

Article

Cytotoxic Flavonoids from the Leaves and Twigs of *Murraya tetramera*

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Abstract: Cytotoxic flavonoids of *Murraya tetramera* were investigated in this study. A novel flavonoid and twelve known flavonoids, including seven flavones (1–7), three flavanones (8–10), and three chalcones (11–13) were isolated from the leaves and twigs of *Murraya tetramera*. Chemical structures were elucidated by NMR combined with MS spectral analysis, and the new compound (6) was confirmed as 3',5'-dihydroxy-5,6,7,4'-tetramethoxyflavone. Furthermore, all the isolated flavonoids were evaluated for their cytotoxicities against murine melanoma cells (B16), and human breast cancer cells (MDA-MB-231) by CCK-8 assay. Among them, compounds 7, 13, and 5 exhibited potent cytotoxic activities against B16 cell lines (IC₅₀ = 3.87, 7.00 and 8.66 μg/mL, respectively). Compounds 5, 13, and 12 displayed potent cytotoxicities against MDA-MB-231 cell lines (IC₅₀ = 3.80, 5.95 and 7.89 μg/mL, respectively). According to the correlation of the structure and activity analysis, 5-hydroxyl and 8-methoxyl substituents of the flavone, 8-methoxyl substituent of the flavanone, and 3',5'-methoxyl substituents of the chalcone could be critical factors of the high cytotoxicity. The results indicated that the active flavonoids have potential to be developed as leading compounds for treating cancers.

Keywords: *Murraya tetramera*; flavonoid; B16; MDA-MB-231; cytotoxicity



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1. Introduction

Cancer is a leading cause of death among most countries in the 21st century, and the incidence and mortality are rapidly growing worldwide [1]. Melanoma of the skin among males and breast cancer among females are two of most prevalent cancers in 2019 [2]. Melanoma arises from epidermal melanocytes and induces 80% of the dermatological cancer-related deaths [3]. Breast cancer is the second major cause of cancer deaths in female worldwide [4]. Serious side effects and drug resistance caused by conventional cancer treatment of chemotherapy and radiotherapy remain the major problems during the treatment [5,6]. Accordingly, plant secondary metabolites have been attracting more attention in drug development owing to multiple factors, and the various compounds discovered in plants made them as a rich source of cancer drug candidates [7–10]. Thousands of flavonoids have been isolated from stems, flowers, fruits, roots, and barks of the plants, moreover, many effective cytotoxic flavonoids from various plants were considered as potential leading compounds for the development of anticancer drugs [8,11–13].

The genus *Murraya* (family Rutaceae) is a common plant source of polymethoxylated and polyhydroxylated flavonoids [14]. *Murraya tetramera* Huang (*M. tetramera*) is a small tree that is widely distributed in Guangxi and Yunnan provinces of China. The folk medicine has been applied for treating coughs, bronchitis, rheumatism, asthma, and traumatic injury, etc. [15,16]. *M. tetramera* contains various flavonoids, coumarins, alkaloids,

and sesquiterpenes [17–20]. Some of the isolated compounds have exhibited significant cytotoxic effects [20–22].

To explore potent cytotoxic flavonoids from *M. tetramera* as potential leading compounds for treating cancers and make comprehensive utilization of its natural resources, a phytochemical investigation was carried out, and the cytotoxicities were evaluated against murine melanoma cells (B16) and human breast cancer cells (MDA-MB-231) by CCK-8 assay.

