LETTER TO EDITOR



WILEY

Clinical characteristics and risk factors for mortality among inpatients with COVID-19 in Wuhan, China

Coronavirus disease 2019 (COVID-19) has become the worldwide pandemic. Currently, COVID-19, caused by the novel coronavirus, has had outbreaks in more than 213 countries and regions around the world and has caused many deaths. As of 24 April 2020, more than 2.62 million people had been diagnosed worldwide, of whom more than 180 000 died from the virus.¹

This cohort included COVID-19 patients admitted to The Central Hospital of Wuhan on 1 January 2020 to 15 February 2020. Of the 660 participating inpatients with COVID-19, 82 died and 578 were discharged. We comprehensively identified the clinical characteristics at the time of admission in nonsurvivors and patients who recovered from it (Table 1), and we reported several novel risk factors of mortality in COVID-19 patients (Figure 1A). Furthermore, dynamic changes in these major markers during hospitalization and the survival outcomes under different conditions were tracked (Figure 1B–E).

Consistent with the previous study, advanced age and high SOFA scores on admission were crucial risk factors for COVID-19 patient mortality.² Intriguingly, according to our multivariable logistic regression analysis, we discovered that a history of cerebral infarction, C-reactive protein (CRP) levels >0.6 mg/dL, and lactate dehydrogenase levels >245 U/L on admission significantly elevated the odds of in-hospital death (Figure 1A).

Our findings show that the history of cerebral infarction, an independent risk factor for inpatient death, caused a significantly poorer outcome for COVID-19 patients. The possible reasons are as follows. First, cerebral infarction may occur mainly in elderly people, and older age was a crucial predictor for COVID-19 mortality.² Second, when cerebral infarction impacts respiratory muscles and respiratory centers or causes hemiplegia, it may cause pulmonary conditions to deteriorate.³ Third, hypercoagulability is associated with cerebral infarction, and cerebral infarction patients may have preexisting hypercoagulability.⁴ Additionally, we observed higher D-dimer in nonsurvivors, suggesting that COVID-19 patients with cerebral infarction may have an aggravating hypercoagulable state, which could contribute to severe dysfunction. Therefore, cerebral infarction patients should recognize that there is a high possibility of severe illness in the event of viral infection and should follow strict precautions.

In nonsurvivors, baseline myocardial injury markers such as lactate dehydrogenase and creatine kinase-MB (CKMB) were elevated; CKMB increased rapidly from day 7 after admission, and lactate dehydrogenase was highest on day 3 after admission. Levels of kidney injury markers such as creatinine were clearly greater in death group, and blood urea nitrogen increased throughout the clinical course. Baseline alanine aminotransferase (ALT) was not quite different in the two groups, but the ALT of nonsurvivors increased rapidly from day 7 and was visibly higher on day 14. D-dimer was elevated in nonsurvivors compared to survivors, but no difference on day 14. As reported, severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) interacted with ACE2 receptors, which are expressed in in most organs of human; on this basis, it is reasonable to hypothesize the involvement and impairment of multiple organs.⁵

The number of white blood cell and neutrophil and CRP levels of die patients were greater than discharged patients throughout the hospital stay, and procalcitonin augmented rapidly begin with day 7, becoming notably larger in die patients by day 14. For nonsurvivors, lymphocyte counts decreased from admission to day 3 and remained low level. Compared with nonsurvivor, eosinophilic granulocyte counts were dramatically lower in surviving group by day 14. Patients with decreased CD4+ T cells (<30%) had dramatically worse outcomes than those with normal CD4+ T cells (<20%) was associated with poor outcomes. Furthermore, patients with higher C-reactive protein had significantly lower survival than those with normal C-reactive protein levels (Figure 1E).

The direct cytopathic effect of the virus and its ability to escape from the host's immune response are considered the main factors of viral disease progression and can even threaten life.⁶ Consistent with changes in complete blood counts in the

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.



