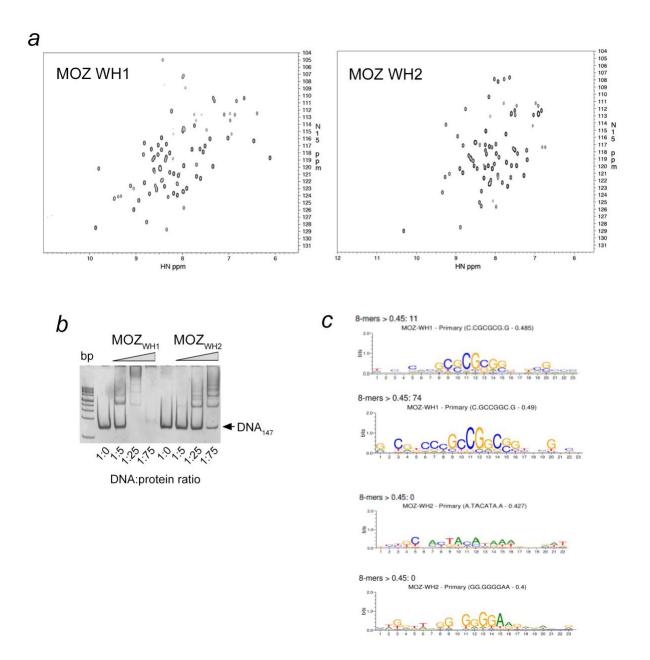
Supplementary Information

MORF and MOZ acetyltransferases target unmethylated CpG islands through the winged helix domain

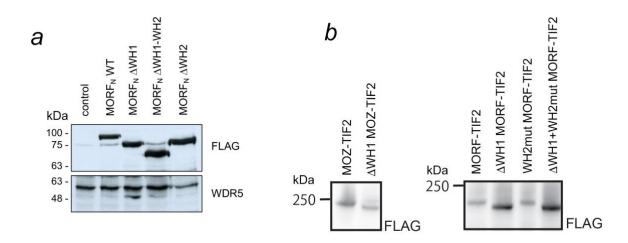
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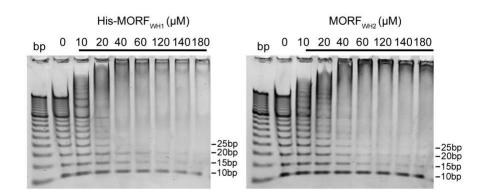
Supplementary Figure 1. Overlay of ¹H, ¹⁵N HSQC spectra of MORF_{WH2} in the absence (black) and presence of 601 DNA (red). Related to Figure 1.

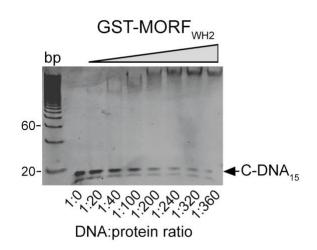


Supplementary Figure 2. (a) 1 H, 15 N HSQC spectra of MOZ_{WH1} and MOZ_{WH2}. (b) EMSA of 147bp 601 DNA in the presence of increasing amounts of MOZ_{WH1} and MOZ_{WH2}. DNA:protein ratio is shown below the gel image. (c) Analysis of DNA binding selectivity of MOZ_{WH1} and MOZ_{WH2} in universal oligonucleotide PBM arrays. Related to Figures 1 and 3. Source data are provided as a Source Data file.