2. Results and Discussions

2.1. Flavonoids Isolated from *M. tetramera*

A novel flavonoid and twelve known flavonoids, including seven flavones (1–7), three flavanones (8–10) and three chalcones (11–13) were isolated from the leaves and twigs of *M. tetramera*. The new one was identified as 3',5'-dihydroxy-5,6,7,4'-tetramethoxyflavone (6) and the others were 3',4',5,5',7-pentamethoxyflavone (1) [23], 5,6,7,3',4',5'-hexamethoxyflavone (2) [24,25], nobiletin (3) [26], 7-hydroxy-3',4',5,5'-tetra methoxyflavone (4) [27], 5-hydroxy-6,7,3',4',5'-pentamethoxyflavone (5) [28], 5,3',5'-trihydroxy-6,7,4'-trimethoxyflavone (7) [29], 5,7,3',4',5'-pentamethoxyflavanone (8) [30], 3',4',5',5,7,8-hexamethoxy flavanone (9) [31,32], 5,6,7,3',4',5'-hexamethoxyflavanone (10) [25], 2'-hydroxy-3,4,5,4',6'-pentamethoxychalcone (11) [23,33], 2'-hydroxy-3,4,5,3',4',6'-hexamethoxychalcone (12) [33], and 2'-hydroxy-3,4,5,4',5',6'-hexamethoxychalcone (13) [34]. Their structures were shown in Figure 1. The ^1H and ^{13}C -NMR data of the twelve known flavonoids were listed in the Supplementary Materials.

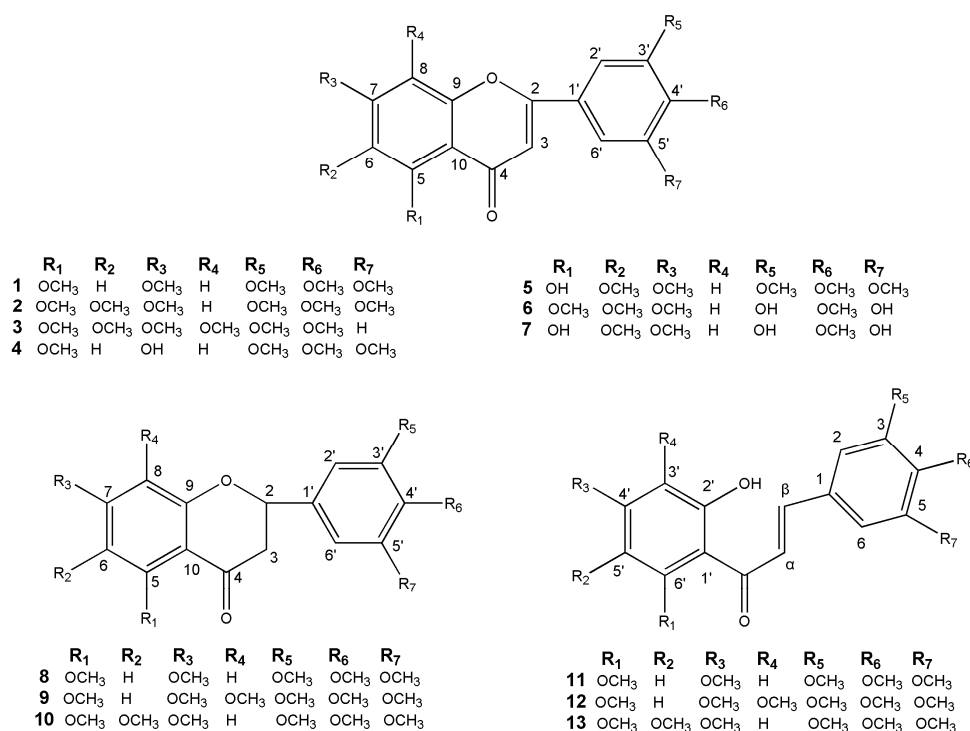


Figure 1. Chemical structures of flavonoids 1–13.

2.2. Structure Elucidation of the New Flavone

Compound 6 was collected as yellow needles. The molecular formula of $\text{C}_{19}\text{H}_{18}\text{O}_8$ was deduced from the peak at m/z 375.1075 $[\text{M}+\text{H}]^+$ (calculated for $\text{C}_{19}\text{H}_{19}\text{O}_8$, 375.1074) in the HR-ESI-MS and the 19 carbon resonances in the ^{13}C -NMR data. The ^{13}C -NMR exhibited the typical flavone signals at δ_{C} 176.0 (C-4), δ_{C} 161.0 (C-2) and δ_{C} 107.2 (C-3). The ^1H -NMR displayed a typical flavone H-3 signal at δ_{H} 6.45 (1H, s), a characteristic flavone signal of H-2' and 6' at δ_{H} 7.00 (2H, s), and an aromatic proton at δ_{H} 7.10 (1H, s).