TABLE 1 Clinical characteristics of survivors and nonsurvivors who were diagnosed with coronavirus disease 2019

		Survivor	Non-survivor	
Characteristics	Total (n = 660)	(n = 578)	(n = 82)	<i>P</i> -value
Median (IQR) age, years	55.0 (34.0-68.0)	54.0 (37.0-66.0)	71.0 (63.0-83.0)	<.0001
≤60	382 (57.9%)	364 (63.0%)	18 (22.0%)	<.0001
>60	278 (42.1%)	214 (37.0%)	64 (78.0%)	
Sex				<.0001
Female	365 (55.3%)	341 (59%)	24 (29.3%)	
Male	295 (44.7%)	237 (41%)	58 (70.7%)	
Comorbidity	326 (49.4%)	262 (45.3%)	64 (78.0%)	<.0001
Chronic obstructive lung disease	43 (6.5%)	32 (5.5%)	11 (13.4%)	.007
Hypertension	230 (34.8%)	177 (30.6%)	53 (64.6%)	<.0001
Diabetes	114 (17.3%)	93 (16.1%)	21 (25.6%)	.033
Myocardial infarction	67 (10.2%)	48 (8.3%)	19 (23.2%)	<.0001
Cerebral infarction	52 (7.9%)	29 (5.0%)	23 (28.0%)	<.0001
Kidney injury	36 (5.5%)	19 (3.3%)	17 (20.7%)	<.0001
Other	38 (5.8%)	28 (4.8%)	10 (12.2%)	.015
Symptoms at disease onset				
Fever	524 (79.4%)	455 (78.7)	69 (84.1%)	.256
Cough	431 (65.3%)	383 (58.5%)	48 (66.3%)	.169
Chest tightness	259 (39.2%)	225 (38.9%)	34 (41.5%)	.660
Thoracalgia	28 (4.2%)	28 (4.8%)	0 (0%)	.081
Myalgia	155 (23.5%)	139 (24.0%)	16 (19.5%)	.364
Fatigue	237 (23.5%)	206 (23.5%)	31 (23.5%)	.702
Diarrhea	53 (8.0%)	47 (8.1%)	6 (7.3%)	.800
Headache	42 (6.4%)	38 (6.6%)	4 (4.9%)	.556
APACHEII	2.5 (1.0-5.0)	2.0 (1.0-4.0)	10.0 (8.0-14.0)	<.0001
SOFA	1.0 (0.0-3.0)	1.0 (0.0-2.0)	5.0 (3.0-6.0)	<.0001
CURB-65	0.0 (0.0-1.0)	0.0 (0.0-1.0)	2.0 (1.0-2.0)	<.0001
0-1	549/642 (85.5%)	519/566 (91.7%)	30/76 (39.5%)	<.0001
2	72/642 (11.2%)	41/566 (7.2%)	31/76 (40.8%)	
3-5	21/642 (3.3%)	6/566 (1.1%)	15/7 (19.7%)	
Median (IQR) time from onset of symptom to hospital admission, days	7.0 (3.0-10.0)	7.0 (4.0-10.0)	6.0 (2.0-9.0)	.960
Evaluation of computed tomography imaging on hospital admission				<.0001
Normal	345/643 (53.6%)	319/567 (56.3%)	26/76 (34.2%)	
Medium	147/643 (22.9%)	138/567 (24.3%)	9/76 (11.8%)	
Severe	151/643 (23.5%)	110/567 (19.4%)	41/76 (54.0%)	
Progression of imaging				<.0001
None	307/579 (53.0%)	305/548 (55.7%)	2/31 (6.5%)	
Yes	272/579 (47.0%)	243/548 (44.3%)	29/31 (93.5%)	
Laboratory Findings				
White blood cell count, $\times 10^9$ per L	4.9 (3.8-6.6)	4.8 (3.8-6.2)	6.3 (4.0-9.7)	.001
<4	193/656 (29.4%)	173/576 (30.0%)	20/80 (25.0%)	<.0001
4-10	419/656 (63.9%)	376/576 (65.3%)	43/80 (53.7%)	
>10	44/656 (6.7%)	27/576 (4.7%)	17/80 (21.3%)	

(Continues)

TABLE 1 (Continued)