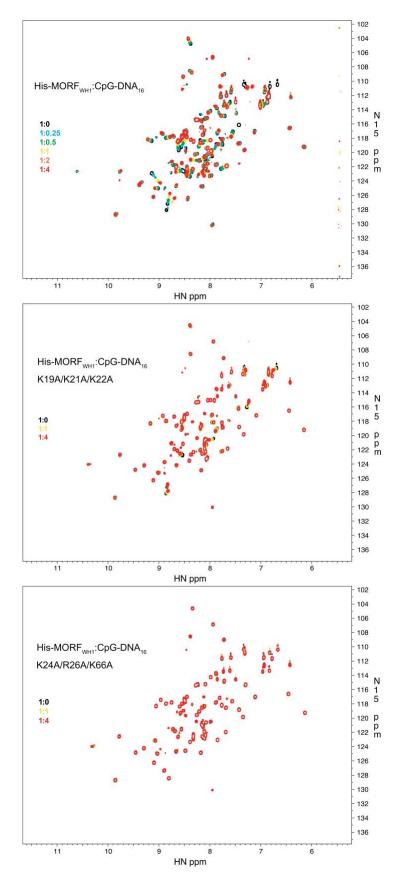


Supplementary Figure 3. Western-Blot showing expression of the indicated MORF and MOZ constructs. (a) FLAG-MORF_{N1-716} WT, Δ WH1, Δ WH2 and Δ WH1-WH2 in the respective K562 cell lines used for ChIP experiments. WDR5 signal is used as loading control. (b) Protein expression of FLAG-tagged MOZ/MORF-TIF2 fusion constructs in plat-E cells. The whole cell extracts were analyzed by Western-Blotting using anti-FLAG antibody. Related to Figures 1 and 6. Source data are provided as a Source Data file.

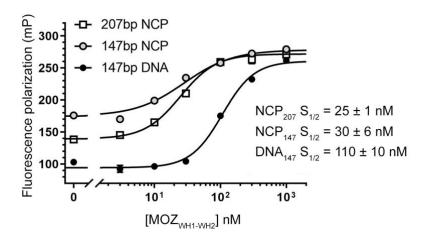




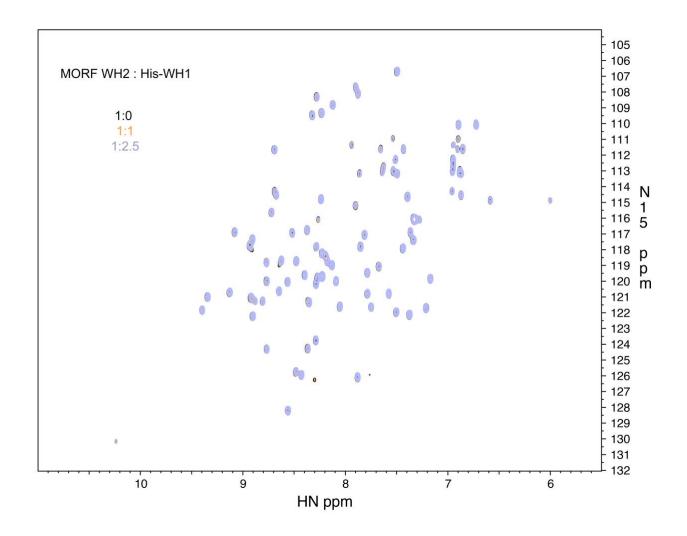
Supplementary Figure 4. EMSAs of a 10bp to 100bp DNA ladder (top) or a 15bp C-rich dsDNA (C-DNA₁₅) (bottom) in the presence of increasing amounts of indicated MORF_{WH1} and MORF_{WH2}. Related to Figures 2 and 3. Source data are provided as a Source Data file.



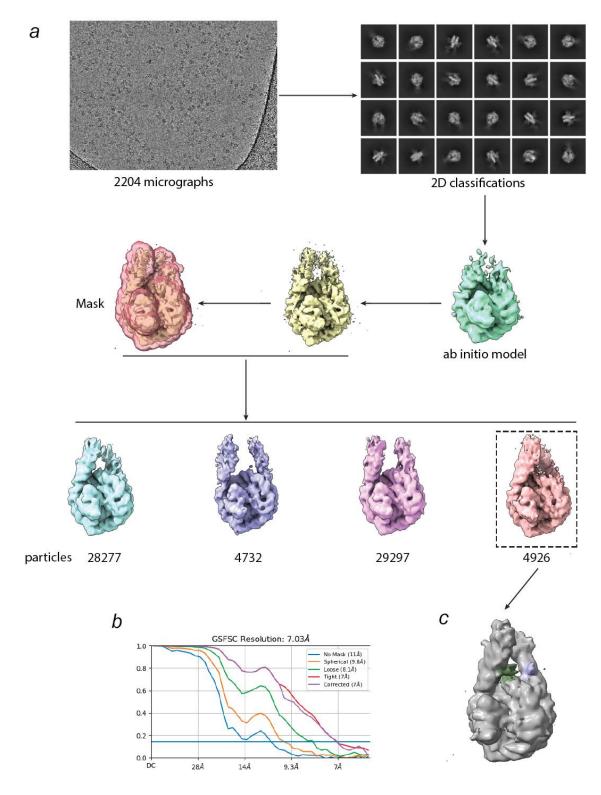
Supplementary Figure 5. Overlay of ¹H, ¹⁵N HSQC spectra of ¹⁵N-labeled His-MORF_{WH1}, WT or K19A/K21A/K22A and K24A/R26A/K66A mutants in the presence of increasing amount of CpG-DNA₁₆. Spectra are color coded according to the protein:DNA molar ratio. Related to Figure 3.