Moreover, the $^1\text{H-NMR}$ exhibited the existence of two hydroxy protons at δ_{H} 9.51 (2H, s), and four methoxyl peaks at δ_{H} 3.96, 3.80, 3.77 and 3.76 (each 3H, s). The HMBC displayed correlations arising from H-8 to C-10/C-6/C-9/C-7, from H-2',6' to C-1'/C-4'/C-3',5'/C-2, from H-3 to C-10/C-1'/C-2/C-4, from 3',5'-OH to C-2',6' / , C-4'/C-3',5', from 7-OCH₃, 5-OCH₃, 6-OCH₃ and 4'-OCH₃ to C-7, C-5, C-6 and C-4', respectively. The key HMBC correlations were indicated in Figure 2. Accordingly, compound 6 was deduced as 3',5'-dihydroxy-5,6,7,4'-tetramethoxyflavone. The ^1H and $^{13}\text{C-NMR}$ data were listed in Table 1. All spectra are available in the Supplementary Materials.

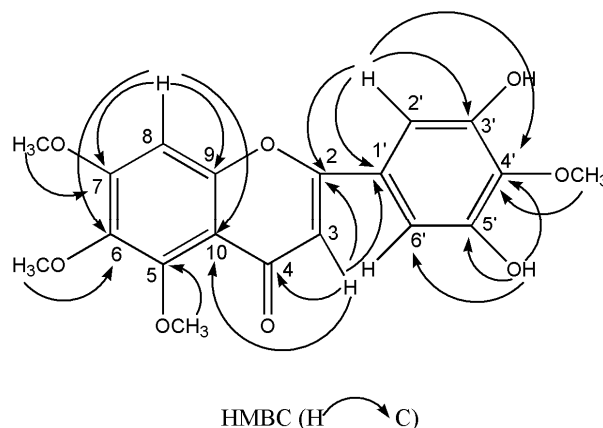


Figure 2. Key HMBC correlations of compound 6.

Table 1. ^1H and $^{13}\text{C-NMR}$ data of flavone 6 in DMSO- d_6 .

Position	δ_{H} (ppm)	δ_{C} (ppm)
2		161.0
3	6.45, s	107.2
4		176.0
5		152.1
6		140.3
7		158.0
8	7.10, s	97.6
9		154.4
10		112.5
1'		126.4
2'	7.00, s	105.9
3'		151.6
4'		138.9
5'		151.6
6'	7.00, s	105.9
5-OCH ₃	3.80, s	62.3
6-OCH ₃	3.77, s	61.5
7-OCH ₃	3.96, s	56.9
4'-OCH ₃	3.76, s	60.3
3', 5'-OH	9.51, s	

2.3. Cytotoxicities of Isolated Flavonoids

Flavonoids 1–13 were evaluated for their cytotoxicities against B16 and MDA-MB-231 cell lines by CCK-8 assay and the results were displayed in Table 2. Among them, compounds 7, 13, and 5 exhibited potent cytotoxic activities against B16 cell lines (IC_{50} = 3.87, 7.00, and 8.66 $\mu\text{g}/\text{mL}$, respectively). Compounds 5, 13, and 12 displayed potent cytotoxicities against MDA-MB-231 cell lines (IC_{50} = 3.80, 5.95 and 7.89 $\mu\text{g}/\text{mL}$, respectively). However, flavonoids 1, 6 and 11 showed weak anticancer efficacy against the two tested tumor cell lines (IC_{50} > 100 $\mu\text{g}/\text{mL}$).