Characteristics	Total (n = 660)	Survivor (n = 578)	Non-survivor $(n = 82)$	<i>P</i> -value
Neutrophil count. $\times 10^9$ per L	3.4 (2.3-4.9)	3.3 (2.2-4.5)	4.7 (3.1-8.7)	<.0001
<1.8	99/656 (15.1%)	89/576 (15.4%)	10/80 (12.5%)	<.0001
1.8-6.3	467/565 (71.2%)	425/576 (73.8%)	42/80 (52.5%)	
>6.3	90/565 (13.7%)	62/576 (10.8%)	28/80 (35.0%)	
Lymphocyte count, $\times 10^9$ per L	1.0 (0.7-1.4)	1.0 (0.7-1.4)	0.6 (0.4-0.9)	<.0001
≥0.8	430/656 (65.5%)	403/576 (70.0%)	27/80 (33.7%)	<.0001
<0.8	226/656 (34.5%)	173/576 (30.0%)	53/80 (66.3%)	
CD4+/CD8+ ratio	1.5 (1.1-2.4)	1.6 (1.1-2.4)	1.5 (0.9-2.4)	.463
<0.95	27/154 (17.5%)	22/132 (16.7%)	5/22 (22.7%)	.742
0.95-2.1	80/154 (51.9%)	70/132 (53.0%)	10/22 (45.5%)	
>2.1	47/154 (30.6%)	40/132 (30.3%)	7/22 (31.8%)	
CD4+ T cells, %	40.0 (20.0-60.0)	50.0 (30.0-70.0)	20.0 (20.0-30.0)	<.0001
<30	50/154 (32.5%)	33/132 (25.0%)	17/22 (77.3%)	<.0001
30-46	34/154 (22.1%)	32/132 (24.2%)	2/22 (9.1%)	
>46	70/154 (45.4%)	67/132 (50.8%)	3/22 (13.6%)	
CD8+ T cells, %	30.0 (20.0-40.0)	30.0 (20.0-40.0)	10.0 (10.0-30.0)	.001
<20	58/154 (37.7%)	43/132 (32.6%)	15/22 (68.2%)	.006
20-33	43/154 (27.9%)	40/132 (30.3%)	3/22 (13.6%)	
>33	53/154 (34.4%)	49/132 (37.1%)	4/22 (18.2%)	
Eosinophilic granulocyte count, $\times 10^9$ per L	0.01 (0.00-0.04)	0.01 (0.00-0.04)	0.00 (0.00-0.02)	.021
<0.02	411/846 (63.4%)	356/574 (62.0%)	55/74 (74.3%)	.039
0.02-0.52	237/648 (36.6%)	218/574 (38.0%)	19/74 (25.7%)	
Basophilic granulocyte count, ×109 per L	0.01 (0.01-0.02)	0.01 (0.01-0.02)	0.01 (0.01-0.02)	.824
0-0.06	641/647 (99.1%)	568/573 (99.1%)	73/74 (98.6%)	1.000
>0.06	6/647 (0.9%)	5/573 (0.9%)	1/74 (1.4%)	
Hemoglobin, g/dL	128.0 (118.0-139.0)	128.0 (120.0-140.0)	122.0 (106.3-137.8)	.002
Platelet count, $\times 10^9$ per L	178.0	182.0	130.0	<.0001
	(135.3-225.8)	(143.0-231.8)	(106.8-183.8)	
<125	118/656 (18.0%)	84/576 (14.6%)	34/80 (42.5%)	<.0001
125-350	507/656 (77.3%)	461/576 (80.0%)	46/80 (57.5%)	
>350	31/656 (4.7%)	31/576 (5.4%)	0/80 (0%)	
C-reactive protein, mg/dL	1.9 (0.5-4.6)	1.5 (0.4-4.1)	5.4 (2.7-8.7)	<.0001
0-0.6	190/640 (29.7%)	186/564 (33.0%)	4/76 (5.3%)	<.0001
>0.6	450/640 (70.3%)	378/564 (67.0%)	72/76 (94.7%)	
Procalcitonin, ng/mL	0.1 (0.0-0.1)	0.1 (0.0-0.1)	0.2 (0.1-0.5)	<.0001
<0.1	444/600 (74.0%)	421/531 (79.3%)	23/69 (33.3%)	<.0001
≥0.1	156/600 (26.0%)	110/531 (20.7%)	46/69 (66.7%)	
PaO ₂ /FiO ₂ , mmHg	346.0 (243.0-510.0)	402.0 (276.0-520.0)	142.0 (92.5-225.8)	<.0001
<100	25/647 (3.9%)	4/567 (0.7%)	21/80 (26.2%)	<.0001
100-300	224/647 (34.6%)	176/567 (31.0%)	48/80 (60.0%)	
>300	398/647 (61.5%)	387/567 (68.3%)	11/80 (13.8%)	
Lactate, mmol/L	1.2 (0.8-1.9)	1.2 (0.8-1.8)	2.1 (1.4-2.8)	<.0001
≥1.5	259/644 (40.2%)	202/565 (35.8%)	57/79 (72.2%)	<.0001