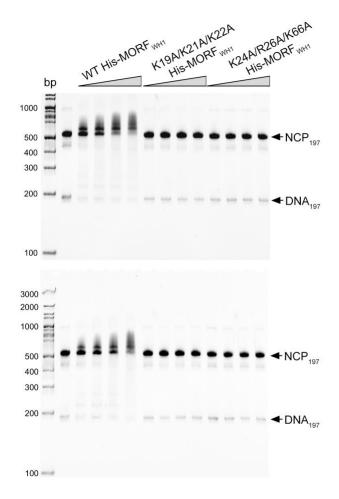
Supplementary Figure 6. Binding curves for the interactions of $MOZ_{WH1-WH2}$ with NCP_{207} , NCP_{147} and DNA_{147} as measured by fluorescence polarization. Data represent mean \pm SD of three independent experiments. n=3 Related to Figure 7. Source data are provided as a Source Data file.



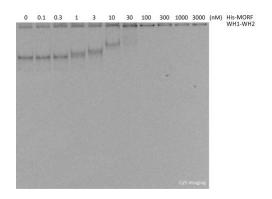
Supplementary Figure 7. Overlay of ¹H,¹⁵N HSQC spectra of ¹⁵N-labeled MORF_{WH2} in the presence of increasing amount of unlabeled His-MORF_{WH1}. Spectra are color coded according to the protein:ligand molar ratio. Related to Figure 7.



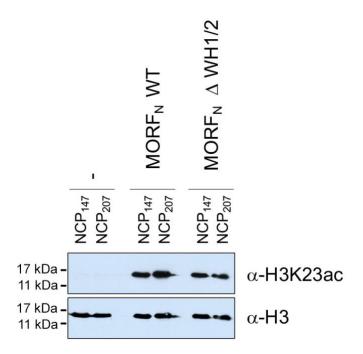
Supplementary Figure 8. Cryo-EM reconstruction of MORF_{WH1-WH2}-Nucleosome-scFv complex. (a) Workflow of the cryo-EM image processing. (b) Fourier Shell Correlations (FSC) of the reconstruction. Gold-standard cut-off (FSC=0.143) marked with a blue line. (c) Final cryo-EM map of the MORF_{WH1-WH2}-nucleosome-scFv complex, the density near the dyad region is colored in green and light blue. Related to Figure 8.



Supplementary Figure 9. Two replicates of EMSA of 197 bp NCP (NCP₁₉₇) in the presence of increasing amounts of indicated WT or mutated His-MORF_{WH1}. Related to Figure 8. Source data are provided as a Source Data file.



Supplementary Figure 10. EMSA of the Cy3-Cy5 labeled NCP_{273} in the presence of increasing amounts of His-MORF_{WH1-WH2}. Related to Figure 8. Source data are provided as a Source Data file.



Supplementary Figure 11. In vitro HAT assays using WT and mutated MORF_N complexes purified from K562 cells on NCP w/o DNA linkers (NCP₁₄₇) and NCP with the linkers (NCP₂₀₇). Related to Figure 9. Source data are provided as a Source Data file.

Supplementary Table 1. NMR and refinement statistics for the $MORF_{WH2}$ structures. Related to Figure 2.