The diverse cytotoxicities might be attributed to the different substituents of the flavonoids. Among flavones 1–7, flavones 5, 7, and 3 exhibited higher cytotoxicities against B16 cells ($IC_{50} = 8.66, 3.87, \text{ and } 11.18 \mu\text{g/mL}$) and MDA-MB-231 cells ($IC_{50} = 3.80, 14.93, \text{ and } 23.46 \mu\text{g/mL}$) than others. Thus, 5-hydroxyl and 8-methoxyl substituents of the flavone were essential for high cytotoxicity, which corresponds to the state of literature [8,35]. In addition, compared with flavone 6, flavone 2 showed a higher cytotoxicity against B16 and MDA-MB-231 cells with IC_{50} values of 14.74 and 34.19 $\mu\text{g/mL}$. Therefore, if the methoxy substituents in position 3' and 4' of flavone was substituted by hydroxyl substituents, as found in flavone 6, the cytotoxicity was significantly reduced. Among the flavanones 8–10, flavanone 9 exhibited the highest cytotoxicity against B16 and MDA-MB-231 cells ($IC_{50} = 12.76 \text{ and } 16.02 \mu\text{g/mL}$). Thus, 8-methoxyl substituent of the flavanone could be a critical factor of the cytotoxic activity, which corresponds to the state of literature [35]. Among chalcones 11–13, chalcones 12 and 13 exhibited higher cytotoxicities against B16 cells ($IC_{50} = 11.53 \text{ and } 7.00 \mu\text{g/mL}$) and MDA-MB-231 cells ($IC_{50} = 7.89 \text{ and } 5.95 \mu\text{g/mL}$) compared with chalcone 11. Hence, 3' and 5'-methoxyl substituents of the chalcone could be a major factor of the cytotoxic activity. Overall, according to the correlation of the structure and activity analysis, the position of methoxy and hydroxyl substituents in the flavonoids may be the major factors of the anticancer efficacy. Further investigation is essential to clarify the structure-active relationships.

Table 2. Cytotoxicities of flavonoids 1–13 from *Murraya tetramera*.

Compound	$IC_{50} \pm SD (\mu\text{g/mL})$	
	B16	MDA-MB-231
1	>100	>100
2	14.74 \pm 3.97	34.19 \pm 3.38
3	11.18 \pm 2.75	23.46 \pm 2.95
4	14.97 \pm 1.96	>100
5	8.66 \pm 1.80	3.80 \pm 1.49
6	>100	>100
7	3.87 \pm 0.68	14.93 \pm 2.71
8	13.03 \pm 1.19	26.46 \pm 2.53
9	12.76 \pm 3.38	16.02 \pm 1.12
10	23.55 \pm 3.51	25.29 \pm 3.84
11	>100	>100
12	11.53 \pm 1.61	7.89 \pm 1.71
13	7.00 \pm 0.64	5.95 \pm 0.65
DOX ¹	0.51 \pm 0.01	2.02 \pm 0.65

¹ Doxorubicin hydrochloride (positive control).

3. Materials and Methods

3.1. General Information

The NMR spectrometer (Bruker Avance III, Bruker, Karlsruhe, Germany) was used to record the NMR spectra at 500 MHz (^1H) and at 125 MHz (^{13}C). The mass spectrometer (Bruker Q-TOF, Bruker, Karlsruhe, Germany) was used to measure the HR-ESI-MS. Preparative HPLC was carried out using a Rainbow Kromasil-C₁₈ column (10 \times 250 mm, 10 μm) on a Waters Delta Prep 4000 instrument with a dual λ absorbance detector (Waters 2487, Waters, Milford, USA). MCI GEL CHP20P of 75–150 μm (Kaiteki Company, Tokyo, Japan) was selected for column chromatography. Silica gel G plates were used for TLC analysis (Qingdao Haiyang Chemical Co., Ltd., Qingdao, China). The deuterated DMSO-*d*₆ and CDCl₃ were supplied by Cambridge Isotope Laboratories, Inc. (Andover, USA). DMEM, RPMI 1640 and fetal bovine serum were supplied by Gibco Inc. (New York, USA). Penicillin and streptomycin were provided by Solarbio science & technology Co., Ltd. (Beijing, China). CCK-8 reagent was obtained from Beyotime Biotechnology (Shanghai, China). All the analytical solvents of analytical grade were supplied by Beijing Chemical Plant (Beijing, China).

3.2. Plant Material

The leaves and twigs of *M. tetramera* were harvested at Xishuangbanna, Yunnan Province, China in May 2014 and were identified by Dr. Liu, Q.R. (College of Life Sciences, Beijing Normal University, Beijing, China). The certificate specimen (BNU-CMH-Dushushan-2014-05-025-001) was stored at the Herbarium of Faculty of Geographical Science, Beijing Normal University.