(Continues)

TABLE 1 (Continued)

		Survivor	Non-survivor	
Characteristics	Total $(n = 660)$	(n = 578)	(n = 82)	<i>P</i> -value
Blood urea nitrogen, mmol/L	4.2 (3.3-5.6)	4.0 (3.2-5.2)	6.5 (4.8-9.9)	<.0001
<2.9	84/653 (12.9%)	82/575 (14.2%)	2/78 (2.5%)	<.0001
2.9-8.2	521/653 (79.8%)	469/575 (81.6%)	52/78 (66.7%)	
>8.2	48/653 (7.3%)	24/575 (4.2%)	24/78 (30.8%)	
Creatinine, μ mol/L	65.1 (52.0-79.5)	63.3 (51.3-77.2)	78.3 (60.0-116.4)	<.0001
<57	227/653 (34.8%)	213/575 (37.0%)	14/78 (18.0%)	<.0001
57-111	389/653 (59.6%)	341/575 (59.3%)	48/78 (61.5%)	
>111	37/653 (5.6%)	21/575 (3.7%)	16/78 (20.5%)	
Total bilirubin, μ mol/L	8.7 (6.4-12.0)	8.8 (6.5-11.6)	8.4 (5.6-14.1)	.504
ALT, U/L	20.0 (13.2-32.5)	19.7 (13.2-33.0)	21.0 (13.5-30.4)	.937
>50	67/648 (10.3%)	64/574 (11.1%)	3/74 (4.1%)	.059
FIB, g/L	3.0 (2.5-3.5)	2.9 (2.5-3.4)	3.5 (3.0-3.8)	<.0001
<2	21/625 (3.4%)	20/550 (3.6%)	1/75 (1.3%)	.001
2-4	542/625 (86.7%)	484/550 (88.0%)	58/75 (77.3%)	
>4	62/625 (9.9%)	46/550 (8.4%)	16/75 (21.4%)	
D-dimer, μ g/L	0.6 (0.3-1.4)	0.5 (0.3-1.1)	1.4 (0.7-5.2)	<.0001
≤1	435/632 (68.8%)	408/563 (72.5%)	27/69 (39.1%)	<.0001
>1	197/632 (31.2%)	155/563 (27.5%)	42/69 (60.9%)	
Lactate dehydrogenase, U/L	191.0	185.5	292.0	<.0001
	(153.0-254.3)	(150.0-237.0)	(211.8-474.8)	
>245	161/598 (26.9%)	116/524 (22.1%)	45/74 (60.8%)	<.0001
Creatine kinase, U/L	75.5 (47.0-133.3)	73.0 (45.8-122.0)	144.3 (65.8-256.2)	<.0001
>190	88/598 (14.7%)	60/524 (11.5%)	28/74 (37.8%)	<.0001
CKMB, U/L	8.0 (6.0-11.9)	7.4 (6.0-11.0)	11.1 (7.8-17.0)	<.0001
>24	25/608 (4.1%)	13/533 (2.4%)	12/75 (16.0%)	<.0001
IL-6, pg/mL	4.7 (2.3-19.0)	3.9 (2.2-9.6)	43.8 (20.1-62.6)	<.0001
HDL-C, mmol/L	1.0 (0.8-1.2)	1.0 (0.8-1.2)	0.9 (0.7-1.1)	.009
LDL-C, mmol/L	2.2 (1.7-2.7)	2.2 (1.7-2.7)	2.0 (1.5-2.7)	.113
Total triglyceride, mmol/L	1.0 (0.8-1.4)	1.0 (0.8-1.4)	1.1 (0.7-1.4)	.881
Total cholesterol, mmol/L	3.8 (3.2-4.4)	3.8 (3.2-4.5)	3.4 (2.9-4.2)	.042
Nonesterified fatty acid, mmol/L	0.5 (0.3-0.6)	0.5 (0.3-0.6)	0.6 (0.4-0.7)	.040
HbA1C, %	7.4 (6.5-8.8)	7.3 (6.4-9.1)	7.4 (6.9-8.0)	.912
>6	63/75 (84.0%)	50/61 (82.0%)	13/14 (92.9%)	.550
Glucose, mmol/L	5.7 (5.0-7.4)	5.6 (4.9-7.0)	7.1 (5.7-9.0)	<.0001
≤6.1	381/653 (58.3%)	358/575 (62.3%)	23/78 (29.5%)	<.0001
>6.1	272/653 (41.7%)	217/575 (37.7%)	55/78 (70.5%)	
Treatments				
Quinotone	420 (63.6%)	368 (63.7%)	52 (63.4%)	.964
Cephalosporins	293 (44.4%)	241 (41.7%)	52 (63.4%)	<.0001
Ribavirin	370 (56.1%)	314 (54.3%)	56 (68.3%)	.017
Oseltamivir	336 (50.9%)	295 (51.0%)	41 (50.0%)	.860
Arbidol	135 (20.5%)	130 (22.5%)	5 (6.1%)	.001
Kaletra	29 (4.4%)	28 (4.8%)	1 (1.2%)	.226
Early corticosteroids	184 (27.9%)	154 (26.6%)	30 (36.6%)	.060
		. ,	. ,	