	MORF _{WH2}
NMR distance and dihedral constraints	
Distance constraints	
Total NOE	1603
Intra-residue	600
Inter-residue	
Sequential $(i-j =1)$	390
Medium-range $(i-j \le 4)$	303
Long-range $(i-j \ge 5)$	280
Hydrogen bonds	40
Total dihedral angle restraints	
φ	54
Ψ	54
Structure statistics	
Violations (mean and s.d.)	
Distance constraints (Å)	0.054 ± 0.001
Dihedral angle constraints (°)	0.842 ± 0.078
Number of distance violation (> 0.35 Å)	0
Number of angle violation (> 10°)	0
Deviations from idealized geometry	
Bond lengths (Å)	0.003 ± 0.000
Bond angles (°)	0.461 ± 0.023
Impropers (°)	0.333 ± 0.017
Average pairwise r.m.s. deviation** (Å)	
Heavy	2.233 ± 0.655
Backbone	1.818 ± 0.743
Heavy (107-118,126-135,140-144,148-162,165-168,171-174)	0.895 ± 0.094
Backbone (107-118,126-135,140-144,148-162,165-168,171-174)	0.309 ± 0.068

^{**}Pairwise r.m.s. deviation to lowest energy structure was calculated among 15 refined structures. Residues 100-182 of MORF were used.

Supplementary Table 2. Taqman probes. Related to Figure 6.

Oligonucleotides		
Taqman probe for murine Gapdh	Life Technologies	Mm99999915_g1
Taqman probe for murine Hoxa9	Life Technologies	Mm00439364_m1

Supplementary Table 3. Custom qPCR probe/primer sequences. Related to Figure 6.

Forward	Reverse	Reporter
		ACTGCCACCATGCC
GATGATATTG	ACTACCA	AAT
GCAGCGGGCAAGA	ACGTTCCCCTCCCT	CCGAGTCTGACCAC
AAGAC	CTCA	CTTAC
GGACTCACTGTCCC	CTTCAAGGCCATGA	TCAGCCTTTCCGCC
TCCTGAA	GGTCTCA	CTC
GCGGTATCTGCTGC	GCATTATGTATGCA	CTGGGTGGAAGGT
TTTGG	CAGCTATCTGGAT	ATCC
CCGGCTAGGGTGG	GAGGCGAAGCCCC	CAGGACGCCCGCA
AAGAG	CTATTC	GCG
GGGTAGGCGCAGG	GGTTTTTCCAAGTC	ATGTGTCCGATTCT
CA	AACGATTCCA	CC
TGGCTGCTTTTTAT	CCGCGTGCGAGTG	CCCCTCACATAAAA
GGCTTCAATT	С	TT
TCACCACCACCCCT	GCAAGCCCGCGAA	CAGGAGCGCATGTA
ACGT	GGA	CC
AGTGGCGGCGTAA	TGATCACGTCTGTG	CCCGCAGCCTCATC
ATCCT	GCTTATTTGAA	
CTCCACAACCGTCT	GCGGGCTTGAGTCT	CAGCTGCCCCAATT
TAAATAACAAACC	GTGA	С
GGCGGCTGCCCTG	CCCGGTGCCACTTT	CTGTGCCCCTTTGC
Т	GC	TG
CTGTGGAGTCGTCA	CGATTCACACGTGA	CCTCGCCTCGCTTT
GAATTCTTCAT	TTATTCAGCAAA	T
	AAGAC GGACTCACTGTCCC TCCTGAA GCGGTATCTGCTGC TTTGG CCGGCTAGGGTGG AAGAG GGGTAGGCGCAGG CA TGGCTGCTTTTTTAT GGCTTCAATT TCACCACCACCCCT ACGT AGTGGCGGCGTAA ATCCT CTCCACAACCGTCT TAAATAACAAACC GGCGGCTGCCCTG T CTGTGGAGTCGTCA	TGTCCGAGCAAGG GATGATATTG GCAGCGGGCAAGA AAGAC GGACTCACTGTCCC TCCTGAA GCGGTATCTGCTGC GCGGTATCTGCTGC TTTGG CCGGCTAGGGTGG AAGAG CCGGCTAGGGTGG CAGCTATCTGGAT CCGGCTAGGGTGG CAGCTATCTGGAT CCGGCTAGGGTGG CAGCTATCTCA GGGTAGCCCCC AAGAG CTATTC GGGTAGCCAGG CA TGGCTGCTTTTTTAT CCGCGTGCGAGTG CCGCGTGCGAGT CCGCGTGCGAA ATCCT CCACACCCCCT ACGT AGTGGCGCGTAA ATCCT CTCCACAACCGTCT TAAATAACAAACC GGCGCGCGCCAC CCGGTGCCACTTT TCGC CCCGGTGCCACTTT CCCCGGTGCCACTTT CCCCCCCTG CCCGGTGCCACTTT CCCCCCCTG CCCGGTGCCACTTT CCCCCCCTG CCCGGTGCCACTTT CCCCCGGTGCCACTTT CCCCGGTGCCACTTT CCCCCGGTGCCACTTT CCCCGGTGCCACTTT CCCCCGGTGCCACTTT CCCCCGGTGCCACTTT CCCCCGGTGCCACTTT CCCCCGGTGCCACTTT CCCCCGGTGCCACTTT CCCCCGGTGCCACTTT CCCCCGGTGCCACTTT CCCCCGGTGCCACTTT CCCCCGGTGCCACTTT CCCCGGTGCCACTTT CCCCCGGTGCCACTTT CCCCGGTGCCACTTT CCCCGGTGCCACTTT CCCCGGTGCCACTTT CCCCCGGTGCCACTTT CCCCCGGTGCCACTTT CCCCCCCCCC