3.3. Extraction and Isolation

The methanol extract of leaves and twigs of *M. tetramera* was obtained from our previous study and 90 fractions were received from the methanol extract by eluting with a stepwise gradient of PE/EtOAc and CHCl₃/CH₃OH [36]. Fr. 55–57 (4.27 g), Fr. 59–60 (3.77 g), Fr. 66–67 (1.96 g), Fr. 68–69 (2.26 g) and Fr. 75 (1.51 g) were separated by MCI column chromatography with a mobile phase of EtOH-H₂O (3:7, 5:5, 7:3 and EtOH), and then further purified by preparative HPLC using a stepwise gradient of MeOH-H₂O (2:8→MeOH) to obtain flavone **1** (20 mg, 0.0008% yield), flavone **2** (150 mg, 0.006% yield), flavone **3** (60 mg, 0.0024% yield), flavone **4** (9.5 mg, 0.0004% yield), flavone **5** (2.1 mg, 0.00008% yield), flavone **6** (2.1 mg, 0.00008% yield), flavone **7** (2.8 mg, 0.0001% yield), flavanone **8** (20 mg, 0.0008% yield), flavanone **9** (200 mg, 0.008% yield), flavanone **10** (15 mg, 0.0006% yield), chalcone **11** (6.2 mg, 0.0002% yield), chalcone **12** (50 mg, 0.002% yield) and chalcone **13** (45 mg, 0.0018% yield), respectively. The compounds were stored at 4 °C in a refrigerator for subsequent experiments.

3.4. Cytotoxicity Assay

The cytotoxicities of flavonoids **1–13** were determined by the standard CCK-8 assay [20,37]. B16 (Number: GDC0039) were originally provided by China Center for Type Culture Collection (Wuhan, China) and MDA-MB-231 (Number: CL0208) were obtained from the Fenghui Biotechnology Co., Ltd. (Changsha, China). Doxorubicin hydrochloride (DOX), the positive control, was purchased from Dalian Meilun Biotechnology Co., Ltd. (Dalian, China). B16 cells were cultured in RPMI 1640 medium and MDA-MB-231 cells were cultured in DMEM medium. The medium supplemented with 10% fetal bovine serum (Gibco Inc.), 100 U/mL penicillin and 0.1 mg/mL streptomycin. The tested cell lines were incubated at 37 °C, 5% CO₂ and 90% humidity in the CO₂ incubator (Binder, Tuttlingen, Germany). Firstly, 100 µL of the cell suspension was seeded into each well of 96-well plates (6 × 10³ per well), and then incubated for 12–24 h to allow cellular attachment. After removing the medium, fresh medium containing seven concentrations of test compounds was added into cultured cells of 100 µL per well and incubated for 48 h. Secondly, 10 µL CCK-8 reagent was added into each well and placed in a CO₂ incubator for 1 h. Finally, the absorbance was recorded using a microplate reader (Bio-Rad, Hercules, CA, USA) at 450 nm. The 50% inhibitory concentration (IC₅₀) values were calculated using Probit analysis (SPSS V20.0).

4. Conclusions

A novel flavonoid and twelve known flavonoids, including seven flavones (**1–7**), three flavanones (**8–10**), and three chalcones (**11–13**) were isolated from the leaves and twigs of *M. tetramera*. The novel one (compound **6**) was identified as 3',5'-dihydroxy-5,6,7,4'-tetramethoxyflavone. Results of cytotoxicity assay indicated that flavones **5** and **7** with 5-hydroxyl substituent, flavones **3** and flavanone **9** with 8-methoxyl substituent, chalcone **12** with 3'-methoxyl substituent and chalcone **13** with 5'-methoxyl substituent exhibited significant cytotoxic activities against B16 and MDA-MB-231 cell lines. According to the correlation of the structure and activity analysis, the position of methoxy and hydroxyl substituents in the flavonoids were the major factors of the high anticancer efficacy. The results indicated that the active flavonoids have potential to be developed as leading compounds for treating cancers.