(Continues)



TABLE 1 (Continued)

		Survivor	Non-survivor	
Characteristics	Total (n = 660)	(n = 578)	(n = 82)	<i>P</i> -value
Early intravenous immunoglobulin	160 (24.2%)	143 (24.7%)	17 (20.7%)	.428
Thymosin	127 (19.2%)	110 (19.2%)	17 (20.7%)	.715
Heparin	213 (32.2%)	169 (29.2%)	44 (53.7%)	<.0001
Median (IQR) time from hospital admission to high-flow nasal cannula oxygen therapy, days	4.0 (1.0-10.0)	4.0 (1.0-7.0)	4.0 (1.0-13.0)	.369
Mechanical ventilation	104 (15.8%)	44 (7.6%)	60 (74.1%)	<.0001
Cause of death				
Respiratory failure	63 (76.9%)	0	63 (76.9%)	
Pulmonary embolism and respiratory failure	1 (1.2%)	0	1 (1.2%)	
Pulmonary infection and respiratory failure	1 (1.2%)	0	1 (1.2%)	
Heart failure	2 (2.4%)	0	2 (2.4%)	
Cardiac arrest	6 (7.3%)	0	6 (7.3%)	
Other	9 (11.0)	0	9 (11.0)	

Note. P-values were calculated by Mann-Whitney U test, χ^2 test, or Fisher's exact test; bold if statistically significant, *P* < .05.

Abbreviations: ALT, alanine aminotransferase; APACHEII, Acute Physiology and Chronic Health Evaluation; CKMB, creatine kinase isoenzyme-MB; FIB, fibrinogen; HbA1C, hemoglobin A1C; HDL-C, high-density lipid cholesterol; IL-6, interleukin-6; IQR, interquartile range; LDL-C, low-density lipid cholesterol; SOFA, Sequential Organ Failure Assessment.



FIGURE 1 Temporal changes in laboratory markers and the death risk factors of COVID-19. A, Association of risk factors of COVID-19. B and C, Temporal changes in lactate dehydrogenase, creatine kinase-MB (CKMB), creatinine, neutrophil count, blood urea nitrogen, alanine aminotransferase, d-dimer, CRP, procalcitonin, white blood cell count, lymphocyte count, and eosinophilic granulocyte. D and E, Cumulative incidence for inpatients with COVID-19 sub-grouped by chronic obstructive pulmonary disease, hypertension, myocardial infarction, cerebral infarction, lymphocyte, CD4+ T cell, CD8+ T cell, and C-reactive protein