Supplementary Table 4. Accession numbers of the NGS data. Related to Figures 4-6.

Sample name	DRA accession number	Sample ID	GEA acce	
Input gDNA (CIRA- seq)	DRA008734		SAMD0018020 8	E-GEAD- 324
Unmethylated CpGs (CIRA-seq)	DRA008734		SAMD0018020 9	E-GEAD- 324
Input chromatin (ChIP)	DRA012473		SAMD0039383 9	E-GEAD- 446
MOZ _{FL} (ChIP)	DRA008732		SAMD0018012 7	E-GEAD- 322
MEAF6 (ChIP)	DRA012473		SAMD0039384 4	E-GEAD- 446
ING4 (ChIP)	DRA012473		SAMD0039384 3	E-GEAD- 446
H3K14ac (ChIP)	DRA014291		SAMD0049558 0	E-GEAD- 497
RNAP2 non-P (ChIP)	DRA012473		SAMD0039384 6	E-GEAD- 446
RNAP2 Ser5-P (ChIP)	DRA012473		SAMD0039384 7	E-GEAD- 446
Vector-FLAG (ChIP)	DRA014290		SAMD0049557 4	E-GEAD- 498
MOZ _{WH1} -FLAG rep1(ChIP)	DRA014290		SAMD0049557 5	E-GEAD- 498
MOZ _{WH1} -FLAG rep2 (ChIP)	DRA014290		SAMD0049557 6	E-GEAD- 498
MOZ _{WH1-WH2-DPF} -FLAG (ChIP)	DRA014290		SAMD0049557 8	E-GEAD- 498
MOZ _{WH2-DPF} -FLAG rep1 (ChIP)	DRA014290		SAMD0049557 7	E-GEAD- 498
MOZ-TIF2-FLAG (ChIP)	DRA010562		SAMD0023783 4	E-GEAD- 381
Vector-FLAG rep2 (ChIP)	DRA015383		SAMD0056746 3	E-GEAD- 584
MOZ _{WH1} -FLAG rep3 (ChIP)	DRA015383		SAMD0056746 4	E-GEAD- 584
MORF _{WH1} -FLAG rep1(ChIP)	DRA015383		SAMD0056746 5	E-GEAD- 584
MORF _{WH1} -FLAG rep2(ChIP)	DRA015383		SAMD0056746 6	E-GEAD- 584

Supplementary Table 5. Cryo-EM data collection statistics. Related to Figure 8.

	$MORF_{WH1-WH2}$ -
	nucleosome - scFv
	(EMDB-27243)
Data collection and processing	
Magnification	56,000
Voltage (kV)	200
Electron exposure (e-/Ų)	43.2
Defocus range (μm)	0.8-1.8
Pixel size (Å)	0.91
Symmetry imposed	C1
Initial particle images (no.)	219,800
Final particle images (no.)	4,926
Map resolution (Å)	7.03
FSC threshold	0.143
Map resolution range (Å)	5.8-20