Supplementary Materials: The following are available online, Figure S1: $^1\text{H-NMR}$ spectrum of compound 6, Figure S2: $^{13}\text{C-NMR}$ spectrum of compound 6, Figure S3: HMBC spectrum of compound 6, Figure S4: HR-ESI-MS spectrum of compound 6, Table S1: $^1\text{H-NMR}$ data of the twelve known flavonoids, Table S2: $^{13}\text{C-NMR}$ data of the twelve known flavonoids.

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Sample Availability: Samples of the compounds are available from the authors.

References

1. Bray, F.; Ferlay, J.; Soerjomataram, I.; Siegel, R.L.; Torre, L.A.; Jemal, A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *Ca-Cancer J. Clin.* **2018**, *68*, 394–424. [[CrossRef](#)] [[PubMed](#)]
2. Miller, K.D.; Nogueira, L.; Mariotto, A.B.; Rowland, J.H.; Yabroff, K.R.; Alfano, C.M.; Jemal, A.; Kramer, J.L.; Siegel, R.L. Cancer treatment and survivorship statistics, 2019. *Ca-Cancer J. Clin.* **2019**, *69*, 363–385. [[CrossRef](#)] [[PubMed](#)]
3. Li, B.W.; Zhang, X.L.; Lu, Y.; Zhao, L.Y.; Guo, Y.X.; Guo, S.S.; Kang, Q.Z.; Liu, J.J.; Dai, L.P.; Zhang, L.G.; et al. Protein 4.1R affects photodynamic therapy for B16 melanoma by regulating the transport of 5-aminolevulinic acid. *Exp. Cell Res.* **2021**, *399*, 112465. [[CrossRef](#)]
4. Han, B.; Sha, L.J.; Yu, X.M.; Yang, M.; Cao, Y.; Zhao, J. Identification of dual therapeutic targets assisted by in situ automatous DNA assembly for combined therapy in breast cancer. *Biosens. Bioelectron.* **2021**, *176*, 112913. [[CrossRef](#)] [[PubMed](#)]
5. Vo, P.H.T.; Nguyen, T.D.T.; Tran, H.T.; Nguyen, Y.N.; Doan, M.T.; Nguyen, P.H.; Lien, G.T.K.; To, D.C.; Tran, M.H. Cytotoxic components from the leaves of *Erythrophleum fordii* induce human acute leukemia cell apoptosis through caspase 3 activation and PARP cleavage. *Bioorg. Med. Chem. Lett.* **2021**, *31*, 127673. [[CrossRef](#)] [[PubMed](#)]
6. Danhier, F.; Feron, O.; Préat, V. To exploit the tumor microenvironment: Passive and active tumor targeting of nanocarriers for anti-cancer drug delivery. *J. Controlled Release* **2010**, *148*, 135–146. [[CrossRef](#)]
7. Pan, L.; Chai, H.; Kinghorn, A.D. The continuing search for antitumor agents from higher plants. *Phytochem. Lett.* **2010**, *3*, 1–8. [[CrossRef](#)] [[PubMed](#)]
8. Taleghani, A.; Tayarani-Najaran, Z. Potent cytotoxic natural flavonoids: The limits of perspective. *Curr. Pharm. Des.* **2018**, *24*, 5555–5579. [[CrossRef](#)] [[PubMed](#)]
9. Rasul, A.R.A.; Ma, T.M.T. In vitro cytotoxic screening of 300 selected Chinese medicinal herbs against human gastric adenocarcinoma SGC-7901 cells. *Afr. J. Pharm. Pharmacol.* **2012**, *6*, 592–600. [[CrossRef](#)]
10. Newman, D.J.; Cragg, G.M. Natural products as sources of new drugs over the 30 years from 1981 to 2010. *J. Nat. Prod.* **2012**, *75*, 311–335. [[CrossRef](#)]
11. Middleton, E. Effect of plant flavonoids on immune and inflammatory cell function. *Adv. Exp. Med. Biol.* **1998**, *439*, 175–182. [[CrossRef](#)] [[PubMed](#)]
12. Passreiter, C.M.; Suckow-Schnitker, A.; Kulawik, A.; Addae-Kyereme, J.; Wright, C.W.; Wätjen, W. Prenylated flavanone derivatives isolated from *Erythrina addisoniae* are potent inducers of apoptotic cell death. *Phytochemistry* **2015**, *117*, 237–244. [[CrossRef](#)]
13. Al-Ashaal, H.A.; El-Sheltawy, S.T. Antioxidant capacity of hesperidin from *Citrus* peel using electron spin resonance and cytotoxic activity against human carcinoma cell lines. *Pharm. Biol.* **2010**, *49*, 276–282. [[CrossRef](#)]
14. Liang, H.Z.; Zhao, M.B.; Tu, P.F.; Jiang, Y. Polymethoxylated flavonoids from *Murraya paniculata* (L.) Jack. *Biochem. Syst. Ecol.* **2020**, *93*, 104162. [[CrossRef](#)]
15. Editorial Committee of Flora of China. *Flora of China*; Science Press: Beijing, China, 1997; p. 145.
16. Lv, H.N.; Wen, R.; Zhou, Y.; Zeng, K.W.; Li, J.; Guo, X.Y.; Tu, P.F.; Jiang, Y. Nitrogen oxide inhibitory trimeric and dimeric carbazole alkaloids from *Murraya tetramera*. *J. Nat. Prod.* **2015**, *78*, 2432–2439. [[CrossRef](#)]