previous study.² our study found that the white blood count and neutrophil counts of the nonsurvivor group during hospitalization were significantly higher and their lymphocyte counts were significantly lower than those of the survivor group, indicating that the nonsurviving patients suffered more serious bacterial or fungal infections during hospitalization. Previous studies have shown that the proportions and absolute counts of lymphocyte populations in severe COVID-19 patients are reduced, especially CD4+ T cells and CD8+ T cells.⁷ Another study also showed that SARS-CoV-2 infection could cause a decrease in the abundance and function of peripheral blood T cells and NK cells.⁸ Consistent with previous research, the present study showed that quantity of CD4+ T cell, lymphocyte, and CD8+ T cell in nonsurviving group was less than that in surviving patients. More interestingly, we found that these three variables were prognostic factors in COVID-19 patients, helping to predict the severity of their condition. These results may have some limitations: bacterial infection or repeated infections in COVID-19 patients will also affect immune indicators. Nonetheless, our findings and those of other researchers indicate that SARS-CoV-2 infection may mainly affect T cells in the lymph, causing cytokine storms in the body; these severe reactions would reduce the body's immunity and trigger multiple organ failure, ultimately leading to the death of the patient.

In conclusion, advanced age, increased SOFA scores a, history of cerebral infarction, CRP larger than 0.6 mg/dL, and lactate dehydrogenase greater than 245 U/L at admission were independent risk factors for in-hospital death in COVID-19. Moreover, these data highlight the greater multiple organ involvement and greater changes in inflammation and immunity markers among nonsurvivors during hospitalization. More intensive attention should be paid to patients with these risk factors, in case of rapid deterioration and bad prognosis.

ACKNOWLEDGMENT

The letter is granted by the National Natural Science Foundation of China (No. 81471781).

DISCLOSURE

The authors declare no conflict of interest.

Fuyang Chen¹ Wenwu Sun¹ Shengrong Sun² Zhiyu Li² Zhong Wang² Li Yu¹

¹Intensive Care Unit, Central Hospital of Wuhan, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China ²Department of Breast and Thyroid Surgery, Renmin Hospital of Wuhan University, Wuhan, China

Correspondence

Zhiyu Li, Department of Breast and Thyroid Surgery, Renmin Hospital of Wuhan University, 99 Zhang Zhidong Road, Wuhan 430060, Hubei Province, China. Email: lizhiyu@whu.edu.cn Zhong Wang, Department of Breast and Thyroid Surgery, Renmin Hospital of Wuhan University, 99 Zhang Zhidong Road, Wuhan 430060, Hubei Province, China. Email: zhongwangchn@whu.edu.cn Li Yu, Intensive Care Unit, Central Hospital of Wuhan, Tongji Medical College, Huazhong University of Science and Technology, 26 Shengli Street, Wuhan 430060, Hubei Province, China. Email: yuliwhzxyy@163.com

Fuyang Chen and Wenwu Sun contributed equally to this work.

ORCID

Zhong Wang (b) https://orcid.org/0000-0001-7072-2410

REFERENCES

- WHO. Coronavirus disease (COVID-19) outbreak situation by WHO. https://www.who.int/emergencies/diseases/novel-corona virus-2019
- Zhou F, Yu T, Du R, et al. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *Lancet*. 2020;395(10229):1054-1062.
- Menezes KK, Nascimento LR, Avelino PR, Alvarenga MTM, Teixeira-Salmela LF. Efficacy of interventions to improve respiratory function after stroke. *Respir Care*. 2018;63(7):920-933.
- Levine SR. Hypercoagulable states and stroke: a selective review. CNS Spectr. 2005;10(7):567-578.
- Abassi ZA, Skorecki K, Heyman SN, Kinaneh S, Armaly Z. Covid-19 infection and mortality—a physiologist's perspective enlightening clinical features and plausible interventional strategies. *Am J Physiol Lung Cell Mol Physiol.* 2020. https://doi.org/10.1152/ajplung.00097.2020
- Channappanavar R, Perlman S. Pathogenic human coronavirus infections: causes and consequences of cytokine storm and immunopathology. *Semin Immunopathol.* 2017;39(5):529-539.
- Chen G, Wu D, Guo W, et al. Clinical and immunological features of severe and moderate coronavirus disease 2019. J Clin Invest. 2020. https://doi.org/10.1172/JCI137244
- Zheng M, Gao Y, Wang G, et al. Functional exhaustion of antiviral lymphocytes in COVID-19 patients. *Cell Mol Immunol.* 2020. https://doi.org/10.1038/s41423-020-0402-