17. Zhou, Y.; Lv, H.N.; Wang, W.G.; Tu, P.F.; Jiang, Y. Flavonoids and anthraquinones from *Murraya tetramera* C. C. Huang (Rutaceae). *Biochem. Syst. Ecol.* **2014**, *57*, 78–80. [[CrossRef](#)]
18. Lv, H.N.; Zhou, Y.; Wen, R.; Shi, M.L.; Zeng, K.W.; Xia, F.; Tu, P.F.; Jiang, Y. Murradiate and murradiol, two structurally unique heterodimers of carbazole-monoterpene and carbazole-phenylethanol from *Murraya tetramera*. *Phytochem. Lett.* **2016**, *15*, 113–115. [[CrossRef](#)]
19. Lyu, H.N.; Zhou, Y.; Wen, R.; Tu, P.F.; Jiang, Y. Nitric oxide inhibitory carbazole alkaloids from the folk medicine *Murraya tetramera* C.C. Huang. *Chem. Biodivers.* **2020**, *17*, e2000490. [[CrossRef](#)] [[PubMed](#)]
20. You, C.X.; Yang, K.; Wang, C.F.; Zhang, W.J.; Wang, Y.; Han, J.; Fan, L.; Du, S.S.; Geng, Z.F.; Deng, Z.W. Cytotoxic compounds isolated from *Murraya tetramera* Huang. *Molecules* **2014**, *19*, 13225–13234. [[CrossRef](#)]
21. Zhou, Y.F.; Wu, M.Z.; Chen, H.P.; Liu, Y.P. Compounds isolated from *Murraya tetramera* Huang and their cytotoxic activity. *Nat. Prod. Res. Dev.* **2019**, *31*, 627–632. [[CrossRef](#)]
22. Zhou, Y.F.; Chen, H.P.; Chen, L.; Liu, Y.P.; Li, Y. Carbazole alkaloids from *Murraya tetramera* Huang and their cytotoxic activity. *Nat. Prod. Res. Dev.* **2019**, *31*, 269–272, 305. [[CrossRef](#)]
23. Mateeva, N.N.; Kode, R.N.; Redda, K.K. Synthesis of novel flavonoid derivatives as potential HIV-integrase inhibitors. *J. Heterocycl. Chem.* **2002**, *39*, 1251–1258. [[CrossRef](#)]
24. Kong, C.H.; Liang, W.J.; Hu, F.; Xu, X.H.; Wang, P.; Jiang, Y.; Xing, B.S. Allelochemicals and their transformations in the *Ageratum conyzoides* intercropped citrus orchard soils. *Plant Soil* **2004**, *264*, 149–157. [[CrossRef](#)]
25. Passador, E.A.P.; Da Silva, M.F.D.G.F.; Fo, E.R.; Fernandes, J.B.; Vieira, P.C.; Pirani, J.R. A pyrano chalcone and a flavanone from *Neoraputia magnifica*. *Phytochemistry* **1997**, *45*, 1533–1537. [[CrossRef](#)]
26. Nagase, H.; Omae, N.; Omori, A.; Nakagawasai, O.; Tadano, T.; Yokosuka, A.; Sashida, Y.; Mimaki, Y.; Yamakuni, T.; Ohizumi, Y. Nobiletin and its related flavonoids with CRE-dependent transcription-stimulating and neuritegenic activities. *Biochem. Bioph. Res. Co.* **2005**, *337*, 1330–1336. [[CrossRef](#)]
27. Facundo, V.A.; Morais, S.M.; Braz Filho, R. Chemical constituents of *Ottionia corcovadensis* Miq. from Amazon Forest: ¹H and ¹³C chemical shift assignments. *Quim. Nova* **2004**, *27*, 79–83. [[CrossRef](#)]
28. Rwangabo, P.C.; Claeys, M.; Pieters, L.; Corthout, J.; Vandenberghe, D.A.; Vlietinck, A.J. Umuhengerin, a new antimicrobially active flavonoid from *Lantana trifolia*. *J. Nat. Prod.* **1988**, *51*, 966–968. [[CrossRef](#)]
29. Kinoshita, T.; Firman, K. Highly oxygenated flavonoids from *Murraya paniculata*. *Phytochemistry* **1996**, *42*, 1207–1210. [[CrossRef](#)]
30. Da Silva, B.F.; Rodrigues-Fo, E. Production of a benzylated flavonoid from 5,7,3',4',5'-pentamethoxyflavanone by *Penicillium griseoroseum*. *J. Mol. Catal. B Enzym.* **2010**, *67*, 184–188. [[CrossRef](#)]
31. Ferracin, R.J.; Da Silva, M.; Fernandes, J.B.; Vieira, P.C. Flavonoids from the fruits of *Murraya paniculata*. *Phytochemistry* **1998**, *47*, 393–396. [[CrossRef](#)]
32. Sherie, E.A.; Gupta, R.K.; Krishnamurti, M. Synthesis of chalcones, flavanones isolated from *Popowia cauliflora* and their analogues. *Agric. Biol. Chem.* **1981**, *45*, 531–533. [[CrossRef](#)]
33. Yao, H.; Jin, Y.R.; Shan, J.; Wang, X.Z.; Zhang, M.C.; Li, X.W. Two new natural methoxyflavonoids from leaves of *Murraya paniculata*(L.) Jack. *Chem. Res. Chin. Univ.* **2013**, *29*, 884–887. [[CrossRef](#)]
34. Gupta, R.K.; Krishnamurti, M.; Parthasarathi, J. Synthesis of some recently isolated chalcones, their analogues and corresponding flavanones. *Agric. Biol. Chem.* **1979**, *43*, 2603–2605. [[CrossRef](#)]
35. Cao, X.D.; Ding, Z.S.; Jiang, F.S.; Ding, X.H.; Chen, J.Z.; Chen, S.H.; Lv, G.Y. Antitumor constituents from the leaves of *Carya cathayensis*. *Nat. Prod. Res.* **2012**, *26*, 2089–2094. [[CrossRef](#)]
36. You, C.X.; Guo, S.S.; Zhang, W.J.; Geng, Z.F.; Liang, J.Y.; Lei, N.; Du, S.S.; Deng, Z.W. Chemical constituents of *Murraya tetramera* Huang and their repellent activity against *Tribolium castaneum*. *Molecules* **2017**, *22*, 1379. [[CrossRef](#)]
37. Wu, S.B.; Ji, Y.P.; Zhu, J.J.; Zhao, Y.; Xia, G.; Hu, Y.H.; Hu, J.F. Steroids from the leaves of Chinese *Melia azedarach* and their cytotoxic effects on human cancer cell lines. *Steroids* **2009**, *74*, 761–765. [[CrossRef](#)]